

氟碳代血液对雄性小鼠生殖细胞的遗传效应

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提要 A系♂小鼠一次静脉输注氟碳乳剂(FCE) 50和10 ml/kg后7 d, 观察精原细胞及初级精母细胞畸变。睾丸光镜及电镜组织形态及成熟精子形态, 以评价其对各期生殖细胞的损伤。结果表明精原细胞数目和结构畸变率, 用药组和对照组均在3%以下。初级精母细胞总畸变率三组间亦无明显差异。成熟精子及睾丸组织结构及超微结构均未见异常。

关键词 氟碳; 精子生成; 精子; 染色体畸变; 电子显微镜检查

从基因及体细胞水平的诱变性检测结果, 未见氟碳代血液 fluorocarbon emulsion (FCE) 的诱变毒性⁽¹⁾。为全面评价其安全性, 进行潜在性危害的检测, 不仅对体细胞而且对生殖细胞的遗传损伤作研究。本文对输注不同剂量 FCE 小鼠睾丸精原细胞和初级精母细胞, 精子进行畸变分析, 并对睾丸组织作光学及电子显微镜检查, 以评价其对遗传影响。

材料和方法

FCE 由中国科学院上海有机化学研究所提供。分子式为(CF₃CF₂CF₂)₃N, 临床输注品, 批号 830241。

小鼠 51 只 A 系♂小鼠, 由上海医科大学实验动物房供应。体重 29 ± SD 6 g。随机分成

FCE 50 ml/kg 组, FCE 10 ml/kg 组及生理盐水 50 ml/kg 对照组。iv 1 次给药后 7 d, 颈椎脱臼处死, 取睾丸, 每组 12 只制备有丝分裂和减数分裂相中期细胞, 5 只取副睾制备精子标本并作睾丸组织病理学观察。

精原细胞和精母细胞制备 按照文献(2)方法略作改进。小鼠 ip 秋水仙素 4 mg/kg 16 h 后处死取睾丸。分离精曲小管。加 0.05 M KCL 低渗 30 min, 充份吹打, 倾去上清液, 加入甲醇: 冰醋酸(3:1)固定, 上层悬液倾入离心管。剩余精曲小管碎片以 2 ml 60%冰醋酸软化 3 min, 加固定液 5 min, 200 × g 离心 10 min, 弃上清。两份液各重复固定 2 次, 放 4°C 保存 2 d 后冰片制片, Giemsa 液染色, 空气干燥, 镜检。

成熟精子制备 取一侧副睾放入生理盐水 2 ml 剪碎, 以 4 层擦镜纸过滤, 滤液以 200 × g 离心 5 min, 取沉淀加适量盐水制成悬液, 涂片, 甲醇固定, 2%伊红染色 1 h, 高倍镜下检查 1000 个精子, 计数畸变率。

结 果

小鼠一次 ip FCE 50 及 10 ml/kg 后精原细

Tab 1. Chromosomal aberrations in spermatogonia from mice 1 wk after infusion of fluorocarbon emulsion (FCE) 10 or 50 ml/kg

Doses ml/kg	Male mice	Cells scored	Type of chromosomal aberrations (%)				
			Aneuploid	Polyploid	Break	Translocations	Total
FCE 50	13	573	2	15(2.9)	2(0.2)	1(0.5)	3.4
FCE 10	13	640	2	15(2.6)	1(0.1)	-	2.8
Saline 50	10	500	3	4(1.4)	1(0.2)	-	1.6

Tab 2. Chromosomal aberrations in spermatocyte I from mice 7 wk after infusion of FCE 10 or 50 ml/kg.

Doses (ml/kg)	Male mice	Cells scored	Types of chromosomal aberrations (%)					
			Tri- ploid	Tetra- ploid	X-Y univalent	Autosomal univalent	Ring quadrivalent	Total
FCE 50	13	571	-	4(0.7)	10(1.7)	-	-	2.4
FCE 10	13	630	1	1(0.1)	7(1.1)	1(0.1)	1(0.1)	1.7
Saline 50	10	500	-	1(0.2)	6(1.2)	-	-	1.4

胞的遗传损伤见表1,各组均有染色体畸变,包括非整倍体及超二倍体(图1A,见图版1,以下各图同)。总畸变率分别为3.4%及2.8%,对照组为1.6%。其中断裂、互换不超过1%,均在正常范围,各组间无明显差异。

初级精母细胞中期分裂相染色体畸变结果见表2。性染色体早熟分离见(图1B),二剂量组与对照组间发生率无明显差异。10 ml/kg剂量组见到1例常染色体早熟分离及1例常染色体易位,具有1环状4价体及18个双价体(图1C)。此外,在各组中尚见到3倍体和4倍体细胞(图1D)。

精子及睾丸光镜检查形态正常。上皮层次正常。各期生殖细胞发育良好。电镜显示精母细胞及次级精子细胞成熟正常(图1E, F)。

讨 论

性成熟后的♂哺乳动物生殖细胞始终处于分裂周期,其中包括不同发育阶段的精原细胞,初级精母细胞,次级精母细胞,精细胞及成熟精子。某些环境因素可引起各期细胞突变,造成配子死亡或子代遗传性状改变,如流产、死胎、不育、先天畸形和肿瘤形成等⁽³⁾。

精原细胞中期分裂相畸变,包括断裂和相互易位(reciprocal translocation)。本实验结果FCE 50 ml/kg输注小鼠仅出现1例易位,断裂频率与对照组无差别,未见FCE对精原细胞的明显损伤。

当精母细胞同源染色体的断片经减数分裂

联会配对时,无着丝点断片可产生遗传性易位而形成环状或链状多价体。易位可发生在常染色体间或常-性染色体间。这种染色体结构异常,可造成配子紊乱及精子生成障碍。本实验结果FCE 10 ml/kg输注组630只细胞中仅出现1个常染色体易位的4价体环。高剂量组及对照组均未检得。尚难评价有遗传损伤。

初级精母细胞畸变的另一指标是观察常或性染色体的单价体。分别显示配对染色体的早熟分离。由于同源断片间缺失合子配对而不联会,亦可影响精子的生成。正常动物X-Y染色体早熟分离发生率有6.7%,常染色体发生率有3.6%⁽⁴⁾。本实验结果均低于2%。

上述结果表明,小鼠经不同剂量FCE输注后,未见对精原细胞,初级精母细胞,成熟精子及睾丸组织的损伤作用。FCE不诱发♂小鼠生殖细胞遗传损伤。

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Genetic effects of fluorocarbon emulsion on germinal cells of male mice

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ABSTRACT The A strain ♂ mice were infused with fluorocarbon emulsion (FCE) 50, 10 ml/kg and NS 50 ml/kg. Metaphase plates of spermatogonia and spermatocytes I were examined on d 7 after infusion and histological damage of the testes was studied by light and electron microscopes.

The frequencies of aneuploid and polyploid of spermatogonia were 2.9, 2.5 and 1.4%, respectively induced by FCE 50, 10 ml/kg and NS 50 ml/kg. While those of breaks and reciprocal translocations were 0.5, 0.1 and 0.2%, respectively. In sper-

matocytes I the frequency of X-Y univalent and autosomal univalent were 2% ($p < 0.05$) and no morphological changes were found in spermatogonia developed from spermatogenic cells.

It may be concluded that no chromosomal aberrations were induced in germinal cells of ♂ mice by FCE.

KEY WORDS fluorocarbons; spermatogenesis; spermatozoa; chromosome aberrations; electron microscopy

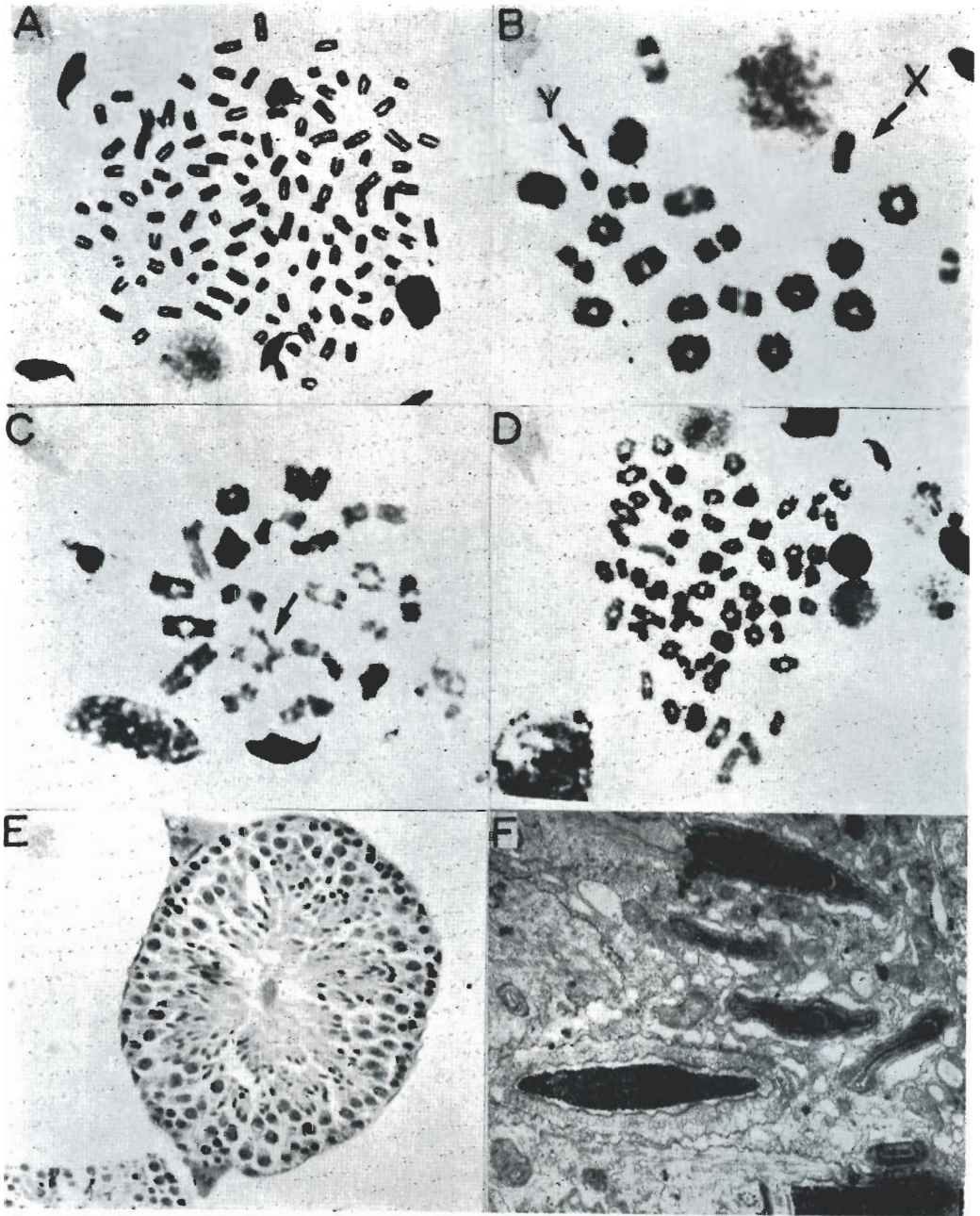


Fig 1. Genetic effects of fluorocarbon emulsion (FCE) infusion in mice. A) Metaphase spermatogonia with polyplody after FCE 10 ml/kg, $\times 1000$. B) Metaphase spermatocyte I with X-Y univalents (arrow) after PCE 50 ml/kg, $\times 1200$. C) Metaphase spermatocyte I with bivalents and ring quadrivalent (arrow) after FCG 10 ml/kg, $\times 1000$. D) Metaphase spermatocyte I with a hexaploid cell after FCE 10 ml/kg, $\times 1000$. E) Immature sperm cells were normal and spermatozoa was abundant in the testis after FCE 50 and 10 ml/kg infusion. F) Normal primary spermatocyte developed from spermatozoa. The chromatin was very abundant and dense mitochondria and endoplasmic reticulum were very prominent (partly dilated).