

the membrane, thus depolarizing the membrane. The decrease in MDP, in turn, reduced the rate and amplitude of 0 phase depolarization, thus decreasing V_{max} , APA, OS, TP. The increase of $[Ca^{2+}]_i$ inactivated the Ca^{2+} channel and activated the K^+ channel⁽⁷⁾, thus shortening the duration of the action potential. Cu and Se might restore the electrical parameters by way of scavenging the free radicals and protecting the cardiac cell membrane.

The results of this experiment demonstrate that Cu and Se played an antioxidative role in scavenging the free radicals. As a result, the heart cells were protected.

REFERENCES

- 1 Wang WJ. Superoxide free radical and superoxide dismutase. *Prog Physiol Sci* 1985; 16 : 196-202.
- 2 Flohe L, Günzer WA, Schock HH. Glutathione peroxidase: A selenoenzyme. *FEBS Lett* 1973; 32 : 132-4.
- 3 Jiang Y, Zhao P, Zhang WJ. Influence of Se on the survival of cultured heart cells. *Chin J Endemiol* 1986; 5 : 163-6.
- 4 Schanne OF, Ceretti E. *Impedance measurement in biological cells* 1 st ed. NY: John Wiley, 1978 : 202-12.

- 5 McCord JM. Oxygen derived free radicals in postischemic tissue injury. *N Engl J Med* 1985; 312 : 158-63.
- 6 Meerson FZ, Kagan VE, Kozlov YP, Belkina LM, Arkhipenko YV. The role of lipid peroxidation in pathogenesis of ischemic damage and the antioxidant protection of heart. *Basic Res Cardiol* 1982; 77 : 465-85.
- 7 Ruiz-Petrich E, de Lorenzi F, Chartier D. Role of the inward rectifier I_{K1} in the myocardial response to hypoxia. *Cardiovasc Res* 1991; 25 : 17-26.

(16)

136-139

铜硒对 X-XO 致损的培养心肌细胞电参数的影响

钟国精、江岩¹、岳刚、李云义、孙晓霞、张文杰 (白求恩医科大学生理教研室, 长春 130021, 中国) R963

提要 向培养基中加入黄嘌呤 0.42 mmol · L⁻¹ 与黄嘌呤氧化酶 5.3 nmol · L⁻¹ (X-XO), 心肌细胞的自由基含量增高; 显著降低动作电位参数, 输入阻抗由 0.34 ± 0.11 降到 0.24 ± 0.1 MΩ. 向培养基中加入 Cu 62.5 ng · ml⁻¹ 与 / 或 Se 173 ng · ml⁻¹, 心肌细胞的所有电参数恢复, 心肌细胞的自由基含量回升. 提示铜、硒都能清除自由基, 对抗 X-XO 所致的心肌细胞损伤.

关键词 培养的细胞; 心肌; 铜; 硒; 自由基; 动作电位; 黄嘌呤氧化酶

BIBLID: ISSN 0253-9756 中国药理学报 *Acta Pharmacologica Sinica* 1992 Mar; 13 (2) : 139-142

Effect of dimethyl sulfoxide on cytosolic calcium in cultured rat hepatocytes injured by D-galactosamine¹

LIU Hua-Ping, CONG Zheng² (Department of Pharmacology, Beijing Medical University, Beijing 100083, China)

ABSTRACT D-galactosamine (Gal 0.5 mmol · L⁻¹) made lactate dehydrogenase (LDH) leakage from the hepatocytes in monolayer-culture increase by

50%. Dimethyl sulfoxide (Me₂SO 2% vol / vol) decreased the LDH leakage (P < 0.05). The cytosolic free Ca²⁺ concentration ([Ca²⁺]_c) of rat hepatocytes exposed to Gal 4 mmol · L⁻¹ for 90 min in suspension culture increased about 2-fold (P < 0.01). Me₂SO (2%) antagonized this [Ca²⁺]_c-increasing effect of Gal. These results verified directly that the [Ca²⁺]_c of hepatocytes was increased in the early stage

Received 1990 Sep 11 Accepted 1991 Dec 26
¹Project supported by the National Natural Science Foundation of China, № 3860305
²To whom correspondence should be addressed

of Gal-induced hepatotoxicity, and suggested that the prevention or lightening of the disturbance in intracellular Ca^{2+} homeostasis may be, at least, one of the mechanisms of the protective action of Me_2SO against Gal-induced hepatocyte injury.

KEY WORDS liver; dimethyl sulfoxide; galactosamine; cultured cells; cytosol; calcium

Me_2SO , usually used in the treatment of dermatologic illness, was used in rheumatism as an analgesic agent during the past years⁽¹⁾. Me_2SO could prolong the survival time and improve the functional state of primary cultured rat hepatocytes^(2,12). It will be significant to study whether Me_2SO has protective effect against Gal-induced injury in primary cultured hepatocytes. There have been different and controversial reports about the changes in cytosolic Ca^{2+} concentration ($[\text{Ca}^{2+}]_c$) in early stage of Gal-induced hepatotoxicity *in vivo*^(3,4), but no direct evidence about the $[\text{Ca}^{2+}]_c$ change in Gal-intoxicated cultured hepatocytes has been seen. Me_2SO could affect the intracellular Ca^{2+} concentration of murine erythroleukemia cells to induce cell differentiation^(5,6). What is the $[\text{Ca}^{2+}]_c$ change in early stage of Gal-intoxicated hepatocytes? Could Me_2SO exhibit its hepato-protective action *via* an influence on the $[\text{Ca}^{2+}]_c$? In present paper, we studied these theoretical problems by direct determination of hepatocyte $[\text{Ca}^{2+}]_c$.

MATERIALS

Wistar rats δ $190 \pm s 10$ g. Collagenase (Type IV), HEPES, DME medium, Percoll (Colloidal PVP coated silica) and digitonin were purchased from Sigma. Me_2SO (AR) was made by Beijing Xudong Chemical Factory. Arsenazo III (AIII, lot No 85062) was made by Shanghai 3rd Reagent Factory. Aminophosphonate resin was a gift of Dr ZHANG Zheng-Pu, Nan-Kai University.

METHODS AND RESULTS

Effect of Me_2SO on Gal-induced cellular injury of rat hepatocytes in primary culture

Hepatocytes were isolated from adult rats by collagenase perfusion method (*Physiol Sci* 1987; 7 : 357), with the yield of $1.5 \times 10^8 - 2 \times 10^8$ cells/liver and viability $> 90\%$ (Trypan blue exclusion). Cells were cultured with DME medium as previously reported (*Physiol Sci* 1987; 7 : 357). The plates were divided into 6 groups, and their media were changed after 8-h culture with DME containing different concentrations of Me_2SO in the presence or absence of Gal $0.5 \text{ mmol} \cdot \text{L}^{-1}$. The lactate dehydrogenase (LDH) activities in medium were determined colorimetrically⁽⁷⁾ after a further 48-h culture. Me_2SO decreased the Gal-induced LDH leakage by about 50% and there was no difference in potency of this action in 0.5–2% Me_2SO (Tab 1). It meant that 0.5% Me_2SO is enough to cause its maximal protective effect, and that 2% Me_2SO is still in the plateau region. Since 2% was said to be the optimal concentration of Me_2SO for its differentiation-promoting action⁽²⁾, we used Me_2SO 2% in all experiments.

Tab 1. Effects of Me_2SO (vol/vol) on lactate dehydrogenase leakage from the Gal-injured monolayer-cultured rat hepatocytes. $n=6$ (tubes), $\bar{x} \pm s$, ** $P < 0.05$ vs Gal group.

Gal/ $\text{mmol} \cdot \text{L}^{-1}$	$\text{Me}_2\text{SO} / \%$	LDH / $\mu\text{mol} \cdot \text{L}^{-1}$	%
0.0	0.0	3.11 ± 0.33	100
0.5	0.0	4.84 ± 0.22	155
0.5	0.5	$3.93 \pm 0.21^{**}$	126
0.5	1.0	$3.78 \pm 0.36^{**}$	123
0.5	1.5	$3.92 \pm 0.26^{**}$	126
0.5	2.0	$3.97 \pm 0.14^{**}$	129

Influences of Gal and Me_2SO on hepatocyte $[\text{Ca}^{2+}]_c$

1 Methodology of null point titration

technique

1.1 Purification of AIII Commercially available AIII was purified by passing through 2 successive columns of aminophosphonate ion-exchange resin in Na⁺ form⁽⁸⁾. We found that the contaminative Ca²⁺ of commercially available AIII (2 nmol · L⁻¹ / mg AIII) used was lower than the value allowed (5 μmol · L⁻¹ / mg AIII).

1.2 Determination of parameters A series of parameters was determined by colorimetric assay⁽⁹⁾. The values were: ε₁ = 8 × 10² · mol⁻¹; ε₂ = 218 × 10² · mol⁻¹; K_d' = 10.75 μmol · L⁻¹, the AIII concentration used for all experiments was 53 μmol · L⁻¹. These data tallied with those in literature (ε₁ = 15 × 10² · mol⁻¹, ε₂ = 235 × 10² · mol⁻¹)⁽¹⁰⁾.

2 Effect of Gal and Me₂SO on [Ca²⁺]_c

After 10 min preincubation at 37°C, the suspension culture tubes were divided into 5 groups. The first group was used to determine [Ca²⁺]_c immediately as a normal value. The other groups were incubated at 37°C for 90 min in the presence or absence of Gal 4 mmol · L⁻¹, with or without Me₂SO 2% vol / vol, respectively. The hepatocytes were separated from the Ca²⁺-containing DME medium and the nonviable cells were removed by low-speed isodensity Percoll centrifugation at 4°C, 50 × g for 10 min⁽¹¹⁾, then hepatocyte [Ca²⁺]_c was determined by null point titration⁽¹⁰⁾. The results showed that in freshly isolated hepatocytes, [Ca²⁺]_c was 0.19 ± 0.07 μmol · L⁻¹, which tallied with that in literature (0.19 ± 0.01 μmol · L⁻¹)⁽¹⁰⁾. Gal 4 mmol · L⁻¹ increased [Ca²⁺]_c about 3-fold as much as the control. Me₂SO itself did not decrease the [Ca²⁺]_c, but prevented the Gal-induced [Ca²⁺]_c increase (Tab 2).

DISCUSSION

In this paper we found that Me₂SO could protect hepatocytes against Gal-induced cytotoxicity (Tab 1). Our previous work

Tab 2. Effects of Me₂SO (2% vol / vol) and Gal (4 mmol · L⁻¹) on cytosolic Ca²⁺ concentration (μmol · L⁻¹) of hepatocytes in suspension culture (90 min). n=8, $\bar{x} \pm s$, **P<0.05, *P<0.01 vs control group. +++P<0.01 vs Gal group. Δ: freshly isolated hepatocytes. §: calculated with the difference in [Ca²⁺]_c between Gal and control as 100%.**

	[Ca ²⁺] _c	Δ[Ca ²⁺] _c % §
FIHΔ	0.19 ± 0.07	
Control	0.26 ± 0.03 ⁺⁺	
Gal	0.75 ± 0.11 ^{***}	100
Me ₂ SO	0.36 ± 0.08 ^{***}	20
Me ₂ SO+Gal	0.40 ± 0.10 ^{***}	30

showed that Me₂SO could prolong the survival time of cultured hepatocytes and improve their ability of maintaining the content of glycogen and cytochrome P-450 during long term (96 h) culture⁽¹²⁾. We concluded that Me₂SO is of benefit to normal hepatocytes and can protect them against Gal-injury.

There have been two different suggestions about the Ca²⁺ changes in hepatocytes in early stage of Gal-induced injury. Some authors thought that first thing in Gal-induced injury is membrane damage, which results in an influx of Ca²⁺. Others considered that redistribution of Ca²⁺ within cells first occurred which made [Ca²⁺]_c increase. They interpreted indirectly that [Ca²⁺]_c increase is one of the causes leading to membrane damage^(4,13). But no direct evidence about the [Ca²⁺]_c change in Gal-injured hepatocytes is seen before.

We observed the change in hepatocyte [Ca²⁺]_c by null point titration technique. In the work of methodology, we verified that null point titration is an economical but reliable method. Our data showed that in the early stage of Gal-induced injury (90 min), the hepatocyte [Ca²⁺]_c did increase about 3-fold. This is a first evidence of the mechanisms of Gal-induced hepatotoxicity in cell level. Because the viability of hepatocytes, observed by both trypan blue exclusion and medium

LDH activity assay, was normal in the early stage of Gal-injury (90 min), we considered that cell membrane damage might be unserious. So, the $[Ca^{2+}]_c$ increase seemed to be the result of Ca redistribution within cells. Recently, it is reported⁽¹⁴⁾ that lipoperoxidation in liver cell membrane increased as early as 30 min after Gal-intoxication *in vivo*, and total liver calcium increased 3 h after Gal administration. So, we think, the dynamic study of $[Ca^{2+}]_c$ change in whole course of Gal-intoxication is needed to clarify the details.

The hepatoprotective action of Me_2SO might be related to its differentiation-promoting action⁽²⁾. Our data in this work showed, Me_2SO itself did not decrease hepatocyte $[Ca^{2+}]_c$, but could prevent or lighten Gal-induced $[Ca^{2+}]_c$ increase (Tab 2). This may be one of mechanisms of the hepato-protective effect of Me_2SO against Gal-induced injury.

REFERENCES

- 1 Jimenez RAH, Willkens RF. Dimethyl sulfoxide: a perspective of its use in rheumatic diseases. *J Lab Clin Med* 1982; **100**: 489-500.
- 2 Isom HC, Secott T, Georgoff I, Woodworth C, Mummaw J. Maintenance of differentiated rat hepatocytes in primary culture. *Proc Natl Acad Sci USA* 1985; **82**: 3252-6.
- 3 Schanne FAX, Pfau RG, Farber JL. Galactosamine-induced cell death in primary cultures of rat hepatocytes. *Am J Pathol* 1980; **100**: 25-38.
- 4 Tran-Thi T-A, Phillips J, Falk H, Decker k. Toxicity of D-galactosamine for rat hepatocytes in monolayer culture. *Exp Mol Pathol* 1985; **42**: 89-116.
- 5 Bridges K, Levenson R, Housman D, Cantley L. Calcium regulates the commitment of murine erythroleukemia cells to terminal erythroid differentiation. *J Cell Biol* 1981; **90**: 542-4.
- 6 Chapman LF. Effect of calcium on differentiation of Friend leukemia cells. *Dev Biol* 1980; **79**: 243-6.
- 7 Wroblewski F, LaDue JS. Lactic dehydrogenase activity in blood. *Proc Soc Exp Bio Med* 1955; **90**: 210-3.
- 8 Kendrick NC. Purification of Arsenazo III, a Ca^{2+} -sensitive dye. *Anal Biochem* 1976; **76**: 487-501.
- 9 Kendrick NC, Ratzlaff RW, Blaustein MP. Arsenazo III as an indicator for ionized calcium in physiological salt solutions: its use for determination of the CaATP dissociation constant. *Anal Biochem* 1977; **83**: 433-50.
- 10 Murphy E, Coll K, Rich TL, Williamson JR. Hormonal effects on calcium homeostasis in isolated hepatocytes. *J Biol Chem* 1980; **255**: 6600-8.
- 11 Kreamer BL, Staecker JL, Sawada N, Sattler GL, Hsia MTS, Pitot HC. Use of a low-speed, iso-density percoll centrifugation method to increase the viability of isolated rat hepatocyte preparations. *In Vitro* 1986; **22**: 201-11.
- 12 Liu HP, Cong ZH. Influence of collagen and DMSO on the survival and functional state of primary cultured rat hepatocytes. *Chin J Pharmacol Toxicol* 1991; **5**: 195-9.
- 13 Lamp RG, Schwertz DW, McCue SB, Tayler D, McGuffin M. Phospholipase C activation by hepatotoxins: a new mechanism of hepatocyte injury. In: Keppler D, Popper H, Bianchi L, Reutter W, editors. *Mechanisms of hepatocyte injury and death*. Lancaster: MTP Press, 1984: 95-106.
- 14 Mourelle M, Meza MA. Colchicine prevents D-galactosamine-induced hepatitis. *J Hepatol* 1989; **8**: 165-72.

139-142

二甲亚砜对 D-半乳糖胺损伤的培养大鼠肝细胞胞液自由钙离子的影响

刘华屏、丛 铮 (北京医科大学药理教研室, 北京 100083, 中国)

12962

提要 D-半乳糖胺(Gal, $0.5\text{mmol} \cdot \text{L}^{-1}$)使培养肝细胞的乳酸脱氢酶外漏增加 55%, 二甲亚砜(Me_2SO)可使此损伤减轻约二分之一($P < 0.05$)。肝细胞接触 Gal $4\text{mmol} \cdot \text{L}^{-1}$ 90 min, 其胞液内自由钙离子浓度($[Ca^{2+}]_c$)增加到对照组的 3 倍($P < 0.01$)。 Me_2SO (2% vol/vol)可拮抗 Gal 引起的 $[Ca^{2+}]_c$ 增加。提示: Gal 损肝早期 $[Ca^{2+}]_c$ 升高, 而 Me_2SO 阻止细胞内钙平衡的紊乱, 可能是其抗 Gal 损伤的机理之一。

关键词 肝脏; 二甲亚砜; D-半乳糖胺; 培养细胞; 细胞溶质; 钙