



Brake adjustment: Ca^{2+} entry pathway provides a novel target for acute pancreatitis therapy

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Provenance: This is an invited article commissioned by the Section Editor Dr. Le Li (Department of Pancreatic and Biliary Surgery, The First Affiliated Hospital of Harbin Medical University, Harbin Medical University, Harbin, China).

Comment on: Son A, Ahuja M, Schwartz DM, *et al.* Ca^{2+} Influx Channel Inhibitor SARAF Protects Mice From Acute Pancreatitis. *Gastroenterology* 2019. [Epub ahead of print].

Submitted Oct 30, 2019. Accepted for publication Nov 13, 2019.

doi: 10.21037/atm.2019.11.119

View this article at: <http://dx.doi.org/10.21037/atm.2019.11.119>

Acute pancreatitis (AP) is accompanied by intense abdominal pain that often requires hospital admission, but no therapies directed at causes of the disease yet exist (1). Evidence accumulated over decades of research revealed that the initial events of AP target the pancreatic acinar cell, where digestive enzymes are produced (1). Especially during the past 25 years, a hypothesis that has gained wide acceptance is that an excess of acinar cell cytosolic Ca^{2+} ($\text{Ca}^{2+}_{\text{cyt}}$) is a key initial trigger of AP pathology (1-3). This $\text{Ca}^{2+}_{\text{cyt}}$ overload likely contributes to premature trypsin activation, a hallmark of AP pathology, as well as a plethora of other responses (4,5). Depending on its extent and persistence, $\text{Ca}^{2+}_{\text{cyt}}$ overload in the acinar cell may perturb cellular energetics, confound complex secretory functions and homeostatic processes like autophagy, or induce cell death.

The basic pathobiology of $\text{Ca}^{2+}_{\text{cyt}}$ overload is well established. Hyperstimulation of surface membrane receptors in acinar cells releases intracellular Ca^{2+} stores such as those within the endoplasmic reticulum (ER). Many receptors couple to ER Ca^{2+} release through activation of the inositol trisphosphate (IP_3) receptors, which are ligand-gated, intracellular Ca^{2+} release channels. Substantial ER Ca^{2+} release triggers entry of extracellular Ca^{2+} , predominantly through a process called store-operated Ca^{2+} entry (SOCE). In SOCE, special transmembrane proteins “sense” lowered ER luminal Ca^{2+} concentration via luminal domains. This information is then transferred

to the cytoplasm, where a distinct domain mediates opening of certain Ca^{2+} permeable plasma membrane channels (designated as calcium release activated calcium, or CRAC) through direct physical contacts. Whereas SOCE is normally required for the Ca^{2+} signaling functions of all electrically excitable and non-excitable cells, its contribution to AP pathology has fueled hopes of developing therapeutic agents that address the root causes of the disease through studies of the mechanisms involved in SOCE and identification of specific targets.

SOCE/CRAC channels were known to exist long before their actual machinery was discovered; only in 2005–2006 were the key mediators of these Ca^{2+} entry pathways elucidated and first characterized. The components of this pathway (summarized briefly in *Table 1*) include the stromal interaction molecule (STIM) family molecules STIM1 and STIM2; these are the ER proteins with Ca^{2+} -sensing motifs directed toward the ER lumen. STIM2 turns out to have two (i.e., STIM2 α and STIM2 β) splice variants with opposite roles as activators and inhibitors of SOCE, respectively (11). Major plasma membrane Ca^{2+} channels that open as a consequence of specific STIM contacts were found to be the Orai family molecules Orai1, Orai2, and Orai3, which form multimeric channels in a combinatorial fashion. Other plasma membrane channels such as transient receptor potential C3, Trpc3 also exhibit SOCE-like ion channel behavior. Release of ER Ca^{2+} and subsequent decrease in ER Ca^{2+} concentration promotes STIM1

Table 1 Major proteins involved in store-operated Ca²⁺ entry and their roles in cells

Reference	Molecule	Promotes (+) or brakes (-) SOCE
Srikanth (6)	CRACR2A	+
Soboloff (7); Hoth (8)	ORAI1	+
Soboloff (7); Hoth (8)	ORAI2	+
Soboloff (7); Hoth (8)	ORAI3	+
Palty (9); Son (10)	SARAF	-
Soboloff (7); Hoth (8)	STIM1	+
Soboloff (7); Hoth (8); Rana (11)	STIM2	+/-
Jing (12)	STIMATE	+

SOCE, store-operated Ca²⁺ entry; SARAF, SOCE-associated regulatory factor; STIM, stromal interaction molecule.

clustering and conformational changes, ER dynamic behavior, and formation of tethered ER-plasma membrane contact sites bridged by STIM-Orai interactions. For excellent early reviews of the main components of SOCE and their functions (7,8).

Although much is now known about STIM behavior and direct interactions with Orai, additional auxiliary molecules also regulate SOCE. In 2010, a novel protein called CRAC regulator 2A (CRACR2A) was hypothesized to stabilize the STIM-Orai channel “ON”-state via interactions with both proteins (6). STIM-Orai-mediated SOCE was shown to be sensitive to levels of Ca²⁺_{cyt}. Thus, after the Ca²⁺ pores switch ON, they subsequently switch OFF in response to the corresponding rise in Ca²⁺_{cyt}, following distinct rapid and slow kinetics. Later, a novel protein termed SOCE-associated regulatory factor (SARAF) (formerly designated TMEM66) was identified in a functional screen that revealed its role in lowering mitochondrial, ER and basal Ca²⁺_{cyt} levels (9). Knocking down SARAF enhanced ON-switching of STIM-Orai channels by removing its interactions with STIM. Thus, SARAF was shown to switch OFF a STIM1-Orai1 channel with slow kinetics (9). Further, its ER localization and structural similarity to STIMs suggested that STIM regulation is its main cellular function. SARAF negatively modulates STIM clustering, a requirement for Orai channel opening (7). A distinct molecule, STIMATE that promotes STIM clustering was found in 2015 by a proteomic approach (12). Thus, regulatory molecules with complementary modes of action participate in SOCE ON- (CRACR2A, STIMATE) and OFF- (SARAF) switching (*Table 1*). Subsequent studies of SARAF have established further aspects of its role as a brake on SOCE via SARAF-STIM contacts (13).

Figure 1 provides an overview of SOCE and the SOCE-braking action of SARAF.

Based on the role of Ca²⁺ overload in AP pathology and its dependence on Ca²⁺ entry, blocking SOCE in acinar cells seems like a good candidate to target for an AP therapy. But which approach will prove most applicable? In recent years a focus of research has been on the chemical blockers (GSK-7975A and CM_128, also known as CM4620) that abrogate Orai channel function during SOCE in acinar cells, pancreatic stellate cells and neutrophils, all of which take part in AP pathology (14-17). Caution is warranted in the use of such Ca²⁺ signaling inhibitors as pancreatitis therapy in humans, as most of the studies have used rodent models and the findings may not translate well (18). In a new study just published in *Gastroenterology*, investigators drill down further into the mechanistic details of the STIM1-Orai mediated Ca²⁺ entry, investigating the use of SARAF as a regulatory brake on STIM-Orai mediated SOCE as an alternative approach (10).

To investigate the role of the SOCE regulatory molecule SARAF in pancreatitis, Son *et al.* manipulated SARAF expression levels in various models and used a fluorescence assay to measure interactions with STIM1. These investigations produce evidence that upon physiologic acinar cell stimulation, a SARAF-STIM1 interaction is enhanced. In contrast, hyperstimulation produces a more biphasic increase-then decrease in this interaction. Overall, the studies recapitulate the concept of SARAF as a brake on SOCE and demonstrate the application of overexpressed SARAF to reduce AP severity.

Roles of Ca²⁺ entry in AP pathology and severity have been appreciated for some time. Detrimental consequences of excessive Ca²⁺_{cyt} levels include impairment of essential

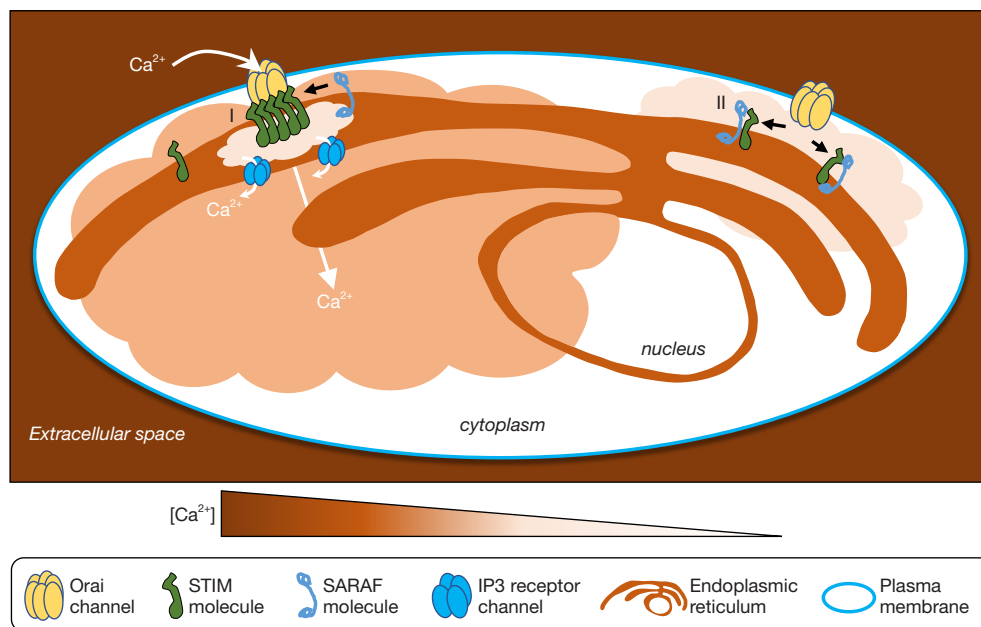


Figure 1 SOCE switched on by STIM-mediated gating of Orai channels, and the aftermath of OFF-switching. (I) Upon sensing a reduced Ca^{2+} level within the ER lumen, STIMs cluster and couple to Orai channels to activate SOCE (left); (II) the regulator SARAF binds to STIM and acts as a “brake” on SOCE (right). SOCE, store-operated Ca^{2+} entry; STIM, stromal interaction molecule; ER, endoplasmic reticulum; SARAF, SOCE-associated regulatory factor.

functions such as maintenance of the mitochondrial electrochemical potential coupled to cellular energy production and the orderly processing and packaging of digestive enzymes into secretory granules in acinar cells. Strategies to abrogate or diminish SOCE were investigated in pancreatitis model systems including those of mice, rats and humans. Until now, the potential of selective Orai channel blockers to attenuate AP severity was at the forefront of such investigations. The new study by Son *et al.* refocuses these studies onto a new target, SARAF that represents an endogenous regulatory mechanism. This study reinvigorates the quest to identify effective therapeutic agents to treat acute pancreatitis.

Acknowledgments

Funding: This work was supported by the National Institutes of Health: [P01DK098108 to SJP; R01 AA019954 to AL; and P50-A11999 to SJP].

Footnote

Conflicts of Interest: The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

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Cite this article as: Waldron RT, Lugea A, Pandol SJ. Brake adjustment: Ca²⁺ entry pathway provides a novel target for acute pancreatitis therapy. *Ann Transl Med* 2019;7(Suppl 8):S284. doi: 10.21037/atm.2019.11.119