

# Effect of tetrandrine on free intracellular calcium in cultured calf basilar artery smooth muscle cells<sup>1</sup>

WANG Bin<sup>2</sup>, XIAO Ji-Gao

*Department of Pharmacology, Nanjing Medical University, Nanjing 210029, China*

**KEY WORDS** tetrandrine; calcium; basilar artery; vascular smooth muscle; cultured cells; caffeine; phenylephrine

## ABSTRACT

**AIM:** To study the effects of tetrandrine (Tet) on extracellular  $\text{Ca}^{2+}$  influx and intracellular  $\text{Ca}^{2+}$  release in cultured calf basilar artery smooth muscle cells. **METHODS:** Free intracellular calcium was examined by a system of measurement of AR-CM-MIC, using Fura 2-AM as a fluorescent indicator. **RESULTS:** In the presence of extracellular  $\text{Ca}^{2+}$  1.3 mmol/L, no significant effect of Tet on resting  $[\text{Ca}^{2+}]_i$  was found. KCl 20, 40, and 60 mmol/L triggered a sustained rise in  $[\text{Ca}^{2+}]_i$ , pretreatment with Tet inhibited the elevation of  $[\text{Ca}^{2+}]_i$  induced by KCl in concentration-dependent manner, Tet at high concentration (100  $\mu\text{mol/L}$ ) almost abolished the rise of  $[\text{Ca}^{2+}]_i$  evoked by KCl. Caffeine 10 mmol/L only produced a transient increase of  $[\text{Ca}^{2+}]_i$ , which spontaneously declined back to resting levels. Tet 10-30  $\mu\text{mol/L}$  had no effect on caffeine-induced  $[\text{Ca}^{2+}]_i$  transient peak. Tet at high concentration (100  $\mu\text{mol/L}$ ), however, reduced the  $[\text{Ca}^{2+}]_i$  transient peak induced by caffeine. Phenylephrine (PE) 10 mmol/L produced a rapid transient peak and a distinct sustained elevation in  $[\text{Ca}^{2+}]_i$  in the presence of extracellular  $\text{Ca}^{2+}$ . In the absence of extracellular  $\text{Ca}^{2+}$  containing egtazic acid (EGTA), PE only produced a rapid transient peak in  $[\text{Ca}^{2+}]_i$ . Pretreatment of Tet (10-100  $\mu\text{mol/L}$ ) inhibited the sustained elevation in  $[\text{Ca}^{2+}]_i$  induced by PE in a concentration-dependent manner. However, only 100  $\mu\text{mol/L}$  of Tet inhibited the transient peak in  $[\text{Ca}^{2+}]_i$  induced by PE both in the presence of extracellular  $\text{Ca}^{2+}$  1.3 mmol/L and in the absence of extracellular  $\text{Ca}^{2+}$  containing EGTA. **CONCLUSION:** Tet inhibited the  $\text{Ca}^{2+}$  influx from the extracellular site via voltage-activated  $\text{Ca}^{2+}$  channel and PE-receptor-operated  $\text{Ca}^{2+}$  channel. At a high concentration, Tet may inhibit the  $\text{Ca}^{2+}$  release from sarcoplasmic reticulum (SR) or refilling of intracellular calcium store in cerebral artery smooth muscle cells.

## INTRODUCTION

Tetrandrine (Tet) is the major active component in the root of *Stephania tetrandra* S. Tet has been used

for the treatment of various diseases, including hypertension, cardiac arrhythmia, angina and inflammation<sup>[1-3]</sup>. Recently, it was reported that Tet had a protective effect on the brain ischemia/reperfusion model in rats and gerbils<sup>[4,5]</sup>. A series of studies have revealed that Tet blocked  $\text{Ca}^{2+}$  channel in cardiac cells, neuroblastoma cells, neurohypophysial nerve terminals, and mesenteric artery, which largely accounted for its antihypertensive, antiarrhythmic and antimyocardial ischemia actions<sup>[1,3]</sup>. In addition to its direct effect on neurons, Tet may relax cerebral artery to protect from

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<sup>2</sup> Correspondence to Dr WANG Bin. Phn/Fax 86-25-666-2884. E-mail Binwang@njmu.edu.cn

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brain ischemia. However, the effect and mechanism of Tet on cerebral artery remain unclear. In the present study, we examined the effect of Tet on  $\text{Ca}^{2+}$  influx from extracellular space and  $\text{Ca}^{2+}$  release from sarcoplasmic reticulum (SR) or refilling of intracellular calcium store in cultured calf basilar artery smooth muscle cells.

## MATERIALS AND METHODS

**Materials** Fura 2-AM, dimethyl sulfoxide, caffeine and phenylephrine were purchased from Sigma Chemical Co. Fura 2-AM was dissolved in dimethyl sulfoxide. Tet was obtained from Jin Hua Pharmaceutical Factory, it was dissolved in HCl 0.1 mol/L, then diluted with D-Hanks' medium to desired concentration. Dulbecco's modified Eagle's medium (DMEM) was obtained from Gibco. The system of measurement of AR-CM-MIC was the product of Spex Co, USA.

**Cell culture** Calf basilar artery smooth muscle cells were cultured according to our previous method<sup>[6]</sup>. Briefly, the basilar artery of newborn calf (Nanjing Weigang Calf Serum Factory) was isolated rapidly. The connective tissue was removed under a dissecting microscope. The endothelium was removed first by erasing when the blood vessels were cut longitudinally, the pieces (2 mm<sup>2</sup>) of the blood vessels were placed endosurface downright in culture dishes and were incubated at 37 °C in a humidified incubator with 95 % air and 5 % CO<sub>2</sub> in DMEM with 10 % (v/v) fetal calf serum (FCS). Cells migrated from the blood vessels in about 7 d. When cells covered the culture dish, small pieces were removed and cells were washed with D-Hanks' solution twice, then incubated at 37 °C for 10 min in D-Hanks' solution with 0.125 % trypsin. The trypsinization was discontinued by adding ice-cold DMEM 10 mL containing 10 % (v/v) FCS. The cell pellets were resuspended in fresh medium with FCS to 1×10<sup>8</sup> cells/L and cultured. The medium was changed every 2-3 d. Cells reached confluence in about 5 d and then passaged every 5-6 d. The single cell was cultured in culture dish with a glass cover slip 2 d before experiment.  $[\text{Ca}^{2+}]_i$  was measured in passages 4 and 5.

**$[\text{Ca}^{2+}]_i$  measurement** Cells grown on glass coverslip were incubated with DMEM containing Fura 2-AM 3 μmol/L at 37 °C for 40 min. The loaded cells were washed twice with Hanks' solution or D-Hanks' solution. The fluorescence was recorded in a system of measurement of AR-CM-MIC at  $\lambda_{\text{ex}}=340$  nm/380

nm,  $\lambda_{\text{em}}=505$  nm, directly after the addition of an agonist.  $[\text{Ca}^{2+}]_i$  of single cell was calculated according to the formula:  $[\text{Ca}^{2+}]_i = K_d (sb_1/sb_2) [(R - R_{\text{min}})/(R_{\text{max}} - R)]$  nmol/L. Where, the dissociation constant  $K_d=224$  nmol/L at 37 °C.  $R$  was fluorescence intensity,  $R_{\text{max}}$  and  $R_{\text{min}}$  were obtained by the addition of ionomycin and egtazic acid respectively.

**Statistical analysis** Data were analyzed with *t*-test. Differences were considered significant when  $P < 0.05$ .

## RESULTS

**Effect of Tet on resting  $[\text{Ca}^{2+}]_i$**  The resting  $[\text{Ca}^{2+}]_i$  was (100±9) nmol/L in the presence of extracellular  $\text{Ca}^{2+}$  1.3 mmol/L. When 10, 30, and 100 μmol/L of Tet were added, the  $[\text{Ca}^{2+}]_i$  was (101±10), (98±8), and (97±10) nmol/L, respectively ( $P > 0.05$  vs 0 μmol/L of Tet). No significant effect of Tet on resting  $[\text{Ca}^{2+}]_i$  was found in cultured calf basilar artery smooth muscle cells.

**Effect of Tet on elevation of  $[\text{Ca}^{2+}]_i$  induced by KCl** KCl 20, 40, and 60 mmol/L triggered a sustained rise in  $[\text{Ca}^{2+}]_i$  without transient component in the presence of extracellular  $\text{Ca}^{2+}$  1.3 mmol/L. In the absence of extracellular  $\text{Ca}^{2+}$  containing egtazic acid, addition of KCl had no effect on  $[\text{Ca}^{2+}]_i$ . This result suggested that KCl increased  $[\text{Ca}^{2+}]_i$  via  $\text{Ca}^{2+}$  channel in the plasma membrane rather than in SR in this condition. Pretreatment with Tet inhibited the elevation of  $[\text{Ca}^{2+}]_i$  induced by KCl in a concentration-dependent manner. High concentration of Tet almost abolished the rise of  $[\text{Ca}^{2+}]_i$  evoked by KCl (Fig 1).

**Effect of Tet on transient elevation of  $[\text{Ca}^{2+}]_i$  induced by caffeine** Caffeine 10 mmol/L only produced a transient increase of  $[\text{Ca}^{2+}]_i$ , which spontaneously declined back to resting levels. To clarify whether the SR stores of  $\text{Ca}^{2+}$  were refilled completely, the repetitive addition and washout of caffeine was performed. When the interval of repetitive addition and washout of caffeine was 5 min, the transient peak in  $[\text{Ca}^{2+}]_i$  evoked by subsequent repetitive addition of caffeine declined, suggesting that the  $\text{Ca}^{2+}$  store in the SR was depleted. With the interval of washout of caffeine was increased to 25 min, the transient peak in  $[\text{Ca}^{2+}]_i$  evoked by repetitive addition of caffeine was kept stable, indicating that the  $\text{Ca}^{2+}$  store in the SR was refilled completely within 25 min. We used 25 min as the interval of washout. Tet 10-30 μmol/L had no effect on caffeine-induced  $[\text{Ca}^{2+}]_i$  transient peak. Tet at a high concentra-

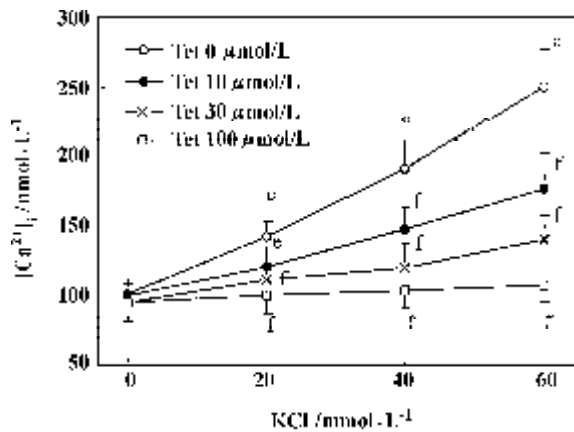


Fig 1. Effect of tetrandrine (Tet) on elevation of  $[Ca^{2+}]_i$  induced by KCl in the presence of extracellular  $Ca^{2+}$  1.3 mmol/L.  $n=6$ . Mean $\pm$ SD.  $^{\circ}P<0.01$  vs KCl 0 mmol/L.  $^{\circ}P<0.05$ ,  $^{\ast}P<0.01$  vs 0 mmol/L of Tet.

tion (100  $\mu$ mol/L), however, reduced the  $[Ca^{2+}]_i$  transient peak induced by caffeine (Fig 2).

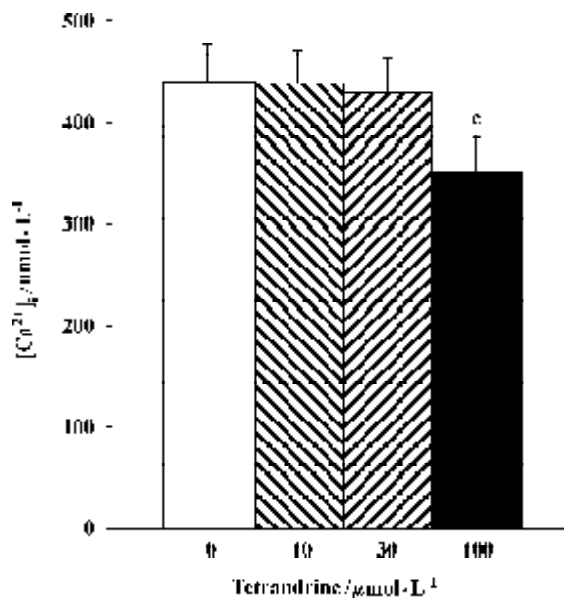


Fig 2. Effect of tetrandrine on transient elevation of  $[Ca^{2+}]_i$  induced by caffeine (10 mmol/L) in the presence of extracellular  $Ca^{2+}$  1.3 mmol/L.  $n=6$ . Mean $\pm$ SD.  $^{\circ}P<0.01$  vs 0 mmol/L of Tet.

**Effect of Tet on PE-induced elevation of  $[Ca^{2+}]_i$**   
PE 10  $\mu$ mol/L produced an increase in  $[Ca^{2+}]_i$ , consisting of a rapid transient peak and a distinct sustained component in the presence of extracellular  $Ca^{2+}$  1.3 mmol/L. In the absence of extracellular  $Ca^{2+}$  containing egtazic acid, PE only produced a rapid transient peak in  $[Ca^{2+}]_i$  without the sustained component, which indi-

cated that the rapid transient peak induced by PE represented  $Ca^{2+}$  release from  $Ca^{2+}$  store. To clarify whether the sustained elevation in  $[Ca^{2+}]_i$  induced by PE represented  $Ca^{2+}$  influx from extracellular site, caffeine was pretreated to deplete  $Ca^{2+}$  store. When PE was added 3 min after pretreatment of caffeine (20 mmol/L), only a sustained elevation in  $[Ca^{2+}]_i$  induced by PE was observed. These data verified that the rapid transient peak and sustained elevation in  $[Ca^{2+}]_i$  induced by PE represented  $Ca^{2+}$  release and  $Ca^{2+}$  influx respectively in cultured calf basilar artery smooth muscle cells.

In the presence of extracellular  $Ca^{2+}$  1.3 mmol/L, pretreatment of Tet (10-100  $\mu$ mol/L) inhibited the sustained elevation in  $[Ca^{2+}]_i$  induced by PE in a concentration-dependent manner and high concentration of Tet (100  $\mu$ mol/L) inhibited the transient peak in  $[Ca^{2+}]_i$  induced by PE (Fig 3). In the absence of extracellular  $Ca^{2+}$  containing egtazic acid, 10 and 30  $\mu$ mol/L of Tet showed no significant effect on the transient peak in  $[Ca^{2+}]_i$  induced by PE and 100  $\mu$ mol/L of Tet inhibited the transient peak in  $[Ca^{2+}]_i$  induced by PE (Fig 4).

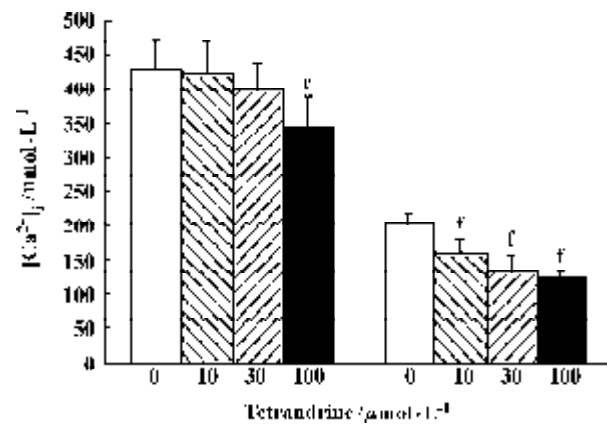


Fig 3. Effect of tetrandrine (Tet) on elevation of  $[Ca^{2+}]_i$  induced by phenylephrine (PE, 10 mmol/L) in the presence of extracellular  $Ca^{2+}$  1.3 mmol/L. Left: transient peak in  $[Ca^{2+}]_i$ , Right: sustained component in  $[Ca^{2+}]_i$ .  $n=6$ . Mean $\pm$ SD.  $^{\circ}P<0.01$  vs Tet 0 mmol/L in  $[Ca^{2+}]_i$  transient peak.  $^{\ast}P<0.01$  vs Tet 0 mmol/L in  $[Ca^{2+}]_i$  sustained component.

## DISCUSSION

The actions of Tet on a series of systems have been investigated, including cardiac cells, cardiac papillary muscle, aortic vessel, mesenteric artery, pulmonary artery, B cells, human leukaemic HL-60 cells, cornea and retina cells<sup>[3, 7-10]</sup>. However little has been reported on the effect of Tet on cerebral artery. Tet was

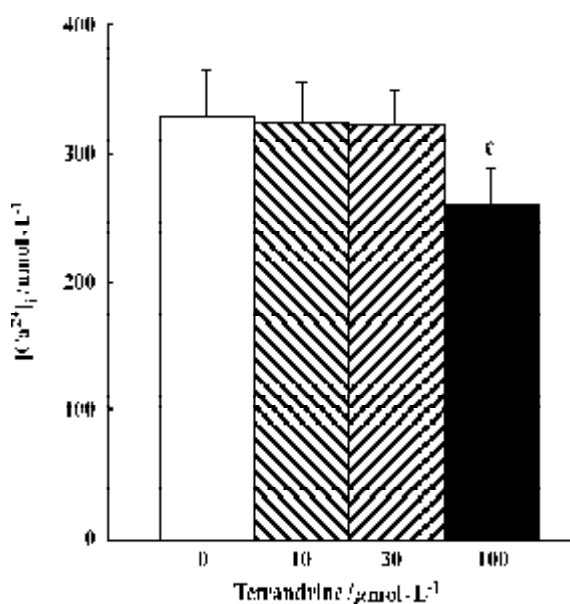


Fig 4. Effect of tetrandrine (Tet) on elevation of  $[Ca^{2+}]_i$  induced by phenylephrine (PE, 10 mmol/L) in the absence of extracellular  $Ca^{2+}$  containing egtazic acid.  $n=6$ . Mean $\pm$ SD. <sup>c</sup> $P<0.01$  vs 0 mmol/L of Tet.

reported to decrease brain injury on brain ischemia/reperfusion model in rats and gerbils<sup>[4,5]</sup>. Neural tissues are highly dependent on glucose and oxygen supplies, an adequate continuous blood supply to the brain is essential to ensure the normal function of the brain. In addition to its direct effect on neurons, Tet was able to inhibit pial artery constriction induced by KCl and PE in rats (unpublished data), which may partially account for its protective effect on cerebral ischemia. However, the effect and mechanism of Tet on cerebral artery remain unclear. In the present study, we demonstrated that Tet blocked voltage-activated  $Ca^{2+}$  channel and PE receptor-gated  $Ca^{2+}$  channel on the plasma membrane in a concentration-dependent manner and Tet at a high concentration may inhibited  $Ca^{2+}$  release or  $Ca^{2+}$  uptake via caffeine- and PE-sensitive  $Ca^{2+}$  store in cerebral basilar artery smooth muscle cells in rats.

Tet relaxed the high  $K^+$ -induced constrictions in isolated arteries, including pulmonary, coronary, renal arteries<sup>[3,11]</sup>. Electrophysiological study revealed that Tet declined a high-voltage-threshold, long-lasting (L-type)  $Ca^{2+}$  channel current in GH3 anterior hypophysial cells, cardiac ventricular muscles, which demonstrated that Tet was an L-type  $Ca^{2+}$  channel blocker<sup>[3,12]</sup>. On the cerebral basilar artery smooth muscle cells (this study), KCl only produced a sustained increase in  $[Ca^{2+}]_i$  without the transient component, indicating that KCl

increased  $Ca^{2+}$  entry but did not influence the  $Ca^{2+}$  store. Tet inhibited markedly KCl-induced rise in  $[Ca^{2+}]_i$  and high concentration of Tet almost abolished the rise of  $[Ca^{2+}]_i$ , suggesting that Tet acted as a voltage-activated  $Ca^{2+}$  channel antagonist on cerebral artery smooth muscle cells.

Kwang *et al*<sup>[13]</sup> reported that Tet inhibited the binding of  $^3H$ -prazosin (an  $\alpha_1$ -adrenoceptor antagonist) to dog aortic muscle plasmalemma-rich membranes and Tet was shown as an  $\alpha_1$ -adrenoceptor antagonist. In addition to inhibiting voltage-activated  $Ca^{2+}$  channel, Tet could inhibit receptor-dependent  $Ca^{2+}$  channel, including PE-, norepinephrine-, histamin-, glutamate-sensitive  $Ca^{2+}$  channel on plasma membrane<sup>[9,10]</sup>. In the present study, Tet concentration-dependently inhibited PE-induced sustained rise in  $[Ca^{2+}]_i$  in  $Ca^{2+}$ -containing medium, indicating that Tet inhibited  $Ca^{2+}$  influx via PE-sensitive  $Ca^{2+}$  channel in cerebral artery smooth muscle cells. Considering the system difference, whether Tet directly interacted with  $\alpha_1$ -adrenoceptor on cerebral artery will require further studies.

The results of studies of Tet on  $Ca^{2+}$  release or  $Ca^{2+}$  uptake sensitive to caffeine are still confusing: Tet facilitated the contraction induced by caffeine in rabbit aortic strip<sup>[14]</sup>, however, elicited no effect on caffeine-induced transient contraction in rat aortic rings<sup>[13]</sup>, it has been also shown that Tet reduced the magnitude of the caffeine-induced  $[Ca^{2+}]_i$  elevation in rat cardiomyocyte<sup>[15]</sup> and the refilling of intracellular calcium stores sensitive to caffeine was inhibited by Tet in rat aortic strips<sup>[16]</sup>. The differences of experimental procedures or preparations used might in part account for the results.

The intracellular  $Ca^{2+}$  store depletion by caffeine elicited  $Ca^{2+}$  entry from extracellular space via specific channels or by passive diffusion or exchange with other ions and intracellular  $Ca^{2+}$  can be uptaken by  $Ca^{2+}$  store. Tet inhibited the  $Ca^{2+}$  uptake to SR in rat cardiomyocyte and the refilling of intracellular calcium stores in rat aortic strips<sup>[15-17]</sup>, Tet might influence the refilling of intracellular calcium stores via affecting plasma membrane channels indirectly or via affecting intracellular  $Ca^{2+}$  store directly, the exact mechanisms remain unclear.

Caffeine inhibited cyclic nucleotide phosphodiesterases and stimulated  $Ca^{2+}$  release<sup>[18]</sup>. The present study showed that the  $Ca^{2+}$  storage sites sensitive to caffeine can not be completely refilled within 5 min with 1.3 mmol/L extracellular  $Ca^{2+}$ . When caffeine was added

again after incubation period of 25 min in extracellular  $\text{Ca}^{2+}$  1.3 mmol/L, the magnitude of  $[\text{Ca}^{2+}]_i$  transient was similar to that of the first one, which indicated that  $\text{Ca}^{2+}$  store was refilled completely. We used 25 min as the interval of washout, and found that 100  $\mu\text{mol/L}$  of Tet had the inhibitory effect on the transient peak in  $[\text{Ca}^{2+}]_i$  evoked by caffeine. Although we can not explain the exact mechanisms, these results also implied that Tet might inhibit refilling of intracellular calcium store in cerebral artery smooth muscle cells.

PE elicited a rapid transient peak accompanied with a more sustained increase in  $[\text{Ca}^{2+}]_i$  in  $\text{Ca}^{2+}$ -containing medium. But in absence of  $\text{Ca}^{2+}$ , only rapid transient peak was induced by PE. After caffeine depleted the SR  $\text{Ca}^{2+}$  store, subsequent addition of PE only elicited a sustained rise in  $[\text{Ca}^{2+}]_i$  without the transient component. These data implied that the caffeine-inducible  $\text{Ca}^{2+}$  pool and PE-inducible  $\text{Ca}^{2+}$  pool might overlap in cerebral artery smooth muscle cells. Tet inhibited the transient peak in  $[\text{Ca}^{2+}]_i$  induced by PE both in presence and absence of  $\text{Ca}^{2+}$ . These result indicated that Tet at a high concentration might inhibit  $\text{Ca}^{2+}$  release or  $\text{Ca}^{2+}$  uptake via caffeine- or PE-inducible  $\text{Ca}^{2+}$  pool. The exact mechanism of Tet on  $\text{Ca}^{2+}$  release or  $\text{Ca}^{2+}$  uptake in cerebral artery smooth muscle cells will warrant further investigation.

In summary, Tet inhibited the  $\text{Ca}^{2+}$  influx from the extracellular site via voltage-activated  $\text{Ca}^{2+}$  channel and PE-receptor-operated  $\text{Ca}^{2+}$  channel. At high concentration, Tet may inhibit the  $\text{Ca}^{2+}$  release from SR or refilling of intracellular calcium store in cerebral artery smooth muscle cells.

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