

# Suppression of monocyte inflammatory and coagulopathy responses in HIV infection

Reena Rajasuriar<sup>1,2,3</sup>, Anna C. Hearps<sup>4,5</sup>, Suzanne M. Crowe<sup>4,6</sup>, Joshua J. Anzinger<sup>7#</sup>, Clovis S. Palmer<sup>4,6,8#</sup>

<sup>1</sup>Department of Pharmacy, Faculty of Medicine, <sup>2</sup>Centre of Excellence for Research in AIDS (CERiA), University of Malaya, Kuala Lumpur, Malaysia; <sup>3</sup>Peter Doherty Institute for Infection and Immunity, University of Melbourne, Melbourne, VIC, Australia; <sup>4</sup>Life Sciences, Burnet Institute, Melbourne, VIC, Australia; <sup>5</sup>Department of Infectious Diseases, <sup>6</sup>School of Public Health and Preventive Medicine, Monash University, Melbourne, VIC, Australia; <sup>7</sup>Department of Microbiology, University of the West Indies, Mona, Kingston, Jamaica; <sup>8</sup>Department of Microbiology and Immunology, University of Melbourne, Melbourne, VIC, Australia

#These authors contributed equally to this work.

Correspondence to: Clovis S. Palmer. 85 Commercial Road, Melbourne, Victoria 3004, Australia. Email: clovis.palmer@burnet.edu.au.

Provenance: This is a Guest Editorial commissioned by the Section Editor Mingzhu Gao (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

Comment on: Schechter ME, Andrade BB, He T, *et al.* Inflammatory monocytes expressing tissue factor drive SIV and HIV coagulopathy. *Sci Transl Med* 2017;9.

Submitted Jun 04, 2018. Accepted for publication Jun 11, 2018.

doi: 10.21037/atm.2018.06.20

View this article at: <http://dx.doi.org/10.21037/atm.2018.06.20>

The increased access globally to effective antiretroviral therapy (ART) among people living with HIV (PLHIV) has led to a change in the demographics of this population (1,2). Once considered a death sentence, HIV infection has now evolved to be a chronic disease with PLHIV achieving near normal life-expectancy (3), despite disparities that still exist between high-income and low-and-middle-income settings (4).

As PLHIV grow older, age-associated comorbidities including cardiovascular disease (CVD) have now become a clinical priority in the routine management of HIV (5). CVD risks are reported to be approximately two times higher amongst PLHIV compared to age-matched uninfected individuals, with CVD risk prediction algorithms not accurately reflecting CVD risk in PLHIV (6). Persistent immune activation is believed to be a major factor contributing to the pathogenesis of CVD-related morbidity and aging in HIV (7). Additionally, increased markers of hypercoagulation including D-dimers have been shown to independently predict mortality and CVD-related morbidity in treated HIV (8-10). In this regard, monocytes have garnered renewed attention as a number of studies examining ART treated PLHIV show associations between coagulation markers and activated monocytes (11), and indeed between monocyte activation and atherosclerosis (12). The study by Schechter *et al.* (13) provides compelling

evidence that a subset of tissue factor-expressing monocytes is central to hypercoagulation and systemic inflammation in PLHIV. Tissue factor (coagulation factor III) is a cell surface glycoprotein highly expressed on monocytes in response to inflammatory stimuli such as LPS that enables the initiation of coagulation cascades. Thus, increased tissue factor expression is associated with cardiovascular complications (14).

In the study, the authors show that compared to HIV uninfected people the proportion of monocytes expressing tissue factor is elevated in both ART naïve and experienced PLHIV, with monocytes from both groups of PLHIV showing inhibition of tissue factor activity when treated *ex vivo* with Ixolaris, a tissue factor inhibitor. Tissue factor-expressing monocytes from PLHIV produced a broader array of inflammatory cytokines than monocytes lacking tissue factor. Furthermore, the authors found HIV infection was associated with a skewing of monocyte inflammatory responses towards a more polyfunctional phenotype, favouring the production of IL-6 and TNF, with this phenotype not being reversed by ART.

Both HIV and pathogenic SIV infection result in gastrointestinal mucosal barrier damage that allows for microbes and microbial products to enter the systemic circulation, a process termed microbial translocation (15).

Monocyte activation and coagulation markers are associated with translocated microbial products but a causal role has not been previously described (16). Given the well-described increase of gastrointestinal microbial translocation products that occur for both ART naïve and ART experienced PLHIV, the authors also investigated monocyte tissue factor expression and functional activity from HIV uninfected people after *in vitro* exposure to LPS, a surrogate for microbial translocated products. In these experiments, monocytes showed robust induction of tissue factor expression and activity with exposure to LPS. Monocyte tissue factor was also increased after exposure to HIV positive serum but this was partially inhibited by blocking type I interferon and TNF receptors, and completely inhibited following the addition of polymyxin B, a Gram-negative bactericidal agent that binds to LPS. In future work, it will be interesting to determine if agents that decrease microbial translocation products also decrease tissue factor expressing monocytes, and thus, limit hypercoagulation and systemic inflammation. Indeed, sevelamer was shown to limit LPS translocation and suppress inflammation in animal models, although clinical trials in HIV-positive persons were disappointing (17).

The authors extended their *in vitro* and *ex vivo* work by using non-human primate models of HIV infection, in which they exploited differences between SIV infection of natural African green monkey (AGM) hosts that do not experience disease and pigtail macaques (PTM) that experience disease similar to HIV infection. The authors show that compared to SIV-infected AGM natural hosts, PTM infected with SIV have increased tissue factor-expressing monocytes and increased polyfunctional inflammatory cytokine production. Notably, previous studies show that PTM experience increased microbial translocation during SIV infection that does not occur in natural AGM hosts (18,19).

Importantly, the authors assessed whether Ixolaris is capable of reducing inflammation and coagulation *in vivo*. Remarkably, SIV-infected PTM treated daily with Ixolaris showed substantial diminution of markers of inflammation and coagulation without significantly affecting SIV viral load. To further validate the effects of Ixolaris treatment on monocyte activation, the team evaluated glucose transporter-1 (Glut1) expression on monocytes, an important monocyte activation marker. Glut1 is a major glucose transporter on monocytes and its level is increased on activated monocytes, reflecting high glucose metabolic demands for these activated cells (20,21). Indeed, the

authors demonstrated that compared to untreated controls, Ixolaris dramatically reduced monocyte Glut1 expression in SIV-infected PTM. Increased Glut1 levels on inflammatory monocytes have previously been shown to be associated with markers of CVD risk in treated HIV-infected men (22), and subclinical CVD in treated HIV-positive women (23). Thus, it remains to be determined whether some of the anti-inflammatory and anti-coagulopathy effects of Ixolaris are due to the cumulative suppression of monocyte tissue factor and Glut1. Yet, it is also unclear if the consequence of Ixolaris treatment was due solely by its action on monocytes alone or other cells that also express tissue factor and Glut1.

Taken together, the study by Schechter *et al.* provides a more detailed understanding of the linkage between microbial translocation, hypercoagulation and systemic inflammation, with tissue factor-expressing monocytes as important intermediaries. These findings also identify a novel target for therapy that may limit CVD and other age-associated diseases in PLHIV, though the feasibility of targeting tissue factor-expressing monocytes is unclear as CVD emerges over the course of many years or decades and would likely require chronic drug treatment. Probiotic restoration of healthy gastrointestinal function has emerged as a possible treatment to limit inflammation caused by microbial translocation (24,25) and would be an interesting strategy to explore in relation to tissue factor-expressing monocytes.

## Acknowledgements

None.

## Footnote

**Conflicts of Interest:** The authors have no conflicts of interest to declare.

## References

1. Smit M, Brinkman K, Geerlings S, et al. Future challenges for clinical care of an ageing population infected with HIV: a modelling study. *Lancet Infect Dis* 2015;15:810-8.
2. Puh R, Kumarasamy N, Ly PS, et al. HIV and Aging: Demographic Change in the Asia-Pacific Region. *J Acquir Immune Defic Syndr* 2017;74:e146-8.
3. Antiretroviral Therapy Cohort Collaboration. Survival of HIV-positive patients starting antiretroviral therapy between 1996 and 2013: a collaborative analysis of cohort

- studies. *Lancet HIV* 2017;4:e349-56.
4. Teeraananchai S, Kerr SJ, Amin J, et al. Life expectancy of HIV-positive people after starting combination antiretroviral therapy: a meta-analysis. *HIV Med* 2017;18:256-66.
  5. Guaraldi G, Palella FJ Jr. Clinical implications of aging with HIV infection: perspectives and the future medical care agenda. *AIDS* 2017;31 Suppl 2:S129-35.
  6. Triant VA, Perez J, Regan S, et al. Cardiovascular Risk Prediction Functions Underestimate Risk in HIV Infection. *Circulation* 2018;137:2203-14.
  7. Lagathu C, Cossarizza A, Bereziat V, et al. Basic science and pathogenesis of ageing with HIV: potential mechanisms and biomarkers. *Aids* 2017;31 Suppl 2:S105-19.
  8. Boulware DR, Hullsiek KH, Puroon CE, et al. Higher levels of CRP, D-dimer, IL-6, and hyaluronic acid before initiation of antiretroviral therapy (ART) are associated with increased risk of AIDS or death. *J Infect Dis* 2011;203:1637-46.
  9. Kuller LH, Tracy R, Belloso W, et al. Inflammatory and Coagulation Biomarkers and Mortality in Patients with HIV Infection. *PLoS Med* 2008;5:e203.
  10. Tenorio AR, Zheng Y, Bosch RJ, et al. Soluble Markers of Inflammation and Coagulation but Not T-Cell Activation Predict Non-AIDS-Defining Morbid Events During Suppressive Antiretroviral Treatment. *J Infect Dis* 2014;210:1248-59.
  11. Anzinger JJ, Butterfield TR, Angelovich TA, et al. Monocytes as regulators of inflammation and HIV-related comorbidities during cART. *J Immunol Res* 2014;2014:569819.
  12. Westhorpe CL, Maisa A, Spelman T, et al. Associations between surface markers on blood monocytes and carotid atherosclerosis in HIV-positive individuals. *Immunol Cell Biol* 2014;92:133-8.
  13. Schechter ME, Andrade BB, He T, et al. Inflammatory monocytes expressing tissue factor drive SIV and HIV coagulopathy. *Sci Transl Med* 2017;9.
  14. Owens AP 3rd, Mackman N. Role of tissue factor in atherothrombosis. *Curr Atheroscler Rep* 2012;14:394-401.
  15. Ortiz AM, Brenchley JM. Microbial translocation: translating simian immunodeficiency virus to HIV. *Curr Opin HIV AIDS* 2018;13:15-21.
  16. Funderburg NT, Mayne E, Sieg SF, et al. Increased tissue factor expression on circulating monocytes in chronic HIV infection: relationship to in vivo coagulation and immune activation. *Blood* 2010;115:161-7.
  17. Sandler NG, Zhang X, Bosch RJ, et al. Sevelamer does not decrease lipopolysaccharide or soluble CD14 levels but decreases soluble tissue factor, low-density lipoprotein (LDL) cholesterol, and oxidized LDL cholesterol levels in individuals with untreated HIV infection. *J Infect Dis* 2014;210:1549-54.
  18. Pandrea IV, Gautam R, Ribeiro RM, et al. Acute loss of intestinal CD4+ T cells is not predictive of simian immunodeficiency virus virulence. *J Immunol* 2007;179:3035-46.
  19. Kristoff J, Haret-Richter G, Ma D, et al. Early microbial translocation blockade reduces SIV-mediated inflammation and viral replication. *J Clin Invest* 2014;124:2802-6.
  20. Palmer CS, Cherry CL, Sada-Ovalle I, et al. Glucose Metabolism in T Cells and Monocytes: New Perspectives in HIV Pathogenesis. *EBioMedicine* 2016;6:31-41.
  21. Palmer CS, Anzinger JJ, Zhou J, et al. Glucose transporter 1-expressing proinflammatory monocytes are elevated in combination antiretroviral therapy-treated and untreated HIV+ subjects. *J Immunol* 2014;193:5595-603.
  22. Anzinger JJ, Butterfield TR, Gouillou M, et al. Glut1 Expression Level on Inflammatory Monocytes is Associated With Markers of Cardiovascular Disease Risk in HIV-Infected Individuals. *J Acquir Immune Defic Syndr* 2018;77:e28-30.
  23. Butterfield TR, Hanna DB, Kaplan RC, et al. Increased glucose transporter-1 expression on intermediate monocytes from HIV-infected women with subclinical cardiovascular disease. *AIDS* 2017;31:199-205.
  24. d'Ettorre G, Rossi G, Scagnolari C, et al. Probiotic supplementation promotes a reduction in T-cell activation, an increase in Th17 frequencies, and a recovery of intestinal epithelium integrity and mitochondrial morphology in ART-treated HIV-1-positive patients. *Immun Inflamm Dis* 2017;5:244-60.
  25. Falasca K, Vecchiet J, Ucciferri C, et al. Effect of Probiotic Supplement on Cytokine Levels in HIV-Infected Individuals: A Preliminary Study. *Nutrients* 2015;7:8335-47.

**Cite this article as:** Rajasuriar R, Hearps AC, Crowe SM, Anzinger JJ, Palmer CS. Suppression of monocyte inflammatory and coagulopathy responses in HIV infection. *Ann Transl Med* 2018;6(13):277. doi: 10.21037/atm.2018.06.20