

Open  
Review

# Yin and Yang of ginseng pharmacology: ginsenosides vs gintonin

Dong-soon IM<sup>1,\*</sup>, Seung-yeol NAH<sup>2,\*</sup>

<sup>1</sup>Molecular Inflammation Research Center for Aging Intervention (MRCA) and College of Pharmacy, Pusan National University, Busan 609–735, Korea; <sup>2</sup>Ginsentology Research Laboratory and Department of Physiology, College of Veterinary Medicine and Bio/Molecular Informatics Center, Konkuk University, Seoul 143–701, Korea

Ginseng, the root of *Panax ginseng*, has been used in traditional Chinese medicine as a tonic herb that provides many beneficial effects. Pharmacologic studies in the last decades have shown that ginsenosides (ginseng saponins) are primarily responsible for the actions of ginseng. However, the effects of ginseng are not fully explained by ginsenosides. Recently, another class of active ingredients called gintonin was identified. Gintonin is a complex of glycosylated ginseng proteins containing lysophosphatidic acids (LPAs) that are the intracellular lipid mitogenic mediator. Gintonin specifically and potently activates the G protein-coupled receptors (GPCRs) for LPA. Thus, the actions of ginseng are now also linked to LPA and its GPCRs. This linkage opens new dimensions for ginseng pharmacology and LPA therapeutics. In the present review, we evaluate the pharmacology of ginseng with the traditional viewpoint of Yin and Yang components. Furthermore, we will compare ginsenoside and gintonin based on the modern view of molecular pharmacology in terms of ion channels and GPCRs.

**Keywords:** ginseng; ginsenoside; gintonin; lysophosphatidic acid; ion channels; G protein-coupled receptors; Ca<sup>2+</sup>; traditional Chinese medicine

Acta Pharmacologica Sinica (2013) 34: 1367–1373; doi: 10.1038/aps.2013.100; published online 14 Oct 2013

## Introduction

Since ginsenosides (ginseng saponins) were first identified as active ingredients in ginseng, many studies have shown that ginsenosides negatively regulate ion channels<sup>[1]</sup>. For example, ginsenoside Rg<sub>3</sub> inhibits not only voltage-gated Ca<sup>2+</sup> and Na<sup>+</sup> channels but also ligand-gated ion channels such as 5-HT<sub>3</sub> receptors<sup>[2–4]</sup>. Furthermore, ginsenosides stimulate anion-gated GABA<sub>A</sub> and glycine receptors<sup>[5, 6]</sup>. Therefore, ginsenosides decrease the excitability of excitable cells by inhibition of cation influx and stimulation of anion influx across the plasma membrane.

Crude ginseng saponin fractions elevate the intracellular Ca<sup>2+</sup> levels in mammalian cells and activate Ca<sup>2+</sup>-activated Cl<sup>-</sup> channels in *Xenopus* oocytes<sup>[7]</sup>. However, this effect was not explained by ginsenosides. Recently, the active ingredients responsible for the Ca<sup>2+</sup> rise were elucidated as novel glycolipoproteins and named gintonins<sup>[8]</sup>. Gintonin is a complex of glycosylated ginseng proteins containing lysophosphatidic

acids (LPAs). LPA is a representative mitogenic lipid mediator<sup>[9]</sup>. Gintonin activates G protein-coupled receptors (GPCRs) for LPA<sup>[8]</sup>. Unexpectedly, the action of ginseng is now linked to LPA and its GPCRs and *vice versa*. This linkage opens new dimensions for ginseng pharmacology and LPA therapeutics.

Ginsenosides tend to attenuate cell excitations by blocking cation Ca<sup>2+</sup> and Na<sup>+</sup> influxes or by enhancing anion Cl<sup>-</sup> influx. However, gintonin induces transient Ca<sup>2+</sup> elevation and activation of MAPK, PI3K, PKC, and Rho kinase via LPA receptors to evoke stimulatory cellular responses<sup>[4, 8, 10, 11]</sup>. Thus, ginsenoside acts as a negative regulator (Yin), and gintonin acts as a positive regulator (Yang) for ginseng pharmacology. The pharmacological actions of ginseng are now explainable and complementary with the opposing actions of gintonin and ginsenoside. Therapeutic application of LPA might be expanded by traditional usages of ginseng and *vice versa*. In this article, both traditional and modern ginseng pharmacology will be discussed from the viewpoint of Yin and Yang.

## Ginseng and ginsenosides

Ginseng, the root of *Panax ginseng* CA Meyer, has been used for thousands of years in Asian countries such as Korea, China, and Japan. *Panax* means 'all heal' in Greek, and the

\* To whom correspondence should be addressed.

E-mail imds@pusan.ac.kr (Dong-soon IM);

synah@konkuk.ac.kr (Seung-yeol NAH)

Received 2013-05-15 Accepted 2013-07-05

Chinese characters of ginseng originated from the human-like shape of the ginseng root<sup>[12]</sup>. The traditional beneficial effects of ginseng are replenishment of vital energy, mood elevation, and longevity. Therefore, in ancient times, ginseng was considered as a panacea that provided eternal youth<sup>[1, 12]</sup>. In addition, modern pharmacological studies have revealed ginseng's adaptogenic activities against stress, fatigue, cardiovascular dysfunction, and various diseases, including cancer and neurodegenerative disorders. Its active components also have been intensively studied over the past decades.

The representative active ingredients of ginseng are ginsenosides (ginseng saponin), which are derivatives of the triterpenoid dammarane<sup>[1]</sup>. More than 100 different types of ginsenosides have been isolated and identified from the roots of Korean and American ginseng<sup>[13]</sup>. Each ginsenoside is composed of three parts, including a hydrophobic four-ring backbone structure, an attached carbohydrate portion, and an aliphatic side chain (Figure 1).

### Ginsenosides and the modulation of ion channels

Because ginsenosides were first identified as the active ingredients in ginseng, many studies have demonstrated negative regulation of ion channels by ginsenosides<sup>[1]</sup>. Their actions are stereoselective, but they lack the specificity and selectivity of other channel inhibitors. The EC<sub>50</sub> values of ginsenosides are in the μmol/L range, and many ion channels are affected. For example, the EC<sub>50</sub> values for the ginsenoside Rg<sub>3</sub> are approximately 0.41–97.3 μmol/L<sup>[4, 14–20]</sup>. The ginsenoside Rg<sub>3</sub> not only inhibits voltage-gated Ca<sup>2+</sup>, K<sup>+</sup>, and Na<sup>+</sup> channels, ligand-gated 5-HT<sub>3</sub>, α7 nicotinic acetylcholine, and NMDA receptors but also activates K<sup>+</sup> channels such as KCNQ K<sup>+</sup>, BK<sub>Ca</sub>, and hERG K<sup>+</sup><sup>[4, 14–21]</sup>. Other ginsenosides enhance the activation of anion GABA<sub>A</sub> and glycine receptors<sup>[22, 23]</sup> (Table 1). Therefore, ginsenosides decrease the cellular excitability of excitable cells by inhibiting cation influx and by stimulating anion influx across plasma membranes.

Site-directed mutagenesis experiments have identified ginsenoside interaction site(s), and homology docking modeling has provided three-dimensional configurations for ginsenosides and channel proteins<sup>[15, 16, 18, 24]</sup> (Figure 1). For example, the hydroxyl groups on the second carbohydrate in ginsenoside Rg<sub>3</sub> form stable hydrogen bonds with the core amino acids of channel proteins (Figure 1). The 1st or 2nd amino acids after the K<sup>+</sup> channel 'signature sequence' (TXGYGD) at the pore entrances have been shown to interact with ginsenoside Rg<sub>3</sub>. That is, K318 in the KCNQ K<sup>+</sup> channel, S631 in the hERG K<sup>+</sup> channel, Y360 in the BK<sub>Ca</sub> K<sup>+</sup> channel, K531 in the K<sub>v</sub>1.4 channel, and K859 in the neuronal Na<sup>+</sup> channel. Similarly, the amino acids I417, N418, and L421 in the Na<sup>+</sup><sub>v</sub>1.2 channel, I433, N434, and L437 in the Na<sup>+</sup><sub>v</sub>1.4 channel, L417, N428, and L431 in the L-type Ca<sup>2+</sup> channel, V291, F292, and I295 in the 5-HT<sub>3</sub> channel, and L247 in the α7 nicotinic acetylcholine receptor were found to interact with ginsenosides<sup>[4, 14–18, 20]</sup> (Table 1 and Figure 1). Therefore, ginsenosides have regulatory effects in a broad range of ion

**Table 1.** Summary of the EC<sub>50</sub> and IC<sub>50</sub> values of ginsenoside-induced inhibitions or stimulations of the activities of various voltage-gated ion or ligand-gated ion channels.

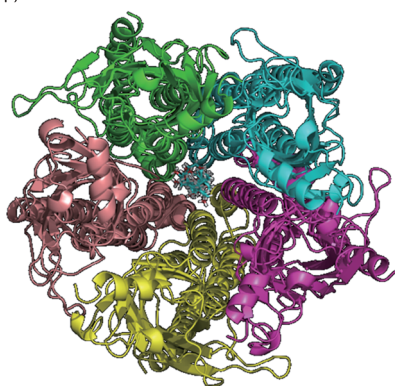
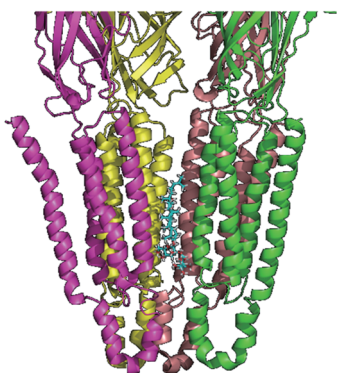
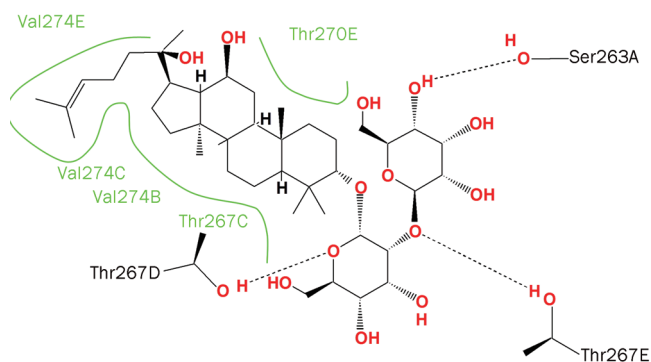
Voltage-gated ion channels	Ginsenoside	EC <sub>50</sub> or IC <sub>50</sub> (μmol/L)	Interacting amino acids
<b>Ca<sup>2+</sup></b>			
L	Rg <sub>3</sub> , Rb <sub>1</sub>	39.9±9.5 <sup>[1, 2]</sup>	L417, N428, L431
N	Rg <sub>3</sub>	64.4±13.6 <sup>[1]</sup>	
P/Q	Rg <sub>3</sub>	29.6±11.3 <sup>[1]</sup>	
R	Rg <sub>3</sub>	57.5±12.5 <sup>[1]</sup>	
T	Rg <sub>3</sub>	97.3±12.4 <sup>[1]</sup>	
<b>K<sup>+</sup></b>			
Kv1.4	Rg <sub>3</sub>	32.6±2.2 <sup>[4]</sup>	K531
BK <sub>Ca</sub>	Rg <sub>3</sub>	15.3±3.1 <sup>[3]</sup>	Y360
hERG	Rg <sub>3</sub>	0.41±0.05 <sup>[3]</sup>	S631
KCNQ	Rg <sub>3</sub>	15.2±8.7 <sup>[4]</sup>	K318
<b>Na<sup>+</sup></b>			
Nav1.2	Rg <sub>3</sub>	32.0±6.0 <sup>[5]</sup>	I417, N418, L421
Nav1.4	Rg <sub>3</sub>	58.5±6.3 <sup>[6]</sup>	I433, N434, L437
Nav1.5	Rg <sub>3</sub>	16.1±2.8 <sup>[7]</sup>	
<b>Ligand-gated ion channels</b>			
GABA <sub>A</sub>	Rc	53.0±12.3 <sup>[8]</sup>	
Glycine	Rf	49.8±9.8 <sup>[9]</sup>	
5-HT <sub>3</sub>	Rg <sub>3</sub>	27.6±4.3 <sup>[10]</sup>	V291, F292, I295
<b>Nicotinic acetylcholine</b>			
α3β4	Rg <sub>2</sub>	60±14 <sup>[11]</sup>	
α1β1δε	Rg <sub>2</sub>	16±9 <sup>[11]</sup>	
α7 (L247A mutant)	Rg <sub>3</sub>	33.1±1.3 <sup>[12]</sup>	L247
NMDA	Protopanaxatriol	48±16 <sup>[13]</sup>	

EC<sub>50</sub> values are shown for BK<sub>Ca</sub>, hERG, and KCNQ K<sup>+</sup> channels and GABA<sub>A</sub> and glycine receptors and IC<sub>50</sub> values are shown for the remainder and were determined in oocytes expressing these ion channels or receptors.

channels with low affinities, but their common factor is that they stabilize membrane potentials and attenuate cellular activities.

### Ginsenosides in ginseng pharmacology

Ginsenosides decrease the excitability of neuronal cells by inhibiting cation influx and/or by stimulating anion influx. These pharmacologic actions of ginsenosides are linked to reductions in the excitabilities of neurons, smooth muscle cells, and cardiomyocytes. Furthermore, the ginsenoside-induced modulation of Ca<sup>2+</sup> and K<sup>+</sup> channels results in the dilation of blood vessels via relaxation of smooth muscles<sup>[25–29]</sup>. In bradycardia, it induces relaxation of the cardiomyocytes, which explains its anti-hypertensive and cardioprotective effects<sup>[30–35]</sup>. In addition, ginsenoside-induced inhibition of the cation-gated NMDA receptor and neuronal Na<sup>+</sup> channels and

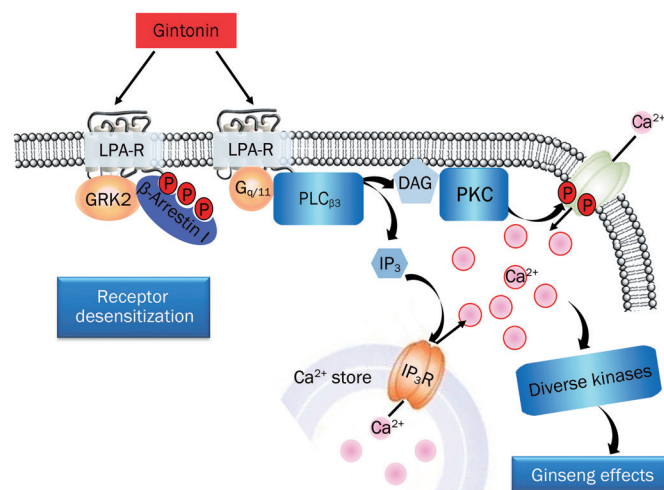
A nACh-Rg<sub>3</sub> (top)B nAChR-Rg<sub>3</sub> (side)C H-bondings: nAChR-Rg<sub>3</sub>

**Figure 1.** Virtual docking model of ginsenoside Rg<sub>3</sub> to chick  $\alpha 7$  nicotinic acetylcholine receptor (nAChR mutant L247T) and hydrogen bonds. Virtual dockings of ginsenoside Rg<sub>3</sub> to chick  $\alpha 7$  nicotinic acetylcholine receptor (AChR, L247T mutant) channel homology models. (A) Top view of the highest-ranked docking model of ginsenoside Rg<sub>3</sub> to chick  $\alpha 7$  nicotinic acetylcholine receptor (nAChR mutant, L247T) channel. The channel is shown as a cartoon diagram, and ginsenoside Rg<sub>3</sub> is represented by a ball and chain model. Subunits are shown in different colors. (B) Side view of the docking model of ginsenoside Rg<sub>3</sub> to nAChR receptor. One of the subunits is omitted in side view for clarity. (C) Poseview analysis of protein-ligand interactions. Hydrogen bonds are denoted by dotted lines. Spline sections indicate hydrophobic contacts, highlight the hydrophobic regions of ginsenoside Rg<sub>3</sub>, and provide the identities of contacting amino acids. The roman numerals in parenthesis indicate subunits of the pentamer. Adapted from Lee et al<sup>[18]</sup>.

its stimulation of anion-gated GABA<sub>A</sub> receptors and glycine receptors explain the neuroprotective and anxiolytic effects of ginseng<sup>[10, 36]</sup>. It is also possible that ginseng attenuation of cisplatin-induced nausea and vomiting is due its inhibition of 5-HT<sub>3</sub> ion channels<sup>[37]</sup>.

### Discovery of gintonin

The crude ginseng total saponin (cGTS) fraction contains approximately 50% ginsenosides by weight. Furthermore, the cGTS fraction increases intracellular Ca<sup>2+</sup> in mammalian cells and activates endogenous Ca<sup>2+</sup>-activated Cl<sup>-</sup> channels in *Xenopus* oocytes, whereas ginsenosides do not. These data imply that some unidentified ginseng component(s) is responsible for the Ca<sup>2+</sup> increase<sup>[7]</sup>. In addition, the cGTS fraction-induced Ca<sup>2+</sup> increases are both reversible and transient. Precise and continuing studies over the past twenty years have elucidated a signaling pathway downstream of cGTS in oocytes, namely the unknown membrane target protein-G<sub>q/11</sub>-PLC $\beta$ <sub>3</sub>-IP<sub>3</sub>-Ca<sup>2+</sup> release<sup>[38]</sup>. Later, homologous desensitization by cGTS was shown to be mediated through GRK2 and  $\beta$ -arrestin I in oocytes, strongly implying GPCR involvement (Figure 2)<sup>[39]</sup>.



**Figure 2.** Schematic diagram of gintonin-mediated signal transduction pathways. For the desensitization, gintonin activates LPA GPCRs, which leads to activation of GRK2. The activated GRK2 phosphorylates the LPA GPCRs and then  $\beta$ -arrestin I is recruited. The recruited  $\beta$ -arrestin I inhibits GPCR-G protein coupling. For cellular responses, gintonin activates LPA GPCRs, which leads to activation of phospholipase C (PLC). The activated PLC produces IP<sub>3</sub> and diacylglycerol (DAG). DAG activates protein kinase C (PKC), which phosphorylates Ca<sup>2+</sup> channels. IP<sub>3</sub> mobilizes Ca<sup>2+</sup> from internal Ca<sup>2+</sup> stores through IP<sub>3</sub> receptors. The increased Ca<sup>2+</sup> levels activate many kinases.

In 2011, the active ingredient in cGTS was finally separated by anion exchange chromatography after ginseng butanol extraction<sup>[40]</sup>. The novel non-saponin ingredient was designated as gintonin, where *gin* was derived from ginseng, *ton* from the tonic effects of ginseng, and *in* from

protein<sup>[41]</sup>. Six different gintonin have been identified, and all six induce intracellular  $\text{Ca}^{2+}$  increases in mammalian cells and *Xenopus* oocytes, confirming that gintonin is responsible for cGTS-induced  $\text{Ca}^{2+}$  mobilization. Gintonin is composed of carbohydrates, lipids, and proteins, such as, ginseng major latex-like protein and ginseng ribonuclease-like storage proteins. Thus, gintonin is part of a novel class of glycolipoproteins in ginseng that induces intracellular  $\text{Ca}^{2+}$  increases in mammalian cells<sup>[8]</sup>. Ginseng contains 0.2% gintonin by weight (Table 2).

**Table 2.** A brief comparison of ginseng components, gintonin, and ginsenosides.

	Gintonin	Ginsenosides
Molecular weight ( $M_w$ )	Native $M_w$ : 67 kDa Apparent $M_w$ : 13 kDa	0.6–1.3 kDa
Composition	Glycolipoprotein: carbohydrates (Glucose), lipids (LPA $\text{C}_{18:2}$ ), and ginseng proteins (GLP and GSP)	Dammarane glycosides
Content in ginseng	0.2%	3%–4% (Sum of individual ginsenosides)
Target protein on cell membrane and signal cascades	LPA receptors, transient [ $\text{Ca}^{2+}$ ] <sub>i</sub> elevation via PTX-sensitive and -insensitive G proteins coupled PLC pathway	Non-selective interac- tions with ion channels and receptors, do not have signal trans- duction pathway
Desensitization after repeated treatment on cells	Induction of rapid desensitization	No desensitization

### Gintonin activation of GPCRs for LPA

During additional studies with the gintonin, it was shown that phospholipase  $A_1$  had attenuating effects on gintonin. This finding highlighted the importance of position 1 esterification on the fatty acid component, which suggests that gintonin contains phospholipids<sup>[8]</sup>. Previously, Tigyi and Miledi demonstrated that LPA bound to serum albumin could sufficiently activate  $\text{Ca}^{2+}$ -activated  $\text{Cl}^-$  channels in *Xenopus* oocytes, and digestion with  $\text{PLA}_1$  prevented this activation<sup>[42]</sup>. Furthermore, using methanol extraction, they demonstrated LPA dissociation from serum albumin<sup>[42]</sup>. Building upon this reference, researchers examined whether gintonin contained LPAs by liquid chromatography-electrospray ionization/multi-stage mass spectrometry (LC-ESI-MS/MS) analysis<sup>[43]</sup>. LC-ESI-MS/MS analysis showed that 9.5% of gintonin is composed of lysophosphatidic acids (LPAs), such as LPA  $\text{C}_{18:2r}$ , LPA  $\text{C}_{16:0r}$  and LPA  $\text{C}_{18:1r}$  indicating that the bioactive component(s) of gintonin (with respect to the  $\text{Ca}^{2+}$  response) is a LPA<sup>[43]</sup>.

LPA is an intercellular lipid mediator that functions as a mitogen with hormone- and growth-factor-like activities in most cell types<sup>[9, 44]</sup>. Currently, three EDG family GPCRs (LPA<sub>1-3</sub>) and three purinergic GPCRs (LPA<sub>4-6</sub>) have been

reported to act as LPA receptors<sup>[9]</sup>. Hwang *et al* demonstrated that gintonin activates LPA receptors in GPCR-expressing B103 mammalian cells and in *Xenopus* oocytes<sup>[45]</sup>. Gintonin only activates LPA receptors specifically, not other GPCRs, such as S1P, fatty acids, 5-HT<sub>1C</sub>, and muscarinic acetylcholine receptor subtypes (m1, m3, and m5)<sup>[8]</sup>. Furthermore, the LPA GPCRs have different affinities for gintonin, which follow a decreasing order of LPA<sub>2</sub>>LPA<sub>5</sub>>LPA<sub>1</sub>>LPA<sub>3</sub>>LPA<sub>4</sub><sup>[8]</sup>. Furthermore, gintonin has 3- to 130-fold greater affinity for LPA GPCRs than free LPA. This might be due to ginseng proteins in gintonin because the protein components of gintonin may function as efficient LPA carriers to effectively carry LPA to LPA receptors and protect LPA from hydrolyzing enzymes<sup>[8]</sup>. Based on the average molecular weight of gintonin (20 kDa), four LPA molecules could most likely bind to one molecule of ginseng protein. The LPA content in ginseng is 80 to 240-fold higher than in other plants, which in part explains its unique pharmacologic properties. Somewhat unexpectedly, the action of ginseng is now linked to LPA and its GPCRs and *vice versa*, and this linkage opens new dimensions for ginseng pharmacology and LPA therapeutics (Table 2).

### Gintonin in ginseng pharmacology

Gintonin induces transient  $\text{Ca}^{2+}$  elevation and activates MAPK, PI3K, PKC, and Rho kinase via LPA receptors to evoke stimulatory cellular responses<sup>[4, 11]</sup> (Figure 2). Gintonin evokes cell proliferation and migration and morphological changes in human umbilical vein endothelial cells and PC12 neuronal cells<sup>[4, 5, 41, 45]</sup>. These effects of gintonin are consistent with those caused by LPAs via GPCRs and diverse G proteins, such as  $\text{G}\alpha_{i/o}$ ,  $\text{G}\alpha_{12/13}$ , and  $\text{G}\alpha_{q/11}$ <sup>[11]</sup>. Gintonin hinders the amyloidogenic pathway and induces non-amyloidogenic pathways that produce beneficial soluble APP $\alpha$  (sAPP $\alpha$ ) in neurons by activating LPA receptors<sup>[41]</sup>. Gintonin also reduces the release of  $\text{A}\beta_{1-42}$  and attenuates  $\text{A}\beta_{1-40}$ -induced cytotoxicity<sup>[41]</sup>. In addition, gintonin has been shown to rescue  $\text{A}\beta_{1-40}$ -induced cognitive dysfunction in mice<sup>[41]</sup>. Furthermore, in a transgenic murine Alzheimer's disease model, long-term oral administration of gintonin effectively attenuated both amyloid plaque deposition and short- and long-term memory impairment<sup>[41]</sup>. Thus, gintonin may contribute to the memory-improving effects of ginseng, which have been proven in human trials.

Autotaxin was found as an autocrine factor released by tumors, which stimulates tumor growth and migration<sup>[44]</sup>. Later, autotaxin was identified as lysophospholipase D, which produces LPA from lysophosphatidylcholine<sup>[46, 47]</sup>. Autotaxin and LPA function as mitogenic and motility signals in various cancers, including neuroblastoma, hepatoma, lung cancer, ovarian cancer, metastatic breast cancer, and melanoma<sup>[44]</sup>. A recent report showed that LPA  $\text{C}_{18:2r}$ , which is highly abundant in gintonin, inhibits autotaxin activity. Furthermore, Hwang *et al* showed that gintonin inhibits autotaxin activity *in vitro* and metastasis of B16/F10 melanoma cells *in vivo*<sup>[45]</sup>. Vessel formation in tumors was also reduced in gintonin-treated mice. These effects may contribute to the anti-cancer effects of

ginseng<sup>[45]</sup>.

### Yin and Yang of ginseng pharmacology

In Chinese medicine, Yin and Yang are opposite and complementary forces that combine to form a whole. Ginseng pharmacology is interesting because the two active components have opposite effects and complement the entire effect of ginseng. For example, ginsenosides stabilize membrane potentials via the dual modulation of ion channels, but gintonin and LPA activate many cellular responses via GPCR activation. Ginsenosides inhibit Ca<sup>2+</sup> influx, but gintonin induces a transient Ca<sup>2+</sup> rise, and ginsenosides have low affinity for calcium but are relatively abundant in ginseng, whereas gintonin has a higher affinity for calcium but is only present at low levels (Table 2). Thus, it is highly likely that ginsenosides act as a Yin component to gintonin's Yang component to produce the effects of ginseng. Therefore, it appears that the healing effects of *Panax* are the result of a harmonious balance between the positive and negative actions of ginsenosides and gintonins, reminiscent of the Yin and Yang forces in Chinese medicine.

### Concluding remark and perspective

Our understanding of ginseng pharmacology has advanced tremendously during the last few decades. Ginsenosides were considered to be the active ingredients of ginseng for 5 decades. However, many researchers used ginseng extract (the butanol fraction) for pharmacologic studies because individual ginsenosides were scarce and difficult to purify. As a result, other components were included in these pharmacology experiments. Gintonin is now considered as a part of the ginseng fraction, and the LPAs in this fraction are considered responsible for a variety of the biological effects of ginseng, which are mediated through GPCRs. Furthermore, it appears that the pharmacological actions of ginseng can now be explained by the complementary opposing actions of gintonin and ginsenoside. One example might be the anticancer effects of the ginseng extract. Many ginsenosides have shown clear anticancer activity in different cancer cell lines by regulation of cell proliferation<sup>[13]</sup>. Gintonin exhibited autotoxin inhibition *in vitro* and inhibition of metastasis of B16/F10 melanoma cells *in vivo*<sup>[45]</sup>. Therefore, both components in ginseng may act synergistically and complementarily for the anti-tumor efficacy of ginseng.

How ginsenosides and gintonin work together in the body or whether ginsenosides affect the actions of gintonin or *vice versa* remains to be established. Future studies are required to elucidate the pharmacological effects of the different actions of ginsenosides and gintonin with respect to their individual contributions and the effects of whole ginseng in biological systems. Future research will undoubtedly expand our knowledge of ginseng pharmacology and the applications of LPA. In addition, the therapeutic application of LPA might be facilitated by knowledge of the traditional usages of ginseng. Although new drug developments based on targeting LPA receptors are in the pipeline, gintonin might provide alterna-

tive therapies for pathologic conditions related to LPA and LPA receptor-related diseases.

### Acknowledgements

This work was supported by the Basic Science Research Program (2011-0021144 and 2011-0021158) and the Priority Research Center Program through the National Research Foundation of Korea (NRF), which is funded by the Ministry of Education, Science, and Technology (2012-0006686), by a grant from the BK21 project funded to Seung-yeol NAH and by the MRC program (Grant No 2009-0083538) of the Korean National Research Foundation funded by the Korean government (MEST).

### References

- 1 Nah SY, Kim DH, Rhim H. Ginsenosides: are any of them candidates for drugs acting on the central nervous system? *CNS Drug Rev* 2007; 13: 381-404.
- 2 Lee JH, Choi SH, Lee BH, Yoon IS, Shin TJ, Pyo MK, et al. Modifications of aliphatic side chain of 20(S)-ginsenoside Rg<sub>3</sub> cause an enhancement or loss of brain Na<sup>+</sup> channel current inhibitions. *Biol Pharm Bull* 2008; 31: 480-6.
- 3 Choi SH, Lee JH, Pyo MK, Lee BH, Shin TJ, Hwang SH, et al. Mutations Leu427, Asn428, and Leu431 residues within transmembrane domain-I-segment 6 attenuate ginsenoside-mediated L-type Ca<sup>2+</sup> channel current inhibitions. *Biol Pharm Bull* 2009; 32: 1224-30.
- 4 Lee BH, Lee JH, Lee SM, Jeong SM, Yoon IS, Choi SH, et al. Identification of ginsenoside interaction sites in 5-HT<sub>3A</sub> receptors. *Neuropharmacology* 2007; 52: 1139-50.
- 5 Jang S, Ryu JH, Kim DH, Oh S. Changes of [3H]MK-801, [3H] muscimol and [3H]flunitrazepam binding in rat brain by the prolonged ventricular infusion of transformed ginsenosides. *Neurochem Res* 2004; 29: 2257-66.
- 6 Kim S, Kim T, Ahn K, Park WK, Nah SY, Rhim H. Ginsenoside Rg<sub>3</sub> antagonizes NMDA receptors through a glycine modulatory site in rat cultured hippocampal neurons. *Biochem Biophys Res Commun* 2004; 323: 416-24.
- 7 Choi S, Rho SH, Jung SY, Kim SC, Park CS, Nah SY. A novel activation of Ca<sup>2+</sup>-activated Cl<sup>-</sup> channel in *Xenopus* oocytes by Ginseng saponins: evidence for the involvement of phospholipase C and intracellular Ca<sup>2+</sup> mobilization. *Br J Pharmacol* 2001; 132: 641-8.
- 8 Hwang SH, Shin TJ, Choi SH, Cho HJ, Lee BH, Pyo MK, et al. Gintonin, newly identified compounds from ginseng, is novel lysophosphatidic acids-protein complexes and activates G protein-coupled lysophosphatidic acid receptors with high affinity. *Mol Cells* 2012; 33: 151-62.
- 9 Choi JW, Chun J. Lysophospholipids and their receptors in the central nervous system. *Biochim Biophys Acta* 2013; 1831: 20-32.
- 10 Kim JH, Cho SY, Lee JH, Jeong SM, Yoon IS, Lee BH, et al. Neuroprotective effects of ginsenoside Rg<sub>3</sub> against homocysteine-induced excitotoxicity in rat hippocampus. *Brain Res* 2007; 1136: 190-9.
- 11 Shin TJ, Kim HJ, Kwon BJ, Choi SH, Kim HB, Hwang SH, et al. Gintonin, a ginseng-derived novel ingredient, evokes long-term potentiation through N-methyl-D-aspartic acid receptor activation: Involvement of LPA receptors. *Mol Cells* 2012; 34: 563-72.
- 12 Choi KT. Botanical characteristics, pharmacological effects and medicinal components of Korean *Panax* ginseng C A Meyer. *Acta Pharmacol Sin* 2008; 29: 1109-18.

- 13 Nag SA, Qin JJ, Wang W, Wang MH, Wang H, Zhang R. Ginsenosides as anticancer agents: *in vitro* and *in vivo* activities, structure-activity relationships, and molecular mechanisms of action. *Front Pharmacol* 2012; 3: 25.
- 14 Lee JH, Jeong SM, Kim JH, Lee BH, Yoon IS, Choi SH, et al. Effects of ginsenosides and their metabolites on voltage-dependent  $Ca^{2+}$  channel subtypes. *Mol Cells* 2006; 21: 52–62.
- 15 Choi SH, Shin TJ, Hwang SH, Lee BH, Kang J, Kim HJ, et al. Ginsenoside Rg(3) decelerates hERG  $K^+$  channel deactivation through Ser631 residue interaction. *Eur J Pharmacol* 2011; 663: 59–67.
- 16 Choi SH, Shin TJ, Lee BH, Chu DH, Choe H, Pyo MK, et al. Ginsenoside Rg<sub>3</sub> activates human KCNQ1  $K^+$  channel currents through interacting with the K318 and V319 residues: a role of KCNE1 subunit. *Eur J Pharmacol* 2010; 637: 138–47.
- 17 Lee JH, Jeong SM, Kim JH, Lee BH, Yoon IS, Choi SH, et al. Characteristics of ginsenoside Rg3-mediated brain  $Na^+$  current inhibition. *Mol Pharmacol* 2005; 68: 1114–26.
- 18 Lee JH, Lee BH, Choi SH, Yoon IS, Pyo MK, Shin TJ, et al. Ginsenoside Rg<sub>3</sub> inhibits human Kv1.4 channel currents by interacting with the Lys531 residue. *Mol Pharmacol* 2008; 73: 619–26.
- 19 Kang DI, Lee JY, Yang JY, Jeong SM, Lee JH, Nah SY, et al. Evidence that the tertiary structure of 20(S)-ginsenoside Rg(3) with tight hydrophobic packing near the chiral center is important for  $Na^+$  channel regulation. *Biochem Biophys Res Commun* 2005; 333: 1194–201.
- 20 Lee BH, Choi SH, Pyo MK, Shin TJ, Hwang SH, Kim BR, et al. A role for Leu247 residue within transmembrane domain 2 in ginsenoside-mediated  $\alpha 7$  nicotinic acetylcholine receptor regulation. *Mol Cells* 2009; 27: 591–9.
- 21 Kim CS, Son SJ, Kim HS, Kim YD, Lee KS, Jeon BH, et al. Modulating effect of ginseng saponins on heterologously expressed HERG currents in *Xenopus* oocytes. *Acta Pharmacol Sin* 2005; 26: 551–8.
- 22 Choi SE, Choi S, Lee JH, Whiting PJ, Lee SM, Nah SY. Effects of ginsenosides on GABA<sub>A</sub> receptor channels expressed in *Xenopus* oocytes. *Arch Pharm Res* 2003; 26: 28–33.
- 23 Noh JH, Choi S, Lee JH, Betz H, Kim JI, Park CS, et al. Effects of ginsenosides on glycine receptor  $\alpha 1$  channels expressed in *Xenopus* oocytes. *Mol Cells* 2003; 15: 34–9.
- 24 Choi SH, Shin TJ, Lee BH, Hwang SH, Lee SM, Lee BC, et al. Ginsenoside Rg<sub>3</sub> enhances large conductance  $Ca^{2+}$ -activated potassium channel currents: a role of Tyr360 residue. *Mol Cells* 2011; 31: 133–40.
- 25 Kwan CY, Kwan TK. Effects of *Panax notoginseng* saponins on vascular endothelial cells *in vitro*. *Acta Pharmacol Sin* 2000; 21: 1101–5.
- 26 Li Z, Chen X, Niwa Y, Sakamoto S, Nakaya Y. Involvement of  $Ca^{2+}$ -activated  $K^+$  channels in ginsenosides-induced aortic relaxation in rats. *J Cardiovasc Pharmacol* 2001; 37: 41–7.
- 27 Kim ND, Kang SY, Park JH, Schini-Kerth VB. Ginsenoside Rg3 mediates endothelium-dependent relaxation in response to ginsenosides in rat aorta: role of  $K^+$  channels. *Eur J Pharmacol* 1999; 367: 41–9.
- 28 Chung I, Kim ND. Ginseng saponins enhance maxi  $Ca^{2+}$ -activated  $K^+$  currents of the rabbit coronary artery smooth muscle cells. *J Ginseng Res* 1999; 23: 230–4.
- 29 Chung I, Lee JS. Ginsenoside Rg3 increases the ATP-sensitive  $K^+$  channel activity in the smooth muscle of the rabbit coronary artery. *J Ginseng Res* 1999; 23: 235–8.
- 30 Jeon BH, Kim CS, Kim HS, Park JB, Nam KY, Chang SJ. Effect of Korean red ginseng on blood pressure and nitric oxide production. *Acta Pharmacol Sin* 2000; 21: 1095–100.
- 31 He H, Xu J, Xu Y, Zhang C, Wang H, He Y, et al. Cardioprotective effects of saponins from *Panax japonicus* on acute myocardial ischemia against oxidative stress-triggered damage and cardiac cell death in rats. *J Ethnopharmacol* 2012; 140: 73–82.
- 32 Li HX, Han SY, Ma X, Zhang K, Wang L, Ma ZZ, et al. The saponin of red ginseng protects the cardiac myocytes against ischemic injury *in vitro* and *in vivo*. *Phytomedicine* 2012; 19: 477–83.
- 33 Bai CX, Takahashi K, Masumiya H, Sawanobori T, Furukawa T. Nitric oxide-dependent modulation of the delayed rectifier  $K^+$  current and the L-type  $Ca^{2+}$  current by ginsenoside Re, an ingredient of *Panax ginseng*, in guinea-pig cardiomyocytes. *Br J Pharmacol* 2004; 142: 567–75.
- 34 Furukawa T, Bai CX, Kaihara A, Ozaki E, Kawano T, Nakaya Y, et al. Ginsenoside Re, a main phytosterol of *Panax ginseng*, activates cardiac potassium channels via a nongenomic pathway of sex hormones. *Mol Pharmacol* 2006; 70: 1916–24.
- 35 Attele AS, Wu JA, Yuan CS. Ginseng pharmacology: multiple constituents and multiple actions. *Biochem Pharmacol* 1999; 58: 1685–93.
- 36 Kim TW, Choi HJ, Kim NJ, Kim DH. Anxiolytic-like effects of ginsenosides Rg3 and Rh2 from red ginseng in the elevated plus-maze model. *Planta Med* 2009; 75: 836–9.
- 37 Kim JH, Yoon IS, Lee BH, Choi SH, Lee JH, Jeong SM, et al. Effects of Korean red ginseng extract on cisplatin-induced nausea and vomiting. *Arch Pharm Res* 2005; 28: 680–4.
- 38 Choi S, Kim HJ, Ko YS, Jeong SW, Kim YI, Simonds WF, et al.  $G\alpha_{q/11}$  coupled to mammalian phospholipase C beta 3-like enzyme mediates the ginsenoside effect on  $Ca^{2+}$ -activated  $Cl^-$  current in the *Xenopus* oocyte. *J Biol Chem* 2001; 276: 48797–802.
- 39 Lee JH, Jeong SM, Lee BH, Noh HS, Kim BK, Kim JI, et al. Prevention of ginsenoside-induced desensitization of  $Ca^{2+}$ -activated  $Cl^-$  current by microinjection of inositol hexakisphosphate in *Xenopus laevis* oocytes: involvement of GRK2 and beta-arrestin I. *J Biol Chem* 2004; 279: 9912–21.
- 40 Pyo MK, Shin TJ, Choi SH, Lee BH, Pyo MK, Lee JH, et al. Novel glycoproteins from ginseng. *J Ginseng Res* 2011; 35: 92–103.
- 41 Hwang SH, Shin EJ, Shin TJ, Lee BH, Choi SH, Kang J, et al. Gintonin, a ginseng-derived lysophosphatidic acid receptor ligand, attenuates Alzheimer's disease-related neuropathies: involvement of non-amyloidogenic processing. *J Alzheimers Dis* 2012; 31: 207–23.
- 42 Tigyi G, Miledi R. Lysophosphatidates bound to serum albumin activate membrane currents in *Xenopus* oocytes and neurite retraction in PC12 pheochromocytoma cells. *J Biol Chem* 1992; 267: 21360–7.
- 43 Yoon HR, Kim H, Cho SH. Quantitative analysis of acyl-lysophosphatidic acid in plasma using negative ionization tandem mass spectrometry. *J Chromatogr B Analyt Technol Biomed Life Sci* 2003; 788: 85–92.
- 44 Houben AJ, Moolenaar WH. Autotaxin and LPA receptor signaling in cancer. *Cancer Metastasis Rev* 2011; 30: 557–65.
- 45 Hwang SH, Lee BH, Kim HJ, Cho HJ, Shin HC, Im KS, et al. Suppression of metastasis of intravenously-inoculated B16/F10 melanoma cells by the novel ginseng-derived ingredient, gintonin: Involvement of autotaxin inhibition. *Int J Oncol* 2013; 42: 317–26.
- 46 Tokumura A, Majima E, Kariya Y, Tominaga K, Kogure K, Yasuda K, et al. Identification of human plasma lysophospholipase D, a lysophosphatidic acid-producing enzyme, as autotaxin, a multifunctional phospholipase.

- phodiesterase. *J Biol Chem* 2002; 277: 39436–42.
- 47 Umezu-Goto M, Kishi Y, Taira A, Hama K, Dohmae N, Takio K, et al. Autotaxin has lysophospholipase D activity leading to tumor cell growth and motility by lysophosphatidic acid production. *J Cell Biol* 2002; 158: 227–33.



**This work is licensed under the Creative Commons Attribution-NonCommercial-No Derivative Works 3.0 Unported License. To view a copy of this license, visit <http://creativecommons.org/licenses/by-nc-nd/3.0/>**