

The association of myelination in the internal capsule with iron deposition in the basal ganglia in macaques: a magnetic resonance imaging study

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Background: Iron plays a vital role in myelin synthesis and maintenance. A tight association between iron concentration and myelin content has been demonstrated in local brain regions; however, whether such a relationship exists between distant brain regions that are anatomically connected is largely unknown.

Methods: We conducted an *in vivo* measurement of iron and myelin content in the brains of 8 young (mean age: 7.7 years) and 8 old (mean age: 24.7 years) macaques by integrating two MRI-based techniques: quantitative susceptibility mapping (QSM) and myelin water fraction (MWF) imaging. We examined the relationship between iron deposition in components of the basal ganglia (BG), and the myelin content of the BG-connecting fiber tract internal capsule (IC) and four more white matter (WM) structures, including the optic tract, and the genu, body, and splenium of the corpus callosum, which are anatomically separate from the BG.

Results: Spearman's correlation analysis revealed a moderate to high (r=0.528–0.808, P<0.05) positive correlation between the magnetic susceptibility of the BG and the MWF of anatomically connected IC structures during myelin production and maintenance, but little significant correlation was found between the susceptibility of the BG and the MWF of WM structures not anatomically connected to the BG.

Conclusions: These results advance the understanding of the relationship between iron and myelin, and suggest that future studies should consider the impact that iron concentration in the BG has on the myelination of WM structures that are anatomically connected to the BG.

Keywords: Iron; myelin; basal ganglia (BG); internal capsule (IC); macaque; MRI

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Introduction

Myelin and iron each play an important role in maintaining normal brain function and metabolic activity. In mammalian central nervous systems, myelin is produced by oligodendrocytes (1). Aggregated evidence has shown that myelination of white matter (WM) continues into adulthood, and myelin degeneration occurs with old age (2-4), which brain iron is essential for myelin synthesis

throughout the lifespan of mammals (5). Iron deficiency early in life can lead to hypomyelination, while excess iron deposition can cause oxidative stress, inflammation, and demyelination (6). The findings of neuroanatomical studies on brain iron and myelin have been mixed, but have consistently suggested that iron is progressively deposited in the brain with age, and accumulates largely in basal ganglia (BG) regions, where pathological changes often result in a wide range of neurological and psychiatric diseases (7-9). The BG, which notably consists of the globus pallidus (GP), the putamen (PU), the substantia nigra, the subthalamic nucleus, and the caudate nucleus (CN), contains the greatest amount of iron in the brain (10,11). The BGconnecting fiber tract internal capsule (IC) is a highly myelinated WM structure which is anatomically connected to the BG. This structure consists of projection fibers that pass through the BG and is located between the lentiform nucleus (which comprises the PU and GP) laterally, and between the thalamus and CN medially (12,13). The IC can be divided into several structures, including the anterior limb of the internal capsule (ALIC) and the posterior limb of the internal capsule (PLIC). Previous research has demonstrated that there is a close association between myelin and iron (14,15). A relatively high correspondence between iron concentration and myelin content has been observed in most gray matter (GM) regions, whereas there is generally a low correspondence between iron and myelin in WM, the thalamus, and BG regions (11). A previous aging study reported a negative correlation between iron and myelin content in the ventral striatum (16). To date, the relationship between iron concentration and myelin content in the BG and its connecting fiber tracts remains essentially unexplored.

It is technically challenging to perform an *in vivo*, noninvasive, quantitative measurement of myelin and iron content in the human brain. Magnetic resonance imaging (MRI)-based technologies are considered to be a promising path in finding a resolution for this issue (17,18). Of these MRI-based technologies, calculating myelin water fraction (MWF) using T_2^* decay signals acquired from a multiple gradient echo (mGRE) sequence is a fast method for wholebrain myelin water imaging (19) in which a three-pool model of WM (20) is used for multi-exponential analysis of T_2^* decay data. This three-pool model assumes that WM is made up of a myelin water pool (short T_2^* component), an axonal water pool (medium T_2^* component), and an extracellular water pool (long T_2^* component) (21,22). MWF is defined as the ratio of myelin water component to total water component (23). Quantitative susceptibility mapping (OSM), on the other hand, measures the bulk magnetic susceptibility of tissue using phase images of mGRE sequence (24) and reveals the quantity of iron, because iron is the main source of magnetic susceptibility in subcortical GM (25,26). Iron can be transported between different areas of the brain along axons and dendrites (27,28). There are two primary forms of transported iron in the brain: transferrin-bound iron and non-transferrinbound iron (6). Disorders such as Alzheimer's disease (AD) and Parkinson's disease (PD) are known to involve myelin degeneration, which leads to the disruption of brain iron homeostasis (5,29). Nevertheless, the relationship between iron concentration and myelin content in distant but anatomically connected brain regions has rarely been examined, even though an understanding of the connection between iron in the BG and myelin in the IC is crucial for investigating the physiological mechanisms of normal aging and some neurodegenerative diseases further.

In the present study, we measured brain iron concentration in young and old adult macaques using QSM, focusing primarily on BG areas, including the CN, GP, and PU. In addition, using the MWF technique, we measured the myelin content of the BG-connecting fiber tracts, including the ALIC and PLIC. The myelin content of four major WM structures that are anatomically separate from the BG, including the optic tract (OT), and the genu, body, and splenium of the corpus callosum (GCC, BCC, and SCC), were also measured as controls.

Methods

Animal preparation

The subjects of the present study were 16 healthy adult macaques (*Macaca fascicularis*). The macaques were divided into 2 groups by age: the young group (n=8, 6 males and 2 females), with a mean age of 7.7 (range: 6.7–8.7) years old; and the old group (n=8, 6 males and 2 females), with a mean age of 24.7 (range: 23.4–26) years old. All 16 macaques were prepared similarly to our previous work (30-32) and maintained in a stable physiological state for MRI scanning. Before each scanning session, the animals were anesthetized with an intramuscular injection of ketamine (10 mg/kg) and atropine sulfate (0.05 mg/kg). After intubation, the animals were ventilated with an MRI-compatible ventilator (CWE Inc., Weston, Wisconsin). To ensure a constant respiratory rate (25–35 breaths/min), the macaques were

maintained with intermittent positive-pressure ventilation. Local anesthetic (5% lidocaine cream) was applied around the macaques' ears to block peripheral nerve stimulation. The macaques were then placed in a custom-built MRIcompatible stereotaxic frame in a sphinx position and their heads were secured before being inserted into the center of the scanner. The animals' vital signs, including blood oxygenation, ECG, rectal temperature (Small Animal Instruments, Inc., Stony Brook, New York), respiratory rate, and end-tidal CO2 (Smiths Medical ASD Inc., Dublin, Ohio) were monitored throughout the experiment. Oxygen saturation was kept at over 95% and body temperature was kept constant (39±1 °C) using a hot water blanket (Gaymar Industries Inc., Orchard Park, New York). During the anesthesia process, lactated Ringer's solution was given at a maximum rate of 10 mL/kg/hour. All experimental procedures in this study were approved by the Institute of Neuroscience Animal Care and Use Committee and by the Shanghai Institute for Biological Sciences Biomedical Research Ethics Committee, and they all conformed to the National Institutes of Health guidelines for the humane care and use of laboratory animals.

MRI acquisition

MRI images of all subjects were acquired using a 3T wholebody scanner (MAGNETOM Trio, Siemens Healthcare A.G., Erlangen, Germany) with an enhanced gradient insert (AC88; 80 mT/m maximum gradient strength, 800 mT/m/ms maximum slew rate) and a customized eight-channel phasedarray coil, which was designed specifically for animal imaging. Image data for QSM and MWF analysis were acquired using a customized 3D mGRE sequence with a bipolar readout gradient. The sequence parameters were as follows: repetition time (TR) 60 ms, the first echo time (TE₁) 2.4 ms, echo spacing (Δ TE) 1.42 ms, number of echoes (#Echo) 32, matrix size 128×128×52, acquisition voxel size 1×1×1 mm³, readout bandwidth 930 Hz/pixel, flip angle 25°, 6 averages, and the acquisition time was 40 minutes.

Computation of parametric maps

The processing procedure for the mGRE data was performed in MATLAB (R2014a, MathWorks, Natick, MA, USA). The QSM computation was conducted with the STISuite v3.0 toolbox (https://people.eecs.berkeley. edu/~chunlei.liu/software.html). The 8 echoes of 1, 5, 9, 13, 17, 21, 25, and 29 acquired from the mGRE sequence

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were chosen to keep the same readout polarity in order to avoid phase fluctuation between even and odd echoes due to echo shift caused by eddy currents. The susceptibility values obtained by this method were verified to strongly correlate with those obtained by traditional QSM protocol (21). Phase images of these echoes were unwrapped using a Laplacian-based phase unwrapping method (33); the V-SHARP method (34) was applied for background field removal; and a fast STAR-QSM algorithm (35) was applied for QSM calculation.

MWF maps were also calculated using mGRE data, employing the proposed method of incorporating local tissue susceptibility to acquire high resolution myelin water imaging (21). A three-component complex model (21) was used to characterize the signal decay in WM as follows:

$$\mathbf{S}(\mathbf{t}) = \left[A_{my} e^{-\left(\frac{1}{T_{2,my}^*} + j2\pi f_{my}\right)t} + A_{ax} e^{-\left(\frac{1}{T_{2,ax}^*} + j2\pi f_{ax}\right)t} + A_{ex} e^{-\left(\frac{1}{T_{2,ax}^*} + j2\pi f_{ex}\right)t} \right] e^{j\varphi_0} \quad [1]$$

In this equation, S(t) represents the complex signal as a function of time, and A and *f* represent signal magnitudes and frequency offsets of each water component, including *my* (myelin water), *ax* (axonal water), and *ex* (extracellular water). φ_0 is the initial phase offset. Subsequently, the phase component of the complex data from the mGRE sequence was replaced by the following:

$$\varphi_i = \frac{4}{3} \pi \chi \gamma B_0 T E_i$$
^[2]

In this equation, χ is the susceptibility value obtained by QSM, γ is the gyromagnetic ratio, and *i* is the echo number. A complex dataset was created by this new phase component and the magnitude component filtered by 3D anisotropic diffusion filter (ADF), the parameter κ of 3D ADF being $\delta\sigma$ (σ is the noise standard deviation in the 3D volumetric image acquired from the first echo image). This synthesized signal was fitted to Eq. [1]. An iterative nonlinear curvefitting method ('lsqcurvefit' function in MATLAB) was used to estimate the complex model parameters. *Table 1* shows the initial values and searching boundaries of each parameter (21,22). Finally, MWF was defined as the fraction of myelin water signal amplitude over the total water signal amplitude; the following equation was used:

$$MWF = A_{my} / (A_{my} + A_{ax} + A_{ex})$$
[3]

Definition of ROIs

Firstly, non-brain tissue was removed using the brain extraction tool (BET) in FSL software (36). To conduct

Table 1 initial values and upper and lower boundary inting to Eq. [1]											
Fitting parameters	A _{my}	A _{ax}	A_{ex}	$T^{*}_{2,my}$ (ms)	$\pmb{T}^{*}_{2,\textit{ax}}$ (ms)	$\pmb{T}^{*}_{2,\textit{ex}}$ (ms)	f _{my} (Hz)	f _{ax} (Hz)	f _{ex} (Hz)	ϕ_0	
Initial value	0.1 S ₁	0.6 S ₁	0.3 S ₁	10	64	48	5	0	0	$\angle \left S_{1} \right $	
Lower bound	0	0	0	3	24	24	-75	-25	-25	-π	
Upper bound	2 S ₁	2 S ₁	2 S ₁	24	1,000	1,000	75	25	25	π	

Table 1 Initial values and upper and lower boundary fitting to Eq. [1]

 S_1 is the complex signal of the first echo.



Figure 1 The registered ROIs from 'INIA19 atlas' overlaid on the first echo image acquired from mGRE sequence of a representative macaque. 1: PLIC, the posterior limb of internal capsule; 2: PU, putamen; 3: ALIC, the anterior limb of internal capsule; 4: CN, caudate nucleus; 5: GP, globus pallidus; 6: OT: optic tract; 7: GCC, the genu of corpus callosum; 8: BCC, the body of corpus callosum; 9: SCC, the splenium of corpus callosum. L: left; R: right.

ROI analysis of MWF values in WM and QSM values in the BG, the first echo image acquired from the mGRE sequence was used for segmentation to obtain WM, GM, and cerebrospinal fluid (CSF) tissue maps by means of FSL FAST module (37). To minimize partial volume artifacts, only voxels that had a WM or GM partial volume fraction larger than 0.95 were included in the final binary WM or GM tissue masks.

To measure the MWF and QSM values in well-defined ROIs, the masks of 10 WM ROIs, being the GCC [left (L) and right (R)], BCC (L and R), SCC (L and R), IC (L and R), and OT (L and R), and six BG ROIs, being the CN (L and R), GP (L and R), and PU (L and R), were derived from the INIA19 atlas (38). The IC is V-shaped on the axial (horizontal) sections, with a knee-bend around the apex of the lentiform nucleus. The IC was manually divided into two sections (ALIC and PLIC) using a line at the inflection point (39). All ROIs from the INIA19 atlas were coregistered to the first echo image from the mGRE sequence by SPM8 (Wellcome Trust Centre for Neuroimaging, London, UK). After the ROIs had been inspected for gross registration errors, MWF or QSM values were acquired for all ROIs in each macaque (as shown in *Figure 1*). Additionally, to further reduce ROI registration errors when applying WM ROIs from the INIA19 atlas, the MWF map was first masked by a WM tissue mask, and when applying BG ROIs from the INIA19 atlas, the QSM was first masked by WM and GM tissue masks, because many regions of BG were attributed as WM by FAST. The mean values of MWF in WM ROIs and the mean values of QSM in BG ROIs were recorded for each macaque.

Statistical analysis

Statistical analysis was performed with SPSS Version 20 (IBM Inc., Armonk, NY, USA). After confirming that MWF and susceptibility data in all ROIs were normally distributed, an independent sample *t*-test was used to



Figure 2 Typical QSM and MWF maps obtained from one of the young macaques (a) and one of the old macaques (b). The black arrows indicate CN (L: left), PU (L), and GP (L) respectively [QSM (a, b)]. The white arrows indicate GCC, BCC, SSC, ALIC (L), PLIC (L) and OT (R: right) respectively [MWF (a, b)]. QSM, quantitative susceptibility mapping; MWF, myelin water fraction; CN, caudate nucleus; GP, globus pallidus; PU, putamen; GCC, the genu of corpus callosum; BCC, the body of the corpus callosum; SCC, the splenium of the corpus callosum; ALIC, the anterior limb of the internal capsule; PLIC, the posterior limb of the internal capsule; OT, optic tract.

investigate differences in MWF and susceptibility between the young and old macaque groups. A paired sample *t*-test was performed to investigate differences in MWF and susceptibility between the left and right hemispheres. Also, to assess the relationship between myelin in WM ROIs and iron in BG ROIs, Spearman's rank-order correlation analysis (two-tailed) was performed on the mean susceptibility of BG ROIs and the mean MWF in WM ROIs both in anatomically connected and anatomically separate WM structures. A difference of P<0.05 for any given *t*-test or Spearman's correlation was considered statistically significant.

Results

ROI analysis results

Figure 2 shows examples of QSM and MWF maps for two representative macaques, one from the young group and one from the old group. The results of the regional MWF analysis and the regional susceptibility analysis between the young and old groups are summarized in detail in *Table 2*

Table 2 Summary of mean values and standard deviations (SD) of the MWF of WM ROIs for the young (N=8) and old (N=8) mac	aques
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MWF (%) -	Left h	nemisphere		Right	hemisphere	Between hemispheres		
	Young (mean ± SD)	Old (mean \pm SD)	P value	Young (mean ± SD)	Old (mean \pm SD)	P value	Young (P value)	Old (P value)
ALIC	10.083±5.184	17.982±4.028	0.004	9.245±3.456	18.901±3.368	<0.001	0.443	0.289
PLIC	13.694±2.940	16.516±2.556	0.060	13.269±2.855	17.793±2.062	0.003	0.291	0.115
GCC	8.160±1.807	10.950±3.198	0.050	8.087±2.008	10.334±3.545	0.141	0.896	0.401
BCC	11.453±2.569	13.589±3.513	0.187	11.262±2.593	13.347±2.654	0.134	0.390	0.708
SCC	11.364±2.576	12.682±3.884	0.437	10.633±2.558	13.444±2.914	0.059	0.460	0.622
ОТ	14.295±3.097	16.461±2.141	0.126	13.712±3.484	16.933±2.736	0.059	0.510	0.640

MWF, myelin water fraction; WM, white matter; ALIC, the anterior limb of internal capsule; PLIC, the posterior limb of internal capsule; GCC, the genu of the corpus callosum; BCC, the body of the corpus callosum; SCC, the splenium of the corpus callosum; OT, optic tract.

Table 3 Summary of mean values and standard deviations (SD) of the susceptibility of BG ROIs for the young (N=8) and old (N=8) macaques

Susceptibility	Left h	emisphere		Right	hemisphere	Between hemispheres		
(ppm)	Young (mean ± SD)	Old (mean ± SD)	P value	Young (mean ± SD)	Old (mean ± SD)	P value	Young (P value)	Old (P value)
GP	0.024±0.017	0.107±0.070	0.012	0.029±0.015	0.101±0.074	0.029	0.093	0.296
PU	0.003±0.007	0.024±0.011	<0.001	0.002±0.006	0.024±0.019	0.016	0.670	0.952
CN	-0.005±0.006	0.009±0.011	0.011	-0.0078±0.004	0.013±0.011	0.001	0.199	0.215

BG, basal ganglia; GP, globus pallidus; PU, putamen; CN, caudate nucleus.

and Table 3, respectively. The mean MWF of the old group was larger than that of the young group for each WM ROI, but significant intergroup differences were only found in the ALIC (L: t=3.403, P=0.004; R: t=5.659, P<0.001) and PLIC (R: t=3.634, P=0.003). Looking at the MWF of WM ROIs in the young group, the highest value was in the OT, followed in descending order by the PLIC, BCC, SCC, ALIC, and GCC. This held true for both the left and right hemispheres. However, considering the MWF of WM ROIs in the old group, the highest value was in the ALIC, followed in descending order by the PLIC, OT, BCC, SCC, and GCC. This held true for both left and right hemispheres. Moreover, no significant intergroup differences in MWF values were found between the left and right hemispheres in all ROIs in both the young and old groups. The mean susceptibility values of the old group were significantly higher than those of the young group in the GP (L: t=3.268, P=0.012; R: t=2.698, P=0.029), PU (L: t=4.564, P<0.001; R: t=2.986, P=0.016) and CN (L: t=3.075, P=0.011; R: t=4.860, P=0.001). In both the left and right hemispheres, the mean susceptibility values in the young and the old groups were highest in the GP, followed by the PU and the CN. There were no significant intergroup

differences in susceptibility values between the left and right hemispheres in either group.

Correlation analysis results

Spearman's correlations between the mean MWF of WM ROIs and the mean susceptibility of BG ROIs are presented in *Table 4. Figure 3* shows the scatterplots of correlations between the mean MWF of WM ROIs (ALIC and PLIC) and the mean susceptibility of BG ROIs for all macaques.

In the young group, a significant correlation (r=-0.766, P=0.027) was only found between the mean MWF of the GCC (R) and mean susceptibility of the CN (L). In the old group, there were significant positive correlations between the mean susceptibility of the CN (L) and the mean MWF of the ALIC (L: r=0.714, P=0.047, *Figure 3A*; R: r=0.810, P=0.015, *Figure 3B*), and there were significant positive correlations between the mean Susceptibility of the CN (R) and the mean MWF of the ALIC (L: r=0.714, P=0.047, *Figure 3E*) and the PLIC (R: r=0.714, P=0.047, *Figure 3H*). There were significant negative correlations between the mean susceptibility of the GP (L) and the mean MWF of the GCC (L: r=-0.929, P=0.001; R: r=-0.738, P=0.037)

Table 4 Results of Spearman correlation analysis between the MWF of WM ROIs and the susceptibility of BG ROIs

	MWF													
r		GCC		BCC		S	SCC		ОТ		ALIC		PLIC	
	ROI	L	R	L	R	L	R	L	R	L	R	L	R	
Susceptibility	CN													
(young), N=8	L	-0.299	-0.766*	-0.323	-0.216	0.228	-0.036	0.240	0.180	0.168	-0.108	0.060	-0.240	
	R	0.108	-0.335	-0.252	0.024	-0.156	-0.587	0.096	-0.395	0.192	0.012	0.299	0.168	
	GP													
	L	0.060	0.216	0.084	0.168	0.252	-0.491	-0.359	-0.371	0.216	0.036	0.204	0.168	
	R	0.395	0.072	0.132	0.263	0.323	-0.299	-0.311	-0.204	0.335	0.108	0.275	0.096	
	PU													
	L	0.156	-0.240	-0.012	0.048	0.443	-0.036	-0.168	0.156	0.383	-0.108	-0.084	-0.048	
	R	0.275	-0.072	-0.228	-0.216	0.036	-0.180	-0.503	-0.132	0.096	-0.132	-0.252	-0.359	
Susceptibility	CN													
(old), N=8	L	-0.167	0.000	0.119	0.238	0.619	-0.024	-0.286	-0.619	0.714*	0.810*	0.690	0.143	
	R	-0.429	-0.095	0.024	-0.119	0.167	-0.286	0.286	-0.167	0.738*	0.619	0.619	0.714*	
	GP													
	L	-0.929**	* –0.738*	-0.262	-0.524	-0.095	-0.738*	0.476	0.262	0.524	0.190	0.286	0.548	
	R	-0.786*	-0.548	-0.357	-0.548	-0.214	-0.524	0.357	-0.048	0.619	0.429	0.190	0.548	
	PU													
	L	-0.667	-0.333	-0.071	-0.286	-0.095	-0.524	0.548	-0.238	0.667	0.500	0.238	0.381	
	R	-0.619	-0.381	-0.500	-0.476	-0.095	-0.095	0.190	0.095	0.429	0.286	0.143	0.762*	
Susceptibility	CN													
(pooled), N=16	L	0.227	0.081	0.233	0.346	0.467	0.330	0.300	0.244	0.776***	0.731**	0.581*	0.468	
	R	0.359	0.160	0.235	0.290	0.155	0.182	0.462	0.262	0.808**	0.808***	0.623*	0.668**	
	GP													
	L	0.174	0.069	0.277	0.278	0.269	0.026	0.391	0.347	0.773***	0.687**	0.537*	0.677**	
	R	0.224	0.069	0.230	0.255	0.231	0.071	0.350	0.265	0.799***	0.723**	0.528*	0.648**	
	PU													
	L	0.353	0.128	0.259	0.293	0.199	0.265	0.403	0.362	0.808***	0.805**	0.458	0.603*	
	R	0.147	-0.013	-0.091	0.004	0.016	0.162	0.121	0.144	0.628**	0.631**	0.260	0.453	

MWF, myelin water fraction; WM, white matter; BG, basal ganglia; GCC, the genu of corpus callosum; BCC, the body of the corpus callosum; SCC, the splenium of the corpus callosum; OT, optic tract; ALIC, the anterior limb of the internal capsule; PLIC, the posterior limb of the internal capsule; CN, caudate nucleus; GP, globus pallidus; PU, putamen; r, correlation coefficient; L, left hemisphere; R, right hemisphere. *P<0.05, **P<0.01, ***P<0.001.



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positively correlated with CN (R: H), GP (L: L, R: P) and PU (L: T). The black lines represent linear regression of pooled data, the blue lines represent linear regression of Figure 3 The scatterplots of correlations between the susceptibilities in BG ROIs [CN (L, R), GP (L, R) and PU (L, R)] and the MWF in anatomically connected WM ROIs [ALIC (L, R) and PLIC (L, R)] for all macaques. ALIC (L) was positively correlated with CN (L: A, R: E), GP (L: I, R: M) and PU (L: Q, R: U). ALIC (R) was also positively correlated with CN (L: B, R: F), GP (L: J, R: N) and PU (L: R, R: V). PLIC (L) was positively correlated with CN (L: C, R: G) and GP (L: K, R: O). PLIC (R) was also the old group, the green lines represent linear regression of the young group. *, P<0.05; **, P<0.01; ***, P<0.001.

and the SCC (R: r=-0.738, P=0.037). A significant negative correlation (r=-0.786, P=0.021) was found between the mean susceptibility of the GP (R) and the mean MWF of the GCC (L). There was also a significant positive correlation between mean susceptibility of the PU (R) and mean MWF of the PLIC (R: r=0.762, P=0.028, *Figure 3X*).

In the pooled group, the MWF of the ALIC (L) was positively correlated with the mean susceptibility of the CN (L: r=0.776, P<0.001, Figure 3A; R: r=0.808, P<0.001, Figure 3E), GP (L: r=0.773, P<0.001, Figure 3I; R: r=0.799, P<0.001, Figure 3M) and PU (L: r=0.808, P<0.001, Figure 3Q; R: r=0.628, P=0.009, Figure 3U), and the MWF of the ALIC (R) was also positively correlated with the mean susceptibility of the CN (L: r=0.731, P=0.001, Figure 3B; R: r=0.808, P<0.001, Figure 3F), GP (L: r=0.687, P=0.003, Figure 37; R: r=0.723, P=0.002, Figure 3N) and PU (L: r=0.805, P<0.001, Figure 3R; R: r=0.631, P=0.009, Figure 3V). The MWF of the PLIC (L) was positively correlated with the mean susceptibility of the CN (L: r=0.581, P=0.018, Figure 3C); R: r=0.623, P=0.01, Figure 3G) and GP (L: r=0.537, P=0.032, Figure 3K); R: r = 0.528, P=0.035, Fig. 30), and the MWF of the PLIC (R) was positively correlated with the mean susceptibility of the CN (R: r=0.668, P=0.005, Figure 3H), GP (L: r=0.677, P=0.004, Figure 3L); R: r=0.648, P=0.007, Figure 3P) and PU (L: r=0.603, P=0.013, Figure 3T). Of all the significantly correlated ROIs in the pooled group, the correlation coefficients between the mean MWF of the ALIC and the mean susceptibility of BG ROIs were higher than those between the mean MWF of the PLIC and the mean susceptibility of BG ROIs.

Discussion

To the best of our knowledge, the present study is the first to investigate non-local associations between iron concentration in the BG and myelin content of WM in non-human primates. Our analyses of 16 (8 young and 8 old) macaques demonstrated significant associations between the MWF of the IC and the susceptibility of the CN, the GP, and the PU, while few significant correlations were observed between the susceptibility of the three abovementioned BG ROIs and the MWF of WM ROIs that are anatomically separate from the BG. These findings deepen the current understanding of the anatomical connection between iron and myelin.

Macaque brains are insusceptible to the psychopathologies that can affect the human brain (40) and are therefore useful models for studying the interactions between myelin and iron under normal conditions, which could better control any biases caused by the potential presence of disease such as AD. Additionally, according to previous human research (2,41), the MWF of WM demonstrates an inverted U-shaped trend with age, with the peak values occurring at approximately the fourth to sixth decade of life. In another study of rhesus macaques (42), the thickness of myelin in layer 4 of the visual cortex was compared between young macaques aged 4 to 9 years and macaques aged 24 years or older, and was found to increase significantly with age. The old macaques, which were comparable to humans of approximately 60 years of age (43), had significantly higher MWF values than the young macaques, which were comparable to humans of approximately 20 years of age (43), in the ALIC (L and R) and PLIC (R). However, no significant differences in MWF were observed between the young and old groups in the GCC, BCC, SCC, and OT, suggesting that myelin is still in the stages of production and maintenance in these WM ROIs. However, previous diffusion tensor imaging studies of brain WM (44) and the optic nerve (45) of macaques showed age-related decreases in fractional anisotropy and increases in radial diffusivity. Another ex vivo diffusion spectrum imaging study (46) found that transcallosal connectivity declined in older macaques, which suggested that brain degeneration had occurred.

Additionally, the mean magnetic susceptibility of the old macaque group was significantly higher than that of the young group in the GP, PU, and CN in both the left and right hemispheres, which suggests that significant deposits of iron occur in these structures with age. This finding is consistent with those of previous human studies (8,9). In the present study, the GP had the highest susceptibility, followed by the PU and CN, in both the young and old macaque groups. This is also consistent with the order in the human brain (10).

Spearman's correlation analysis demonstrated that the susceptibility of the BG is positively correlated with the MWF of anatomically connected IC structures during myelin production and maintenance. The correlation between the MWF of the ALIC and the susceptibility of the BG is stronger than the correlation between the MWF of the PLIC and the susceptibility of the BG. There was less of a correlation, however, between BG susceptibility and the MWF of anatomically separate WM structures.

The BG is a cluster of neurons located deep beneath the cerebral cortex with a high concentration of iron. The results of this study suggest that iron in the BG is involved in the synthesis and maintenance of myelin in IC structures. A genetic mutant mouse model study found that iron loading influences the expression of myelin-related genes, and that there is preferential iron loading in myelinated tracts of the BG (47). It is interesting to note that the nucleus accumbens, CN, and PU, which are regions with low iron levels, have a high density of transferrin receptors. These regions connect to the GP and the substantia nigra, which are regions with high iron levels. This suggests a possible existing mechanism of iron uptake and delivery through axonal transportation (48). Unfortunately, it is difficult to verify any existing mechanisms between iron in the BG and WM myelin *in vivo*, and the pathway that iron takes from the BG to regions producing myelin remains unclear. This requires further histological study.

The results of the present study demonstrate no significant relationship between the susceptibility of BG ROIs and the MWF in WM ROIs in the young group. This could be attributed to the small sample size (N=8). The young macaques also exhibited similar or less iron deposition in the BG region than their older counterparts. On the other hand, in the old group there was a significant positive relationship between the susceptibility of the CN and the MWF of the IC (both the ALIC and PLIC), but no significant correlation existed between the MWF values of the IC and the susceptibility of the GP. These results suggest that iron in the CN is more tightly linked with IC myelin than that of the GP and PU, even though iron content in the CN is the least among these three regions. A past tractography study (49) showed that, compared to the GP and PU, most of the CN is connected to the ALIC and has significant connections with the distant PLIC, which might explain the significant correlations between the MWF of IC and the susceptibility of the CN in the present study's old group.

By using a customized mGRE sequence and proper data post-processing methods, MWF and susceptibility could be measured in the brain of each macaque with a single MRI scan. In the present study, we obtained 32 echoes with 1.42 ms echo spacing. There were 16 echoes with TE $\leq T_{2,my}^*$, since the first echo time was 2.4 ms, and $T_{2,my}^*$ was in the range of 3–24 ms at 3T (22). However, for the T₂-decay based method (23,50), which obtained 32 echoes with 10–15 ms echo spacing, there were up to 4 echoes with TE < T_{2,my}, due to T_{2,my} was in the range of 15–40 ms at 3T. Therefore, the MRI signals we obtained by our method had a greater contribution from myelin water. However, possible errors due to the sensitivity of mGRE sequences to magnetic

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inhomogeneity must be taken into consideration. The MWF values obtained in the study are biased towards the BG, where there are large deposits of iron. A recent study (51) found a significant decrease in MWF values in WM areas after iron extraction, demonstrating that MWF is possibly affected by factors other than changes in myelin content.

QSM is considered to be a good biomarker for paramagnetic materials in the subcortical GM. However, the susceptibility of WM is partially dependent on the orientation of axons and myelin (52,53); therefore, only the MWF of WM and the susceptibility of the BG were used as valid data sources in the present study, which hindered investigation into the relationship between iron and myelin in local brain regions.

There were several other limitations to this study. First, the sample size was relatively small and the signal-to-noise ratio of images was affected by iron and myelin levels, meaning that subtle changes in the ratio may have been lost in areas with low myelin and iron levels. Also, because the registration tools used are optimized for humans brains and not macaques, when ROIs from the INIA19 atlas were registered to each macaque, registration errors may have occurred, resulting in potential errors in statistical analysis.

Conclusions

In this study, we demonstrated that moderate to high levels of positive correlations exist between the magnetic susceptibility of the BG and the MWF of IC structures that are anatomically connected to the BG. This suggests that future studies should consider the impact of iron concentration in the BG on myelin development in these anatomically connected WM structures. Further physiological studies are needed to understand the underlying processes of iron transportation.

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Footnote

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at http://dx.doi. org/10.21037/gims-19-1014). The authors have no conflicts of interest to declare.

Ethical Statement: All experimental procedures in this study were approved by the Institute of Neuroscience Animal Care and Use Committee and by the Shanghai Institute for Biological Sciences Biomedical Research Ethics Committee, and they all conformed to the National Institutes of Health guidelines for the humane care and use of laboratory animals.

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