

# Mining of single nucleotide polymorphisms in the 3' untranslated region of liver cancer-implicated miR-122 target genes

Padmanaban S. Suresh<sup>1</sup>, Thejaswini Venkatesh<sup>2</sup>, Rie Tsutsumi<sup>3</sup>

<sup>1</sup>Department of Biosciences, Mangalore University, Mangalagangotri, Mangalore 574 199, Karnataka, India; <sup>2</sup>Nitte University Centre for Science Education and Research, Nitte University, Derlakatte, Mangalore, Karnataka, India; <sup>3</sup>Department of Nutrition and Metabolism, Institute of Biomedical Science, Tokushima University, Tokushima, Japan

Correspondence to: Padmanaban S. Suresh, Department of Biosciences, Mangalore University, Mangalagangotri, Mangalore 574 199, Karnataka, India. Email: surepadman@rediffmail.com.

Submitted Feb 05, 2016. Accepted for publication Feb 29, 2016.

doi: 10.21037/atm.2016.02.13

View this article at: <http://dx.doi.org/10.21037/atm.2016.02.13>

Currently there has been a lot of focus on the microRNA (miR)-122 to understand the genetics of hepatocellular carcinoma (HCC), pathology of hepatitis B virus (HBV) and hepatitis C virus (HCV) infections, hepatic insulin resistance, and lipid metabolism (1-4). Interestingly, miR-122 expression levels were significantly reduced than the normal levels in HBV-associated liver cancer, but not in HCV related liver cancer (5). Also, there is an overwhelming amount of data suggesting the role of single nucleotide polymorphisms (SNPs) in hepatic genes and its association with the altered risk/development of hepatic cancer and its progression. Polymorphisms in the miRNA-binding sites of the target genes are more frequent than SNPs in miRNA genes and therefore, it is considered that polymorphisms in the cytokines and other genes have correlations with chronic HBV or HCV infections. These SNPs in the miRNA binding sites of the target genes can potentially enhance or weaken the interaction between the miRNAs and the target transcripts. Therefore, it is important to study the SNPs in miR-122 binding sites of the target genes to understand the genetic basis of HCC.

Tumor suppressor miR-122 levels are relatively different in both HBV and HCV infections although both types of infection ultimately can lead to HCC. This is an interesting evidence to look for alternate mechanisms involved in hepatic carcinogenesis. The use of *in silico* prediction and experimental validation will only be the beginning steps in a large scale effort to analyze a variety of clinical liver

cancer samples associated with altered miR-122 levels. Therefore, in this report by using bioinformatics tool we catalogued SNPs in the 3' untranslated region (UTR) of hepatic cancer implicated genes that can affect miR-122 regulation (1, 6). The list of genes in the current study was chosen based on the previous study (1). These genes have miR-122 binding sites and also participate in pathogenesis of HCC as reported by Tsai *et al.*, 2009 (1). Analyses of SNPs and INDELs in miR-122 target sites were performed by using PolymiRTS Database 3.0 that can be accessed at <http://compbio.uthsc.edu/miRSNP/> (6). The Percentage of SNPs in 3' UTR of all the studied genes were obtained by using dbSNP database that can be accessed at <https://www.ncbi.nlm.nih.gov/snp>. The results are shown in *Table 1*. The table provides information about SNPs/INDELs in miR-122 target sites of hepatic cancer implicated genes that are predicted by the PolymiRTS database.

We believe that our report provides the first step towards integrating SNP analysis with studies on miRNA-122 and global de-repression of host miR-122 targets in hepatic cancer cells. This will provide a suitable base for future research that can improve our understanding of 3' UTR polymorphisms and the failure of miR122 regulation in varied clinical samples. Therefore, integrating SNP analysis with studies on miR-122 regulation in liver cancer cells by framing suitable hypotheses and experimental designs can result in the development of novel cancer-targeting therapeutics.

**Table 1** Summary of SNPs in miR-122 target sites of studied genes

Gene symbol	Transcript ID	SNPs (PolymiRTS Database 3.0) and INDELS in miR-122 target sites	% of SNPs in 3' untranslated region (dbSNP database)	Minor allele frequency (MAF)/minor allele count
<i>NUMBL</i>	NM_004756	Nil	6.20	–
<i>FOXJ3</i>	M_001198851	Nil	1.94	–
<i>XPO6</i>	NM_015171	rs28574753	0.75	MAF/minor allele count (A=0.0164/82)
<i>SLC7A1</i>	NM_003045	rs35196293	5.65	NA
<i>STX6</i>	NM_005819	rs190938353	5.72	MAF/minor allele count (A=0.0078/39)
<i>AP3M2</i>	NM_001134296	rs190451219	10.36	MAF/minor allele count (A=0.0020/10)
<i>G6PC3</i>	NM_138387	Nil	8.89	–
<i>GALNT10</i>	NM_198321	Nil	1.68	–
<i>TPD52L2</i>	NM_001243891	Nil	7.05	–
<i>FUNDC2</i>	NM_023934	Nil	6.19	–
<i>MAPK11</i>	NM_002751	rs200443930	14.49	NA
<i>ATP11A</i>	NM_015205	Nil	3.47	–
<i>SORT1</i>	NM_001205228	Nil	6.23	–
<i>ATP1A2</i>	NM_000702	rs78930771	5.15	MAF/minor allele count (T=0.0240/120)
<i>ADAM17</i>	NM_003183	Nil	1.69	–
<i>DUSP2</i>	NM_004418	Nil	10.90	–
<i>OSMR</i>	NM_001168355	Nil	6.61	–
<i>RABIF</i>	NM_002871	Nil	18.34	–
<i>PALM</i>	NM_002579	Nil	3.37	–
<i>AACS</i>	NM_023928	Nil	1.28	–
<i>TBX19</i>	NM_005149	Nil	4.03	–
<i>UBAP2</i>	NM_018449	Nil	0.83	–
<i>EGLN3</i>	NM_022073	Nil	6.75	–
<i>NCAM1</i>	NM_181351	Nil	1.37	–
<i>MECP2</i>	NM_001110792	Nil	8.03	–
<i>CS</i>	NM_004077	rs193099996	3.35	MAF/minor allele count (C=0.0002/1)
<i>FOXP1</i>	NM_001244808	Nil	0.83	–
<i>RAB11FIP1</i>	NM_001002814	Nil	7.77	–
<i>RAB6B</i>	NM_016577	rs200553268	5.89	NA
<i>TRIB1</i>	NM_025195	Nil	15.01	–
<i>TTYH3</i>	NM_025250	Nil	7.92	–
<i>ALDOA</i>	NM_000034	rs1138624, rs12831	2.86	NA
<i>ANXA11</i>	NM_145869	Nil	6.38	–
<i>ENTPD4</i>	NM_001128930	Nil	11.07	–

**Table 1** (continued)

**Table 1** (continued)

Gene symbol	Transcript ID	SNPs (PolymiRTS Database 3.0) and INDELS in miR-122 target sites	% of SNPs in 3' untranslated region (dbSNP database)	Minor allele frequency (MAF)/minor allele count
<i>NFATC2IP</i>	NM_032815	Nil	11.24	–
<i>ANK2</i>	NM_001148	Nil	0.42	–
<i>MEP1A</i>	NM_005588	Nil	1.95	–
<i>NFATC1</i>	NM_006162	Nil	1.83	–
<i>SLC7A11</i>	NM_014331	Nil	5.01	–

SNP, single nucleotide polymorphism.

## Acknowledgements

PS Suresh would like to thank Faculty Recharge Programme of UGC for the support.

## Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

## References

1. Tsai WC, Hsu PW, Lai TC, et al. MicroRNA-122, a tumor suppressor microRNA that regulates intrahepatic metastasis of hepatocellular carcinoma. *Hepatology* 2009;49:1571-82.
2. Yang YM, Seo SY, Kim TH, et al. Decrease of microRNA-122 causes hepatic insulin resistance by inducing protein tyrosine phosphatase 1B, which is reversed by licorice flavonoid. *Hepatology* 2012;56:2209-20.
3. Li C, Wang Y, Wang S, et al. Hepatitis B virus mRNA-mediated miR-122 inhibition upregulates PTTG1-binding protein, which promotes hepatocellular carcinoma tumor growth and cell invasion. *J Virol* 2013;87:2193-205.
4. Luna JM, Scheel TK, Danino T, et al. Hepatitis C virus RNA functionally sequesters miR-122. *Cell* 2015;160:1099-110.
5. Spaniel C, Honda M, Selitsky SR, et al. microRNA-122 abundance in hepatocellular carcinoma and non-tumor liver tissue from Japanese patients with persistent HCV versus HBV infection. *PLoS One* 2013;8:e76867.
6. Bhattacharya A, Ziebarth JD, Cui Y. PolymiRTS Database 3.0: linking polymorphisms in microRNAs and their target sites with human diseases and biological pathways. *Nucleic Acids Res* 2014;42:D86-91.

**Cite this article as:** Suresh PS, Venkatesh T, Tsutsumi R. Mining of single nucleotide polymorphisms in the 3' untranslated region of liver cancer-implicated miR-122 target genes. *Ann Transl Med* 2016;4(5):102. doi: 10.21037/atm.2016.02.13