

## Monoclonal antibodies specific for ohmefentanyl<sup>1</sup>

LU Yi-Feng, TONG Chun-Xiang<sup>2</sup>, WEI Ai-Li, LIN Sheng-Yin<sup>2</sup>, CHEN Xin-Jian, ZHOU De-He, CHI Zhi-Qiang

(Shanghai Joint Laboratory of Life Sciences, Shanghai Institute of Materia Medica, <sup>2</sup> Shanghai Institute of Cell Biology, Chinese Academy of Sciences, Shanghai 200031, China)

**ABSTRACT** Ohmefentanyl (Ohm, *N*[1-( $\beta$ -hydroxy- $\beta$ -phenylethyl)-3-methyl-4-piperidyl]-*N*-phenylpro-pronamide), designed and synthesized by our laboratory, is a highly selective  $\mu$  receptor agonist. After somatic cell fusion between splenocytes of BALB c mice, immunized by Ohm-BSA conjugate and NS-1 myeloma cells, 2 lines of hybridoma (D<sub>2</sub> and F<sub>4</sub>) secreting monoclonal anti-Ohm antibodies (MAb) were obtained. Saturation and competition experiments showed that MAb-D<sub>2</sub> and MAb-F<sub>4</sub> bound to Ohm-BSA with high affinity and high specificity. Using radioligand binding assay and bioassay, we also found that MAb-D<sub>2</sub> and MAb-F<sub>4</sub> inhibited [<sup>3</sup>H] Ohm binding to rat brain opioid receptors in a dose-dependent manner and antagonized the effect of Ohm on the contraction of guinea pig ileum induced by electric field stimulation. These results suggested that MAb-D<sub>2</sub> and MAb-F<sub>4</sub> were 2 monoclonal antibodies specific for Ohm and could be useful as functional antagonists of Ohm.

**KEY WORDS** ohmefentanyl; mu receptors; monoclonal antibodies

Ohmefentanyl (Ohm, *N*[1-( $\beta$ -hydroxy- $\beta$ -phenylethyl)-3-methyl-4-piperidyl]-*N*-phenylpro-pronamide), designed and synthesized by our laboratory, is a highly selective  $\mu$  receptor agonist<sup>1,2</sup>. At present, we wish to produce monoclonal antibodies oriented to

Ohm, as a possible immunogen for the production of anti-Ohm anti-idiotypic antibodies which may carry an internal image of Ohm and may be useful in the isolation and identification of  $\mu$  receptor binding protein.

### MATERIALS AND METHODS

**1 Animals and drugs** BALB c ♀ mice weighing 20 ± 2 g (Shanghai Institute of Cell Biology, Chinese Academy of Sciences). Sprague-Dawley rats ♂ weighing 250 ± 30 g and guinea pigs ♂ weighing 380 ± 50 g (Shanghai Institute of Materia Medica, Chinese Academy of Sciences). The myeloma cell line NS-1 (Cell Banks of Shanghai, Chinese Academy of Sciences).

Ohm,  $\beta$ -hydroxylfentanyl, 3-methylfentanyl, fentanyl, [*D*-Ala<sup>2</sup>, *D*-Leu<sup>5</sup>]enkephalin (DADLE), and etorphine (Shanghai Institute of Materia Medica). Morphine (Qinhai Pharmaceutical Factory). Naloxone (Shanghai Medical University). Dihydroetorphine (Chinese Academy of Military Medical Sciences). [*D*-Ala<sup>2</sup>, Mephe<sup>3</sup>, Glyol<sup>5</sup>]enkephalin (DAGO), [*D*-Pen<sup>2</sup>, *D*-Pen<sup>5</sup>]enkephalin (DPDPE), and *N*-methyl-*N*-[7-(1-pyrrodidinyl)-1-oxaspiro(4,5)dec-8-yl]benzeneacetamide (U-69593) (Sigma Co.). [<sup>3</sup>H]Ohm (2.02 PBq · mol<sup>-1</sup>) (Shanghai Institute of Nuclear Research, Chinese Academy of Sciences).

### 2 Production of monoclonal anti-Ohm antibodies (MAb)

**Immunization** Ohm-6-hemisuccinate was prepared and conjugated to bovine serum albumin (BSA) (molar ratio, 47:1). BALB c mice were injected intrasplenically (20  $\mu$ g of Ohm-BSA) and ip (100  $\mu$ g of Ohm-BSA) alternatively at 3-4 wk intervals. Sera were tested periodically for anti-Ohm antibodies by radioimmunoprecipitation test and enzyme-linked immunosorbent assay (ELISA). By the end of immunization period, 100  $\mu$ g of the Ohm-BSA was injected

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both ip and iv. After 3 d the mouse splenocytes were harvested for cell fusion.

**Fusion** About  $1 \times 10^6$  splenocytes were fused with  $5 \times 10^7$  myeloma cells in the presence of 0.7 ml 40 % polyethylene glycol 4000, washed in Dulbecco's modified Eagle medium (DMEM), and resuspended in DMEM-fetal calf serum. On d 1, 3, 6, and 10, a fresh medium containing hypoxanthine, aminopterin, and thymidine (HAT) was added. After d 15, the culture supernatants were tested for anti-Ohm antibodies by ELISA.

**Subcloning of positive hybridoma** Hybridoma cell populations secreting anti-Ohm antibodies were subcloned for 4–5 times to obtain the monoclonal cell line by limiting the dilution on feeder cell layers in microtiter polystyrene plates.

**Amplification in the ascites** Monoclonal hybridoma cell lines were incubated in Petri dishes and  $2 \times 10^6$  cells of each line were injected ip into mice previously primed with 0.5 ml of pristane. Ascitic fluid was centrifuged at  $110 \times g$  to remove cells.

**3 Purification of MAb** The anti-Ohm MAb were purified from ascitic fluids by 2-step precipitation using 50 % and 33 % saturation of  $(\text{NH}_4)_2\text{SO}_4$ , respectively or by protein A — Sepharose column chromatography. MAb were assessed by SDS-PAGE and immunoprecipitation to identify the purity and Ig subgroups.

**4 Radioimmunoprecipitation test**  $[^3\text{H}]$  Ohm ( $0.1 \text{ ml}$  of  $2 \text{ nmol} \cdot \text{L}^{-1}$ ), serial dilutions of antiserum ( $0.05 \text{ ml}$ ), and Tris-HCl buffer ( $0.35 \text{ ml}$  of  $50 \text{ mmol} \cdot \text{L}^{-1}$ ) were mixed and incubated at  $4 \text{ }^\circ\text{C}$  for 16 h. Rabbit anti-mouse IgG antibodies ( $0.03 \text{ ml}$ ) and 1 % normal rabbit serum ( $0.03 \text{ ml}$ ) were added at  $4 \text{ }^\circ\text{C}$  for 4 h.  $[^3\text{H}]$  Ohm-antibody compounds were isolated by centrifugation and counted by a liquid scintillation counter (YSJ-80, Shanghai Institute of Nuclear Research).

**5 ELISA** Ohm-BSA ( $0.1 \text{ ml}$  of  $50 \mu\text{g} \cdot \text{ml}^{-1}$ ) was coated on immunoplates. After washing and blocking with PBS containing 10 % bovine serum,  $0.1 \text{ ml}$  of anti-Ohm MAb at different concentrations were added and incubated 2 h at  $37 \text{ }^\circ\text{C}$ . After washing,  $0.1 \text{ ml}$  of sheep anti-mouse IgG antibodies coupled with peroxidase were mixed for 2 h further. Peroxidase activity was revealed by 0.04 %  $\text{H}_2\text{O}_2$ -OPD substrate.  $A_{490}$  was measured by an automatic microplate reader

(BIO-TEK EL311).

**6 Competitive ELISA** Serial dilutions of opioid ligands ( $0.1 \text{ ml}$ ) and anti-Ohm MAb ( $0.1 \text{ ml}$ ) were mixed before adding to the well-plates previously coated with Ohm-BSA  $50 \mu\text{g} \cdot \text{ml}^{-1}$ , and then incubated for 2 h at  $37 \text{ }^\circ\text{C}$ . Other conditions were specified for the normal ELISA.

**7 Radioligand binding assay** Rat brain membranes were prepared<sup>(17)</sup>.  $[^3\text{H}]$  Ohm ( $0.1 \text{ ml}$  of  $2 \text{ nmol} \cdot \text{L}^{-1}$ ), rat brain membranes ( $0.3 \text{ ml}$ ,  $1 \text{ mg}$  protein), and increasing concentrations of anti-Ohm MAb ( $0.1 \text{ ml}$ ) were mixed at  $30 \text{ }^\circ\text{C}$  for 45 min. Nonspecific binding was measured in the presence of  $0.1 \text{ ml}$  of unlabeled Ohm  $1 \mu\text{mol} \cdot \text{L}^{-1}$ . Binding was terminated by rapid filtration and quantitated using a liquid scintillation counter.

**8 Bioassay** The segment of guinea pig ileum was prepared<sup>(23)</sup>. The contraction of ileum was induced by electric field stimulation ( $40 \text{ V}$ ,  $1 \text{ ms}$ ,  $15 \text{ s}$  intervals). Electric field stimulating contraction (ESC) under such conditions could be inhibited by Ohm, and the effects of anti-Ohm MAb on inhibition of Ohm were observed.

## RESULTS

**1 Detection of anti-Ohm antisera** After repeatedly immunizing the BALB c mice with Ohm-BSA for more than 10 times, the titers of anti-Ohm antisera lay between 1:1600 and 1:3200 at 30 % of the ratio of  $[^3\text{H}]$  Ohm-antibody to  $[^3\text{H}]$  Ohm (Fig 1).

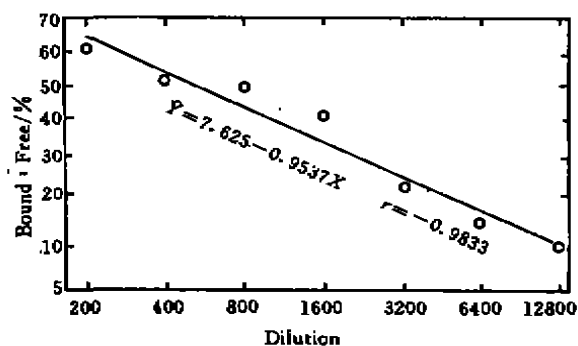


Fig 1. Titers of mouse anti-Ohm antiserum detected by radio-immunoprecipitation test. Each point is mean of three independent experiments.

Preincubated the antisera with 10% bovine serum at 37°C for 2 h, the titers of anti-Ohm antisera by ELISA was assayed. The titers lay between 1:2000 and 1:4000 while  $A_{50}$  was 1.00, which was identical with the titers detected by radioimmuno-precipitation test (Fig 2).

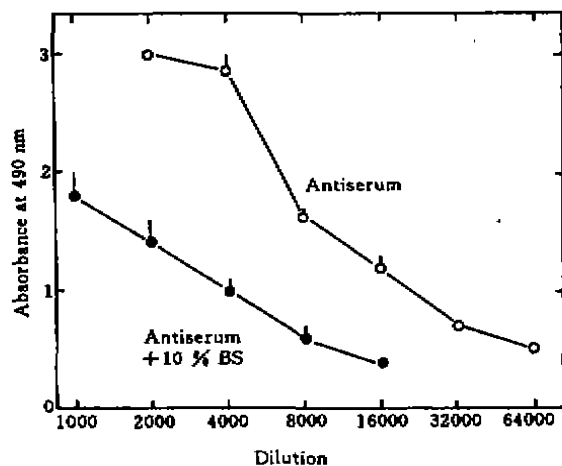


Fig 2. Titers of mouse anti-Ohm antiserum detected by ELISA.  $n=4$ ,  $\bar{x} \pm s$ .

### 2 Identification of immunoglobulins

After cell fusion, 2 lines of hybridomas ( $D_2$  and  $F_1$ ) secreting monoclonal anti-Ohm antibodies were obtained. Immunoprecipitation study showed that the purified immunoglobulins Ig of MAb- $D_2$  and MAb- $F_1$  were all of the IgG class. Further analysis showed that the subclasses of MAb- $D_2$  and MAb- $F_1$  belonged to IgG<sub>1</sub>.

Electrophoresis was performed using SDS-15% PAGE in the presence of 1% mercaptoethanol. The 2 bands with molecular weights of 56 300 and 24 200 were shown to be of heavy and light chain, respectively (Fig 3).

**3 Binding of MAb to antigen** Anti-Ohm MAb bound to Ohm-BSA in a saturation and dose-dependent manner. The  $A_{50}$  of

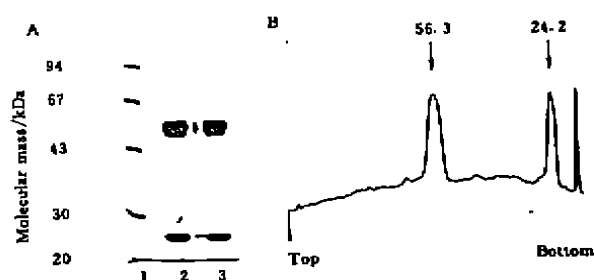


Fig 3. A. SDS-PAGE of monoclonal anti-Ohm antibodies. 1) Markers of indicated molecular mass. 2) MAb- $D_2$  and 3) MAb- $F_1$  purified by 2-step precipitation using 50% and 33%  $(NH_4)_2SO_4$  saturation. B. Scanning graph of lane 2.

MAb- $D_2$  and MAb- $F_1$  were  $1.1 \pm 0.3$  and  $1.3 \pm 0.7$  nmol  $\cdot$  L<sup>-1</sup> ( $n=5$ ,  $\bar{x} \pm s$ ), respectively. In the same experiments, MAb did not bind to BSA and BSA nor inhibited the binding of MAb to Ohm-BSA (Fig 4).

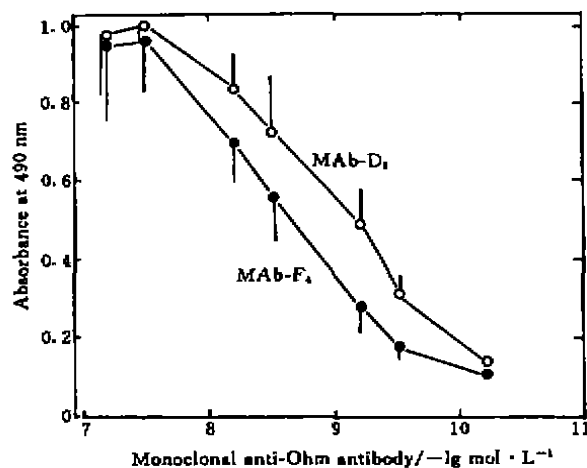


Fig 4. Saturation of MAb binding to Ohm-BSA.  $n=5$ ,  $\bar{x} \pm s$ .

**4 Cross reactivity** The cross reactivity of anti-Ohm MAb with a number of selected opioid agonists and antagonists were studied using competitive ELISA. Results showed that Ohm and its analogs inhibited the binding of MAb to Ohm-BSA in a dose-dependent manner, but DAGO, morphine, etorphine, DPDPE, DADLE, and U-69593 showed no

effect. Although dihydroetorphine and naloxone slightly inhibited the binding of MAb to Ohm-BSA in concentrations of  $>10 \mu\text{mol} \cdot \text{L}^{-1}$ , the rates of cross reactivity were  $<1.0 \%$  (Tab 1).

Tab 1. Cross reactivity of MAb-D<sub>2</sub> and MAb-F<sub>4</sub> with opioid ligands.  $n=3-6, \bar{x} \pm s$ .

Ligands	IC <sub>50</sub> ( $\mu\text{mol} \cdot \text{L}^{-1}$ )		Rate of cross reactivity (%)	
	D <sub>2</sub>	F <sub>4</sub>	D <sub>2</sub>	F <sub>4</sub>
Ohmefentanyl	0.8±0.5	0.5±0.3	100	100
β-Hydroxyfentanyl	14±4	12±6	5.7	4.2
3-Methylfentanyl	21±3	18±13	3.7	2.7
Fentanyl	53±11	64±30	1.5	<1.0
Dihydroetorphine	>100	>100	<1.0	<1.0
Naloxone	>100	>100	<1.0	<1.0

**5 Radioligand binding assay** Anti-Ohm MAB inhibited the specific binding of [<sup>3</sup>H]Ohm to rat brain opioid receptors in a dose-dependent manner. IC<sub>50</sub> values of MAb-D<sub>2</sub> and MAb-F<sub>4</sub> were  $0.18 \pm 0.09$  and  $0.45 \pm 0.10 \mu\text{mol} \cdot \text{L}^{-1}$  ( $n=5, \bar{x} \pm s$ ), respectively (Fig 5).

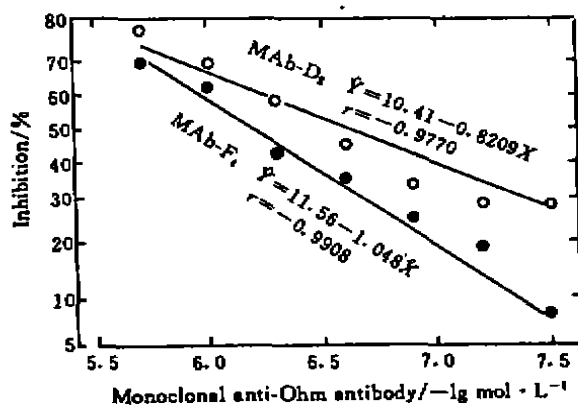


Fig 5. Inhibition of [<sup>3</sup>H]Ohm binding to rat brain opioid receptors by MAb. Each point is mean of five independent experiments.

**6 Bioassay** Although anti-Ohm MAB did not influence the ESC of guinea pig ileum

directly, it antagonized the inhibition of Ohm on the ESC of guinea pig ileum. MAb-D<sub>2</sub> and MAb-F<sub>4</sub> ( $1 \mu\text{mol} \cdot \text{L}^{-1}$  each) respectively antagonized  $81 \pm 9$  and  $61 \pm 13 \%$  ( $n=4, \bar{x} \pm s$ ) of Ohm ( $0.25 \text{ nmol} \cdot \text{L}^{-1}$ ) inhibition on the ESC (Fig 6).

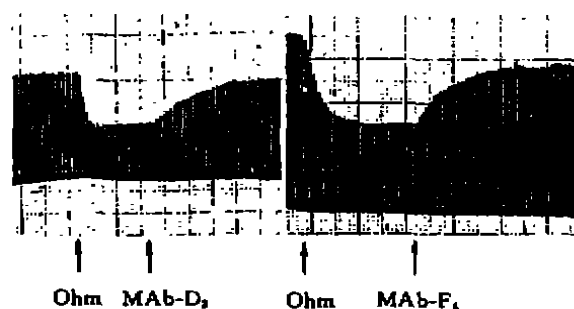


Fig 6. MAb ( $1 \mu\text{mol} \cdot \text{L}^{-1}$ ) antagonized the effect of Ohm ( $0.25 \text{ nmol} \cdot \text{L}^{-1}$ ) on electric field stimulating contraction of guinea pig ileum.

**DISCUSSION**

Because of its size, Ohm may not be immunogenic on itself. It should be coupled to a carrier protein before immunization. In our study, Ohm-6-hemisuccinate was prepared and was conjugated to BSA. During immunization with Ohm-BSA, however, antibodies to BSA will also be produced. By simple reasoning, 2 antigens with different carriers should be used, one for immunization, the other for screening of hybridomas secreting anti-hapten antibodies. Ji and Xu reported that using one antigen, they also detected the anti-hapten antibodies in screening of lymphocyte hybridomas<sup>4</sup>. According to their method modified by us, we also detected anti-Ohm antibodies in screening of the hybridomas by blocking the anti-BSA antibodies with 10% bovine serum by ELISA.

In preliminary studies, the titers of anti-Ohm antisera of mice, rat, and rabbit immunized with conventional methods were very low ( $<1:100$ ). Opioid agonists (eg Ohm)

have immunosuppressive effect<sup>15,6</sup>. Hence, we chose intrasplenic injection for immunization, an effective method for low immunocompetent antigen<sup>17</sup>, and finally a high-titer anti-Ohm antiserum in BALB/c mice was obtained.

Saturation and competition experiments showed that MAb-D<sub>2</sub> and MAb-F<sub>1</sub> were 2 high affinity and highly specific monoclonal antibodies against Ohm. The relative potencies of Ohm analogs inhibiting the binding of MAb to Ohm-BSA were in the order of Ohm > β-hydroxylfentanyl > 3-methylfentanyl > fentanyl. This closely paralleled that their binding capabilities to the opioid receptors<sup>2</sup>.

MAb<sub>1</sub> interacted with Ohm, inhibited the specific binding of [<sup>3</sup>H]Ohm to rat brain opioid receptors and antagonized the effect of Ohm on guinea pig ileum. Ohm was a highly selective ligand of μ receptors. Anti-Ohm MAb<sub>1</sub> functioned as antagonists, may be useful in studying the effect and mechanism of Ohm on the μ receptors.

Anti-Ohm MAB were also used in the anti-Ohm anti-idiotypic antibodies (anti-Id) inducing. According to the network theory of the immune system<sup>9</sup>, among the anti-Ohm anti-Id certain anti-Id bearing the "internal image" of Ohm would be identified. These anti-Id antibodies have bright prospect in the research of opioid receptors such as in the isolation of μ receptor binding protein, screening of cDNA clone encoding μ receptors, and so on.

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**抗羟甲芬太尼单克隆抗体**

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陆亦凤, 童春香, 魏爱丽, 林盛荫<sup>1</sup>, 陈新建, 周德和, 池志强 (上海生命科学联合开放实验室, 中国科学院上海药物研究所, 中国科学院上海细胞生物学研究所, 上海 200031, 中国)

**A摘要** 采用细胞融合和单克隆技术, 获得抗羟甲芬太尼(ohmefentanyl, Ohm)单克隆抗体 MAb-D<sub>2</sub>和 MAb-F<sub>1</sub>。MAb-D<sub>2</sub>和 MAb-F<sub>1</sub>与 Ohm-BSA 结合亲和力高、专一性强。MAb-D<sub>2</sub>和 MAb-F<sub>1</sub>还能抑制 [<sup>3</sup>H]Ohm 与大鼠脑匀浆膜蛋白上的阿片受体的结合, 并能对抗 Ohm 抑制电场刺激引起豚鼠回肠收缩的作用。以上结果表明 MAb-D<sub>2</sub>和 MAb-F<sub>1</sub> 是特异性的抗 Ohm 单克隆抗体, 作为 Ohm 的拮抗剂具有广泛用途。

**关键词** 羟甲芬太尼; μ受体; 单克隆抗体