Inhibitory effects of ONO-3708 and S-145 on shape change and aggregation of rabbit platelets induced by STA₂

LI Bai-Yan, QIAO Guo-Fen, SUN Jian-Ping, GAO Yun-Rui, LI Wen-Han (Department of Pharmacology, Harbin Medical University, Harbin 150086, China)

KEY WORDS blood platelets; platelet aggregation; platelet aggregation inhibitors; thromboxane A_2 ; calcium; STA₂; ONO-3708; S-145

AIM: To study the mode of inhibition of ONO-3708 and S-145, 2 antagonists of thromboxane A₂ (TXA₂) receptors, against the rabbit platelet shape change and aggregation induced by stable analogue of TXA_2 (STA₂). METHODS: The platelet shape change and aggregation were quantified by the light transmission through platelet-rich plasma (PRP) and the intracellular calcium concentration ($[Ca^{2+}]_i$) was measured by fluorescence and imaging. RESULTS: (1) In PRP, STA₂(3 μ mol·L⁻¹)-induced aggregation was inhibited by egtazic acid 3 mmol·L⁻¹, ONO-3708 300 μ mol·L⁻¹, and S-145 1 μ mol·L⁻¹(P < 0.01), but not by indometacin (Ind) 3 μ mol·L⁻¹. The shape change induced by STA₂ was inhibited only by S-145 in a concentration-dependent manner. S-145 1 and 3 μ mol \cdot L⁻¹ were required to inhibit the shape change and aggregation. (2) The inhibitory effect of S-145, but not ONO-3708, was along with the prolongation of increased preincubation. (3) ONO-3708 lost the Inhibitory effect on STA2~induced aggregation after washing, while the inhibitory effect of S-145 was enhanced by prolongation of preincubation and remained after washing. (4) STA₂ 3 µmol · L⁻¹-induced [Ca²⁺], mobilization was unaffected by Ind, partially reduced by ONO-3708 and egtazic acid 3 mmol·L⁻¹(P < 0.01), but completely inhibited by S-145 (P < 0.01). CONCLUSION: S-145 and ONO-3708 were bound to a different site of the TXA₂ receptor.

Platelet activation by thromboxane $A_2(TXA_2)$ results in a shape change in platelets or centralization of granules, the exposure of adhesive

Received 1995-06-20 Accepted 1996-03-15

molecules, such as $GP \coprod b/\coprod a$, responsible for platelet aggregation and finally the release of dense granules and further aggregation. These sequential responses coincide with the activation – of phospholipase C and the release of 1, 4, 5-inositol triphosphate (IP_3) and diaceglycerol (DG)⁽¹⁾ and are mediated by subsequent increase in cytosolic calcium and protein kinase C activation. The number of TXA₂ receptors in human was 1500 -2500 sites per platelet^(2,3). In addition, the TXA_2 /prostaglandin (PG) endoperoxide (PGH₂) receptor was purified by affinity chromatography using TXA₂ antagonist (\pm) -5 | Z-7-[3-indophenylsulfonylamino(2.2.1) bicyclohept-2-exo-yl]heplanoic acid, sodium salt $(S-145)^{(4)}$ and the presence of 2 TXA₂/PGH₂ receptors was reports in terms of reversible and irreversible binding sites of TXA_2 antagonists⁽⁵⁾. The present study was performed to investigate the possibility that 2 types of TXA₂ inhibitors may bind to the receptors in a different mode in terms of platelet shape change and aggregation.

MATERIALS AND METHODS

Agents S-145 was gifted by Shionogi Pharmaceutical Co Ltd, Osaka. ONO-3708 17-[2a, 4a-(methylmethano)-6β- $(2-cyclopentyl-2\beta-hydroxyactami-no)-1\alpha-cyclohexyl]-5 (Z)$ heptanoic acid, sodium salt (ONO Pharmaceutical Co, Osaka)| was dissolved in buffered saline. Indometacin (Ind. Merck, Sharp & Dohme Research Lab, Rahway NJ) was dissolved in absolute ethanol (10 $g \cdot L^{-1}$) and kept at 4 °C, and diluted by buffered saline before use. STA2 (9, 11epithio-11, 12-methano-TXA2, a stable analogue of TXA2 kindly provided by ONO Pharmaceutical Co, Osaka) was dissolved in ethanol and stored at - 20 $^{\circ}\mathrm{C}$. Egtazic acid (Sigma Co, St Louis) was dissolved in distilled water with NaOH 1 mol \cdot L⁻¹ and stored at room temperature after the pH was adjusted to 7.4 with HCl 1 mol· L^{-1} . Fura-2 and Fura 2-AM (Wako Pure Chemicals, Osaka) were used for the Ca²⁺ calibration curve and platelet loading.

Preparation of washed platelets^[6]

Preincubation of platelets with TXA₂ antagonists The

washed platelets $(4 \times 10^{11} \cdot L^{-1})$ were preincubated with S-145 (10 and 30 nmol·L⁻¹), ONO-3708 (3 and 10 μ mol·L⁻¹), or buffer at 37 °C for 2. 5, or 10 min. The washed platelets were stimulated by STA₂ 3 μ mol·L⁻¹ at 4 min after the addition of CaCl₂ 1 mmol·L⁻¹(Fig 1A).



Fig 1. Aggregatory inhibition by preincubation with ONO-3708 and S-145 (A) and effects of washing (B). n = 11 - 15 samples from at least 6 rabbits, $\vec{x} \pm s$. ^aP > 0.05, ^cP < 0.01, ^gP > 0.05 and ⁱP < 0.01 vs S-145/C, ^dP > 0.05, ^cP < 0.05 vs S-145/W. C: control (without

washing); W: the addition of STA₂ after washing.

Washing experiments After the washed platelets were preincubated with S-145 10 or 30 nmol·L⁻¹ plus ONO-3708 10 μ mol·L⁻¹ for 2 min, they were centrifuged at 700 × g for 20 min. The platelet pellets were suspended in HEPESbuffer and the final platelet counts were adjusted to 4 × 10¹¹ ·L⁻¹. The rewashed platelets were aggregated by STA₂ 3 μ mol·L⁻¹ at 4 min after the addition of CaCl₂ 1 mmol·L⁻³. The control was run without washing.

Piatelet shape change and aggregation^(6,7)

Fura-2 loading and [Ca²⁺]₁ measurement^[6,8]

Data analysis Statistical analysis was performed by either paired or unpaired t-test.

RESULTS

Inhibitory effects of ONO-3708 and S-145 on STA₂-induced shape change and aggregation in PRP STA₂-induced aggregation was completely inhibited by ONO-3708 and S-145, and also inhibited by egtazic acid 3 mmol·L⁻¹, but not by Ind 3 μ mol·L⁻¹. While the shape change was inhibited only by S-145. S-145 1 and 3 μ mol·L⁻¹ were required respectively to completely inhibit platelet shape change and aggregation (Tab 1).

Tab 1.	 Effects of 	indometaci	in (Ind),	egtazic acid,	S-145
and O	NO-3708 on	STA ₂ (3	µmool•L	-1)-induced	platelet
shape	change and s	ggregation	in rabbit)	PRP.	
n = 7 -	- 11 samples	from 6 rai	bbits, $\mathbf{x} \pm \mathbf{x}$	s.	
*P>0	.05, °P<0	01 vs STA	2 -		

Groups	Shape change, %	Aggregation. %
STA2	-8.4 ± 08	61 ± 6
Ind 3 μ mol·L ⁻¹	-8.3 ± 0.9^{4}	65 ± 7*
Egtazic acid 3 mmol·L ⁻¹	$=10.4\pm0.9^{\circ}$	1.1 ± 0.8^{c}
S-145 1 μ mol·L ⁻¹	$-5.7\pm0.8^{\circ}$	1.9±1.4°
$3 \ \mu mol \cdot L^{-1}$	$-1.0 \pm 0.4^{\circ}$	0.0 ± 0.0
ONO-3708 100 μ mol·L ⁻¹	-8.1 ± 0.9	22 ± 4°
300 µmol•L ⁻¹	-9.8 ± 0.9	0.0 ± 0.0

The dose-response curve of $STA_2(3 \ \mu mol \cdot L^{-1})$ induced aggregation was diminished by ONO-3708 and the maximal response was not affected, in case of S-145, the dose-response was diminished, but the maximal response was reduced markedly (data not shown).

Effect of washing and preincubation on STA_2 induced aggregation The inhibition of S-145 on STA_2 -caused aggregation was increased (P < 0.01) along with the prolongation of preincubation time. In contrast, STA_2 -induced aggregation was not affected by ONO-3708, even preincubating platelets with ONO-3708 for 10 min (Fig 1A).

S-145 concentration-dependently inhibited STA_{2} induced aggregation. This inhibitory effect was enhanced by the administration of ONO-3708 together with S-145. The effect of S-145 remained after washing, but ONO-3708 lost its inhibitory effect (Fig 1B).

Intracellular calcium mobilization The cytosolic-Ca²⁺ concentration $[Ca^{2+}]_i$ was measured in single platelets. The STA₂ 1 and 3 µmol·L⁻¹-induced elevation of $[Ca^{2+}]_i$ was not influenced by Ind. The calcium mobilization by STA₂ 3 µmol·L⁻¹ was partially depressed by ONO-3708 30 µmol·L⁻¹ and egtazic acid 3 mmol·L⁻¹, and completely inhibited by S-145 100 nmol·L⁻¹(P < 0.01) (Fig 2).

DISCUSSION

It has been considered that platelets will undergo the morphological change in shape and then



Fig 2. A) Effect of indometacin 3 μ mol on STA₂-induced [Ca²⁺], mobilization in single platelets. B) Effects of egtazic acid 3 mmol, S-145 100 nmol, and ONO-3708 30 μ mol on STA₂-stimulated [Ca²⁺]₁ mobilization in single platelets. n = 41 - 57 cells from 6 rabbits, $\bar{x} \pm s$. "P > 0.05 vs STA₂.

cell to cell aggregation once platelets are activated by aggregating agents, but, whether or not there are 2 different TXA_2 receptors or 2 different binding sites for TXA_2 antagonists to regulate the platelet shape change and aggregation, it is still unknown.

In this experiment, STA_2 (the specific agonist of TXA_2 receptor) was used to cause the platelet shape change and aggregation, which directly activated the TXA_2 receptors and its effects were not associated with the metabolic products of arachidonic acid because STA_2 -induced platelet shape change and aggregation were not influenced by Ind, so the pharmacological effects of STA_2 and its antagonists on the shape change and aggregation of platelets were easily analysed.

Some evidences from present experiments supported the fact that TXA₂ receptors exist 2 binding sites for ONO-3708 and S-145, and these 2 binding portions are considered to be relative to the regulation of platelet shape change and aggregation. Firstly, the effect of ONO-3708 and S-145 on platelet shape change was exactly different from each other even though the aggregation was completely inhibited by both ONO-3708 and S-145. The dose-response curve STA₂-induced of aggregation was shifted parallely by ONO-3708 and unparallely by S-145 with the decrease in the

Secondly, the inhibitory effect maximal response. of S-145 was not affected by the procedure of washing platelets, but the procedure rendered ONO-In addition, the effect of 3708 lost its effect. S-145 was enhanced by the prolongation of preincubation time, the effect of ONO-3708 under the same condition was not changed. These results suggested that S-145 tightly and irreversibly bound to its binding portion and the combination of ONO-3708 with its binding site was untight and Finally, we compared the action of reversible. both antagonists on STA_2 -induced [Ca^{2+}], mobilization, the results indicated that S-145 abolished both calcium influx and intracellular calcium release and ONO-3708 only inhibited STA2induced calcium influx without the effect on the intracellular calcium release, this effect of ONO-3708 was similar to that of egtazic acid but due to the different mechanisms. All together it can easily be explained that why S-145 completely inhibited STA2-induced platelet shape change and aggregation and that the reason why ONO-3708 and egtazic acid only inhibited STA2-induced aggregation without the effect on the platelet shape change. In other words, the platelet shape change is dependent on the intracellular calcium release while the aggregation is extracellular calcium related. So far, the relationship between intracellular calcium release and extracellular calcium influx during platelet shape change and aggregation and the molecular mechanisms connected these 2 cellular events are still unclear.

In summary, according to the evidences mentioned above and the biochemical data from other reports^[4,9], it has been suggested that S-145 may be tightly and irreversibly bound to its binding portion different from that for ONO-3708 in quite opposite way on the same TXA₂ receptor, therefore they regulated the different cellular event. But the possibility of more than one TXA₂ receptors on the platelets is still remained although the single cDNA cloned was detected^[10], which did not exclude a single protein moiety with different glycoprotein.

REFERENCES

Siess W, Boehlig B, Weber PC, Lapetina EG.
 Prostaglandin endoperoxide analogues stimulate phospholipase C

and protein phosphorylation during platelet shape change. Blood 1985; **65**: 1141 - 8.

- Halushka PV, Mais DE, Mayeux PR, Mormelli TA.
 Thromboxane, prostaglandin and leukotrine receptor.
 Annu Rev Pharmacol Toxicol 1989; 29: 213 39.
- 3 Narutniya S, Okuma M, Ushikubi F. Binding of a radio iodinated 13-azapinane thromboxane antagonist to platelets: correlation with antiaggregatory activity in different species. Br J Pharmacol 1966; 68: 323 - 31.
- Takahara K, Murray R, Fitzgerald GA, Fitzgerald DJ.
 The response to thromboxane A₂ analogues in human platelets.
 J Biol Chem 1990; 265: 6836 44.
- 5 Ushikubi F, Nakajima M, Hirata M, Okuma M, Fujiwara M, Narutniya S. Purification of the thromboxane A₂/prostaglandin H₂ receptor from human blood platelets.
 J Biol Chem 1989; 264: 16496 - 501.
- 6 Li BY, Bai Y, Li GZ, Li WH, Katori M. Effects of MK-447 on thrombin-induced aggregation, secretion of ATP. and $\{Ca^{2^+}\}$, mobilization in rabbit platelets. Acta Pharmacol Sin 1995; 16: 108 – 13.
- 7 Born GVR. Aggregation of blood platelets by adenosine diphosphate and its reversal. Nature 1962; 194: 927 9.
- B Grynkiewicz G, Poenie M, Tsien RY. A new generation of Cs²⁺ indicators with greatly improved fluorescence properties.
 J Biol Chem 1985; 260: 3440 50.
- 9 Furci L, Fitzgerald DJ, Fitzgerald GA. Heterogeneity of prostaglandin H₂/thromboxane A₂ receptors: Distinct subtypes mediate vascular smooth muscle contraction and platelet aggregation. J Pharmacol Exp Ther 1991; 258: 74 - 81.
- 10 Hirata M, Hayashi Y, Ushikubi F, Yokota Y, Kageyama R,

Nakanishi S₁ et al. Cloning and expression of cDNA for a human thromboxane A_2 receptor.

345-348 Nature 1991; 349: 617-20.

ONO-3708 和 S-145 对 STA2 介导的 家兔血小板变形和聚集反应的抑制作用

李柏岩,乔国芬,孙建平,高云瑞,李文汉 (哈尔滨医科大学药理教研室,哈尔滨 150086,中国)

关键词 血小板; <u>血小板聚集;</u> 血小板聚集抑制剂 剂; 血栓素 A₂; 钙; STA₂; ONO-3708; S-145

A 目的: 评价 ONO-3708 和 S-145 对血小板变形和 聚集反应的不同抑制模式.方法:以透光度法测 量血小板变形和聚集反应,荧光图像分析法测量 单细胞内游离钙的变化.结果:(1) STA2 的聚集 反应可被依他酸,ONO-3708 和 S-145 抑制(P< 0.01),血小板变形仅被 S-145 抑制.(2) S-145 的抑制作用随孵育时间延长而增强,ONO-3708 不 变.(3)洗脱后 ONO-3708 的作用消失,而 S-145 抑制作用依然存在.(4) STA2 的细胞内游离钙 动员部分被 ONO-3708 和依他酸取消(P<0.01), 但可被 S-145 完全抑制.结论: S-145 和 ONO-3708 分别作用于血小板 TXA2 受体的不同结合位点.

BIBLID: ISSN 0253-9756

Acta Pharmacologica Sinica 中国药理学机

1996 Jul; 17 (4): 348 - 350

R 965.1 R963

Protective effects of tetrandrine on CCl₄-injured hepatocytes

CHEN Xiao-Hong, HU You-Mei, LIAO Ya-Qin (Department of Pharmacology, The Third Military Medical College, Chongging 630038, China)

KEY WORDS tetrandrine; liver; cultured cells; carbon tetrachloride poisoning; malondialdehyde; calcium; membrane fluidity; lactate dehydrogenase

AIM: To study the protective effects of tetrandrine (Tet) on CCI₄-injured hepatocytes. **METHODS:** The cultured rat liver cells were poisoned by CCI₄ (10 mmol \cdot L⁻¹). The membrane fluidity was detected by 1,6-diphenyl-1,3,5-hexatriene (DPH), a

lipid probe. The Ca²⁺ concentration was assayed with Fura 2-AM, a sensitive calcium indicator. **RESULTS:** Tet $(1 - 1000 \text{ nmol} \cdot \text{L}^{-1})$ increased viability of liver cell (from 71 % to 72 % - 89 %), reduced lactate dehydrogenase (LDH) release, and malondialdehyde (MDA) formation. Tet prevented the heightening of the intracellular Ca²⁺ concentration and the attenuation of the membrane fluidity of liver cells (P < 0.05). CONCLUSION: Tet had a protective effect on CCl₄-injured hepatocytes by inhibiting the lipid peroxidation,