Erratum to epigenetics of colorectal cancer: emerging circulating diagnostic and prognostic biomarkers

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Erratum to: Ann Transl Med 2017;5:279

In the article entitled "Epigenetics of colorectal cancer: emerging circulating diagnostic and prognostic biomarkers" that appeared on page 279 of Vol 5, No 13 of *Annals of Translational Medicine (ATM)*, there are some errors. *Table 1* should be replaced by the following content.

The publisher regrets the error.

Study	Country	Sample type	Discovery phase (method and sample population)	Validation phase(s) (method and sample population)	Candidate miRNAs (expression)	AUC (95% CI)	SE	SP
Ghanbari R, <i>et al.</i> 2015 (48)	Iran	Plasma	miRNA Microarray system (Agilent); 37 CRC, 8 controls	qRT-PCR;	miR-142-3p (↓)	0.71	-	-
				61 CRC, 24 controls	miR-26a-5p (↓)	0.67	-	-
Giráldez MD, <i>et al.</i> 2013 (49)	Spain	Plasma	SAM-Bead Array platform (Illumina); 21 CRC, 20 advanced adenoma, 20 controls	qRT-PCR; 42 CRC, 40	miR19a, miR19b (all ↑)	miR19a, miR19b 0.82 78 (all ↑) (0.73–0.90)	78.57%	77.36%
				advanced adenomas, 53 controls	miR19a, miR19b, miR15b (all ↑)	0.84 (0.76–0.92)	78.57%	79.25%
Hofsli E, <i>et al.</i> 2013 (50)	Norway	Serum	miRCURY LNA Universal RT microRNA PCR (Exiqon) (375 miRNAs); 30 CRC (stage IV), 10 controls	miRCURY LNA Universal RT microRNA PCR (34	miR-423-5p, miR- 210, miR-720, miR-320a and miR-378 (all ↑)	-	-	-
				miRNAs); 40 CRC (stage I, II), 10 controls	miR-106a, miR- 143, miR-103, miR-199a-3p, miR-382 and miR- 151-5p (all ↓)	-	_	-
Imaoka H, <i>et al.</i> 2016 (51)	Japan	Serum	Microarray analysis; 3 CRC, 3 adenoma, 3 controls	qRT-PCR; 211 CRCs, 56 adenomas, 57	miR-1290 (↑)	CRC <i>vs.</i> controls 0.83	7.01%	91.2%
				controls		Adenoma vs. controls 0.72	46.4%	91.2%
Study	Country	Sample type	Discovery phase (method and sample population)	Validation phase(s) (method and sample population)	Candidate miRNAs (expression)	AUC (95% CI)	SE	SP

Table 1 Studies aimed to identify diagnostic miRNA for CRC through high-throughput screening and subsequent qPCR validation

Table 1 (continued)

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Table 1 (continued)

Table I (continu	ea)							
Study	Country	Sample type	Discovery phase (method and sample population)	Validation phase(s) (method and sample population)	Candidate miRNAs (expression)	AUC (95% Cl)	SE	SP
Kanaan Z, <i>et al.</i> 2013 (52)	Kentucky (USA)	Plasma	TaqMan low-density array card (life technologies) 380 miRNAs; 20 CRC, 9 adenoma, 12 controls	qRT-PCR; 45 CRC, 16 adenoma, 26 controls	miR-139-3p, miR- 431 (all ↑)	CRC <i>vs.</i> controls; 0.83 (0.73–0.93)	91%	57%
Li J, <i>et al.</i> 2015 (53)	China	Serum	TaqMan low-density array (applied biosystems) (749 miRNAs); pooled 20 CRC, 20 controls	qRT-PCR; 175 stage II/III CRC patients, 130 controls	miR-145 (↓)	0.87 (0.84–0.91)	-	-
					miR-17-3p (↑)	0.78 (0.73–0.83)	-	-
					miR-106a (↑)	0.81 (0.77–0.86)	-	-
					miR-145 (↓), miR- 17-3p (↑), miR106a (↑)	0.89 (0.85–0.92)	78.5%	82.8%
Luo X, <i>et al.</i> 2013 (54)	Germany	Plasma	TaqMan microRNA array (applied biosystems) 667 miRNAs; 5 pools of CRC patients (N=50), 5 pools of controls (N=50)	qRT-PCR; 80 CRC, 50 advanced adenoma, 194 controls	miR-18a, miR-20a, miR21, miR-29a, miR-92a, miR- 106b, miR-133a, miR-143, miR-145 (all ↑)	CRC vs. controls; 0.745 (0.708– 0.846)	-	-
Ng EK, <i>et al.</i> 2009 (55)	China	Plasma	Cancer microRNA array (system biosciences) 95 miRNAs; 5 CRC, 5 controls	qRT-PCR; (I) 25 CRC,20 controls; (II) 90 CRC 50 controls	miR-17-3p (↑)	(I) 0.717 (0.633– 0.800)	(I) 64%	70%
					miR-92 (↑)	(II) 0.885 (0.828– 0.942)	(II) 89%	70%
Sun Y, et al. 2016 (56)	China	Plasma	TaqMan array (life technologies) (754 miRNAs); 40 CRC, 10 controls	qRT-PCR; 187 CRC, 47 controls	miR-96 (↑)	0.740 (0.650– 0.831)	65.4%	73.3%
Vychytilova-	Czech Republic	zech Serum epublic	Illumina small RNA sequencing; pooled 144 CRC, 96 controls	qRT-PCR; 203 CRC, 100 controls	miR-23a-3p (↑)	0.89	-	-
Faltejskova P,					miR-27a-3p (†)	0.70	-	-
et al. 2016 (57)					miR-142 5p (↑)	0.81	-	-
					miR-376c-3p (↑)	0.65	-	-
					miR-23a-3p, miR- 27a-3p, miR-142- 5p, miR-376c-3p (all ↑)	0.92	89%	81%

Table 1 (continued)

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Table 1	(continued)
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Study	Country	Sample type	Discovery phase (method and sample population)	Validation phase(s) (method and sample population)	Candidate miRNAs (expression)	AUC (95% CI)	SE	SP
Wang Q, <i>et al.</i> 2012 (58)	China	Plasma	miRCURY LNA [™] universal RT microRNA PCR system (exiqon, demark) (742 miRNAs); 10 CRC, 10 controls	qRT-PCR; 90 CRC, 43 advanced adenoma, 58 controls	miR-601 (↓)	CRC <i>vs.</i> controls; 0.747 (0.666– 0.828)	69.2%	72.4%
					miR-760 (↓)	CRC <i>vs.</i> controls; 0.788 (0.714– 0.862)	80%	72.4%
Wang S, <i>et al.</i> 2015 (59)	China	Plasma	Human microRNA microarrays Agilent technologies (723 miRNAs); 47 CRC, 33 controls	qRT-PCR; (I) 55 CRC, 57 controls; (II) 22 CRC, 27 controls	miR-409-3p (†), miR-7 (↓), miR-93 (↓)	(I) 0.866 (0.788– 0.923); (II) 0.897	(I) 91%; (II) 82%	(I) 88%; (II) 89%
Xu L, <i>et al.</i> 2014 (60)	China	Plasma	TaqMan low density MiRNA arrays [applied biosystems (754 miRNAs)]; 6 CRC, 6 controls	qRT-PCR; 88 CRC, 40 controls	miR-375 (↓)	0.749 (0.654- 0.844)	76.92%	64.63%
					miR-206 (↑)	0.705 (0.612- 0.798	-	-
					miR-375 (↓) miR- 206 (↑)	0.846 (0.775- 0.917)	-	-
Zheng G, <i>et al.</i> 2014 (61)	China	Serum	Miseq sequencing; pooled serum samples (30 CRC, 25 adenoma, 30 controls)	qRT-PCR; (l) 160 CRC, 66 CA patients,	qRT-PCR; (I) miR-19a-3p (↑) 160 CRC, 66 CA patients,	CRC <i>vs.</i> controls (I) 0.849	-	-
				94 controls;	miR-92a-3p (↑)	(I) 0.890	-	-
				(II) 117 CRC, 73 adenoma, 102 controls	miR-223-3p (↑)	(l) 0.871	-	-
					miR-422a (↓)	(l) 0.843	-	_
					miR-19a-3p, miR- 223-3p, miR-92a- 3p and miR-422a	(II) 0.951 (0.907– 0.978	-	-
Zhu M, et al. 2017 (62)	China	Serum	Exiqon miRCURY- Ready-to-Use-PCR- Human-panel-I + II- V1 (168 miRNAs); 30 CRC, 30 controls	qRT-PCR; (I)136 CRC, 90 controls; (II) 30 CRC, 18 controls	miR-19a-3p, miR- 21-5p, miR-425- 5p (all ↑)	0.830 (0.708– 0.952)	-	-

(†), miRNAs upregulated in CRC patients compared to healthy subjects; (↓), miRNAs downregulated in CRC patients compared to healthy subjects. CRC, colorectal cancer; AUC, area under curve; SE, sensitivity; SP, specificity.

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