

Probiotics and synbiotics targeting the intestinal microbiome attenuate non-alcoholic fatty liver disease

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Non-alcoholic fatty liver disease (NAFLD) presents a chronic condition that links the overarching influence of the intestinal microbiome to a disease state with the multisystem manifestations of obesity, type 2 diabetes (T2D), chronic kidney disease (CKD) and cardiovascular disease (CVD) (1). NAFLD is an increasingly common disease with a current prevalence of 25% worldwide (2). The proportion of severe presentations of NAFLD is also increasing and these trends will lead to an increased burden from NAFLD sequelae, including cirrhosis, liver failure and hepatocellular cancer (2).

The diverse community of microbes encountered in the intestines highlights the complexity confronting research of the gut-liver axis. Interrogating the intestinal microbiome while building an evidenced based approach for novel treatment modalities that target the gut cohort of bacteria focuses research expectations at reversing or inhibiting the progression of NAFLD pathogenesis. Preclinical studies employing gut microbiota transplants in germ-free animal models have demonstrated that the development of fatty liver disease is substantially determined by gut bacteria (3). Subsequently, clinical studies report that pro-inflammatory intestinal bacteria from the phylum Proteobacteria and the genus Escherichia coli predominate in patients diagnosed with NAFLD (4). In contrast, beneficial bacteria that can attenuate fatty liver pathology such as Faecalibacterium prausnitzii were reported decreased in NAFLD patients (4). Additional studies (5) that investigated the severity of liver disease in NAFLD correlated with shifts in intestinal microbiome patters such as increased *Bacteroides*, decreased *Prevotella* abundance and with increased abundance of *Ruminococcus* in NAFLD with fibrosis (5).

One mechanism postulated to link the gut microbiome with NAFLD is disruption of the gut epithelial barrier allowing microbial products to enter the portal circulation. Endotoxins from Gram-negative bacteria [e.g., lipopolysaccharides (LPS) from Escherichia coli] can induce intestinal inflammation and increase permeability, facilitating its own access to the liver where it can directly cause NAFLD and liver fibrosis (Figure 1) (1). In contrast, other bacterial products are needed to prevent inflammation via the nucleotide-binding oligomerisation domaincontaining protein 2 (a pattern recognition receptor involved in innate bacterial immunity) (6). Intestinal derived bacterial metabolites [such as butyrate and other short chain fatty acids (SCFA)] are also important signalling molecules for maintaining the epithelial cell-to-cell tight junction protein network and secretion of mucin proteins. Intestinal dysbiosis can lead to decreases in these metabolites and disruption of the intestinal epithelial barrier (7).

NAFLD can be considered a link between dysbiosis and a multisystem disease that includes CVD, CKD, T2D and obesity. Choline metabolism has been postulated as one mechanism behind this. Overgrowth of cholinemetabolising bacteria can cause choline deficiency, a known cause of NAFLD (8). A bacterial metabolite of



Figure 1 Intestinal mucosa inflammation links gut dysbiosis to a fatty liver [modified from Saltzman et al., 2018 (1)].

choline, trimethylamine is metabolised by the liver into trimethylamine-N-oxide, elevated levels of which is associated with CVD (8). LPS and SCFA also provide a link between the multisystem manifestations of NAFLD. LPS has been implicated as an inductor of a pro-inflammatory environment which is conducive to metabolic syndrome, insulin resistance and T2D. SCFA are important signalling molecules with systemic effects on metabolic regulation (7).

A compromised intestinal barrier can be associated with bacterial translocation from the gut into the systemic circulation increasing the risk of sepsis. LPS, a constituent of Gram-negative bacteria, is found to be increased in the systemic circulation, indicative of dysbiosis (9). LPS has been associated with inducing apoptosis of lymphocytes under *in vivo* conditions (10) demonstrating an immune-modulatory effect. Studies have suggested that a loss of lymphocytes in the intestinal mucosa is a consequence of intestinal epithelial dysbiosis and subsequent release of metabolic endotoxins (11). Gram-negative bacteria containing LPS are therefore hypothesized to contribute to NAFLD development. In addition dysbiosis and elevated systemic LPS can be predicted as markers of intestinal toxicity (12). Intestinal toxicity driven dysbiosis supports gut mucosal inflammatory responses that are concomitant with an increase in gut permeability. This combined disturbance of the intestinal barrier and local mucosal immune system activity can support and mediate NAFLD pathogenesis via the gut-liver axis (1).

The idea that a probiotic bacterium could improve inflammatory responses in the gut was demonstrated in a study that reported a decrease in hepatic steatosis with a probiotic that resulted in an analogous anti-inflammatory effect as anti-TNF antibody treatment (13). This result has provided a biologically plausible posit for the administration of probiotics to ameliorate NAFLD.

Numerous clinical trials have ensued, and Sharpton and colleagues' have provided a recent high quality systematic and meta-analysis study (14), published in the American Journal of Clinical Nutrition investigating the feasibility of probiotics and synbiotics administered as microbiome targeted therapies for the attenuation of NAFLD. The strength of the systematic review rests in the most current evidence of the included studies, which consisted of twentyone randomized clinical trials of probiotics (n=9) or synbiotics (i.e., a formulation that comprises probiotics + prebiotics; n=12) versus a comparator treatment. The most common comparator treatment was the use of a placebo in 14 trials, usual care in 5 and placebo + sitagliptin (both groups) in 1 and placebo + metformin (both groups) in 1 other study. There was high variability in the constituents of the formulations and clinical studies investigated.

Furthermore, studies were grouped according to the intervention and study outcome that allowed for clinical study heterogeneity assessment. Consequently, statistical heterogeneity was clearly evident and appropriately the I² statistic was measured to indicate the percentage of variance attributed to the clinical studies that were included in the meta-analysis. Probiotics and synbiotics administration showed significant reduced liver stiffness measurements by elastography (weighed mean difference of -0.70 kPa, 95% CI: -1.0, -0.4 kPa); and alanine aminotransferase activity (weighted mean difference of -11.23 U/L, 95% CI: -15.02, -7.44 U/L) with high pooled study heterogeneity (I²: 93.4%) and 90.6%, respectively). Both probiotics and synbiotics had increased odds for improvement of hepatic steatosis (i.e., reduced) by graded ultrasound (odds ratio: 2.40, 95% CI: 1.5, 3.84). Overall the meta-analysis and metaregression showed that the use of probiotics or synbiotics presented with improvements in liver specific markers of hepatic inflammation, liver stiffness measurements and liver steatosis in patients diagnosed with NAFLD. Unquestionably, the results of Sharpton and colleagues' meta-analysis study (14) should prompt and warrant more robust and well-defined trials.

Probiotics or synbiotics are not a panacea for the overall amelioration of chronic diseases but rather an evolving intestinal targeted therapy. The further development of probiotic/synbiotic based therapies for the prevention of progression and treatment of NAFLD requires a further focused molecular understanding of the multiple actions of probiotics and synbiotics in the intestines. Namely, such as the preservation of intestinal epithelia tight junctions as demonstrated in animal models of obesity induced NAFLD (15); the attenuation of intestinal inflammation in NAFLD (15); and the intestinal microbiome as the metaanalysis by Sharpton *et al.* (14) has reported.

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Footnote

Conflicts of Interest: Both authors have completed the ICMJE uniform disclosure form (available at http://dx.doi. org/10.21037/hbsn.2019.11.24). LV reports in addition, LV has a patent 2017101478 issued. LV and JDH participate in the Medlab Clinical's research on probiotics.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

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