# Effect of morphine on interleukin-1 and tumor necrosis factor \alpha production from mouse peritoneal macrophages in vitro1

BIAN Tong-Hua, WANG Xiao-Feng, LI Xiao-Yu<sup>2</sup> (Shanghai Institute of Materia Medica, Chinese Academy of Sciences, Shanghai 200031, China)

AIM: To study the influence of morphine on lymphokine production from mouse peritoneal macrophages  $(PM \varnothing)$ . METHODS: The productions of interleukin-1 (IL-1) and tumor necrosis factor (TNF-a) from PMO under various concentrations of morphine with or without naloxone were measured by testing thymocyte proliferation and L929 cell lysis in vitro. **RESULTS**: Both morphine  $(1 \times 10^{-8} 1 \times 10^{-3} \,\mathrm{mol} \, \mathrm{L}^{-1}$ ) and naloxone (50  $\mu\mathrm{mol} \, \mathrm{L}^{-1}$ ) inhibited IL-1 production from mouse PM Ø primed by sodium thioglycolate. Naloxone did not block the inhibitory effect of morphine. TNF-a production from PMO was inhibited by only high concentration (1 mmol L<sup>-1</sup>) of morphine which was not blocked by CONCLUSION: Morphine had a naloxone. direct inhibitory effect on PM Ø, which was not mediated by opioid receptor of PMO.

KEW WORDS morphine; naloxone; interleukin-1; tumor necrosis factor

Addiction to morphine is accompanied by an increased incidence of infections (1). Reticuloendothelial system plays an important role in the host defense function. Morphine drastically reduces reticuloendothelial system activity and then exacerbates infections. This paper is to study the direct effect of morphine on the function of peritoneal macrophages (PMO) in vitro.

Received 1994-11-15 Accepted #995-05-12

### MATERIALS AND METHODS

Morphine was produced by Qinhai Reagents Pharmaceutics and recrystallized by Dr YlN Dun-Xiang, Department of Synthetic Chemistry of our Institute. Naloxone, lipopolysaccharides (LPS), dactinomycin (Dac), and deoxyribonucleases (DNase) were from Sigma Co. Crystal violet (AR) was from Shanghai Chongming Chemical Reagent Factory. [H]TdR was from Shanghai Institute of Nuclear Research, Chinese Academy of Sciences. RPMI 1640 medium (Gibco Co) was supplemented with HEPES buffer 10 mmol L-, penicillin 100 kU L-1, streptomycin 100 mg L<sup>-1</sup>, L-glutamine 2 mmol L<sup>-1</sup>, 2-mercaptoethanol 50 mol L-1, and 10 % newborn bovine serum, pH 7.2.

Mice ICR mice,  $\frac{c}{2}$ , weighing 18-22 g,: BALB/c mice, \(\frac{1}{4}\), weighing 16-20 g, were purchased from Shanghai Animal Center, Chinese Academy of

Cell line L929 cell line (tumorigenic murine fibroblast) was purchased from Shanghai Institute of Cell Biology, Chinese Academy of Sciences. The cell line was subcultured with 10 % newborn bovine serum RPMI 1640 medium.

Production of interleukin-1 (IL-1) by PM Ø 133 PMØ were obtained from mice ip injected with 10 % thioglycolate broth (Difco, I mL/mouse, 4 d later). The perironeal cavities were lavaged with RPMI 1640 media. The cells were washed 3 times in RPMI 1640 media, placed in 24-well plates at 2 × 106 cells/well, and allowed to adhere at 37 °C in a 5 % CO2 for I h. Nonadherent cells were washed out. IL-1 was induced with LPS (5 mg L-1) from PM Ø. Indometacin (10 mg L-1) and morphine (1×10-6-1×  $10^{-1}$  mol  $L^{-1}$ ) with or without naloxone (50  $\mu mol$ L<sup>-1</sup>) were added, then the plates were incubated at 37 U in 5 %: CO2 atmosphere for 24 h. After incubation, the supernatant was collected by centrifugation (600  $^\circ$  g, 10 min) and stored at -25 C until assay.

<sup>1</sup> Project supported by the Fund for the 8th Five-Year Plan Key Project of Chinese Academy of Sciences, № S852015.

Correspondence to Prof Ll Xiao-Yu.

IL-1 assay IL-1 activity was evaluated by thymocyte proliferation assay. Thymocytes  $(1.5 \times 10^6 \text{ cells/well})$  isolated from BALB/c mice, were cocultured with PM $\odot$  supernatant in 96-well microtiter plates for 72 h in the presence of Con A  $(5 \text{ mg L}^{-1})$ . The cultures were added with [ $^3$ H]TdR (24.5 kBq/well) for the final 6-h incubation and harvested onto glass fiber filter paper. IL-1 activity was evaluated by [ $^3$ H]TdR uptake.

BIBLID: ISSN 0353-9756

Tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ) production <sup>[4,5]</sup>. The PM  $\otimes$  (1 mL, 2 × 10<sup>5</sup> cells/well) was cultured with LPS (10 mg L<sup>-1</sup>). Morphine (1 × 10<sup>-4</sup> – 1 × 10<sup>-3</sup> mol L<sup>-1</sup>) with or without naloxone (50  $\mu$ mol L<sup>-1</sup>) was added in 24-well plate at 37 °C in a 5 % CO<sub>2</sub> incubator for 6 h, each sample duplicated. After incubation, all the supernatants containing TNF- $\alpha$  were collected by centrifugation (600 × g, 10 min). The supernatants were stored at -25 °C until assay.

TNF- $\alpha$  assay The TNF- $\alpha$  activity was determined by the cytotoxocity assay against L929 cells. L929 cells (100  $\mu$ L,  $5 \times 10^4$  cells/well) were seeded into the 96-well plate and incubated at 37 C in 5 % CO<sub>2</sub> atmosphere for 24 h. Culture medium was removed. TNF- $\alpha$  supernatant (100  $\mu$ L) and RPMI 1640 medium containing Dac (final concentration 1 mg L<sup>-1</sup>) were added. The plates were reincubated for 20 h. The cells were stained with 0.5 % crystal violet (containing 3.2 % formaldehyde isotonic solution) for 15 min. The plates were rinsed and dried. The residue was dissolved in 47.5 % ethanol (100  $\mu$ L/well) which was then measured at 570 nm on an ELISA autoreader. TNF- $\alpha$  activity was calculated: Cytotoxicity (%) =

(Control - Test)/Control < 100 %.

Effect of morphine on L929 cell activity As TNF- $\alpha$  assay. L929 cells (100  $\mu$ L, 5 × 10<sup>4</sup> cells/well) were seeded into the 96-well plate. After 24-h incubation at 37 °C in 5 % CO<sub>2</sub> atmosphere, Dac (final concentration 1 mg L<sup>-1</sup>) and morphine (1 × 10<sup>-3</sup> – 1 × 10<sup>-4</sup> mol L<sup>-1</sup>) were added. L929 cells activity was measured by ELISA autoreader after 20-h reincubation.

# RESULTS

Morphine  $(1 \times 10^{-8} - 1 \times 10^{-3} \text{ mol } L^{-1})$  inhibited IL-1 production from PM  $\varnothing$ . Naloxone (50  $\mu$ mol  $L^{-1}$ ) also inhibited it and did not block the inhibitary effect of morphine (Tab 1).

Morphine had no influence on TNF- $\alpha$  production of PM  $\varnothing$  at low concentration (1 ×  $10^{-8}-1\times10^{-5}$  mol L<sup>-1</sup>) while 1 mmol L<sup>-1</sup> inhibited TNF- $\alpha$  production obviously and was not blocked by naloxone (50  $\mu$ mol L<sup>-1</sup>). Moreover, morphine  $1\times10^{-8}-1\times10^{-4}$  mol L<sup>-1</sup> had no influence on L929 activity (Tab 1).

## DISCUSSION

The present study demonstrated that in vitro morphine  $(1 \times 10^{-8} - 1 \times 10^{-3} \text{ mol } L^{-1})$  inhibited IL-1 production, and morphine 1

Tab 1. Effects of morphine on L929 cell activity (n=4 wells/sample). LPS (6. 25 mg L<sup>-1</sup>)-induced IL-1 activity (dpm/2 × 10° thymocytes) and TNF- $\alpha$  production (L929 cell specific lysls  $\frac{9}{10}$ ) of peritoneal macrophages with or without naloxone (50  $\mu$ mol L<sup>-1</sup>). n=3 wells for 1 homogenate (pooled from 10 mice),  $\overline{x}\pm s$ .  $^{4}P>0.05$ .  $^{4}P<0.05$ ,  $^{4}P<0.01$  vs control group.

Morphine/ mol L <sup>-1</sup>	L929 cell activity As70 ma	IL-1 activity/10 <sup>-4</sup> > dpm		TNF-α activity/specific lysis %	
		— naloxone	+naloxone	—naloxone	+naloxone
O	0.53±0.06	7. 0±1. 2	1.9±0.5	53.8±2.6	51.3±2.6
10 '9	$0.53 \pm 0.10$	$3.6 \pm 0.8^{b}$	$1.8 \pm 0.5$	53.8±2.6"	48. $7\pm 2.6^{\circ}$
$10^{-7}$	$0.53 \pm 0.13$ "	3. $4 \pm 0$ . 4'	$1.7 \pm 0.1$	59.0±2.6	51.3±2.6
10-1	0. $54 \pm 0.04$ °	$3.1\pm1.2^{b}$	$2.1 \pm 0.5^{\circ}$	56.4±2.6°	46.1±7.7
10-,	0.50±0.02°	3.2±0.7	2.4=0.4"	55, $1\pm 2$ , 6'	46.1±2.6
10~4	$0.55 \pm 0.06$ °	_		_	+0.1 <u>1</u> 2.0
10-:	<u> </u>	1. $8 \pm 0$ . $2^r$	1.3±0.4	23. $1 \pm 7$ . 7'	7.7±5.1°

mmol L $^{-1}$  also inhibited the TNF- $\alpha$  production from mouse PMO induced by LPS. Since IL-1 and TNF-α are important cytokines produced by PM O and play key roles in immune responses15.71, our results suggested that the direct inhibitory effect of morphine on the function of macrophages might be one of the mechanisms for the increase of incidence of infection in morphine addicts.

Our previous work (to be published) had 449 - 45 1 demonstrated that the effects of morphine on Con A-induced T-lymphocyte proliferation were blocked by naloxone (50  $\mu$ mol L<sup>-1</sup>), a  $\mu$ opioid receptor antagonist. Now the inhibitory effects of morphine on the IL-1 and TNF-α production of PM Ø were not antagonized by naloxone 50 µmol L<sup>-1</sup>, which alone inhibited IL-1 production. This suggested that unlike the effect on T-lymphocyte, the inhibitory effect of morphine on PMØ was not mediated by opioid receptor.

#### REFERENCES

- J McDonough RJ, Madden JJ, Falek A, Shafer DA, Pline M. Gordon D. et al. Alteration of T and null lymphocyte frequencies in the pempheral blood of human opiate addicts; in vivo evidence for opiate receptor sites on lymphocytes. J Immunol 1980; 125: 2539-43.
- 2 Tubaru E. Borelli G. Croce C. Cavallo G. Santiangeli C. Effect of morphine on resistance to infection. J Infect Dis 1983; 148: 656-66.
- 3 Yang SX. Li XY. A simple and practical method for IL-I assay. Shanghai J Immunol 1989: 9: 363-5.
- Flick DA, Gifford GE. Comparison of in vitro cell cytotoxic assays for tumor necrosis factor. J Immunol Methods 1984: 68: 167-75.

- Kiener PA, Marek F, Rodgers G, Lin PF, Warr G. Desiderio J. Induction of tumor necrosis factor, IFN-7, and acute lethality in mice by toxic and non-toxic forms of lipid A. J Immunol 1988; 141: 870-4.
- Dinarello CA. Interleukin-I and its biologically related cytokines. In; Cohen S. editor. Lymphokines and the immune response.
  - Boca Raton: CRC Press, 1990; 145-79.
- 7 Beutler B. Cerami A. The biology of cachectin/TNF a primary mediator of the host response.

Annu Rev Immunol 1989; 7: 625-55.

吗啡对腹腔巨噬细胞产生白细胞介素-1 及肿瘤坏死因子α的体外影响

卞同华, 王晓风, 李晓玉 (中国科学院上海药物研究所,上海200031,中国)

目的:研究吗啡对腹腔巨噬细胞产生细胞因子 的体外作用. 方法:通过测定胸腺细胞的增殖 及 L929细胞的杀伤率观察存在或不存在纳洛 酮的条件下不同浓度的吗啡对腹腔巨噬细胞产 生 IL-1及 TNF-α 的影响. 结果: 吗啡(1× 10-8-1×10-3 mol L-1)及纳洛酮(50 μmol L-1)都能抑制由巯基乙酸钠启动的腹腔巨噬细 胞产生 IL-1, 但纳洛酮不能阻断吗啡的上述抑 制作用. 对腹腔巨噬细胞 TNF-α 的产生,只 有高浓度的吗啡(1 mmol L-1)具有抑制作用, 也不能被纳洛酮阻断. 结论: 吗啡能直接抑制 腹腔巨噬细胞的功能,而这种作用不是由腹腔 巨噬细胞上阿片受体介导的.

关键词 吗啡: 肿瘤坏死因子