

Editorial

Calcium signaling in physiology and pathophysiology

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Abstract

Calcium ions are the most ubiquitous and pluripotent cellular signaling molecules that control a wide variety of cellular processes. The calcium signaling system is represented by a relatively limited number of highly conserved transporters and channels, which execute Ca^{2+} movements across biological membranes and by many thousands of Ca^{2+} -sensitive effectors. Molecular cascades, responsible for the generation of calcium signals, are tightly controlled by Ca^{2+} ions themselves and by genetic factors, which tune the expression of different Ca^{2+} -handling molecules according to adaptational requirements. Ca^{2+} ions determine normal physiological reactions and the development of many pathological processes.

Ja, Kalzium das ist alles...

Otto Loewi

(1936 Nobel Laureate)

Experimental indications, demonstrating the role of calcium as a universal signalling molecule, controlling a huge variety of very different physiological functions appeared at the end of 19th century. First, Sydney Ringer showed that calcium ions were indispensable for fish survival, muscle contraction, the development of fertilized eggs and tadpole and for cells adhesion^[1–5]. Several years later, Locke^[6] and Overton^[7] demonstrated the critical importance of Ca^{2+} for signal transduction between nerve and muscle. The general theory of calcium as a universal second messenger, however, appeared half a century later, when Lewis Victor Heilbrunn concluded that “the reaction of this calcium with the protoplasm inside the cell is the most basic of all protoplasmic reactions”^[8]. This theory, although almost completely ignored at the time of its appearance, brilliantly withstood the test of time and experimental efforts (Figure 1), and today, the calcium signalling is generally regarded as the most ubiquitous and the most pluripotent system, involved in regulation of almost all known cellular processes^[9].

The universality of calcium as a signaling molecule is manifested on many levels. First, Ca^{2+} ions act as intracellular messengers throughout phylogenetic history, from early

prokaryotes to eukaryotic cells.

Second, within every cell, Ca^{2+} exerts its action through several very different levels, which are executed in different spatial and temporal domains. Indeed, Ca^{2+} ions control localized processes, (eg, exocytosis) and global responses (eg, myocyte contraction) with equivalent efficacy and ease (Figure 2). Similarly, Ca^{2+} -dependent cellular responses occur in an amazingly wide time scale, from microseconds (eg, activation of ion channels) to many hours, weeks, months or even years (eg, synaptic plasticity, memory, long-term adaptation or neuronal ageing).

Third, the Ca^{2+} signaling system is constructed with an incredible intrinsic versatility. The actual molecular cascades controlling Ca^{2+} movements through cellular membranes (Figure 3) are limited to several families of relatively similar pumps (plasmalemmal and intracellular PMCA, SERCA or SPCA^[10–12]), sodium-calcium exchangers (NCX, residing in plasmalemma or in mitochondria^[13,14]) and plasmalemmal^[15–18] and intracellular^[13,19–21] Ca^{2+} channels. Yet these cascades are very tightly regulated, which determines their great adaptability and versatility. Importantly, calcium signalling molecules are subject to a control by Ca^{2+} ions themselves: changes in Ca^{2+} gradients or local concentration control the availability of Ca^{2+} channels and regulate the activity of Ca^{2+} pumps^[22–24]. On a different level, the expression of various molecules, controlling Ca^{2+} movements is responsive to the

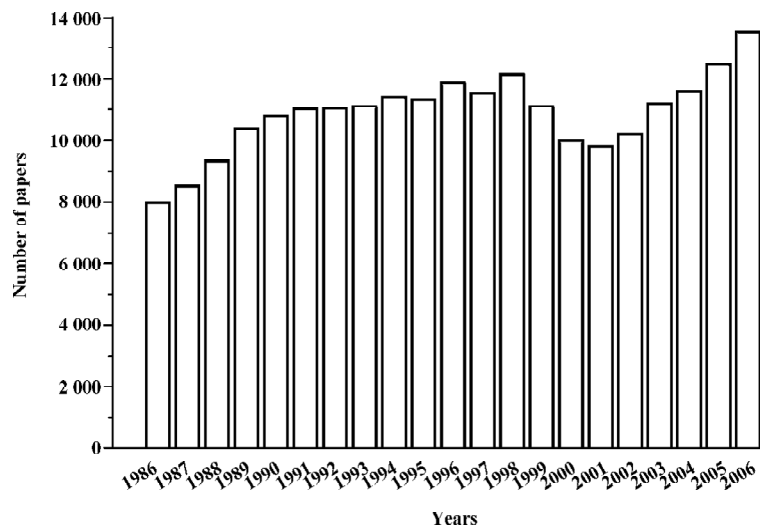


Figure 1. Publication of papers, dedicated to calcium signalling, according to the PubMed.

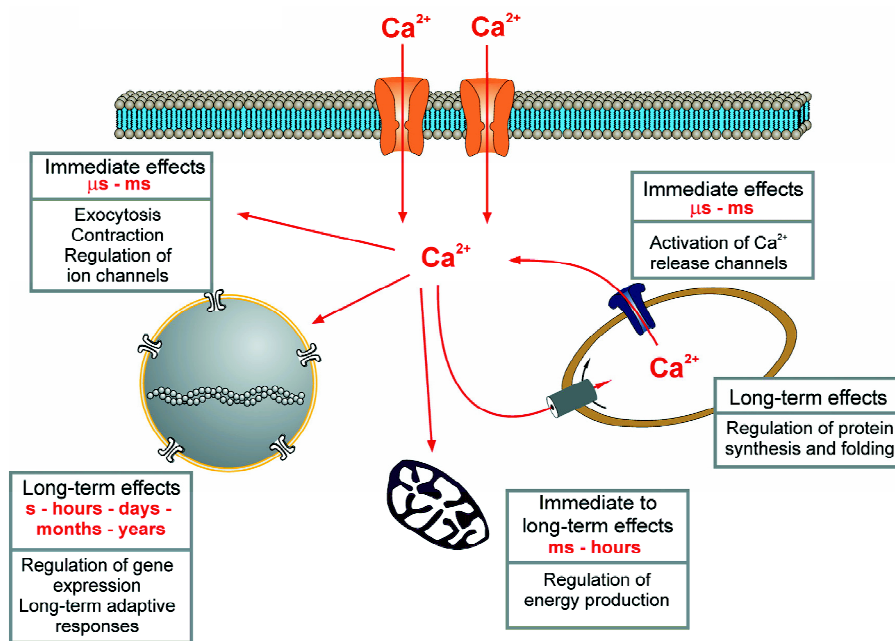


Figure 2. Temporal domains of calcium signalling.

changes in the environment, and therefore the combinations of calcium signaling molecules (or “Ca²⁺ signalling toolkits”^[25]) can be rapidly modified, thus adapting the system to the external demands.

Fourth, the effector part of the calcium signalling system, the Ca²⁺ sensors, is represented by thousands of proteins, which have different affinity to Ca²⁺ ions, with the dissociate constant spanning seven orders of magnitude (Figure 4), and different cellular location. This host of Ca²⁺ sensors

determines the ubiquity and promiscuity of Ca²⁺ signaling: expression of specific Ca²⁺ sensors commands specific Ca²⁺-regulatory function (eg, expression of Ca²⁺-sensitive contractile in muscle cells determines the excitation contraction coupling), whereas different affinity/localization of Ca²⁺ sensors will allow precise regulation of very different processes within a single cell.

The specificity and precise localization of calcium signalling machinery is also supported by an existence of

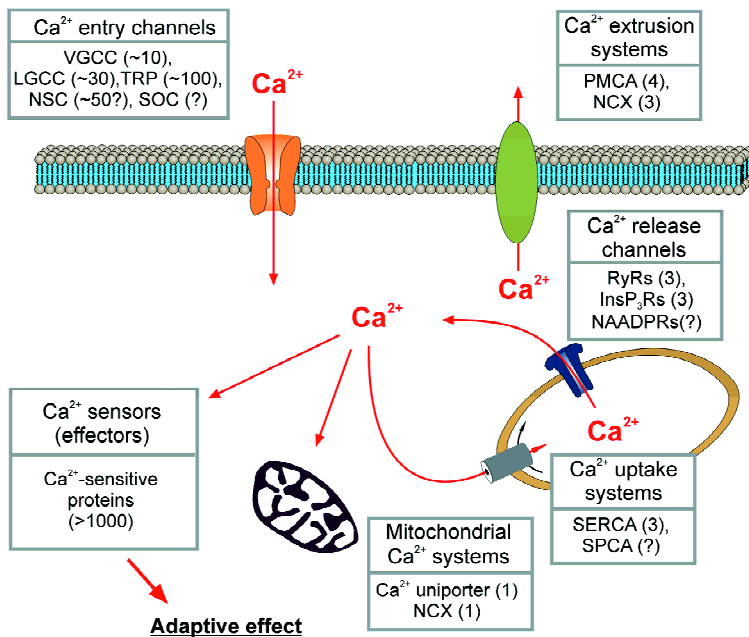


Figure 3. Simplicity and complexity of calcium signalling. Calcium signals are controlled by a relatively limited quantity of molecules (approximate number of these molecules is indicated in the parentheses), which include Ca²⁺ channels [voltage-gated Ca²⁺ channels (VGCC), ligand-gated Ca²⁺ channels (LGCC); transient receptor potential Ca²⁺ permeable channels (TRP); non-selective channels; SOC-store-operated Ca²⁺ channels (NSC)]; plasmalemmal Ca²⁺ extrusion systems [plasmalemmal Ca²⁺ ATPase (PMCA); Na⁺/Ca²⁺ exchanger (NCX)]; intracellular Ca²⁺ release channels [ryanodine receptors (RyRs); InsP₃ receptors (InsP₃Rs); NAADP receptors (NAADPRs)]; intracellular Ca²⁺ pumps [sarco(endo)plasmic reticulum Ca²⁺ ATPase (SERCA); Ca²⁺ ATPases of Golgi complex (SPCA)] and mitochondrial Ca²⁺ transporting systems (Ca²⁺ uniporter; and mitochondrial Na⁺/Ca²⁺ exchanger). The calcium signalling system exerts physiological effects through Ca²⁺ sensors (effectors), which are represented by approximate thousands of enzymes and Ca²⁺-binding proteins.

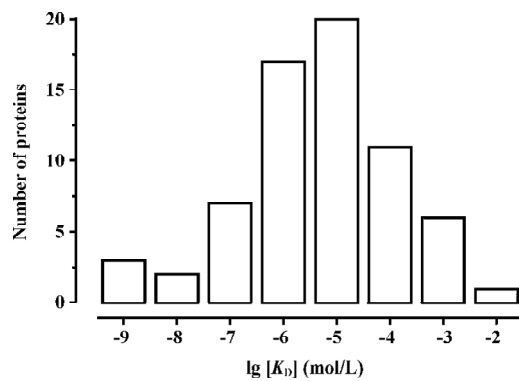


Figure 4. Diversity of calcium binding affinity of Ca²⁺-sensors. The dissociation constants (K_d) of Ca²⁺ binding proteins (68 K_d values obtained by text search and manual curation from the literature) vary over 7 logarithmic units, ranging from nmol/L to 10 mmol/L with a broad mode around 10 μmol/L.

several intracellular compartments, characterized by a clearly distinct Ca²⁺ homeostasis. These compartments are represented by the cytosol, by endoplasmic reticulum (ER) and mitochondria. In the cytosol the concentration of free Ca²⁺ ([Ca²⁺]_i) is very low, approximately 50–100 nmol/L, which is achieved by continuous activity of Ca²⁺ extruding systems and by high-affinity cytosolic calcium buffers^[14,26,27]. As a consequence, activation of Ca²⁺ entry channels results in rapid elevation of [Ca²⁺]_i, yet the strong Ca²⁺ buffering favours localisation of Ca²⁺ signals and the creation of Ca²⁺ microdomains. This is very important for regulation of focal

cellular responses, such as exocytosis^[28,29].

The ER, in contrast, provides for a very different Ca²⁺ handling environment. The intra-ER, or intraluminal free Ca²⁺ concentration ([Ca²⁺]_L), is set at a rather high level, 100–800 μmol/L^[30–36], which is achieved by a continuous activity of SERCA pumps. In addition, the affinity of intra-ER Ca²⁺ buffers is rather low, being in the range of 0.5–1.0 mmol/L, which favours Ca²⁺ diffusion through the continuous ER lumen. The latter therefore forms a nanoscopic “Ca²⁺ tunnel”, which allows long-range Ca²⁺ transport in polarised cells^[37–40]. Importantly, numerous intra-ER Ca²⁺-dependent enzymatic systems require high (>50 μmol/L) [Ca²⁺]_L for normal functioning^[41,42]. The ER acts as a very powerful intracellular signalling organelle, which integrates various incoming signals with cellular biochemistry (through regulation of protein synthesis and posttranslational folding). Furthermore, the ER produces numerous output signals, which regulate cell function and determine adaptive responses. Particularly important is the role of ER in the generation of cytoplasmic Ca²⁺ signals because the ER acts as a dynamic Ca²⁺ store able to rapidly release Ca²⁺ through intracellular Ca²⁺ channels^[19,21] and to terminate Ca²⁺ signals through SERCA-dependent Ca²⁺ pumping. As a consequence, the ER appears simultaneously as a source and sink for [Ca²⁺]_i^[43–45], while the balance between Ca²⁺ release and Ca²⁺ uptake is regulated by [Ca²⁺]_L and [Ca²⁺]_i dynamics in a vicinity of Ca²⁺ release channels^[46,47].

The third intracellular compartment with specific Ca²⁺ homeostasis is represented by mitochondria, which are able

to accumulate (via Ca^{2+} uniporter) and release (via $\text{Na}^+/\text{Ca}^{2+}$ exchanger) Ca^{2+} [13]. Mitochondrial Ca^{2+} signalling links cellular activity to ATP production and ROS metabolism; in addition mitochondria can participate in $[\text{Ca}^{2+}]_i$ regulation, especially in pathological conditions [48–50].

Finally, the signalling system mediated by Ca^{2+} ions operates in two modes: the digital and analogue. The digital mode is determined by a discrete character of Ca^{2+} entry through the membrane, which is controlled by opening and closing of Ca^{2+} permeable channels. Yet, when inside the intracellular compartments, Ca^{2+} ions diffuse, and they diffuse with a different velocity and anisotropy, thus creating a complex concentration gradients, which represents an analogue signalling, coded in amplitude, space and time.

All these features make the Ca^{2+} signaling system absolutely unique among other cellular signaling pathways. Ca^{2+} ions are fundamentally different from other signalling molecules in a sense that they are subjected to neither catabolism nor anabolism; they can be merely bound to calcium buffers or accumulated into Ca^{2+} stores, yet they remain readily available for mobilisation. This makes the signalling system quite economical. Huge Ca^{2+} gradients, existing between extracellular space, intracellular organelles and the cytoplasm contribute to an exceedingly high signal-to-noise ratio of the whole signalling system. Further, the promiscuity of Ca^{2+} ions as intracellular messengers provides for a remarkable versatility; the variety of Ca^{2+} sensor proteins together with temporal and spatial heterogeneity of Ca^{2+} fluctuations, make the signalling system both context and history-specific. As a consequence, Ca^{2+} ions often play very opposite effects even within the same cell. One of the best examples of such a dualism exists in arterial smooth muscle cells, where subsurface calcium sparks relax the myocyte by activating Ca^{2+} -dependent K^+ channels [51–53], whereas global calcium signals trigger cell contraction.

Not surprisingly, the omnipotence of Ca^{2+} signaling makes it an important player not only in normal conditions but also in pathological cellular reactions. Here the dualism of Ca^{2+} ions transpires even more illustriously, as indeed Ca^{2+} ions are the ions of life and death. Depriving the cells from Ca^{2+} ions by the removal of extracellular Ca^{2+} , or artificial chelating of intracellular Ca^{2+} , or depletion of cellular free Ca^{2+} , all of these interventions result in rapid and inevitable cell death [42,54]. At the same time excess of Ca^{2+} is absolutely toxic, and cell death from Ca^{2+} overload represents probably the most general mechanism of cell demise [55,56]. Similarly, chronic disruptions of Ca^{2+} homeostatic machinery may cause development of various diseases, such as ischemic-induced cell death [57–63], neurodegeneration [42,54,64],

heart failure [65,66] or underlying cognitive deficits in senescence [67–69].

When compiling this special issue we tried to cover all of the important parts of calcium signaling machinery and its role in physiology and disease. We hope that this collection of articles will spark further interest in various aspects of Ca^{2+} and inspire further developments into the functions and importance of this truly magnificent ion of life.

References

- 1 Ringer S. A further contribution regarding the influence of different constituents of the blood on the contractions of the heart. *J Physiol (Lond)* 1883; 4: 29–43.
- 2 Ringer S. The influence of saline media on fishes. *J Physiol (Lond)* 1883; 4: vi–viii.
- 3 Ringer S. Further experiments regarding the influence of small quantities of lime, potassium and other salts on muscular tissue. *J Physiol (Lond)* 1886; 7: 291–308.
- 4 Ringer S. Concerning experiments to test the influence of lime, sodium and potassium salts on the development of ova and growth of tadpoles. *J Physiol (Lond)* 1890; 11: 79–84.
- 5 Ringer S Sainsbury H. The action of potassium, sodium and calcium salts on *Tubifex rivulorum*. *J Physiol (Lond)* 1894; 16: 1–9.
- 6 Locke FS. Notiz uber den Einfluss, physiologischer Kochsalzlosung auf die Eregerbarkeit von Muskel und Nerve. *Zentralb Physiol* 1894; 8: 166–7.
- 7 Overton E. Beitrage zur allgemeinen Muskel- und Nervenphysiologie. III. Mittheilung. Studien uber die Wirkung der Alkali- und Erdkali-salze auf Skeletalmuskeln und Nerven. *Pflugers Arch* 1904; 105: 176–290.
- 8 Heilbrunn LV. An outline of general physiology. Philadelphia: Saunders; 1943.
- 9 Petersen OH, Michalak M, Verkhratsky A. Calcium signalling: past, present and future. *Cell Calcium* 2005; 38: 161–9.
- 10 Wuytack F, Raeymaekers L, Missiaen L. PMR1/SPCA Ca^{2+} pumps and the role of the Golgi apparatus as a Ca^{2+} store. *Pflugers Arch* 2003; 446: 148–53.
- 11 Vanoevelen J, Dode L, Van Baelen K, Fairclough RJ, Missiaen L, Raeymaekers L, *et al*. The secretory pathway $\text{Ca}^{2+}/\text{Mn}^{2+}$ -ATPase 2 is a Golgi-localized pump with high affinity for Ca^{2+} ions. *J Biol Chem* 2005; 280: 22800–8.
- 12 Vangheluwe P, Raeymaekers L, Dode L, Wuytack F. Modulating sarco(endo)plasmic reticulum Ca^{2+} ATPase 2 (SERCA2) activity: cell biological implications. *Cell Calcium* 2005; 38: 291–302.
- 13 Nicholls DG. Mitochondria and calcium signaling. *Cell Calcium* 2005; 38: 311–7.
- 14 Guerini D, Coletto L, Carafoli E. Exporting calcium from cells. *Cell Calcium* 2005; 38: 281–9.
- 15 Triggler DJ. L-type calcium channels. *Curr Pharm Des* 2006; 12: 443–57.
- 16 Parekh AB, Putney JW Jr. Store-operated calcium channels. *Physiol Rev* 2005; 85: 757–810.
- 17 Perez-Reyes E. Molecular physiology of low-voltage-activated t-type calcium channels. *Physiol Rev* 2003; 83: 117–61.
- 18 Pedersen SF, Owsianik G, Nilius B. TRP channels: an overview.

- Cell Calcium 2005; 38: 233–52.
- 19 Bezprozvanny I. The inositol 1,4,5-trisphosphate receptors. *Cell Calcium* 2005; 38: 261–72.
 - 20 Galione A, Ruas M. NAADP receptors. *Cell Calcium* 2005; 38: 273–80.
 - 21 Hamilton SL. Ryanodine receptors. *Cell Calcium* 2005; 38: 253–60.
 - 22 Morad M, Soldatov N. Calcium channel inactivation: possible role in signal transduction and Ca²⁺ signaling. *Cell Calcium* 2005; 38: 223–31.
 - 23 Burdakov D, Verkhratsky A. Biophysical re-equilibration of Ca²⁺ fluxes as a simple biologically plausible explanation for complex intracellular Ca²⁺ release patterns. *FEBS Lett* 2006; 380: 463–8.
 - 24 Burdakov D, Petersen OH, Verkhratsky A. Intraluminal calcium as a primary regulator of endoplasmic reticulum function. *Cell Calcium* 2005; 38: 303–10.
 - 25 Berridge MJ, Bootman MD, Roderick HL. Calcium signalling: dynamics, homeostasis and remodelling. *Nat Rev Mol Cell Biol* 2003; 4: 517–29.
 - 26 Petersen OH, Petersen CC, Kasai H. Calcium and hormone action. *Annu Rev Physiol* 1994; 56: 297–319.
 - 27 Burnashev N, Rozov A. Presynaptic Ca²⁺ dynamics, Ca²⁺ buffers and synaptic efficacy. *Cell Calcium* 2005; 37: 489–95.
 - 28 Barclay JW, Morgan A, Burgoyne RD. Calcium-dependent regulation of exocytosis. *Cell Calcium* 2005; 38: 343–53.
 - 29 Jarvis SE, Zamponi GW. Masters or slaves? Vesicle release machinery and the regulation of presynaptic calcium channels. *Cell Calcium* 2005; 37: 483–8.
 - 30 Alonso MT, Barrero MJ, Michelena P, Carnicero E, Cuchillo I, Garcia AG, *et al*. Ca²⁺-induced Ca²⁺ release in chromaffin cells seen from inside the ER with targeted aequorin. *J Cell Biol* 1999; 144: 241–54.
 - 31 Alvarez J, Montero M. Measuring [Ca²⁺] in the endoplasmic reticulum with aequorin. *Cell Calcium* 2002; 32: 251–60.
 - 32 Mogami H, Tepikin AV, Petersen OH. Termination of cytosolic Ca²⁺ signals: Ca²⁺ reuptake into intracellular stores is regulated by the free Ca²⁺ concentration in the store lumen. *EMBO J* 1998; 17: 435–42.
 - 33 Solovyova N, Verkhratsky A. Neuronal endoplasmic reticulum acts as a single functional Ca²⁺ store shared by ryanodine and inositol-1,4,5-trisphosphate receptors as revealed by intra-ER [Ca²⁺] recordings in single rat sensory neurones. *Pflugers Arch* 2003; 446: 447–54.
 - 34 Solovyova N, Veselovsky N, Toescu EC, Verkhratsky A. Ca²⁺ dynamics in the lumen of the endoplasmic reticulum in sensory neurons: direct visualization of Ca²⁺-induced Ca²⁺ release triggered by physiological Ca²⁺ entry. *EMBO J* 2002; 21: 622–30.
 - 35 Tse FW, Tse A, Hille B. Cyclic Ca²⁺ changes in intracellular stores of gonadotropes during gonadotropin-releasing hormone-stimulated Ca²⁺ oscillations. *Proc Natl Acad Sci USA* 1994; 91: 9750–4.
 - 36 Verkhratsky A. Physiology and pathophysiology of the calcium store in the endoplasmic reticulum of neurons. *Physiol Rev* 2005; 85: 201–79.
 - 37 Mogami H, Nakano K, Tepikin AV, Petersen OH. Ca²⁺ flow via tunnels in polarized cells: recharging of apical Ca²⁺ stores by focal Ca²⁺ entry through basal membrane patch. *Cell* 1997; 88: 49–55.
 - 38 Mogami H, Gardner J, Gerasimenko OV, Camello P, Petersen OH, Tepikin AV. Calcium binding capacity of the cytosol and endoplasmic reticulum of mouse pancreatic acinar cells. *J Physiol* 1999; 518: 463–7.
 - 39 Petersen OH, Tepikin A, Park MK. The endoplasmic reticulum: one continuous or several separate Ca²⁺ stores? *Trends Neurosci* 2001; 24: 271–6.
 - 40 Verkhratsky A. The endoplasmic reticulum and neuronal calcium signalling. *Cell Calcium* 2002; 32: 393–404.
 - 41 Michalak M, Robert Parker JM, Opas M. Ca²⁺ signaling and calcium binding chaperones of the endoplasmic reticulum. *Cell Calcium* 2002; 32: 269–78.
 - 42 Verkhratsky A, Toescu EC. Endoplasmic reticulum Ca²⁺ homeostasis and neuronal death. *J Cell Mol Med* 2003; 7: 351–61.
 - 43 Friel DD, Tsien RW. A caffeine- and ryanodine-sensitive Ca²⁺ store in bullfrog sympathetic neurones modulates effects of Ca²⁺ entry on [Ca²⁺]_i. *J Physiol* 1992; 450: 217–46.
 - 44 Shmigol A, Kostyuk P, Verkhratsky A. Role of caffeine-sensitive Ca²⁺ stores in Ca²⁺ signal termination in adult mouse DRG neurones. *Neuroreport* 1994; 5: 2073–6.
 - 45 Usachev Y, Shmigol A, Pronchuk N, Kostyuk P, Verkhratsky A. Caffeine-induced calcium release from internal stores in cultured rat sensory neurons. *Neuroscience* 1993; 57: 845–59.
 - 46 Hongpaisan J, Pivovarova NB, Colegrove SL, Leapman RD, Friel DD, Andrews SB. Multiple modes of calcium-induced calcium release in sympathetic neurons II: a [Ca²⁺]_i- and location-dependent transition from endoplasmic reticulum Ca accumulation to net Ca release. *J Gen Physiol* 2001; 118: 101–12.
 - 47 Albrecht MA, Colegrove SL, Hongpaisan J, Pivovarova NB, Andrews SB, Friel DD. Multiple modes of calcium-induced calcium release in sympathetic neurons I: attenuation of endoplasmic reticulum Ca²⁺ accumulation at low [Ca²⁺]_i during weak depolarization. *J Gen Physiol* 2001; 118: 83–100.
 - 48 Toescu EC, Verkhratsky A. Neuronal ageing from an intraneuronal perspective: roles of endoplasmic reticulum and mitochondria. *Cell Calcium* 2003; 34: 311–23.
 - 49 Toescu EC. Hypoxia sensing and pathways of cytosolic Ca²⁺ increases. *Cell Calcium* 2004; 36: 187–99.
 - 50 Toescu EC. Hypoxia response elements. *Cell Calcium* 2004; 36: 181–5.
 - 51 Nelson MT, Cheng H, Rubart M, Santana LF, Bonev AD, Knot HJ, *et al*. Relaxation of arterial smooth muscle by calcium sparks. *Science* 1995; 270: 633–7.
 - 52 Wellman GC, Nathan DJ, Saundry CM, Perez G, Bonev AD, Penar PL, *et al*. Ca²⁺ sparks and their function in human cerebral arteries. *Stroke* 2002; 33: 802–8.
 - 53 Wellman GC, Nelson MT. Signaling between SR and plasmalemma in smooth muscle: sparks and the activation of Ca²⁺-sensitive ion channels. *Cell Calcium* 2003; 34: 211–29.
 - 54 Paschen W, Mengesdorf T. Endoplasmic reticulum stress response and neurodegeneration. *Cell Calcium* 2005; 38: 409–15.
 - 55 Berliocchi L, Bano D, Nicotera P. Ca²⁺ signals and death programmes in neurons. *Philos Trans R Soc Lond B Biol Sci* 2005; 360: 2255–8.
 - 56 Leist M, Nicotera P. Apoptosis versus necrosis: the shape of neuronal cell death. *Results Probl Cell Differ* 1998; 24: 105–35.
 - 57 Kristian T. Metabolic stages, mitochondria and calcium in hypoxic/ischemic brain damage. *Cell Calcium* 2004; 36: 221–33.

- 58 Pringle AK. In, out, shake it all about: elevation of $[Ca^{2+}]_i$ during acute cerebral ischaemia. *Cell Calcium* 2004; 36: 235–45.
- 59 Yao H, Haddad GG. Calcium and pH homeostasis in neurons during hypoxia and ischemia. *Cell Calcium* 2004; 36: 247–55.
- 60 Starkov AA, Chinopoulos C, Fiskum G. Mitochondrial calcium and oxidative stress as mediators of ischemic brain injury. *Cell Calcium* 2004; 36: 257–64.
- 61 Pisani A, Bonsi P, Calabresi P. Calcium signaling and neuronal vulnerability to ischemia in the striatum. *Cell Calcium* 2004; 36: 277–84.
- 62 Yamashita T. Ca^{2+} -dependent proteases in ischemic neuronal death: a conserved 'calpain-cathepsin cascade' from nematodes to primates. *Cell Calcium* 2004; 36: 285–93.
- 63 Kahlert S, Reiser G. Glial perspectives of metabolic states during cerebral hypoxia-calcium regulation and metabolic energy. *Cell Calcium* 2004; 36: 295–302.
- 64 Mattson MP, Chan SL. Neuronal and glial calcium signaling in Alzheimer's disease. *Cell Calcium* 2003; 34: 385–97.
- 65 Scoote M, Williams AJ. Myocardial calcium signalling and arrhythmia pathogenesis. *Biochem Biophys Res Commun* 2004; 322: 1286–309.
- 66 Sipido KR, Eisner D. Something old, something new: changing views on the cellular mechanisms of heart failure. *Cardiovasc Res* 2005; 68: 167–74.
- 67 Toescu EC, Verkhratsky A. Ca^{2+} and mitochondria as substrates for deficits in synaptic plasticity in normal brain ageing. *J Cell Mol Med* 2004; 8: 181–90.
- 68 Verkhratsky A, Toescu EC. Calcium and neuronal ageing. *Trends Neurosci* 1998; 21: 2–7.
- 69 Toescu EC, Verkhratsky A, Landfield PW. Ca^{2+} regulation and gene expression in normal brain aging. *Trends Neurosci* 2004; 27: 614–20.

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