Long noncoding RNAs and their roles in skeletal muscle fate determination

Mackenzie Hagan¹, Mi Zhou², Muhammad Ashraf³, Il-man Kim¹, Huabo Su¹, Neal L. Weintraub¹, Yaoliang Tang¹

¹Vascular Biology Center, Medical College of Georgia, Augusta University, Augusta, GA, USA; ²Department of cardiac surgery, Ruijin Hospital, Shanghai Jiao Tong University, Shanghai 200000, China; ³Department of Emergency Medicine, College of Medicine, The Ohio State University, Columbus, OH, USA

Contributions: (I) Conception and design: None; (II) Administrative support: None; (III) Provision of study materials or patients: None; (IV) Collection and assembly of data: None; (V) Data analysis and interpretation: None; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors

Correspondence to: Yaoliang Tang, MD, PHD, FAHA. Professor of Medicine, Vascular Biology Center, Medical College of Georgia at Augusta University, 1459 Laney Walker Blvd, Augusta, GA 30912, USA. Email: yaotang@augusta.edu.

Abstract: Myogenic fate determination is important in skeletal muscle development, growth and repair. A variety of factors regulate myogenic cell determination via transcriptional and non-transcriptional mechanisms. Amongst these factors, long noncoding RNAs (lncRNAs) have gained considerable attention for their important roles in regulating myogenic differentiation and function. Many classes of lncRNAs have been discovered; various lncRNAs have been implicated in the regulation of myogenic cell fate determination and are the subject of this brief review.

Keywords: MicroRNA; long noncoding RNA (lncRNA); competing endogenous RNA (ceRNA); myogenesis; muscle stem cells

Received: 24 November 2017; Accepted: 10 December 2017; Published: 13 December 2017. doi: 10.21037/ncri.2017.12.01 View this article at: http://dx.doi.org/10.21037/ncri.2017.12.01

Introduction

Myogenic fate determination refers to the process of stem cell commitment towards skeletal muscle progenitor cells which, in turn, promote growth and repair of skeletal muscles. This process is orchestratedly mediated by the synchronized expression and repression of key transcription factors, gene products and various proteins. Approximately 24–48 hours following muscle injury, the muscular stem cells known as satellite cells (SCs) will become activated. This activation results in either the repair of damaged tissues via the generation of muscular progenitor cells (1), or the expansion of SCs which maintains the SC reserve pool. SCs will accomplish this in one of two ways, either by asymmetrical division, which results in one SC and one muscular progenitor cell, or symmetrical division which will result in either the generation of two SCs or two committed progenitor cells. Understanding the mechanisms that control activation and division of muscle SCs is thus crucial to the field of cell-mediated skeletal muscle repair. It has been suggested that long noncoding RNAs (lncRNAs) can mediate the cell fate decision upon SC activation, resulting in either an increase or a reduction in the rate of muscle regeneration (2). LncRNAs include several types of RNA products that can regulate and coordinate gene expression through epigenetic mechanisms. Here, we will review what is known about regulation of myogenic fate determination by lncRNAs.

Biology of noncoding RNAs (ncRNAs)

The RNA family is much larger and more complex than scientists could have ever imagined upon their initial

Page 2 of 5

discovery. Amongst the least understood are the class of lncRNAs thought to function as genetic modifiers rather than protein encoding machinery. It has been estimated that only 2% of the genome actually encodes proteins, this mean that the vast majority of the transcriptome encodes lncRNAs (3). Implications for the various classes of lncRNAs are widespread and scientists have only just begun to scratch the surface of their potential functions. LncRNAs have been demonstrated to function in diverse ways, including as base-pairing partners, protein ligands, and enzymes (4). This review will focus on the lncRNAs currently under investigation, which are ill-understood compared to microRNAs (miRNAs).

LncRNAs localize in both the nucleus and the cytoplasm to regulate gene expression post-transcriptionally or through chromatin remodeling (5). LncRNAs are noncoding RNAs longer than 200 nucleotides in sequence (6) (as opposed to miRNAs which are roughly 21–25 nucleotides in length), they encompass the largest and most diverse group of ncRNAs.

Biogenesis of IncRNAs

The biogenesis of lncRNAs is not as well understood as that of the miRNAs. At its core, the process is quite similar to that of miRNAs, in that RNase II polymerase is involved in the initial transcription and splicing must occur for processing into the mature form. The main distinction between the two processes lies in the absence of the open reading frame in lncRNAs. In addition to this, lncRNAs appear to be regulated in large groups, wherein the knockdown of Dicer results in the reduced expression of hundreds of lncRNAs (7). LncRNAs are often processed like mRNAs, where polyadenylation adds a poly(A) tail to the 3' end of the transcript. A 5' cap is also added to the immature transcript, allowing for generation of the mature lncRNA structure. In some cases, however, lncRNAs lack the poly(A) tail, and instead are processed as described above in the miRNA section (7).

There are a number of subclasses of lncRNAs, including circular RNAs, small nucleolar RNAs, and lncRNAs which contain a miRNA, are examples (7). LncRNAs can be classified as overlapping, divergent (or bidirectional), intronic, intergenic, sense, or antisense. The bidirectionality of certain lncRNAs is an important distinction which sets it apart from most other RNA classes (8).

SCs in skeletal muscle regeneration: regulation by ncRNAs

SCs are quiescent under resting conditions; they become activated upon insult and proceed to generate committed progenitor cells as noted above. Expression of Pax3 and/or Pax7 is crucial for maintenance of the SC reserve pool, both also play vital roles in regulating *Myf5* and *MyoD* (9).

The Myf5 and MyoD genes are responsible for myogenic determination, and expression of Myf5 is indicative of commitment to a myogenic cell fate (10). Pax7 in SCs is also responsible for activation of MyoD, which is a skeletal muscle transcription factor, however its mRNA transcript does not undergo translation. The same is true for mRNA transcripts of $M\gamma f5$ (11). It is important to note that in mammalian cells, proliferation must stop before the cell can enter the G0 phase of the cell cycle. The G0 phase occurs directly before, or concurrently with differentiation. To summarize the process of SC fate specification: Pax3 and Pax7 function to maintain the undifferentiated SC pool, MyoD and Myf5 function as primary myogenic regulatory factors (MRF) to promote myoblast differentiation, and myogenin and MRF4 act as secondary MRFs in terminal cell differentiation (12).

LncRNAs in myogenesis (Table 1)

LncRNAs have recently emerged as potent regulators of myogenesis; many of these lncRNAs act through miRNA regulation. Circular RNAs (cirRNAs) for example, function as miRNA "sponges"; and their unique structures enable long-term regulation of miRNAs (11). One mechanism lncRNAs use to counter miRNA actions is to competitively bind to miRNA targets (3). Another regulatory method is to restrict expression of the target miRNA. The YY1associated lncRNA Yam-1 acts to reduce the expression of a number of the myomiRs during myoblast proliferation. Through activation of miR-715, Yam-1 induces myogenic repression via the Wnt7b pathway (13). There are multiple forms of Yam lncRNAs, Yam-1 and -4 appear to inhibit muscle differentiation, while Yam-2 and -3 appear to promote differentiation (13,14). Additionally, linc-YY1 also has been shown to promote differentiation (15).

Linc-MD1 promotes skeletal muscle differentiation via inhibition of miR-133, which acts to prevent terminal differentiation (16). In other studies, linc-MD1 was shown to

Table 1	Functions	of lncRNAs ir	regulating	myogenesis
	runctions	01 1110101 115 11	i i cguiaung	myogenesis

Names	Upstream	Target	Mechanism	Myogenesis	Ref.
Yam-1	YY1	MiR-715	Wnt7b signaling	Inhibit	(13)
Yam-2, 3	YY1	-	-	Promote	(13,14)
Yam-4	YY1	-	-	Inhibit	(13,14)
Linc-YY1	-	YY1	De-repression YY1/PRC2	Promote	(15)
Lnc-MD1	-	CeRNA for miR-133	MAML1 & MEF2c	Promote	(16)
H19	Imprinted	MiR-675-5p/3p	BMP/SMAD, CDC6	Promote	(17)
IncMyoD	МуоD	IGF2-mRNA-binding protein 2 (IMP2)	IMP2 mediated translation of N-Ras & c-myc	Promote	(18)
Dum	MyoD	Dnmt1, 3a, 3b	Silence neighboring Dppa2	Promote	(19)
Lnc-mg		CeRNA for miR-125b	IGF2 signaling	Promote	(20)
Malat1	MiR-181a	Suv39h1/HP1β/HDAC-1 repressive complex	H3k9me3 mediated MyoD suppression	Inhibit	(21)
Linc-RAM	-	Directly binding MyoD	Assembly of MyoD-Baf60c-Brg1 complex to enhance transcription of miR-206 and MyoG	Promote	(22)
MUNC	-	Trans, indirectly	Increased endogenous MyoD, myogenin, and Myh3 mRNAs	Promote	(23)
Sirt1-AS	-	Sirt1 3' UTR	Rescue Sirt1 transcriptional suppression by competing with miR-34a	Inhibit	(24)
Lnc-31	Nuclear precursor of miR-31	-	Cyclin D1, cyclin E and cdc25a	Inhibit	(25)

generate miR-133, or to function as a miR-133 target (26). A common target of lncRNA is the MRF MyoD. Interestingly, MyoD facilitates its own de-repression via lncRNA H19. Encoded within H19 are miRNA 675-3p and -5p, which both have roles in myogenic differentiation (17). H19 has also shown activity in regulating Let-7 (27). LncMyoD is a downstream target of MyoD, this lncRNA regulates cell-cycle exit and has an important role in myogenesis (18). MyoD induces the lncRNA Dum following myoblast differentiation, and it has been known to be a pro-myogenic factor in muscle regeneration (19).

Other lncRNAs which have been found to influence myogenesis include: lnc-mg, Malat1, linc-RAM, MUNC, Sirt-1, and linc-31. Lnc-mg is necessary for myogenic differentiation. Through interaction with miR-125b, lnc-mg promotes myoblast differentiation (20). Malat1 is important for the transition between the proliferation and differentiation stages of myogenesis (21). Linc-RAM also acts during the differentiation stage of myogenesis, enhancing transcriptional initiation of miR-206 and MyoG (22). MUNC has been known to inhibit mRNA for common myogenic markers such as myogenin and Myh3 in the early and late stages of differentiation, respectively (23). Sirt1 antisense lncRNA (AS lncRNA), through interaction with miR-34a, negatively regulates myoblast proliferation and differentiation (24). Lastly, lnc-31 acts to inhibit differentiation and maintain proliferation (25).

Conclusion remarks

Much has been learned regarding lncRNA in the past quarter century, from the discovery of lncRNA to the use of antisense for genetic knockdown. It is evident that the biology of lncRNAs is highly complex and regulates cell fate and differentiation via multiple mechanisms. It is likely however that we have only just begun to scratch the surface of how these molecules function and interact with each other. Future research is required to elucidate the

Non-coding RNA Investigation, 2017

Page 4 of 5

mechanism through which these RNAs act on the various stages of myogenesis.

Acknowledgments

Funding: This work was supported by the American Heart Association Grant-in-Aid 16GRNT31430008 to Y Tang; NIH grant AR070029 to Y Tang, M Ashraf and NL Weintraub; NIH grant 2RHL086555 to Y Tang; NIH grant HL134354 to Y Tang, M Ashraf and NL Weintraub, and NIH grant R01 HL124251 to IM Kim.

Footnote

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at http://dx.doi. org/10.21037/ncri.2017.12.01). YT serves as an unpaid editorial board member of *Non-coding RNA Investigation* from August 2017 to July 2019. The other authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Open Access Statement: This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: https://creativecommons.org/licenses/by-nc-nd/4.0/.

References

- 1. Chang NC, Rudnicki MA. Satellite cells: the architects of skeletal muscle. Curr Top Dev Biol 2014;107:161-81.
- 2. Yin H, Price F, Rudnicki MA. Satellite cells and the muscle stem cell niche. Physiol Rev 2013;93:23-67.
- Wilusz JE, Sunwoo H, Spector DL. Long noncoding RNAs: functional surprises from the RNA world. Genes Dev 2009;23:1494-504.
- 4. Wang KC, Chang HY. Molecular mechanisms of long noncoding RNAs. Mol Cell 2011;43:904-14.
- 5. Nie L, Wu HJ, Hsu JM, et al. Long non-coding RNAs:

versatile master regulators of gene expression and crucial players in cancer. Am J Transl Res 2012;4:127-50.

- Rinn JL, Chang HY. Genome regulation by long noncoding RNAs. Annu Rev Biochem 2012;81:145-66.
- Quinn JJ, Chang HY. Unique features of long non-coding RNA biogenesis and function. Nat Rev Genet 2016;17:47-62.
- 8. Spurlock CF 3rd, Crooke PS 3rd, Aune TM. Biogenesis and Transcriptional Regulation of Long Noncoding RNAs in the Human Immune System. J Immunol 2016;197:4509-17.
- Collins CA, Gnocchi VF, White RB, et al. Integrated functions of Pax3 and Pax7 in the regulation of proliferation, cell size and myogenic differentiation. PLoS One 2009;4:e4475.
- Crist CG, Montarras D, Buckingham M. Muscle satellite cells are primed for myogenesis but maintain quiescence with sequestration of Myf5 mRNA targeted by microRNA-31 in mRNP granules. Cell Stem Cell 2012;11:118-26.
- Sohi G, Dilworth FJ. Noncoding RNAs as epigenetic mediators of skeletal muscle regeneration. FEBS J 2015;282:1630-46.
- Chen Y, Melton DW, Gelfond JA, et al. MiR-351 transiently increases during muscle regeneration and promotes progenitor cell proliferation and survival upon differentiation. Physiol Genomics 2012;44:1042-51.
- Lu L, Sun K, Chen X, et al. Genome-wide survey by ChIP-seq reveals YY1 regulation of lincRNAs in skeletal myogenesis. EMBO J 2013;32:2575-88.
- 14. Shiekhattar R. The Yin and Yang of enhancer-like RNAs. EMBO J 2013;32:2533-4.
- Zhou L, Sun K, Zhao Y, et al. Linc-YY1 promotes myogenic differentiation and muscle regeneration through an interaction with the transcription factor YY1. Nat Commun 2015;6:10026.
- Cesana M, Cacchiarelli D, Legnini I, et al. A long noncoding RNA controls muscle differentiation by functioning as a competing endogenous RNA. Cell 2011;147:358-69.
- Dey BK, Pfeifer K, Dutta A. The H19 long noncoding RNA gives rise to microRNAs miR-675-3p and miR-675-5p to promote skeletal muscle differentiation and regeneration. Genes Dev 2014;28:491-501.
- Gong C, Li Z, Ramanujan K, et al. A long non-coding RNA, LncMyoD, regulates skeletal muscle differentiation by blocking IMP2-mediated mRNA translation. Dev Cell 2015;34:181-91.
- 19. Wang L, Zhao Y, Bao X, et al. LncRNA Dum interacts with Dnmts to regulate Dppa2 expression during

Non-coding RNA Investigation, 2017

myogenic differentiation and muscle regeneration. Cell Res 2015;25:335-50.

- Zhu M, Liu J, Xiao J, et al. Lnc-mg is a long noncoding RNA that promotes myogenesis. Nat Commun 2017;8:14718.
- Chen X, He L, Zhao Y, et al. Malat1 regulates myogenic differentiation and muscle regeneration through modulating MyoD transcriptional activity. Cell Discov 2017;3:17002.
- 22. Yu X, Zhang Y, Li T, et al. Long non-coding RNA Linc-RAM enhances myogenic differentiation by interacting with MyoD. Nat Commun 2017;8:14016.
- Mueller AC, Cichewicz MA, Dey BK, et al. MUNC, a long noncoding RNA that facilitates the function of MyoD in skeletal myogenesis. Mol Cell Biol 2015;35:498-513.

doi: 10.21037/ncri.2017.12.01

Cite this article as: Hagan M, Zhou M, Ashraf M, Kim IM, Su H, Weintraub NL, Tang Y. Long noncoding RNAs and their roles in skeletal muscle fate determination. Non-coding RNA Investig 2017;1:24.

- 24. Wang GQ, Wang Y, Xiong Y, et al. Sirt1 AS lncRNA interacts with its mRNA to inhibit muscle formation by attenuating function of miR-34a. Sci Rep 2016;6:21865.
- Ballarino M, Cazzella V, D'Andrea D, et al. Novel long noncoding RNAs (lncRNAs) in myogenesis: a miR-31 overlapping lncRNA transcript controls myoblast differentiation. Mol Cell Biol 2015;35:728-36.
- 26. Legnini I, Morlando M, Mangiavacchi A, et al. A feedforward regulatory loop between HuR and the long noncoding RNA linc-MD1 controls early phases of myogenesis. Mol Cell 2014;53:506-14.
- Kallen AN, Zhou XB, Xu J, et al. The imprinted H19 lncRNA antagonizes let-7 microRNAs. Mol Cell 2013;52:101-12.