

Original Research

Effect of U-73122 on calcium channel activity in porcine myometrial cells and pancreatic beta-cell line RINm5F

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KEY WORDS Bay-K-8644; calcium; calcium channels; Fura-2; islets of Langerhans; myometrium; patch-clamp techniques; phospholipase C; U-73122; U-73343

AIM: To study the effect of U-73122 putative phospholipase C (PLC) inhibition on cytosolic Ca2+ concentration ([Ca2+]i) and voltage dependent calcium channels (V). METHODS: To record whole cell calcium current (ICa) by perforated patch and to measure cytosolic free calcium concentration ([Ca2+]i) using Fura-2. RESULTS: U-73122 inhibited alpha-induced and Bay-K-8644-induced increase in [Ca2+]i dose-dependently. U-73122 increased markedly depolarization-induced ICa in RINm5F and porcine myometrial cells in a concentration-dependent manner. The effect of U-73122 was more potent in porcine myometrial cells than that in RINm5F cells. U-73343 (an analog of U-73122) at does not inhibit PLC also reduced ICa by 50% and Vmax of VDCC in myometrial cells.

phospholipase C (PLC) by activation and neurotransmitters is an important signal transduction pathway. The role of PLC-mediated signals plays an important role in various cellular functions. U-73122, a novel inhibitor of PLC, has been applied in the studies of the PLC-dependent process in many cell types such as platelets and neutrophils. These hamster ovary cells, pancreatic acinar cells, Chi

cells insulin-secreting cell lines RINm5p2) and TC3(3) pancreatic AR42J cells a neuroblastoma cell line neuronal cells pituitary gonadotrophs rat chondrochondral chondrocyt and porcine myometrial cells(4)

U-73122 has additional actions that are independent of the inhibition of PLC activity. For example U-73122 stimulates the release of Ca2+ from endoplasmic reticulum (ER) of the rat liver through inhibition of Ca2+ uptake by Ca2+-ATPase(5). U-73122 blocks KCl-induced (50 mmol.L-1) Ca2+ influx in NG108-15 cells but not in dorsal root ganglion neurons. U-73122 also inhibits the Ca2+ influx induced by psigargin a blocker of ER Ca2+-ATPase in mouse fibroblasts(6).

We have used U-73122 as a PLC inhibitor in porcine myometrial cells to study the mechanisms of oxytocin receptors(4) and in RINm5F cells to study the mechanisms of muscarinic and arginine-vasopressin receptors(2). In this study we explored the effect of U-73122 on Ca2+ influx via VDCC in RINm5F cells and endogenous porcine myometrial cells to demonstrate the effect of U-73122 on these endocrine cells and smooth muscle cells. The major VDCC is the L-type channel in the pancreatic cells(7) and smooth muscle cells. Besides we used one of L-type VDCC agonist Bay-K-8644(8) 14-dihydro-26-dimethyl-5-4-[2-(4-fluoromethyl)-phenyl]-3-pyridinecarboxylic acid and an analog of U-73122 U-73343(9) 1-[6-[[17-methoxy-1,3,5-triazol-4-yl]amino]-2,5-pyrrolidinedione] which should not effect on PLC.

MATERIALS AND METHODS

Cell culture RINm5F cells were maintained in Roswell Park Memorial Institute medium 1640 containing 10% fetal bovine serum and aerated with 5% CO2 at 37°C.

All experiments were performed with cells from passages 45-54.

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preparation of myometrial cells Porcine uteri were obtained from a local slaughter house; the myometrial cells from pregnant sows in late pregnancy (d 107-112) were isolated (IO).

Recordings of  $[Ca^{2+}]_i$  The perforated patch-clamp (U) was used to record whole cell  $[Ca^{2+}]_i$  with an Axopatch 2B voltage-clamp amplifier (Axon Instruments Foster City CA). Patch pipettes (3-5 Mn) were made of disposable glass pipettes (VWR Scientific West Chester PA) by a two-stage pull and tip polished. The liquid junction was nullified with an offset circuit before the formation of gigaseals. Linear resistance and capacitive currents were electronically compensated by the P/N protocol. Currents were low-pass filtered at 1 kHz. The data were acquired and analyzed using an IBM computer with the pClamp software (Axon Instruments). The patch pipette was filled with a solution containing L-glutamic acid 12 mmol-L<sup>-1</sup>, CsOH 130 mmol-L<sup>-1</sup>, alliphotericin B 250 mg-L<sup>-1</sup>, HEPES 10 mmol-L<sup>-1</sup> (pH 7.2). The bath solution contained tetraethylammonium chloride 140 mmol-L<sup>-1</sup>, 4-aminopyridine 5 mmol-L<sup>-1</sup>, CaCl<sub>2</sub> 2 mmol-L<sup>-1</sup>, dextrose 1.67 mol-L<sup>-1</sup>, KOH 5.4 mmol-L<sup>-1</sup> and HCl 0 mmol-L<sup>-1</sup>. U-73122 was applied to the bath solution directly and all experiments were performed at room temperature. In RINm5F cells the concentrations of U-73122 were actual (but not cumulative). In myometrial cells the concentrations of U-73122 were cumulative due to technical difficulty in perforating patch-clamp experiments in myometrial cells.

Measurement of  $[Ca^{2+}]_i$  in RINm5F cells To measure  $[Ca^{2+}]_i$  in all cell suspensions 30 x 10<sup>6</sup> cells were loaded with Fura-2 acetoxymethyl ester (AM) 2 μmol-L<sup>-1</sup> in KRB at 37 °C for 30 min. The loaded cells were washed and kept at 24 °C until use. Cells were resuspended at a concentration of 10<sup>7</sup> L<sup>-1</sup> and a 1.5 mL aliquot was used for  $[Ca^{2+}]_i$  determination at 24 °C. The 340 nm fluorescence ratio was monitored in an SLM-8000 fluorescence spectrophotometer (SLMUA ANAL). The  $[Ca^{2+}]_i$  was calibrated after the cell lysis (12). When needed U-73122 was given 1 s before the administration of KO or Bay-K-8644.

Statistical analysis Results were analyzed by paired and unpaired t test.

Materials Fura-2-AM (Molecular Biology Eugene OR) U-73122 and U-73143 (Biomol Research Laboratories Meeting PA) Bay-K-8644 (Research Biochemicals Inc Natick MA). Fura-2-AM was dissolved in Me<sub>2</sub>SO Bay-K-8644 was dissolved in 95% ethanol. U-73122 and U-73143 were dissolved in chloroform and dispensed in aliquots (10 nmol/20 μL) and then the chloroform was evaporated under a stream of N<sub>2</sub>. This left a dry film of U-73122 or U-73143 in the container which was stored in a desiccator. The dry film was dissolved in Me<sub>2</sub>SO immediately before use. When ethanol or Me<sub>2</sub>SO was used as a solvent the final concentration of the organic solvent was 1:10 in aqueous solution.

RESULTS

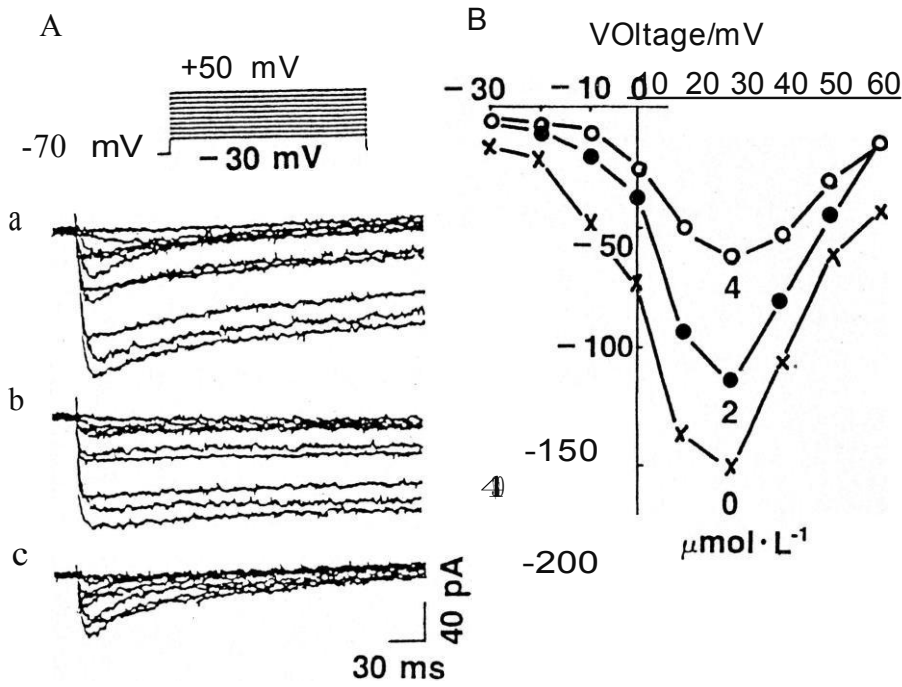
Effect of U-73122 on KCl- and Bay-K-8644 induced increase in  $[Ca^{2+}]_i$  of RINm5F cells The  $[Ca^{2+}]_i$  rise after KCl (5 mmol-L<sup>-1</sup>) usually reached a maximum by 25 s and stayed at the plateau level for 3-4 min. Pretreatment with U-73122 (1-8 μmol-L<sup>-1</sup>) for 1 min inhibited KCl (15 nmol-L<sup>-1</sup>) induced increases in  $[Ca^{2+}]_i$  dose dependently. U-73122 8 μmol-L<sup>-1</sup> inhibited KCl (15 mmol-L<sup>-1</sup>) induced increase in  $[Ca^{2+}]_i$  by 46% (Tab 1).

Table 1. Effect of U-73122 on KCl- and Bay-K-8644 induced peak  $[Ca^{2+}]_i$  in RINm5F cells. U-73122 given 1 s before KCl (15 mmol-L<sup>-1</sup>) or Bay-K-8644 (1 μmol-L<sup>-1</sup>). n=4. \*P<0.05, \*\*P<0.01.

U-73122 / μmol-L <sup>-1</sup>	KO-induced $[Ca^{2+}]_i$ / nmol-L <sup>-1</sup>	Bay-K-8644-induced $[Ca^{2+}]_i$ / nmol-L <sup>-1</sup>
	47.2 ± 1.7	25.1 ± 1.0
1	46.7 ± 1.0*	19.6 ± 0.6*
2	37.2 ± 2.0b	16.2 ± 0.8b
4	31.2 ± 1.3b	14.1 ± 1.0b
8	25.3 ± 2.0b	10.9 ± 0.1*

When Bay-K-8644 (1 μmol-L<sup>-1</sup>) was applied RINm5F cells caused a rapid increase in  $[Ca^{2+}]_i$  which was sustained for 3-4 min. The effect of Bay-K-8644 (1 μmol-L<sup>-1</sup>) was abolished by the L-type VDCC blocker nimodipine (1 μmol-L<sup>-1</sup>). To examine the effect of U-73122 was on PLC we investigated its effect on Bay-K-8644 induced increase in  $[Ca^{2+}]_i$ . Pretreatment with U-73122 (1-8 μmol-L<sup>-1</sup>) for 1 min inhibited Bay-K-8644 (1 μmol-L<sup>-1</sup>)-induced increase in  $[Ca^{2+}]_i$  dose dependently (Fig 1B). U-73122 8 μmol-L<sup>-1</sup> inhibited Bay-K-8644 (1 μmol-L<sup>-1</sup>)-induced increase in  $[Ca^{2+}]_i$  by 57%.

Inhibition by U-73122 of depolarization induced  $[Ca^{2+}]_i$  in RINm5F cells At the holding potential (HP) of -70 mV currents were elicited by 3 min depolarization steps from -30 mV to +50 mV. The maximal  $[Ca^{2+}]_i$  was evoked by a depolarizing pulse to +20 mV. U-73122 (2 and 4 μmol-L<sup>-1</sup>) reduced the peak  $[Ca^{2+}]_i$  but did not shift membrane potential value of the initial or peak  $[Ca^{2+}]_i$  (Fig 1).



**Fig 1.** Inhibition by U-73122 of depolarization-induced I<sub>Ca</sub> in RINm5F cells. Currents were elicited by 3 ms steps from -30 mV to +50 mV at the holding potential of -70 mV. A control traces obtained under control condition (a) and in the presence of U-73122 (2 μmol · L<sup>-1</sup> and 4 μmol · L<sup>-1</sup>) B corrected (n=4) showing are represented in Fig 1 of 4 experiments.

The control I<sub>Ca</sub> was established by depolarizing a HP of -70 mV to +20 mV. This pulse was applied every 10 s which elicited I<sub>Ca</sub>. U-73122 (8 μmol · L<sup>-1</sup>) abolished I<sub>Ca</sub> 2 min of administration (Fig 2A). The effect of U-73122 on I<sub>Ca</sub> was reversed by a washout and application of U-73122 again produced inhibition of I<sub>Ca</sub>. The I<sub>Ca</sub> evoked in RINm5F cells was inhibited by U-73122 (1 - 8 μmol · L<sup>-1</sup>) in a dose-dependent manner (Fig 2C). However U-73343 (8 μmol · L<sup>-1</sup>) an analog of U-73122 does not inhibit I<sub>Ca</sub> by 50% ± 4% (n = 4) (Fig 2B).

Inhibition of depolarization-induced I<sub>Ca</sub> in porcine myocyte cells. At the holding potential of -50 mV, I<sub>Ca</sub> was evoked by the depolarizing pulse to +20 mV. U-73122 (30 and 60 μmol · L<sup>-1</sup>) reduced peak I<sub>Ca</sub> but did not shift the potential value of the initial and peak I<sub>Ca</sub> (Fig 3).

We selected 6 cells from 3 different tissues to study the cumulative dose-response of U-73122. The I<sub>Ca</sub> was evoked by a 3 ms depolarizing step from an HP of -50 mV to +20 mV. This depolarizing step was applied every 20 s. Because the inhibitory effect of each dose of U-73122 reached a stable level within 1 min each dose of U-73122 was administered every 1 min. U-73122 caused a concentration-dependent inhibition of I<sub>Ca</sub>. At the highest concentration of U-73122 studied (1 μmol · L<sup>-1</sup>) I<sub>Ca</sub> was nearly abolished (Fig 4).

**DISCUSSION**

U-73122 has been used to study the PLC-mediated physiological events and mechanisms underlying these events (13). Activation of PLC leads to increased inositol 1,4,5-trisphosphate (IP<sub>3</sub>) formation which in turn promotes the release of Ca<sup>2+</sup> from the ER and Ca<sup>2+</sup> influx through opening of VOCC and receptor-operated Ca<sup>2+</sup> channels (14). U-73122 specifically inhibits PLC-mediated Ca<sup>2+</sup> release and influx (2, 5). However U-73122 indirectly

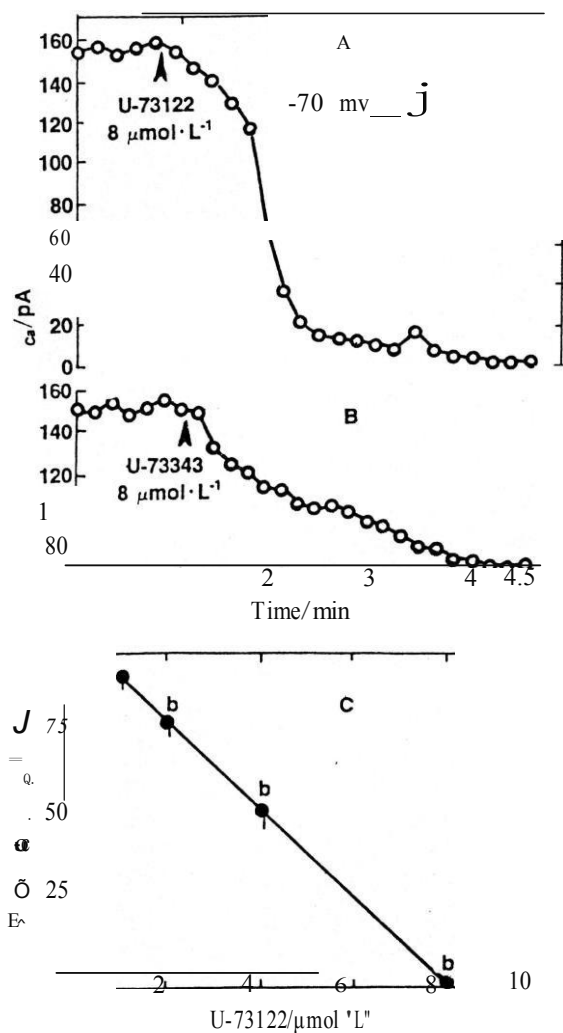


Fig2. Inhibition by U-73122 of  $I$  generated by one-step depolarizations in RINm5F cells. Currents were elicited from a holding potential of  $-70$  mV to  $+20$  mV every 10 s. A: The course of inhibition by U-73122. B: The course of inhibition by U-73343. Data shown in A and B are representative of 4 experiments. C: Dose-response to U-73122 ( $1 - 8 \mu\text{mol} \cdot \text{L}^{-1}$ ). Maximum current was recorded 10 min after application of U-73122 when the effect reached a stable level.  $n = 4$ ,  $s.e.p. < 0.05$  vs control.

inhibit  $\text{Ca}^{2+}$  influx which is independent of PLC activation. The effect of U-73122 has been demonstrated in mouse fibroblasts where U-73122 reduced the  $\text{Ca}^{2+}$  influx induced by apsigargin, an inhibitor of the ER  $\text{Ca}^{2+}$ -ATPase and in differentiated NG100-15 cells where U-73122 blocks completely the KCl-induced  $\text{Ca}^{2+}$  influx(4). KCl increases  $\text{Ca}^{2+}$

influx by depolarizing the membrane potential and opening VIC. Administering a VDCC agonist such as Bay-K-8644 is another method of opening VOCC. In the present study we showed that U-73122 at effective concentrations for inhibiting PLC decreased KCl- and Bay-K-8644-induced  $\text{Ca}^{2+}$  influx in RINm5F cells. In addition U-73122 at these concentrations inhibited dilation-induced  $I$  in RINm5F cells and porcine myometrial cells. U-73343, the inactive analog of U-73122, on P also inhibited  $t_a$  of RINm5F cells but was less effective than U-73122. The results suggest that the inhibitory effect of U-73122 on VIC is independent of the inhibition of PLC. The difference between U-73122 and U-73343 could be due to the slight difference in chemical structure.

U-73122 may have different sensitivity in different cell types with regard to its inhibitory effect on  $\text{Ca}^{2+}$  influx because it blocks KCl-induced  $\text{Ca}^{2+}$  influx in NG100-15 cells but not in dorsal root ganglion neurons(4). Our results further suggest that porcine myometrial cells were more sensitive to U-73122 inhibitory effect on VOCC than RINm5F cells. These results are interesting because: 1) the dose responses of U-73122 in inhibiting  $I_{Ca}$  and PLC-mediated  $\text{Ca}^{2+}$  release parallels each other in both porcine myometrial cells and RINm5F cells and 2) U-73122 is more effective in inhibiting both the  $I_{Ca}$  and PLC-mediated  $\text{Ca}^{2+}$  release in porcine myometrial cells than RINm5F cells(2,35). However, it is not clear why U-73122 is more effective in porcine myometrial cells than RINm5F cells.

In the present study U-73122 was more potent in inhibiting  $I_{Ca}$  than KCl- or Bay-K-8644-induced increase in  $[\text{Ca}^{2+}]_i$ . It is probable that the data obtained at patch clamp experiments were performed on single cells whereas the experiments on  $[\text{Ca}^{2+}]_i$  were performed in a cluster of cells. The cluster would affect uptake of the drug into individual cells.

The inhibitory effect of U-73122 on VIC might be attributed to a possible mechanism. U-73122 is an antinostrioid which is lipophilic and readily diffuses through the plasma membrane or adheres to the plasma membrane thereby potentially changing the conformation and/or activity of VDCC. We found the binding of U-73122 to cells is

also reversible

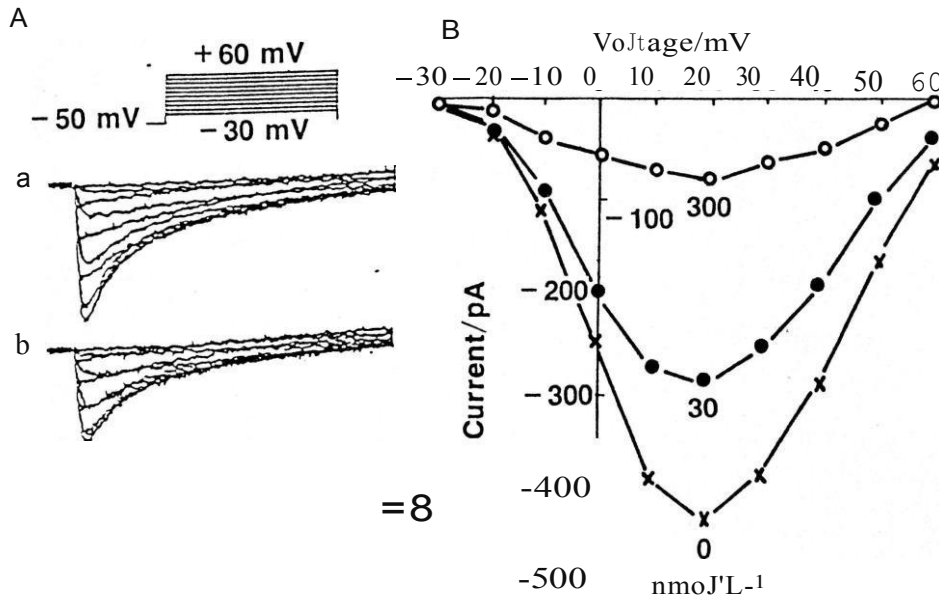


Fig. 3. Inhibition of depolarization-induced current by U-73122 in porcine myometrial cells. Currents were elicited by depolarization steps from -30 mV to +60 mV at the holding potential of -50 mV. Current tracings obtained under control solution (a) and in the presence of U-73122 (b 30 nmol L<sup>-1</sup> and c 300 nmol L<sup>-1</sup>). B, comparing I-V relationship.

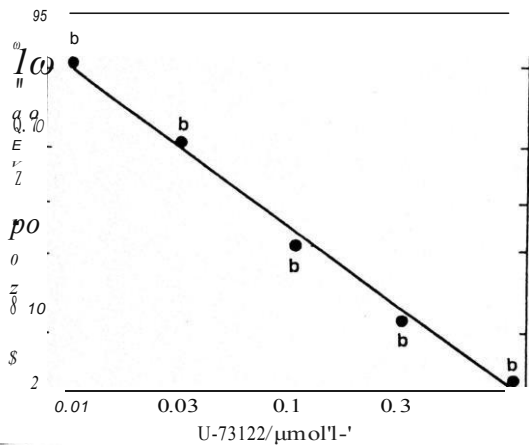


Fig. 4. Inhibition of current by U-73122 in porcine myometrial cells. Currents were elicited from a holding potential of -50 mV to +20 mV. U-73122 was administered in a cumulative manner at the indicated concentrations. *bp* < 0.05 vs control.

its effect on  $I_{Ca}$  can be washed out by the change of medium. Further studies are needed to elucidate the mechanisms underlying the inhibitory effect of U-73122 on VDCC.

U-73122 has additional effects that independent of PLC inhibition. In rat liver microsomes(6) and rabbit pancreatic acinar cells(15) U-73122 releases  $Ca^{2+}$  through inhibition of  $Ca^{2+}$ -ATPase on the ER. However only high concentrations of U-73122 ( $>10\mu mol L^{-1}$ ) increase  $Ca^{2+}$  release noticeably. These concentrations exceed the dose range for inhibiting PLC-mediated  $Ca^{2+}$  release(6,15).

In conclusion U-73122 inhibits VDCC in RINm5F cells and porcine myometrial cells. It is likely that the inhibition of VDCC by U-73122 may not be attributed to its inhibitory effect on PLC. Further studies are needed to determine whether the inhibitory effect of U-73122 on PLC contributes to its inhibition of VDCC. Because the concentration range for producing the inhibitory effect on VDCC is similar to that for inhibition of PLC activation caution must be exercised in interpreting the results from studies of PLC-mediated changes in  $Ca^{2+}$  channel activity.

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U-73122  
RINm5F

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Bay-K-8538 ; ; Fura-2 ;  
; ; C; U-73122

U-73343  
: U-73122

: U-73122 RINm5F

KCl Bay-K-44  
U-73122  
RINm5F C  
U-73122 U-73343 50 %.  
: U-73122 RINm5F