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氟伏沙明抑制中国青年人体内米帕明去甲代谢许振华, 黄松林, 周宏灏¹

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关键词 米帕明; 地昔帕明; 氟伏沙明; 药物相互作用; 药动动力学**目的:** 研究氟伏沙明(Flu)对健康中国志愿者体内米帕明(lmi)去甲代谢的影响。 **方法:** 7名青年志

愿者连续口服 Flu (100 mg/天) 2 周, 在服用 Flu 的第 8 天受试者口服 lmi (100 mg), 将此时 lmi 及其去甲代谢产物地昔帕明(Des)的药动学参数与 lmi 单独口服时进行比较。 **结果:** Flu 使口服单剂量 lmi 的体内消除过程发生显著改变, 服用 Flu 导致 lmi C_{max} 和 AUC 约增高 1 倍, $T_{1/2}$ 显著延长, Cl_0 降低 60 %。同时, lmi 的主要代谢产物 Des 的生成减少, 表现在 C_{max} 和 AUC 均明显降低。 **结论:** Flu 能明显抑制 lmi 的去甲基代谢, CYP1A2 被抑制可能是 Flu 和 lmi 发生相互作用的生化基础。

Doxorubicin cellular pharmacokinetics plays no role in chemosensitizing effect of verapamil on Swiss-3T3 cellsWANG Nan, TANG Zhong-Ming, ZHANG Yang-Pei, DING Qing-Ming
(Institute of Radiation Medicine, Beijing 100850, China)**KEY WORDS** doxorubicin; verapamil; drug resistance; pharmacokinetics; fibroblasts; cultured tumor cells

AIM: To find whether or not the doxorubicin (Dox) cellular pharmacokinetics plays a role in chemosensitizing effect of verapamil (Ver) on drug sensitive cells. **METHODS:** Cytotoxicity and cellular Dox contents (during accumulation and retention periods) were measured in the absence and presence of verapamil in Swiss-3T3 cells and compared with those in multidrug resistant (MDR) MCF-7^{Adr} cells and drug sensitive MCF-7^{WT} cells. *mdr-1* mRNA expression in Swiss-3T3 cells was analyzed. **RESULT:** Dox cytotoxicity was enhanced 2.0-fold in Swiss-3T3 cells by Ver ($3 \mu\text{mol}\cdot\text{L}^{-1}$) and 3.6-fold in MCF-7^{Adr} cells by Ver ($6 \mu\text{mol}\cdot\text{L}^{-1}$), but not in MCF-7^{WT} cells (Ver $6 \mu\text{mol}\cdot\text{L}^{-1}$). Cellular accumulation of equi-effective concentrations of Dox increased at 6-h incubation in the presence of Ver in Swiss-3T3 (1.5-fold) and MCF-7^{WT} cells (2.1-fold) but decreased rapidly in MCF-7^{Adr} cells by 20 % to

50 % compared to that in the absence of Ver. Cellular retention of Dox decreased after 10-min increase in the presence of Ver in Swiss-3T3 cells compared to that in the absence of Ver, that was similar to that in MCF-7^{WT} cells, while the retention was augmented by Ver in MCF-7^{Adr} cells. Slot blot analysis of RNA revealed no *mdr-1* gene expression in Swiss-3T3 cells. **CONCLUSION:** Changes in cellular accumulation and retention of Dox did not account for the chemosensitizing effect of Ver on Swiss-3T3 cells.

Multidrug resistance (MDR) was associated with the expression of multidrug resistance genes encoding a family of integral membrane glycoproteins known as P-glycoproteins (Pgp)^[1]. These transporting proteins bound various chemotherapeutic agents and pumped them out of the cells. Verapamil (Ver), a well-known resistant modifier or chemosensitizer, competitively bound to Pgp and increased the cellular uptake and cytotoxicity of drugs in a calcium-independent manner^[2]. Sensitizing effect of Ver was occasionally noted in drug sensitive cells^[3] or non-

Pgp expressed MDR cells⁽⁴⁻⁷⁾. To find if any role of cellular pharmacokinetic changes in Ver sensitization, we observed the cellular doxorubicin (Dox) accumulation and retention in the presence of Ver in Swiss-3T3 cells, and compared with that in MDR and drug sensitive MCF-7 cells.

MATERIALS AND METHODS

Drugs and chemicals Dox, Ver and 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-tetrazolium bromide (MTT) were purchased from Sigma (USA). Me₂SO was from E Merck (Germany). Dulbecco's modified Eagle's medium (DMEM) was obtained from Gibco/BRL (USA). [α -P³²]dATP was from YaHui Biomed Co (Beijing, China).

Cell culture The human breast cancer cell line MCF-7^{WT} and mouse fibroblast cell line Swiss-3T3 were cultured in DMEM supplemented with 10 % calf serum in a 5 % CO₂ humidified incubator at 37 °C. The MCF-7^{Adr}(8) (a gift from Dr KH Cowan, NCI, NIH, USA) was maintained in the medium containing Dox (10 $\mu\text{mol}\cdot\text{L}^{-1}$) which was removed 20 d before the experiments.

Chemosenstivity testing Cytotoxicity was determined by MTT assay⁽⁹⁾. Cells were plated into 96-well tissue culture plates and exposed to drugs continuously. After incubation at 37 °C for 4 d, 50 μL of MTT dye (2 $\text{g}\cdot\text{L}^{-1}$ in Hanks' solution) was added to each well and incubated for 4 h. The medium was then aspirated and 100 μL of Me₂SO was added. The plates were agitated for 5 min and absorbance at 540 nm was read on a Titertek Multiscan MCC/340 plate reader (Flow Laboratories, UK). In all experiments 4 replicate wells were used for each drug concentration. IC₅₀ values of Dox were obtained from dose-response curves fitted by logistic model⁽⁹⁾.

Intracellular Dox assay and protein assay Fluorescent measurement of Dox was performed as described⁽¹¹⁾ with slight modification. Drug-treated cells cultured in 100 or 90 mm dishes were washed 4 times rapidly with 5 mL ice-cold PBS and scraped into Eppendorf tubes. After centrifugation (3000 \times g, 4 °C, 5 min), the cell pellet was resuspended in distilled water and left overnight in the dark to lyse the cells. Aliquots of 0.1 mL were suspended in 0.9 mL methanol and centrifuged 12 000 \times g at 4 °C for 10 min, the supernatant was read on a MPF-4 fluorescence spectrophotometer (Hitachi, Japan) (λ_{ex} = 468 nm, λ_{em} = 585 nm, slit 10 nm, and sensitivity at 3.5). The concentration of Dox was determined in comparison with a standard Dox curve. Cell lysates 5 - 10 μL were used for protein assay according to dye-binding method⁽¹²⁾, modified to 96-well microplate scale.

Drug accumulation and retention Cell monolayer cultures were incubated with Dox at the concentrations of IC₅₀ values in the absence or presence of Ver. At appropriate

times the cells were harvested and intracellular accumulations of Dox were determined. For drug retention tests, the cell monolayers incubated with Dox and Ver for 6 h was further incubated in the absence or presence of Ver after withdrawal of Dox, cells were harvested at various time points for Dox measurements. The Dox concentration was corrected to the protein content in the cell sample (as nmol of Dox/g protein).

RNA analysis RNA was isolated from Swiss-3T3 cells by single-step method⁽¹³⁾. Slot blot analysis of total cellular RNA was carried out as described⁽¹⁴⁾ using a full length cDNA probe of *mdr-1* (a gift from Dr MM Gottesman, NCI, NIH, USA). *mdr-1* Gene-transfected Swiss-3T3 cells were included as positive control. The blots were reprobed with mouse β -actin cDNA probe for comparison of sample loading.

Statistics The differences of Dox accumulations or retentions between Ver treatment were compared by *t* test.

RESULTS

Cytotoxicity The cytotoxicity of Dox to Swiss-3T3 cells was enhanced after treatment with Ver. The IC₅₀ for Swiss-3T3 cells was 141 (95 % confidence limits 14 - 268) $\text{nmol}\cdot\text{L}^{-1}$ and 72 (95 % confidence limits 41 - 103) $\text{nmol}\cdot\text{L}^{-1}$ in the absence and presence of Ver 3 $\mu\text{mol}\cdot\text{L}^{-1}$, respectively. MCF-7^{Adr} showed a high level of resistance to Dox (IC₅₀ = 25.50 $\mu\text{mol}\cdot\text{L}^{-1}$, 95 % confidence limits 14.11 - 36.89), compared with its parent cell line MCF-7^{WT} (IC₅₀ = 73 $\text{nmol}\cdot\text{L}^{-1}$, 95 % confidence limits 46 - 100). Ver partially reversed the resistance of Dox in MCF-7^{Adr} (IC₅₀ = 7049 $\text{nmol}\cdot\text{L}^{-1}$ in the presence of Ver 6 $\mu\text{mol}\cdot\text{L}^{-1}$, 95 % confidence limits 5387 - 8711). While Ver 6 $\mu\text{mol}\cdot\text{L}^{-1}$ did not enhance Dox cytotoxicity against MCF-7^{WT} cells (IC₅₀ = 76 $\text{nmol}\cdot\text{L}^{-1}$, 95 % confidence limits 43 - 109). At concentrations of 3 $\mu\text{mol}\cdot\text{L}^{-1}$ and 6 $\mu\text{mol}\cdot\text{L}^{-1}$ for Ver alone, no effect on cell viability or growth was observed for Swiss-3T3 and MCF-7 cells (Fig 1).

Drug accumulation When incubated with equi-effective (IC₅₀) concentration of Dox in the presence of Ver (3 $\mu\text{mol}\cdot\text{L}^{-1}$), intracellular accumulation of Dox in Swiss-3T3 cells did not change significantly at 1 h and 3 h, until at 6 h ($P < 0.01$), compared with that without Ver. Dox accumulation in MCF-7^{WT} was influenced by Ver (6 $\mu\text{mol}\cdot\text{L}^{-1}$) in a similar manner to that in Swiss-3T3 cells (Fig 2). While in MCF-7^{Adr} accumulation of Dox in the presence of Ver

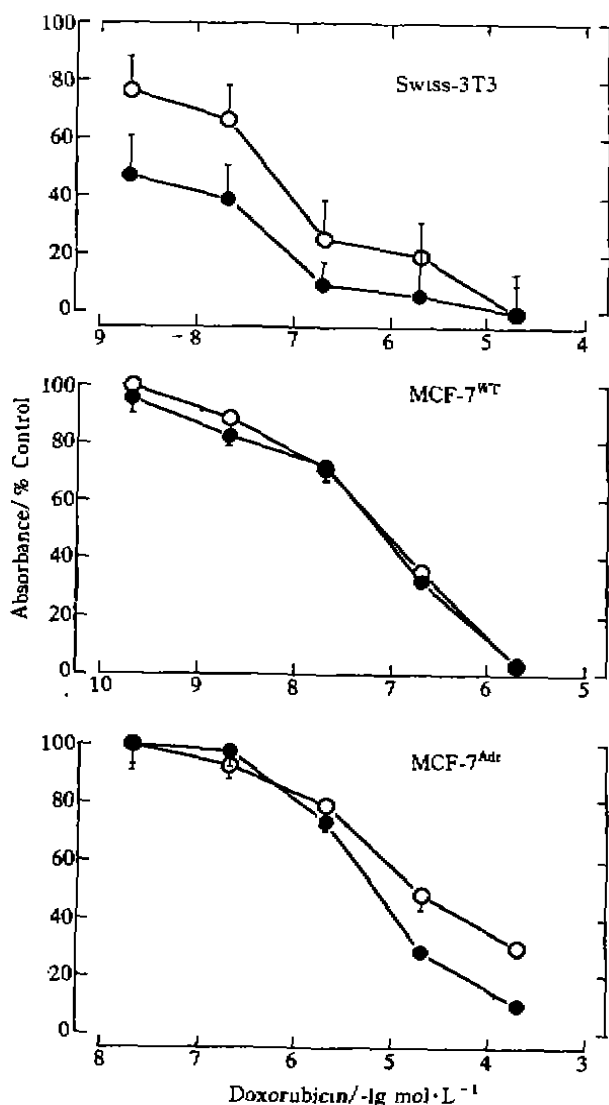


Fig 1. Response of cells to Dox in the absence (○) and presence (●) of Ver ($3 \mu\text{mol} \cdot \text{L}^{-1}$ for Swiss-3T3, and $6 \mu\text{mol} \cdot \text{L}^{-1}$ for the other two). $n = 4$. $\bar{x} \pm s$.

($6 \mu\text{mol} \cdot \text{L}^{-1}$) reduced by 20 %, 40 %, and 50 %, compared to that without Ver, at 1 h, 3 h, and 6 h incubation of Dox of IC_{50} .

Drug retention Continuing treatment with Ver after withdrawal of Dox increased the retention in MCF-7^{Adr}. The retention of Dox in Swiss-3T3 cells increased at 10 min and then decreased in the continuing presence of Ver compared to absence of Ver, which showed a dramatically similar pattern as that in MCF-7^{WT} (Fig 3).

RNA slot blot Slot blot analysis revealed no evidence of *mdr-1* message in the Swiss-3T3 cells

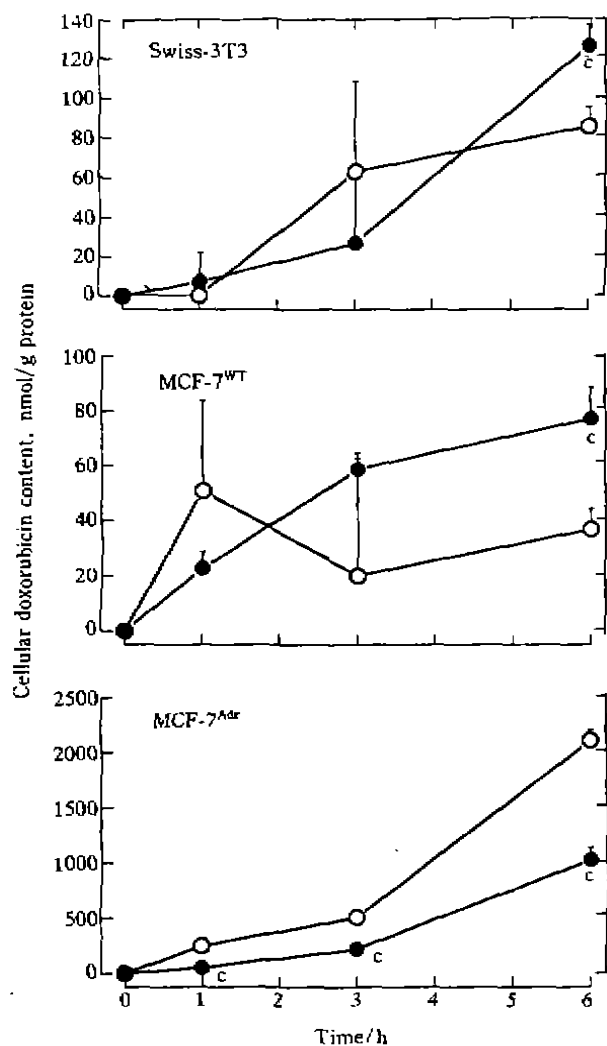


Fig 2. Accumulation of Dox by cells in the absence (○) and presence (●) of Ver ($3 \mu\text{mol} \cdot \text{L}^{-1}$ for Swiss-3T3, $6 \mu\text{mol} \cdot \text{L}^{-1}$ for the other two). The cells were loaded with IC_{50} of Dox. $n = 4$. $\bar{x} \pm s$. * $P < 0.01$ vs absence of Ver.

(Fig 4). *mdr-1* Gene transfected Swiss-3T3 cells were included as positive control which showed *mdr-1* mRNA expression.

DISCUSSION

Changes in cellular drug levels have been accounted for the expression of drug resistance and its reversal with calcium antagonists in Pgp expressed MDR cells^[1]. In a typical MDR cell line MCF-7^{Adr}, diminished Dox accumulation and retention could be seen when these values were expressed as ratio of intracellular to extracellular

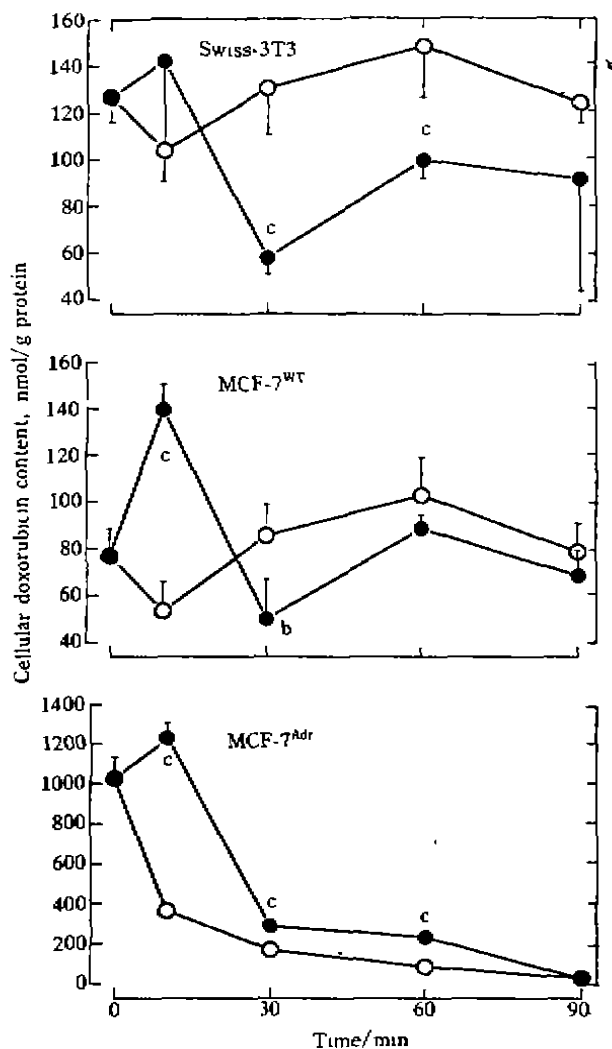


Fig 3. Dox retention in cells preloaded with IC_{50} of Dox in the absence (○) or presence (●) of Ver. $n = 4$. $\bar{x} \pm s$. ^b $P < 0.05$, ^c $P < 0.01$ vs (○).

drug concentrations (data not shown) compared with those in its parent cell line MCF-7^{WT}, which was partially restored by Ver. In our study IC_{50} concentrations of Dox were used in order to compare the pharmacokinetic processes at similar levels of biological action of the drug on these cells. The chemosensitizing effect of Ver on Dox cytotoxicity was observed in Swiss-3T3 cells in which *mdr-1* gene expression was not detected. The cellular Dox level was altered in the presence of Ver during the accumulation and/or retention phase in this cell, the pattern and magnitude of alterations were strikingly similar to that in drug sensitive MCF-7^{WT} on which no sensitizing effect of Ver was observed. While

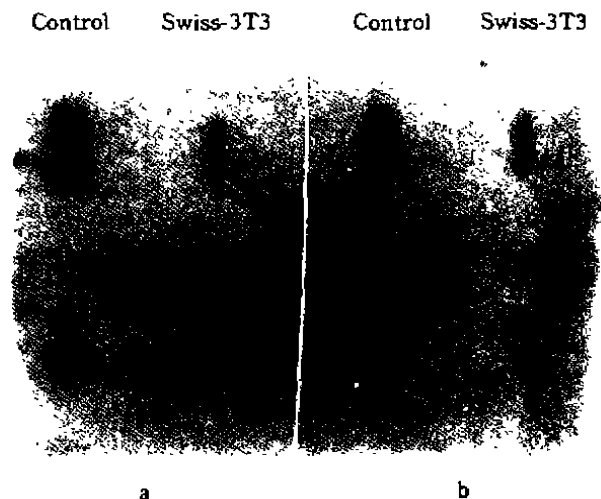


Fig 4. Slot blot analysis of Swiss-3T3 RNA. Total cellular RNA was applied to the nylon membrane by 1/7 dilution (20 μ g at upper and 3 μ g at lower). *mdr-1* Gene transfected Swiss-3T3 cells were used as positive control. Hybridization with *mdr-1* cDNA probe (a) or β -actin cDNA probe (b).

the alteration in cellular Dox pharmacokinetics in MCF-7^{Adr} was quite different from Swiss-3T3 and MCF-7^{WT} cells. Ver enhanced drug accumulation and retention was observed in other sensitive cells and was attributed to the nonspecific action of Ver on plasma membrane^[15]. Our results suggested that sensitizing effect of Ver on Swiss-3T3 cells was not attributed, at least mainly, to cellular pharmacokinetic changes. Intracellular Ver enhanced sensitizing mechanisms could be involved in such effect as it has been reported that Ver potentiated cytotoxicity of Dox was associated with an increase in DNA-breakage in two human ovarian tumor cell lines^[7]. There could be great difference between cell lines in mechanism of response to Ver, as shown in Swiss-3T3 cells and MCF-7 cells.

REFERENCES

- Gottesman MM, Pastan I. Biochemistry of multidrug resistance mediated by the multidrug transporter. *Annu Rev Biochem* 1993; **62**: 385 - 427.
- Ford JM, Hait WN. Pharmacology of drugs that alter multidrug resistance in cancer. *Pharmacol Rev* 1990; **42**: 155 - 99.
- Beck WT. Modulators of P-glycoprotein-associated multidrug resistance. In: Ozols RF, editor. *Molecular and clinical advances in anticancer drug resistance*. Boston: Kluwer Academic Publishers, 1991: 151 - 70.

4 Nygren P, Larsson R, Gruber A, Peterson C, Bergh J. Doxorubicin selected multidrug-resistant small cell lung cancer cell lines characterised by elevated cytoplasmic Ca²⁺ and resistance modulation by verapamil in absence of P-glycoprotein overexpression. *Br J Cancer* 1991; **64**: 1011 - 8.

5 Slovak ML, Hoeltge GA, Dalton WS, Trent, JM. Pharmacological and biological evidence for differing mechanisms of doxorubicin resistance in two human tumor cell lines. *Cancer Res* 1988; **48**: 2793 - 7.

6 Coley HM, Workman P, Twentymen PR. Retention of activity by selected anthracyclines in a multidrug resistant human large cell lung carcinoma line without P-glycoprotein hyperexpression. *Br J Cancer* 1991; **63**: 351 - 7.

7 Delvaeye M, Verovski V, De Neve W, Storme G. DNA breakage, cytotoxicity, drug accumulation and retention in two human ovarian tumor cell lines AZ224 and AZ364 treated with adriamycin, modulated by verapamil. *Anticancer Res* 1993; **13**: 1533 - 8.

8 Batist G, Tulpule A, Sinha BK, Katki AG, Myers CE, Cowan KH. Overexpression of a novel anionic glutathione transferase in multidrug-resistant human breast cancer cells. *J Biol Chem* 1986; **261**: 15544 - 9.

9 Carmichael J, DeGraff WG, Gazdar AF, Minna JD, Mitchell JB. Evaluation of a tetrazolium-based semiautomated colorimetric assay: assessment of chemosensitivity testing. *Cancer Res* 1987; **47**: 936 - 42.

10 Wang N, Zhao DH, Sheng BH. A program for analysis of dose-response relationship with logistic model. *Acta Pharmacol Sin* 1990; **11**: 187 - 92.

11 Radel S, Bankusli I, Mayhew E, Rustum YM. The effects of verapamil and a tiapamil analogue, DMDP, on adriamycin-induced cytotoxicity in P388 adriamycin-resistant and -sensitive leukemia *in vitro* and *in vivo*. *Cancer Chemother Pharmacol* 1988; **21**: 25 - 30.

12 Bradford MM. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem* 1976; **72**: 248 - 54.

13 Chomczynski P, Sacchi N. Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction.

Anal Biochem 1987; **162**: 156 - 9.

14 Sambrook J, Fritsch EF, Maniatis T, editors. *Molecular cloning: a laboratory manual*. 2nd ed Cold Spring Harbor: Cold Spring Harbor Laboratory Press, 1989: 7 54

15 Cano-Gauci DF, Riordan JR. Action of calcium antagonists on multidrug resistant cells. Specific cytotoxicity independent of increased cancer drug accumulation. *Biochem Pharmacol* 1987; **36**: 2115 - 23.

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阿霉素细胞水平药物动力学在维拉帕米对 Swiss-3T3 细胞的化学增敏效应中不起作用

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关键词 阿霉素; 维拉帕米; 抗药性; 药物动力学; 成纤维细胞; 培养的肿瘤细胞

A 目的: 确定维拉帕米(Ver)对药物敏感细胞化学增敏效应中, 阿霉素(Dox)细胞药物动力学是否起作用。 **方法:** 测定合并 Ver 与否时, Dox 对 Swiss-3T3 细胞的毒性, 蓄积和残留期细胞内 Dox 含量, 并与多药抗药细胞 MCF-7^{Adr} 及敏感细胞 MCF-7^{WT} 比较。 **分析** Swiss-3T3 中 *mdr-1* mRNA 的表达 **结果:** Ver 增强 Dox 对 Swiss-3T3 和 MCF-7^{Adr} 的毒性, 而不增强 MCF-7^{WT} 的毒性。 存在 Ver 与反之相比, Swiss-3T3 和 MCF-7^{WT} 对等效浓度 Dox 的蓄积于孵育 6 h 增高, 残留量在 10 min 内升高, 而后下降; 而 MCF-7^{Adr} 对 Dox 的蓄积迅速降低, 残留则被 Ver 提高。 狭缝杂交未见 Swiss-3T3 有 *mdr-1* 基因表达。 **结论:** Dox 在细胞内蓄积与残留的变化, 不能说明 Ver 对 Swiss-3T3 的化学增敏作用。