

# Mesenchymal stem cells in pathogenesis of myelodysplastic syndromes

## Jingya Wang<sup>1</sup>, Zhijian Xiao<sup>1,2</sup>

<sup>1</sup>MDS and MPN Centre, <sup>2</sup>State Key Laboratory of Experimental Hematology, Institute of Hematology and Blood Diseases Hospital, Chinese Academy of Medical Sciences & Peking Union Medical College, Tianjin 300020, China

*Correspondence to:* Zhijian Xiao, MD. The MDS and MPN Centre, Institute of Hematology and Blood Diseases Hospital, Chinese Academy of Medical Sciences, 288 Nanjing Road, Tianjin 300020, China. Email: zjxiao@medmail.com.cn.

**Abstract:** Myelodysplastic syndromes (MDS) are clonal malignant stem cell disorders characterized by inefficient hematopoiesis. The role of the marrow microenvironment in the pathogenesis of the disease has been controversial. Emerging evidence indicated that mesenchymal stem cells (MSC) derived from MDS patients were cytogenetically abnormal, and they showed a deficient hematopoietic-supportive capacity and increased production of cytokine such as tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ), interleukin 6 (IL-6), interferon  $\gamma$  (IFN- $\gamma$ ). From the point of some evidence, the abnormal microenvironment seems to participate in the progression of the disease by contributing to the selective expansion of the malignant clone. In this review, we will discuss the most recent progress related to identification of normal MSC and the importance of the stem cell niche in development and maintenance of MDS.

Keywords: Mesenchymal stem cell (MSC); myelodysplastic syndromes (MDS); pathogenesis

Received: 15 June 2014; Accepted: 18 August 2014; Published: 21 August, 2014. doi: 10.3978/j.issn.2306-9759.2014.08.02 **View this article at:** http://dx.doi.org/10.3978/j.issn.2306-9759.2014.08.02

Myelodysplastic syndromes (MDS) comprise a heterogeneous group of clonal hematopoietic disorders characterized by inefficient hematopoiesis, peripheral blood cytopenia and risk of progression to acute leukemia (1). The clinical heterogeneity of MDS is a reflection of the various pathogenic mechanisms responsible for their development. It is commonly thought that MDS are a neoplasm caused by the neoplastic transformation of hematopoietic stem or myeloid progenitor cells (2,3). However, this hematopoietic cell-centered approach to explain deregulation in MDS has some limitations. It has been very difficult to duplicate disease in in vivo models by transplanting hematopoietic cells from MDS patients into immunodeficient mice (4). This observation has developed a long-standing debate about a potential causative or facilitating role for marrow microenvironment in the pathogenesis of this disease.

Mesenchymal stem cells (MSC), the most important component of marrow microenvironment, have been defined as primitive, non-hematopoietic cells residing primarily in bone marrow, and capable of giving rise to different cell lineages, including fat, muscle, cartilage, bone, and fibroblasts (5-7). Although there are no unique cell surface markers for the identification of MSC, minimal criteria to define human MSC, including plasticadherent;  $\geq$ 95% of the adherent population must express CD105, CD90 and CD73; they must lack expression of CD45, CD34 and CD14; and they must show *in vitro* differentiation capabilities into osteoblasts, adipocytes and chondroblasts have been published (8).

Is the MSC biologically normal in MDS? In terms of morphology, as well as the expression of certain cell markers, no differences were observed between MSC from MDS patients and those derived from normal marrow (9). Initial studies looking at adherent cell layers in Dextertype long-term cultures indicated that the composition and function of such cells were similar to those of their normal counterparts, suggesting that the MDS-derived MSC is normal (10,11). Soenen-Cornu *et al.* (12) tested the osteoblastic differentiation potential of MDS-derived MSC and found that such cells have intact differentiation ability for that particular lineage. Using co-culture studies they also found that MDS-derived MSC have the ability to sustain the growth of MDS-derived CD34<sup>+</sup>CD38<sup>-</sup> and CD34<sup>+</sup>CD38<sup>+</sup> cells, with either normal or abnormal karyotypes. In another study Flores-Figueroa *et al.* (13) have demonstrated that MDS-derived MSC had the capability of differentiating towards osteoblasts, chondrocytes and adipocytes; secreting normal levels of tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ), interleukin 6 (IL-6), SCF, GM-CSF and VEGF; secreting normal levels of collagen and fibronectin; supporting the growth of normal UCB-derived early progenitors (CD34<sup>+</sup>CD38<sup>-</sup>Lin<sup>-</sup> cells) and sustaining the adhesion of progenitor cells.

In contrast several studies have shown a significant proportion (44-55%) of cytogenetic alterations in MDSderived MSC (9,14), and more recent studies involving the use of genomic tools have demonstrated alterations at the gene expression level in these cells (15). Both the hematopoietic cells and MSC can probably be altered in response to damage-inducing factors and harbor independent chromosomal aberrations. Several additional alterations lead to the development of a leukemic clone, which is known to exhibit genomic instabilities and a DNA repair deficiency (16).

Additional studies by different groups showed a deficient hematopoietic-supportive capacity by MDS-derived MSC. Aizawa *et al.* (17) found that in some cases of MDSrefractory anemia, the stromal cells lack a hematopoietic supportive function.

Moreover, several defects in the bone marrow microenvironment may contribute to the pathogenesis of MDS including the aberrant secretion of angiogenic cytokines by monocyte and MSC. Numerous studies, both *in vitro* and *in vivo*, have demonstrated increased levels of cytokine production in the marrow microenvironment of MDS patients, such as TNF $\alpha$ , IL-6, interferon  $\gamma$  (IFN- $\gamma$ ) (18-22). Furthermore, a recent data have provided a diseaseprogression model in which overproduction of IFN- $\gamma$  and TNF $\alpha$  in the microenvironment is the primary event. This causes B7-H1 molecule expression on MDS blasts, which generates a bifunctional signal inducing T-cell apoptosis and enhancing blast proliferation (23).

MSC have immunomodulatory properties, exerted by cell-to-cell contact and in a paracrine pattern (24,25). Several clinical and immunological studies suggest a significant deregulation of the immune system in the complex pathogenesis of MDS. This deregulation may even promote the progression of early MDS to advanced MDS (26). This raised the question of whether MSC play a definite role in the immune abnormalities of MDS. One previous study by Zhao et al. (27) demonstrated that the immunoregulatory function of MDS-MSC were impaired, suggesting the involvement of MSC in the pathogenesis of MDS. In their recent study the MSC from low-risk MDS patients and high-risk MDS patients were compared, and they demonstrated that high-risk MDS-MSC exhibited lower inhibition effect on T cells apoptosis and higher immunosuppressive rate on T-cell proliferation than that of low-risk MDS-MSC. The secretion of cytokines as well as the ability of hematopoietic support was different between the two groups (28). These results illuminated the different immunoregulatory role of MSC in low- and highrisk MDS, which may be important for understanding the pathogenesis of MDS and develop the immunomodulatory treatment of MDS.

Recently, other evidence was provided that primary alterations in the bone marrow microenvironment can drive oncogenesis in hematopoietic system. These findings suggest that a specific deletion of DICER1 in MDS-derived osteoprogenitors triggers blood cytopenias, high apoptosis rate, myelodysplasia and subsequent AML development in a murine model (29). Then Santamaría et al. (30) demonstrated an impaired microRNA biogenesis in human MSC from MDS patients, where DICER1 and DROSHA gene and protein downregulation correlated to a gene and microRNA abnormal expression profile, validating the animal model results previously described. These findings support the mechanism of malignancy resulting from the interaction of cell autonomous and microenvironmentally determined events, and point to the microenvironment as the site of the initiating event that leads to secondary genetic changes in other cells.

Overall, there still remains one key question: what is the origin of such abnormalities of MSC? Three explanation of MSC contribution in MDS initiation and progression are possible. First, it may be hypothesized that marrow microenvironment acts as an initiator of oncogenesis. The disruption of hematopoiesis integrity induced by the microenvironment may enable the hematopoietic progenitor cells initiating genetic events or clonal transformation (4). A second possible explanation is that the same agent that caused the initial genetic damage in myelodysplastic hematopoietic cells (radiation exposure, aromatic compounds, viruses, etc.) also affected MSC, although not necessarily inducing the same type of alterations. And the abnormal microenvironment may provide a selective milieu

#### Stem Cell Investigation, August 18, 2014

for clonal expansion of hematopoietic stem cells. Thirdly, recent data support the view that in MDS, reciprocal heterotypic signaling between disease-propagating hematopoietic cells and MSC within the bone marrow environment is required to drive disease initiation and progression. "Reprogramming" of MSC by hematopoietic cells may be the initiative step and subsequently followed by a cascade of yet-to-be-defined events facilitating disease progression (31,32). Up to now, we do not yet have the precise answers to the above question; however, there is no doubt that actual explanation to this issue will come out in the near future.

### Acknowledgements

None.

### Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

#### References

- Malcovati L, Porta MG, Pascutto C, et al. Prognostic factors and life expectancy in myelodysplastic syndromes classified according to WHO criteria: a basis for clinical decision making. J Clin Oncol 2005;23:7594-603.
- Nimer SD. MDS: a stem cell disorder--but what exactly is wrong with the primitive hematopoietic cells in this disease? Hematology Am Soc Hematol Educ Program 2008:43-51.
- Tefferi A, Vardiman JW. Myelodysplastic syndromes. N Engl J Med 2009;361:1872-85.
- Raaijmakers MH. Niche contributions to oncogenesis: emerging concepts and implications for the hematopoietic system. Haematologica 2011;96:1041-8.
- Caplan AI. Mesenchymal stem cells. J Orthop Res 1991;9:641-50.
- Pittenger MF, Mackay AM, Beck SC, et al. Multilineage potential of adult human mesenchymal stem cells. Science 1999;284:143-7.
- Dazzi F, Ramasamy R, Glennie S, et al. The role of mesenchymal stem cells in haemopoiesis. Blood Rev 2006;20:161-71.
- Dominici M, Le Blanc K, Mueller I, et al. Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position

statement. Cytotherapy 2006;8:315-7.

- Flores-Figueroa E, Arana-Trejo RM, Gutiérrez-Espíndola G, et al. Mesenchymal stem cells in myelodysplastic syndromes: phenotypic and cytogenetic characterization. Leuk Res 2005;29:215-24.
- Coutinho LH, Geary CG, Chang J, et al. Functional studies of bone marrow haemopoietic and stromal cells in the myelodysplastic syndrome (MDS). Br J Haematol 1990;75:16-25.
- Hirayama Y, Kohgo Y, Matsunaga T, et al. Cytokine mRNA expression of bone marrow stromal cells from patients with aplastic anaemia and myelodysplastic syndrome. Br J Haematol 1993;85:676-83.
- Soenen-Cornu V, Tourino C, Bonnet ML, et al. Mesenchymal cells generated from patients with myelodysplastic syndromes are devoid of chromosomal clonal markers and support short- and long-term hematopoiesis in vitro. Oncogene 2005;24:2441-8.
- Flores-Figueroa E, Montesinos JJ, Flores-Guzmán P, et al. Functional analysis of myelodysplastic syndromes-derived mesenchymal stem cells. Leuk Res 2008;32:1407-16.
- Blau O, Hofmann WK, Baldus CD, et al. Chromosomal aberrations in bone marrow mesenchymal stroma cells from patients with myelodysplastic syndrome and acute myeloblastic leukemia. Exp Hematol 2007;35:221-9.
- Roela RA, Carraro DM, Brentani HP, et al. Gene stagespecific expression in the microenvironment of pediatric myelodysplastic syndromes. Leuk Res 2007;31:579-89.
- Krämer A, Neben K, Ho AD. Centrosome replication, genomic instability and cancer. Leukemia 2002;16:767-75.
- Aizawa S, Nakano M, Iwase O, et al. Bone marrow stroma from refractory anemia of myelodysplastic syndrome is defective in its ability to support normal CD34-positive cell proliferation and differentiation in vitro. Leuk Res 1999;23:239-46.
- Kitagawa M, Saito I, Kuwata T, et al. Overexpression of tumor necrosis factor (TNF)-alpha and interferon (IFN)-gamma by bone marrow cells from patients with myelodysplastic syndromes. Leukemia 1997;11:2049-54.
- 19. Wetzler M, Kurzrock R, Estrov Z, et al. Cytokine expression in adherent layers from patients with myelodysplastic syndrome and acute myelogenous leukemia. Leuk Res 1995;19:23-34.
- Deeg HJ, Beckham C, Loken MR, et al. Negative regulators of hemopoiesis and stroma function in patients with myelodysplastic syndrome. Leuk Lymphoma 2000;37:405-14.
- 21. Shetty V, Mundle S, Alvi S, et al. Measurement of apoptosis,

#### Wang and Xiao. MSC in pathogenesis of MDS

#### Page 4 of 4

proliferation and three cytokines in 46 patients with myelodysplastic syndromes. Leuk Res 1996;20:891-900.

- 22. Allampallam K, Shetty V, Hussaini S, et al. Measurement of mRNA expression for a variety of cytokines and its receptors in bone marrows of patients with myelodysplastic syndromes. Anticancer Res 1999;19:5323-8.
- Kondo A, Yamashita T, Tamura H, et al. Interferongamma and tumor necrosis factor-alpha induce an immunoinhibitory molecule, B7-H1, via nuclear factorkappaB activation in blasts in myelodysplastic syndromes. Blood 2010;116:1124-31.
- Tse WT, Pendleton JD, Beyer WM, et al. Suppression of allogeneic T-cell proliferation by human marrow stromal cells: implications in transplantation. Transplantation 2003;75:389-97.
- Aggarwal S, Pittenger MF. Human mesenchymal stem cells modulate allogeneic immune cell responses. Blood 2005;105:1815-22.
- Epperson DE, Nakamura R, Saunthararajah Y, et al. Oligoclonal T cell expansion in myelodysplastic syndrome: evidence for an autoimmune process. Leuk Res 2001;25:1075-83.
- 27. Zhao ZG, Li WM, Chen ZC, et al. Immunosuppressive

### doi: 10.3978/j.issn.2306-9759.2014.08.02

**Cite this article as:** Wang J, Xiao Z. Mesenchymal stem cells in pathogenesis of myelodysplastic syndromes. Stem Cell Investig 2014;1:16. properties of mesenchymal stem cells derived from bone marrow of patient with hematological malignant diseases. Leuk Lymphoma 2008;49:2187-95.

- Zhao Z, Wang Z, Li Q, et al. The different immunoregulatory functions of mesenchymal stem cells in patients with low-risk or high-risk myelodysplastic syndromes. PLoS One 2012;7:e45675.
- 29. Raaijmakers MH, Mukherjee S, Guo S, et al. Bone progenitor dysfunction induces myelodysplasia and secondary leukaemia. Nature 2010;464:852-7.
- Santamaría C, Muntión S, Rosón B, et al. Impaired expression of DICER, DROSHA, SBDS and some microRNAs in mesenchymal stromal cells from myelodysplastic syndrome patients. Haematologica 2012;97:1218-24.
- Medyouf H, Mossner M, Jann JC, et al. Myelodysplastic cells in patients reprogram mesenchymal stromal cells to establish a transplantable stem cell niche disease unit. Cell Stem Cell 2014;14:824-37.
- Raaijmakers MH. Disease progression in myelodysplastic syndromes: do mesenchymal cells pave the way? Cell Stem Cell 2014;14:695-7.