Inhibitory effects of triazolodiazepine on mouse splenocytes and peritoneal macrophages in vitro

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ABSTRACT Effects of triazolodiazepine (Tri) on mouse splenocyte proliferation and macrophage phagocytosis were studied. Tri 0. 1 – 10 µmol·L⁻¹ significantly inhibited concanavalin A (Con A) 5 µg·ml⁻¹ and lipopolysaccharides (LPS) 10 µg·ml⁻¹-induced splenocyte proliferation and phagocytic activity of activated peritoneal macrophages. Tri 10 µmol·L⁻¹ reduced Con A induced colony stimulating factor release from mouse splenocytes and LPS-challenged intracellular and extracellular interleukin-1 production from peritoneal macrophages. These results partially explain the wide and strong anti-inflammatory effects of Tri.

KEY WORDS triazolodiazepine: interleukin-1; colony-stimulating factors; macrophages; phagocytosis; spleen

The participation of platelet-activating factor (PAF) in inflammatory and immunological responses has been studied intensively. PAF antagonists inhibited most of PAF effects in vitro and in vivo^(1,2). Triazolodiazepine (Tri, WEB 2086) is a hetrazepine PAF antagonist. Previous work emphasized on its antagonism of PAF effects and paid little attention to its effect on immune process⁽²⁾. We speculated that its strong and wide antiinflammatory effects might be linked to its inhibition on immunological reactions. In the present study, we observed the effects of Tri on mitogen-induced mouse splenocyte proliferation and colony stimulating factor (CSF) secretion. Effects of Tri on phagocytic activity and interleukin-1 (IL-1) production of activated peritoneal macrophages were also studied.

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 $Triazolodiazepine (WEB 0286) \\ 3-[4-(2-Chlorophenyl)-9-methyl-6H-thieno[3,2-f] \\ [1,2,4]triazolo-[4,3-a][1,4]-diazepin-2-yl]-1- \\ (4-morpholinyl)-1-propanone \\ C_{22}H_{22}ClN_5O_2S \qquad M_r = 455.97 \\ \end{cases}$

MATERIALS AND METHODS

Chemicals and mice Tri was gifted from Boehringer Ingelheim KG, Ingelheim am Rhein, Germany. Concanavalin A (Con A), lipopolysaccharides (LPS, *E coli* 055:B5), neutral red, and RPMI-1640 medium were purchased from Sigma Chemical Co, USA. [¹H] TdR (814 TBq \cdot mol⁻¹) was obtained from Shanghai Institute of Nuclear Research, Chinese Academy of Sciences. Fetal calf serum (FCS) was purchased from Department of Pathology, Second Military Medical University. BALB c and ICR mice of either sex weighting 16-24 g were supplied by Animal Center, Second Military Medical University.

Splenocyte proliferation BALB e mice were killed by cervical dislocation and spleen cells were prepared¹⁴. Splenocytes at 5×10^5 /well in 0.1 ml RPMI-1610 containing 10^{16} heat-inactivated FCS were seeded into 96-well flat-hottom microtiter plates. Tri was added to the cells in the presence or abstrace of Con χ 5 gg+ml⁻¹ or LPS 10 µg+ml⁻¹ and much red in a homodefied 5^{1-1} CD₁ atomosphere or 57^{+} . J H J fdR 1850 Fe[/well was added for the her 12 b of alture periodtells were honested at 52 b and cell associated radioactivity was measured in a FJ-2107 scintillation counter (Xi-an, China).

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CSF assay Con A 5 µg·ml⁻¹ alone or in combination with Tri were added to murine splenocytes at 5 $\times 10^6$ cells per well per ml to induce CSF. After 96 h of incubation in a 5% CO2 incubator at 37 C . supernatants were harvested by centrifugation $(1000 \times g, 10)$ min). CSF content was assessed using mouse bone marrow cell (BMC) proliferation assay⁽⁵⁾. Briefly. BMC from ICR mice was seeded at 2×10^4 cells per well in 0.1 ml RPMI-1640 with 20% FCS and 20% horse serum. Another 0.1 ml of RPMI-1640 containing serial dilutions of test samples was added to each well. After 90 h of incubation the cells were pulsed with ['H]TdR 1850 Bq per well for 6 h. The cells were harvested using a microplate cell harvester and the incorporation of [¹H]TdR was determined by scintillation counting.

IL-1 measurement Peritoneal exudate cells were obtained from thioglycollate-primed ICR mice⁽⁶⁾. The cells were added at 2 × 10⁶ cells per well in 1 ml RPMI-1640 countaining 10% FCS for 2 h in a humidified incubator at 37 (with 5% CO2 to facilitate cell adherence. The media was discarded and LPS 10 μ g ml⁻¹ were added to each well in a final volume of 1 ml RP-MI-1640 alone or together with different concentrations of Tri to induce IL-1. After 24 h of culture the media were collected and centrifugated to get supernatants. One ml of fresh medium was added to the cell monolayer and the samples were fruzen. After 3 cycles of freezing/thawing, the IL-1 activity in these samples was referred as the cell-associated one. IL-1 content in these samples and the supernatants was assessed using Con A (2.5 μ g • ml⁻¹)-induced ICR thymocyte proliferation assay⁽¹⁾. Intracellular and extracellular IL-1 activities were expressed as dpm of ['H] TdR incorporated by thymocytes at 1+8 dilution.

Phagocytic activity determination Phagocytosis was determined employing the method of neutral red⁽⁷⁾ with minor modifications. Murine exudate cells primed 4 d previously were added at 2×10^5 /well. Macrophages were separated by adherence on the plates. Various concentrations of Tri were added to the adherent macrophages in a volume of 0.1 RPMI-1640 supplemented with 10% FCS. After 24 h of incubation, the media was discarded and 0.1 ml of 0.1% neutral red in PBS was added to each well. The cells were incubated at 37 C for 30 min. The neutral red was removed and the cells were washed twice by PBS. Extraction solution 0. I ml (EtOH: HAc = 1:1) was added to each well to extract the dye taken by macrophages and absorbance of the color was read on a DG 3022 microplate spectrophotometer (Xi-an, China) at 550 nm. The phagocytic activity was expressed as the amount of neutral red phagocytized by macrophages.

Statistical analysis All data were expressed as $\bar{x} \pm s$ and analyzed by t test.

RESULTS

Mitogen-induced mouse splenocyte proliferation The levels of proliferation were determined by measuring the radioactivity of [^sH]TdR. Tri 0.01-10 μ mol·L⁻¹ inhibited the splenocyte proliferation induced by Con A 5 μ g·ml⁻¹ or LPS 10 μ g·ml⁻¹ concentrationdependently (Tab 1).

Tab 1. Effect of triazolodiazepine (Tri) on mouse splenocyte proliferation induced by concanavalin A (Con A 5 μ g·ml⁻¹) or lipopolysaccharides (LPS 10 μ g·ml⁻¹). n=6 samples. $\overline{x}\pm s$. ${}^{\circ}P > 0.05$, ${}^{\circ}P < 0.05$, (P < 0.01 vs 0.

Radioactivity/dpm		
Con A	LPS	
106 037±9 443	7654 ± 615	
91 407±7 142 ^b	7 074±627°	
88 205±14 02l ^b	$6\ 212\pm566^{\circ}$	
$68554\pm 5605'$	4 617 \pm 543°	
63 458±4 241′	$4\ 221\pm 606^{\circ}$	
28 378±3 851°	$1504 \pm 214^{\circ}$	
	Con A 106 037±9 443 91 407±7 142 ^b 88 205±14 021 ^b 68 554±5 605' 63 458±4 241'	

Production of CSF by mouse splenocytes CSF activity was determined by BMC proliferation assay. Con A 5 μ g·ml⁻¹-induced CSF activity at different dilutions was obviously reduced by Tri at 10 μ mol·L⁻¹ but not at lower concentrations (Fig 1).

IL-1 release and synthesis When added to LPS-stimulated adherent peritoneal macrophages, Tri markedly inhibited the

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Fig 1. Effect of Tri on production of colony-stimulating factor (CSF) from mouse splenocytes induced by Con A 5 ug \cdot ml⁻¹. n = 6, $\overline{x} \pm s$. "P > 0.05, "P < 0.05, "P < 0.05, "P < 0.05, "P < 0.01 us 0.

release of IL-1 in a concentration-dependent manner. Similar decrease was obtained in the cell-associated IL-1 activity (Tab 2). The intracellular production of IL-1 was less than extracellular one.

Tab 2. Inhibitory effect of Tri on LPS-induced production of IL-1 from mouse peritoneal macrophages, expressed as [³H] TdR incorporation by mouse thymocytes at 1:8 dilution. n=3 samples. $\overline{x}\pm s$. "P>0.05, "P<0.05, "P<0.05, "P<0.01 us 0.

Tri/µmol•1'	Radioactivity/dpm	
	Intracellular	Extracellular
U	13852 ± 1758	20.954 ± 4.231
0.001	$11\ 645 \pm 1\ 696^{\circ}$	$16\ 885 \pm 2\ 003^{\circ}$
0.01	$10.692 \pm 2.546^{ m b}$	$13473\pm523^{\text{b}}$
0.1	$10.421 \pm 878^{\circ}$	$11\ 596\pm 2\ 664^{b}$
1	$4.823 \pm 1.390^{\circ}$	$8.832 \pm 921^{\circ}$
10	$5824 \pm 1343^{\circ}$	$5770 \pm 265^{\circ}$

Phagocytosis Phagocytic activity of activated macrophages was presented as amount of neutral red taken by macrophages. Tri (> 0. 1 μ mol·L⁻¹) exerted an inhibitory effect on phagocytosis of mouse peritoneal macrophages (Tab 3).

DISCUSSION

Tri markedly inhibited most of PAF

Tab 3. Effect of Tri on phagocytic activity of activated mouse peritoneal macrophages, expressed as the amount of neutral red phagocytized by macrophages. n = 6 samples. $\overline{x} \pm s$. "P > 0.05. "P < 0.05, "P < 0.01 us 0.

Γri/µmol•L ^{−1}	Neutral red/µg
• 0	 11. 26±1. 78
0.001	$11.06 \pm 1.58^{\circ}$
0.01	$10.46 \pm 2.57^{\circ}$
0.1	9.04 ± 0.99^{b}
1	7.83±1.57°
10	7.45 \pm 0.77°

response and elicited strong anti-inflammatory effects. In this paper Tri inhibited Con A and LPS-induced murine spleen cell proliferation. Con A and LPS induced the proliferation of T and B lymphocytes, respectively. Ward SG et al reported Tri had no effects on mitogen or interleukin-2-induced human T lymphocyte proliferation¹⁸. Different cell resources may be explained as the cause of this contradiction. CSF not only stimulates the proliferation and differentiation of both myelomonocytic and erythroid stem cells, but also induces a significant leukotrienes synthesis and enhances r arachidonic acid release, amplifying the inflammatory response⁽⁹⁾. IL-1 is also an important mediator in inflammatory reactions. In this study Tri reduced Con A-induced CSF release from splenocytes and LPS-stimulated IL-1 production from peritoneal macrophages. Phagocytic activity is one of the important characteristics of macrophages.⁷. Phagocytosis takes part in the beginning of various immunological response. Tri inhibited the phagocytic activity of activated macrophages. This article does not determine the precise mechanism of this action. The inhibition of immunological process including T, B lymphocyte proliferation, macrophage phagocytosis and cytokine production may partially explain

its wide and strong anti-inflammatory effects.

PAF-acether increased release, synthesis total production of IL-1 hy and rat macrophages. this effect of PAF was inhibited in the presence of the specific antagonist, BN Challenged by LPS, macrophages 52021. produced both IL-1 and PAF^{(3,10}. These results suggest that PAF plays a key role in the production of IL-1. Our previous data showed that Tri decreased the production of tumor necrosis factor (TNF) by macrophages⁶¹. PAF is a major inflammatory mediator which may interact with other mediators like IL-1, CSF and TNF, thereby resulting in an autocatalytic augmentation of the inflammatory response². Tri, being able to inhibit the production of these cytokines, may be of great importance in anti-inflammatory theraру٠

REFERENCES

- 65-68 Prepol B, Henanc S, Mencia-Huerta J-M, Rola Pleszczynski, M., Braquet P., Effect of platelet-activating factor (PAF-acether) and its specific receiping antagoms), BN 52021, on interleukin J (IL1) release and synthesis by rat spleen adherent monocytes. Prostaglandins 1987: 33 : 431-9.
- 2 Koltar M, Hosford D, Ginnot P, Esamu A, Braquet P. Platelet activating factor (PAF), a review of its effects, A antagonists and possible future clinical implications (part 1). Drugs 1991; 42 ; 9-29.
- Э Mencia-Huerta J-M. Benveniste J. Platelet-activating factor and macrophages. I. Evidence for the release from rat and mouse peritoneal macrophages and not from mastocytes.

Enr J Immunol 1979; 9: 409-53.

Zhang JP, Qian DH, Qi I.H. Effects of cantharidin on interleukin-2 and intertenkin-1 production in mice m vivo. Acta Pharmacol Sin 1992: 13 ; 263-4-

- 5. Wang HB, Zheng QY. Comparison of 4 methods for the assay of colony-stimulating factor. Chin J Haemstol 1991; 12: 323-4-
- lu DW, Zheng QY, Wang HB, Wang XF, Fong J. Efñ fect of WEB 2056 on the production of tumor recrosis factor from mirme peritoneal macrophages. Acta Pharm Sm. - in press.
- Wang XJ, Ding GF, Fan SG. The effects of different 7 opiate peptides and ACTH on the phagorytosis of mouse macrophages. Chin J Immonol 1987; 3: 211-3.
- Ward SG, Lewis GP, Westwick J. Differential effect of platelet-activating factor receptor antagonists on human T lymphocyte probleration-

Prostaglandins 1987; 34 : 149.

- 9 Dipersio JF, Billing P, Williams R, Gasson JC, Human granulocyte-macrophage colony-stimulating factor and other cytokines prime human neutrophils for enhanced arachidome and release and lookotnene B₄ synthesis. J Immunol 1988; 140 : 4315-22.
- (1) Engelherts I, von Asmuth EJU, van der Linden CJ, Baurman WA. The interrelation between TNF, IL-6, and PAF secretion induced by LPS in an in priviland in pitro. murane model.

Lymphokine Cytokine Res 1991; 10: 127-31. 10

Triazolodiazepine 在体外对小鼠脾细胞和腹腔 R965.2 巨噬细胞的抑制作用

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摘要 Triazolodiazepine (Tri)体外対Con A 和 LPS 透 导的小鼠脾细胞增殖以及巯基乙醇硫酸盐激击的腹腔 巨噬细胞吞噬功能呈浓度依赖性抑制作用。 Tri 还明 显抑制 Con A 诱导的脾细胞释放集落刺激因子和巨噬 细胞释放白细胞介素-1,结果提示,Tri抑制小鼠脾细 胞和巨唾细胞功能可能与其广泛而强大的抗炎作用有 关.

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关键词 triazolodiazepine; 白细胞介素-1; 集落刺激因 子;巨噬细胞;吞噬作用; <u>脾</u>脏