

Inhibition of 11 β -hydroxysteroid dehydrogenase in guinea pig kidney by three bioflavonoids and their interactions with gossypol

WANG Mao-Shan¹, SHI Hong, WANG Ke-Shen, Marcus M REIDENBERG²

(Department of Biochemical Pharmacology, Family Planning Research Institute, Zhejiang Academy of Medical Sciences, Hangzhou 310013, China)

KEY WORDS drug interactions; flavones; gossypol; hydroxysteroid dehydrogenases; hypokalemia

ABSTRACT

AIM: To study the effects of some bioflavonoids on the gossypol-induced hypokalemia. **METHODS:** The 11 β -hydroxysteroid dehydrogenase (11 β -OHS) protein was prepared from guinea pig kidney. The activity of 11 β -OHS with NAD as the coenzyme was measured by HPLC. The drug interaction was analysed by isobolographic method. **RESULTS:** The 11 β -OHS can be inhibited by some bioflavonoids. The IC₅₀ (95 % confidence limits) values were: quercetin 164 (79-341) μ mol/L, morin 913 (385-2173) μ mol/L, and naringenin 2193 (1114-4315) μ mol/L. When the 11 β -OHS was treated with quercetin, tangeretin, morin, naringenin plus gossypol, the combination index (CI) values were 0.92, 0.85, 0.98, and 1.01 respectively. **CONCLUSION:** The interaction of some bioflavonoids with gossypol might be one of the factors for gossypol-induced hypokalemia.

INTRODUCTION

The 11 β -hydroxysteroid dehydrogenase (11 β -OHS, EC 1.1.1.146), a critical enzyme to control the level of glucocorticoid in kidney, was studied vastly in recent twenty years^[1]. Its activity is lower in cases of apparent mineralocorticoid excess that the hypokalemia and paralysis is mainly clinical symptoms^[2,3].

The antifertility activity of gossypol for male was

first found by Chinese scientists^[4]. It is not used in clinical because it induces hypokalemia in some subjects who take the gossypol as contraceptive for a long period of time^[5,6]. It is proved that gossypol has a potent competitive inhibitory effect on the enzyme activity of 11 β -OHS^[7].

Bioflavonoid is a large category of compounds and they are naturally present in the diet and medical plant widely. We know there are similar structures between gossypol and bioflavonoids, and the inhibitory activity of some bioflavonoids on 11 β -OHS *in vitro* was reported^[8]. In this paper, we tried to determine whether there were interactions between these compounds and gossypol on the enzyme.

MATERIALS AND METHODS

Chemicals and solutions Gossypol acetate (purity >99 %) was a gift from Shanghai Institute of Materia Medica, Chinese Academy of Sciences. Cortisol, cortisone, corticosterone, quercetin, morin, naringenin, and NAD were purchased from Sigma Co (USA), and tangeretin was a gift from Prof M REIDENBERG (Cornell University Medical College, USA). Gossypol acetate and bioflavonoids were dissolved in methanol. NAD (5 mmol) was dissolved in Tris-HCl buffer (0.1 mol/L, pH 8.0) before use.

Preparation of enzyme protein The kidneys were collected from 30 δ Hartley guinea pigs (Zhejiang Laboratory Animal Center, Certificate number 96001) weighing 300-350 g. The kidney cortex microsome was prepared^[7].

Enzyme reaction and extraction of steroids

Enzyme activity of 11 β -OHS was measured and steroid was extracted as described previously^[9], except that in these experiments the compounds evaluated were added to the mixture before cortisol.

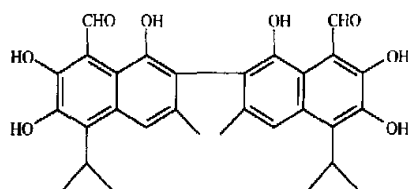
Steroid measurement by HPLC^[9] A ZORBAX Bondclone 10 C18 column (250 mm \times 4.6 mm,

¹ Correspondence to Dr WANG Mao-Shan. Phn 86-571-8807-6765, ext 8156. Fax 86-571-8807-5447.

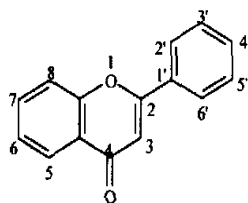
² Now in Department of Pharmacology and Medicine, Cornell University Medical College, 1300 York Ave, New York, NY 10021, USA.

Received 2001-03-05

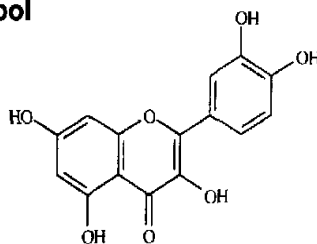
Accepted 2001-10-14



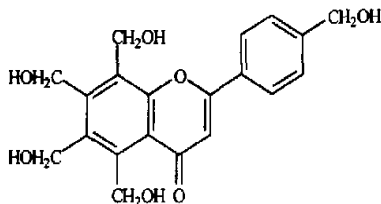
Gossypol



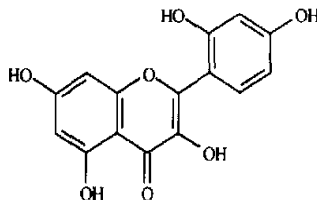
Flavone



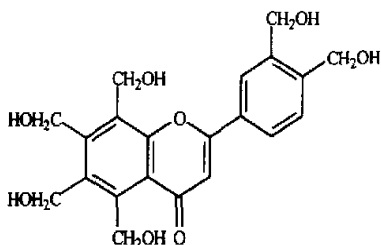
Quercetin



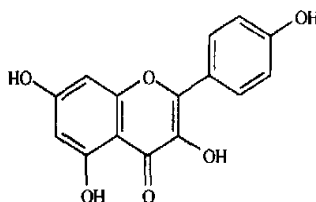
Tangeretin



Morin



Nobiletin



Naringenin

DuPont Co, USA) was used at 30 °C for all assays in a CTO-10A column oven. Components of the HPLC assembly (SHIMADZU Co, Japan) included a 7125 injector, a SPE-10A UV-vis detector, and LC-10AD liquid chromatograph pump. Output from the absorbance unit was recorded with a C-R 6A recorder. A wavelength of 245 nm and a maximal sensitivity of 0.005 absorbance units, full scale, were used. The mobile phase consisted of methanol-water (3 : 2, v/v). The retention time (RT) for cortisol, cortisone, and corticosterone was 8.55, 7.24, and 13.20 min, respectively at a flow rate of 1.0 mL/min. The coefficient variation of cortisol assay was 0.79 % using 8 simultaneous replicate

samples when cortisol 5.5 nmol was incubated with boiled microsomes. The recovery rates by methylene chloride extraction were 102.5 %, 98.1 %, 99.3 %, and 96.1 % for cortisol and 117.3 %, 115.2 %, 117.7 %, and 102.8 % for cortisone when cortisol 1.10, 3.31, 5.52, and 11.03 μmol and cortisone 1.11, 3.33, 5.55, and 11.10 μmol were added into 40 μL microsomes homogenate, respectively.

Analysis of data For determining the two-drug interaction of bioflavonoids with gossypol in inhibiting 11β-OHSD, the mixture of these compounds was made at a constant ratio of 3/4, 1/2, 1/4 of IC₅₀ values of gossypol with 1/4, 1/2, 3/4 of IC₅₀ values of

bioflavonoids, respectively. Isobolographic method was used⁽¹⁰⁾ and the Combination Index (CI) values was calculated based on the median-effect equation:

$$CI = [D]_1/[DX]_1 + [D]_2/[DX]_2$$

CI values of < 1, 1, and > 1 indicate synergism, additivity, and antagonism, respectively. $[DX]_1$ and $[DX]_2$ in the denominators are the concentrations (drug 1 and drug 2, respectively) that are required to produce X % effect alone^(11,12).

RESULTS

Effects of bioflavonoids on 11 β -OHSD *in vitro* The 11 β -OHSD from guinea pig kidney cortex could be inhibited by tested compounds *in vitro* when NAD as the coenzyme and cortisol as the substratum. The IC₅₀ values of gossypol, quercetin, morin, and naringenin alone were shown in Tab 1. The sequence for inhibiting 11 β -OHSD in IC₅₀ values was:

gossypol > quercetin > morin > naringenin

Interaction of bioflavonoids with gossypol in inhibiting 11 β -OHSD *in vitro* The results of two-

Tab 1. Effects of some bioflavonoids and gossypol on 11 β -OHSD.

Compounds	Concentration / $\mu\text{mol}\cdot\text{L}^{-1}$	Inhibition/%	IC ₅₀ (95 % CL)/ $\mu\text{mol}\cdot\text{L}^{-1}$
Gossypol	20.6	37.0 \pm 0.3	38.2 (11.8 - 123.0)
	41.3	51.3 \pm 0.5	
	82.6	66.6 \pm 0.7	
Quercetin	84	27.4 \pm 0.6	164 (79 - 341)
	169	52.4 \pm 1.0	
	338	74.0 \pm 1.0	
Morin	473	30.2 \pm 0.6	913 (385 - 2173)
	945	53.1 \pm 0.4	
	1891	70.0 \pm 0.9	
Naringenin	1049	24.6 \pm 0.7	2193 (1114 - 4315)
	2089	42.9 \pm 0.5	
	4177	76.4 \pm 0.7	

drug interaction of some bioflavonoids with gossypol were shown in Fig 1. The CI values of quercetin, tangeretin, morin, naringenin plus gossypol were 0.92, 0.85, 0.98, and 1.01 respectively. These results indicated that there were moderate or slight synergism or

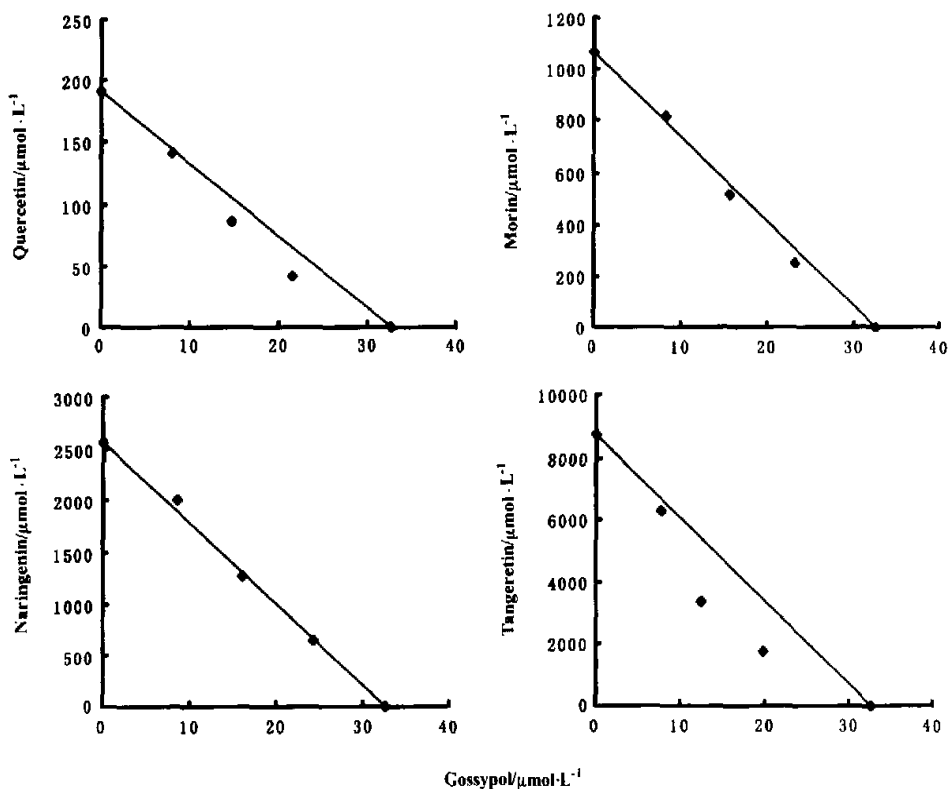


Fig 1. Isobologram for the interaction of some bioflavonoids with gossypol.

additive effects between gossypol and three bioflavonoids.

DISCUSSION

The previously studies have demonstrated that the 11β -OHSD can be inhibited by gossypol *in vivo* and *in vitro*^[7], the NAD-dependent 11β -OHSD that is localized in the distal renal tubules is a more critical physiologic mechanism in regulating the renal glucocorticoid levels^[9], and the gossypol caused the lower dosage of corticosteroid to elicit higher K^+ / Na^+ ratio in 24-h urine and lower serum potassium level^[13], and hypothesis remained that inhibition of NAD-dependent 11β -OHSD (11β -OHSD2) might be one of the mechanism of gossypol-induced hypokalemia.

The bioflavonoids widely existing in diet and medical plant is a large catalog of compounds. The previous and present study demonstrated that the 11β -OHSD can be inhibited by some bioflavonoids, and now we find there is synergism or additive effect when the enzyme was treated by these compounds with gossypol. From these results, we could ascribed the hypokalemia induced by gossypol to the bioflavonoid partially, and the interaction of some bioflavonoids with gossypol may be one of the factors for gossypol-induced hypokalemia. Whether variation in intake of dietary inhibitors of 11β -OHSD such as bioflavonoid in different geographic locations accounts for variation incident of gossypol-induced hypokalemia remains to be determined.

REFERENCES

- 1 Monder C, Funder J, editors. First Symposium on 11β -hydroxysteroid dehydrogenase. Steroid 1994; 59: 65-169.
- 2 Monder C, Shackleton CHL, Bradlow HL, New MI, Stoner E, Iohan F, *et al.* The syndrome of apparent mineralocorticoid excess; its association with 11β -dehydrogenase and 5β -reductase deficiency and some consequences for corticosteroid metabolism. J Clin Endocr Metab 1986; 63: 550-7.
- 3 Stewart PM, Krozowski ZS, Gupta A, Milford DV, Howie AJ, Sheppard MC, *et al.* Hypertension in the syndrome of apparent mineralocorticoid excess due to mutation of the 11β -hydroxysteroid dehydrogenase type 2 gene. Lancet 1996; 347: 88-91.
- 4 National Coordination Group on Male Antifertility Agents. Gossypol — a new antifertility agent for males. Chin Med J 1978; 4: 417-28.
- 5 Wang C, Yeung RTT. Gossypol and hypokalemia. Contraception 1985; 32: 237-52.
- 6 Qian SZ, Wang ZG. Gossypol: A potential antifertility agent

- for males. Ann Rev Pharmacol Toxicol 1984; 24: 329-60.
- 7 Sang GW, Lorenzo JB, Reidenberg MM. Inhibitory effects of gossypol on corticosteroid 11β -hydroxysteroid dehydrogenase from guinea pig kidney; a possible mechanism for causing hypokalemia. J Steroid Biochem Mol Biol 1991; 39: 169-76.
- 8 Song D, Lorenzo JB, Reidenberg MM. Inhibition of 11β -hydroxysteroid dehydrogenase by gossypol and bioflavonoids. J Lab Clin Med 1992; 120: 792-7.
- 9 Wang MS, Lorenzo JB, Reidenberg MM. NAD- and NADP-dependent 11β -hydroxysteroid dehydrogenase isoforms in guinea pig kidney with gossypol inhibition. Acta Pharmacol Sin 1997; 18: 481-5.
- 10 Gessner PK. The isobolographic method applied to drug interactions. In: Morselli PL, Cohen SN, Garattini S, editors. Drug interactions. New York: Raven Press; 1974. p 349-62.
- 11 Chou TC, Talalay P. Quantitative analysis of dose-effect relationship: The combined effects of multiple drugs or enzyme inhibitors. Adv Enzyme Regul 1989; 22: 27-55.
- 12 Chou TC. The median-effect principle and the combination index for quantitation of synergism and antagonism. In: Chou TC, Rideout D, editors. Synergism and antagonism in chemotherapy. San Diego: Academic Press; 1991. p 61-102.
- 13 Ge RS, Wang MS, Sang GW. The antinatriuretic and kaliuretic effects of gossypol acetic acid in combination with glucocorticoids in rat. Proceedings of the 2nd International Symposium on Fertility Regulation; 1995 May 21-25; Beijing, China. Beijing: National Research Institute for Family Planning; 1995. p 65.

三种生物黄酮类化合物及其与棉酚相互作用抑制豚鼠肾脏 11β -羟甾脱氢酶

R96 A

王茂山¹, 史红, 王恪申, Marcus M REIDENBERG (浙江省医学科学院计划生育研究所生化药理研究室, 杭州 310013, 中国)

关键词 药物相互作用; 黄酮类; 棉酚; 羟甾类脱氢酶类; 低钾血症

目的: 探讨三种生物黄酮类化合物在棉酚诱发低血钾中的作用。 **方法:** 从豚鼠肾脏制备 11β -羟甾脱氢酶, NAD 为辅酶, 加入待测化合物, 37°C 反应 1 小时后提取甾体, 反相高压液相测定酶活性, 热辐射图像法分析药物相互作用。 **结果:** 所试生物黄酮类化合物均能抑制 11β -羟甾脱氢酶, 其 IC_{50} (95% 可信限) 分别为: 槲皮素 $164 (79-341) \mu\text{mol/L}$, 桑色素

913 (385 - 2173) $\mu\text{mol/L}$, 柑桔素 2193 (1114 - 4315) $\mu\text{mol/L}$. 当 11 β -羟甾脱氢酶分别用槲皮素, 红桔素, 桑色素, 柑桔素加上棉酚处理时, 其 CI 值分别为: 0.92、0.85、0.98 和 1.01. **结论:** 三种生物黄酮类化合物与棉酚在抑制 11 β -羟甾脱氢酶中具有协同或相加作用. (责任编辑 朱倩蓉)

Papers are welcome

Acta Pharmacologica Sinica publishes monthly original researches on all life sciences, both experimental and clinical. Reviews based primarily on the author's own research of international importance with 3 - 10 key words are also welcome. Manuscripts in English of full-length articles from any part of the world are welcome.

The article should be prepared in accordance with the "Information for authors" in Acta Pharmacol Sin 2002 Jan; 23 (1); I - VII or the "Uniform requirements for manuscripts submitted to biomedical journals" in Ann Intern Med 1997 Jan 1; 126 (1); 36 - 47.

For more information please visit

<http://www.chinaphar.com>

Address: 294 Tai-yuan Road, Shanghai 200031, China

E-mail aps@mail.shcnc.ac.cn

Fax 86-21-6474-2629 or 86-21-6437-0269

Pbn 86-21-6474-2629 (direct) or 86-21-6431-1833, ext 200.