

## Effects of allitridi on intracellular $\text{Ca}^{2+}$ concentration in isolated rat brain cells

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**KEY WORDS** brain; cultured cells; allitridi; intracellular fluid; calcium; Fura-2; norepinephrine; sodium glutamate

### ABSTRACT

**AIM:** To study actions of allitridi extracted from garlic on intracellular calcium in isolated rat brain cells. **METHODS:** Brain cells were isolated from newborn rat brain with Fura 2-AM measurements of intracellular  $\text{Ca}^{2+}$  concentration ( $[\text{Ca}^{2+}]_i$ ). **RESULTS:** Allitridi 1 - 100  $\mu\text{mol} \cdot \text{L}^{-1}$  concentration-dependently blocked increases of  $[\text{Ca}^{2+}]_i$  caused by potassium chloride and sodium glutamate (Glu) with  $\text{IC}_{50}$  of 59.7 and 69.9  $\mu\text{mol} \cdot \text{L}^{-1}$  respectively. Allitridi 100  $\mu\text{mol} \cdot \text{L}^{-1}$  blocked norepinephrine (Nor)-induced  $[\text{Ca}^{2+}]_i$  elevation. **CONCLUSION:** Allitridi is an effective agent for blocking the  $[\text{Ca}^{2+}]_i$  increase caused by potassium chloride, Nor and Glu.

### INTRODUCTION

Allitridi extracted from *Allium sativum* L. has traditionally been used for the treatment of infections and cardiovascular diseases, etc.<sup>[1]</sup>. Allitridi ameliorated cerebral infarction, especially in atherosclerotic cerebral arteries<sup>[2]</sup>. We previously showed that allitridi protected cerebral ischemia in animals<sup>[3]</sup>, and it improved the rat cerebral microcirculation. Allitridi might be a calcium antagonist<sup>[4]</sup>. The present study was to explore the mechanism of  $[\text{Ca}^{2+}]_i$  reduction induced by allitridi.

### MATERIALS AND METHODS

**Materials** Allitridi was purchased from Tianqing Pharmaceutical Factory (Jiangsu Province). Ionomycin, trypsin, norepinephrine bitartrate (Nor), sodium glutamate (Glu), and Fura 2-AM were obtained from Sigma Chemical Co (St Louis MO).

**Cell isolation** Brain cells were isolated as described<sup>[5]</sup> with some modifications. Newborn (1-3 d) Sprague Dawley rats (Jiangsu Laboratory Animal Center) were decapitated in a Petri dish on ice. The isolated brain was rinsed with ice-cold  $\text{Ca}^{2+}$ - and  $\text{Mg}^{2+}$ -free Hanks' solution containing NaCl 137, KCl 5, glucose 5.6, and HEPES 10  $\text{mmol} \cdot \text{L}^{-1}$  (pH 7.2-7.4). Having been washed with  $\text{Ca}^{2+}$ - and  $\text{Mg}^{2+}$ -free Hanks' solution, the brain was cut into 3-mm<sup>3</sup> piece, and placed in a 10-mL flask containing 0.125 % trypsin and egtazic acid 0.5  $\text{mmol} \cdot \text{L}^{-1}$  in  $\text{Ca}^{2+}$ - and  $\text{Mg}^{2+}$ -free Hanks' solution. The flask was shaken at 37 °C for 20 min. Trypsinization was discontinued by adding ice-old Hanks' solution 10 mL containing 10 % bovine serum. Tissue pieces were mechanically dissociated by gently triturating 10-15 times with a polished pipette. The isolated brain cells were filtered through nylon mesh (200 mesh, hole width 95  $\mu\text{m}$ ) and collected in a flask. Cells were centrifuged twice at 100 × g for 3 min each. The supernatant was decanted and the cells were resuspended in Hanks' solution containing NaCl 137,  $\text{CaCl}_2$  1.3,  $\text{MgCl}_2$  0.5, KCl 5, glucose 5.6, and HEPES 10  $\text{mmol} \cdot \text{L}^{-1}$  (pH 7.4). Trypan blue staining showed a 90 % - 95 % cellular viability rate. The cell suspension was further diluted to 8 mL with Hanks' solution and placed at 37 °C for 5 min. A final concentration of Fura 2-AM 5  $\mu\text{mol} \cdot \text{L}^{-1}$  dissolved in  $\text{Me}_2\text{SO}$  was added. The cells were loaded with Fura 2-AM in water bath for 30 min. The Fura 2-AM-loaded cells were centrifuged at 100 × g for 30 min. The cells were resuspended in Hanks'

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solution 4 mL containing 0.2 % bovine serum albumin resulting in approximately  $5 \times 10^7$  cells  $\cdot$  L<sup>-1</sup> which were stored in ice. Prior to measurement, suspensions of cells were incubated at 37 °C for 5 min.

[Ca<sup>2+</sup>]<sub>i</sub> was measured with the ratio metric fluorescent dye Fura 2-AM using AR-CM-MIC cation measurement system (Spex Corp, USA) at λ<sub>ex</sub> 340 nm and 380 nm, λ<sub>em</sub> 505 nm, as described by Grynkiewicz *et al*<sup>(6)</sup>. [Ca<sup>2+</sup>]<sub>i</sub> was calculated according to the formular with K<sub>d</sub> of 224 nmol  $\cdot$  L<sup>-1</sup>:

[Ca<sup>2+</sup>]<sub>i</sub> = K<sub>d</sub> × (sb<sub>1</sub>/sb<sub>2</sub>) × (R - R<sub>min</sub>)/(R<sub>max</sub> - R) (nmol  $\cdot$  L<sup>-1</sup>). R = F<sub>340</sub>/F<sub>380</sub>; R<sub>min</sub> = F<sub>340min</sub>/F<sub>380min</sub>; R<sub>max</sub> = F<sub>340max</sub>/F<sub>380max</sub>; sb<sub>1</sub>/sb<sub>2</sub> = F<sub>380max</sub>/F<sub>380min</sub>; F refers to fluorescence, R<sub>min</sub> and R<sub>max</sub> were obtained by adding ionomycin and egtazic acid, the parameters mentioned above were given, [Ca<sup>2+</sup>]<sub>i</sub> was automatically calculated by the computer. Allitridi was added 15 min before the fluorescence measurement while KCl, Nor, and Glu were added immediately prior to the fluorescent determinations.

**Statistical analysis** Data were expressed as  $\bar{x} \pm s$  and compared using *t* test.

## RESULTS

**Allitridi on resting [Ca<sup>2+</sup>]<sub>i</sub>** In a Ca-free medium containing egtazic acid 0.1 mmol  $\cdot$  L<sup>-1</sup>, rat neuronal cells contained (76 ± 12) nmol  $\cdot$  L<sup>-1</sup> of [Ca<sup>2+</sup>]<sub>i</sub>. The levels were (105 ± 14), (136 ± 16), (172 ± 23), and (254 ± 18) nmol  $\cdot$  L<sup>-1</sup> in the presence of extracellular Ca<sup>2+</sup> 0.01, 0.1, 1, and 2 mmol  $\cdot$  L<sup>-1</sup>, respectively. [Ca<sup>2+</sup>]<sub>i</sub> was dependent on the extracellular Ca<sup>2+</sup> concentration. Preincubation with allitridi 100 μmol  $\cdot$  L<sup>-1</sup> for 15 min in Hanks' solution containing Ca<sup>2+</sup> 0, 0.01, 0.1, 1, and 2 mmol  $\cdot$  L<sup>-1</sup> had no significant effect on the passive diffusive flux through the cytoplasmic membrane of the brain cells (Tab 1).

### Allitridi on KCl-induced [Ca<sup>2+</sup>]<sub>i</sub> elevation

In a Ca-free medium, KCl 20, 40, and 80 mmol  $\cdot$  L<sup>-1</sup> did not affect the [Ca<sup>2+</sup>]<sub>i</sub>, while the brain cells were exposed to high K<sup>+</sup> in Hanks' solution containing Ca<sup>2+</sup> 1.3 mmol  $\cdot$  L<sup>-1</sup>, the [Ca<sup>2+</sup>]<sub>i</sub> increased rapidly and concentration-dependently. KCl 20, 40, and 80 mmol  $\cdot$  L<sup>-1</sup> increased the [Ca<sup>2+</sup>]<sub>i</sub> by 16 %, 53 %, and 153 % respectively, allitridi 1 μmol  $\cdot$  L<sup>-1</sup> did not

**Tab 1. Effects of allitridi (100 μmol  $\cdot$  L<sup>-1</sup>) on resting [Ca<sup>2+</sup>]<sub>i</sub> in isolated rat brain cells in different extracellular Ca<sup>2+</sup> concentration medium. n = 5.  $\bar{x} \pm s$ .**

[Ca <sup>2+</sup> ] <sub>e</sub> /mmol $\cdot$ L <sup>-1</sup>	Control	Allitridi-treated
0	76 ± 12	67 ± 17
0.01	105 ± 14	107 ± 38
0.1	136 ± 16	143 ± 35
1.0	172 ± 23	156 ± 29
2.0	254 ± 18	245 ± 37

change the [Ca<sup>2+</sup>]<sub>i</sub>, while pretreatment with allitridi 10, 25, 50, 100 μmol  $\cdot$  L<sup>-1</sup>, could decrease extracellular high KCl (80 mmol  $\cdot$  L<sup>-1</sup>)-induced [Ca<sup>2+</sup>]<sub>i</sub> elevation with IC<sub>50</sub> of 59.7 μmol  $\cdot$  L<sup>-1</sup> (Tab 2).

**Tab 2. Effects of allitridi on KCl actions on [Ca<sup>2+</sup>]<sub>i</sub> of isolated rat brain cells in normal Ca-medium. n = 5.  $\bar{x} \pm s$ . \*P < 0.01 vs no KCl. <sup>o</sup>P < 0.05, <sup>f</sup>P < 0.01 vs KCl alone.**

KCl/ mmol $\cdot$ L <sup>-1</sup>	[Ca <sup>2+</sup> ] <sub>i</sub> / nmol $\cdot$ L <sup>-1</sup>	Allitridi/ μmol $\cdot$ L <sup>-1</sup>	Allitridi-treated [Ca <sup>2+</sup> ] <sub>i</sub> / nmol $\cdot$ L <sup>-1</sup>
0	220 ± 19	100	218 ± 45
20	256 ± 18	100	242 ± 19
40	337 ± 23 <sup>e</sup>	100	276 ± 27 <sup>e</sup>
80	556 ± 34 <sup>e</sup>	1	544 ± 35
80	556 ± 34 <sup>e</sup>	10	535 ± 26
80	556 ± 34 <sup>e</sup>	25	500 ± 23 <sup>e</sup>
80	556 ± 34 <sup>e</sup>	50	387 ± 29 <sup>f</sup>
80	556 ± 34 <sup>e</sup>	100	307 ± 24 <sup>f</sup>

### Allitridi on Nor-induced [Ca<sup>2+</sup>]<sub>i</sub> elevation

In Ca<sup>2+</sup>-free solution, Nor 0.01, 0.1, 1, and 10 μmol  $\cdot$  L<sup>-1</sup> increased the [Ca<sup>2+</sup>]<sub>i</sub> by 22 %, 34 %, 68 %, and 80 %, respectively, allitridi 100 μmol  $\cdot$  L<sup>-1</sup> inhibited the Nor (0.01 - 10 μmol  $\cdot$  L<sup>-1</sup>)-induced [Ca<sup>2+</sup>]<sub>i</sub> elevation by -11 %, 7 %, 7 %, and 21 %, respectively. When exposed to extracellular Ca<sup>2+</sup> of 1.3 mmol  $\cdot$  L<sup>-1</sup>, Nor (0.01 - 10 μmol  $\cdot$  L<sup>-1</sup>) increased the [Ca<sup>2+</sup>]<sub>i</sub> by 43 %, 79 %, 98 %, and 114 %, respectively, allitridi 100 μmol  $\cdot$  L<sup>-1</sup> inhibited the Nor-induced [Ca<sup>2+</sup>]<sub>i</sub> elevation by 10 %, 23 %, 33 %, and 44 %, respectively. Low concentration of allitridi (1 - 10 μmol  $\cdot$  L<sup>-1</sup>) did not show apparent

effects on Nor-induced  $[Ca^{2+}]_i$  elevation (Tab 3).

Tab 3. Effects of allitridi ( $100 \mu\text{mol} \cdot \text{L}^{-1}$ ) on Nor actions on  $[Ca^{2+}]_i$  of isolated rat brain cells in normal Ca vs Ca-free medium.  $n=5$ ,  $\bar{x} \pm s$ .  $^aP < 0.01$  vs no Nor.  $^bP < 0.05$ ,  $^cP < 0.01$  vs Nor alone.

Nor/ $\text{mmol} \cdot \text{L}^{-1}$	Normal Ca-medium	Allitridi- treated	Ca-free medium	Allitridi- treated
0	$220 \pm 19$	$218 \pm 45$	$76 \pm 12$	$67 \pm 17$
0.01	$314 \pm 25^c$	$280 \pm 26$	$92 \pm 9$	$88 \pm 13$
0.1	$394 \pm 20^c$	$340 \pm 31^c$	$102 \pm 7^c$	$93 \pm 14$
1.0	$435 \pm 22^c$	$360 \pm 31^f$	$127 \pm 19^c$	$107 \pm 22$
10	$471 \pm 22^c$	$371 \pm 26^f$	$138 \pm 12^c$	$108 \pm 17^c$

#### Allitridi on Glu-induced $[Ca^{2+}]_i$ elevation

In  $Ca^{2+}$ -free solution, Glu had no obvious effect on  $[Ca^{2+}]_i$ , while when exposed to extracellular  $Ca^{2+}$  of  $1.3 \text{ mmol} \cdot \text{L}^{-1}$ , Glu  $10 \mu\text{mol} \cdot \text{L}^{-1}$  increased the  $[Ca^{2+}]_i$  from  $(220 \pm 19)$  to  $(477 \pm 47) \text{ nmol} \cdot \text{L}^{-1}$ , indicating that the  $[Ca^{2+}]_i$  elevation induced by Glu was mainly dependent on extracellular  $Ca^{2+}$ ,  $MgCl_2$   $5 \text{ mmol} \cdot \text{L}^{-1}$  inhibited the  $[Ca^{2+}]_i$  elevation by 93% [to  $(238 \pm 12) \text{ nmol} \cdot \text{L}^{-1}$ ], Glu  $0.01 - 10 \mu\text{mol} \cdot \text{L}^{-1}$  concentration-dependently elevated the  $[Ca^{2+}]_i$ . Allitridi  $1 - 100 \mu\text{mol} \cdot \text{L}^{-1}$  dose-dependently inhibited the  $[Ca^{2+}]_i$  with  $IC_{50}$  of  $69.9 \mu\text{mol} \cdot \text{L}^{-1}$  (Tab 4).

Tab 4. Effects of allitridi on Glu actions on  $[Ca^{2+}]_i$  of isolated rat brain cells in normal Ca-medium.  $n=5$ ,  $\bar{x} \pm s$ .  $^aP < 0.01$  vs A.  $^bP < 0.05$ ,  $^cP < 0.01$  vs E.

Group	Glu/ $\text{mmol} \cdot \text{L}^{-1}$	Allitridi/ $\mu\text{mol} \cdot \text{L}^{-1}$	$[Ca^{2+}]_i$ / $\text{nmol} \cdot \text{L}^{-1}$
A	0	0	$220 \pm 19$
B	0.01	0	$234 \pm 31$
C	0.1	0	$298 \pm 20^c$
D	1	0	$382 \pm 22^c$
E	10	0	$477 \pm 47^c$
F	100	0	$554 \pm 29^c$
G	10	1	$456 \pm 17$
H	10	10	$438 \pm 39$
I	10	25	$404 \pm 34^c$
J	10	50	$351 \pm 39^f$
K	10	100	$323 \pm 23^f$

#### DISCUSSION

Our results demonstrate that allitridi reduced

$[Ca^{2+}]_i$  via 2 mechanisms; a) by blocking  $Ca^{2+}$ -influx from the extracellular space, and b) by blocking  $Ca^{2+}$ -release from the intracellular  $Ca^{2+}$  stores.

Cytosolic calcium is a very important second messenger, which mediates a lot of physiological or pharmacological reactions and regulates cellular functions. Abnormality of calcium metabolism in brain tissues can produce vasospasm, seizures, edema, etc<sup>17,8</sup>. Calcium overload plays a central role in both acute and especially delayed neuronal damage in ischemia brain tissues.  $Ca^{2+}$  fluxes from the extracellular fluid into the intracellular space may trigger irreversible injury<sup>9-11</sup> and enhance the breakdown of protein and lipids thus results in cell damage. Ischemia causes increased  $Ca^{2+}$  fluxes into cells because the depolarization and transmitter release could open the voltage-sensitive  $Ca^{2+}$  channels and the receptor-operated  $Ca^{2+}$  channels. Our study suggested that allitridi had inhibitory effects on the  $[Ca^{2+}]_i$  elevation induced by KCl, Nor, and Glu. In conclusion, allitridi was shown to reduce the  $Ca^{2+}$  fluxes via either voltage-sensitive  $Ca^{2+}$  channels blockade or receptor-operated  $Ca^{2+}$  channels blockade at a higher dosage, the inhibitory effects of allitridi on  $[Ca^{2+}]_i$  elevation are beneficial to its protection of brain against cerebral ischemia.

#### REFERENCES

- 1 Kandler BS. Garlic (*Allium sativum*) and onion (*Allium cepa*): a review of their relationship to cardiovascular disease. *Prev Med* 1987; 16: 670-85.
- 2 Shi ZX, Du JX, Wu ZM, Wang LH, Chen GD, Zhang TZ, et al. Clinical and experimental researches on cerebral infarction treated with allicin. *Acta Med Sin* 1991; 6: 33-5.
- 3 Ma XH, Pan XX, Liu TP. Protective effect of allitridi on the acute experimental cerebral ischemia. *Chin J Pharmacol Toxicol* 1998; 12: 151-2.
- 4 Chen SH, Yin ZZ, Ma BB, Shi ZX. Calcium antagonism of allitridi. *Acta Pharmacol Sin* 1988; 9: 533-5.
- 5 Dildy JE, Leslie SW. Ethanol inhibits NMDA-induced increases in free intracellular  $Ca^{2+}$  in dissociated brain cells. *Brain Res* 1989; 499: 383-7.
- 6 Grynkiwicz G, Poenie M, Tsien RY. A new generation of  $Ca^{2+}$  indicators with greatly improved fluorescence properties. *J Biol Chem* 1985; 260: 3440-50.
- 7 Tsien RW, Lipscombe D, Madison DV, Bley KR, Fox

601 - 612

8

AP. Multiple types of neuronal calcium channels and their selective modulation. Trends Neurosci 1988; 11: 431-8.

8 Siesjö BK, Bengtsson F. Calcium fluxes, calcium antagonists, and calcium-related pathology in brain ischemia, hypoglycemia, and spreading depression: a unifying hypothesis. J Cereb Blood Flow Metab 1989; 9: 127-40.

9 Siesjö BK. Calcium-mediated processes in neuronal degeneration. Ann N Y Acad Sci 1994; 747: 140-61.

10 Choi DW. Calcium and excitotoxic neuronal injury. Ann N Y Acad Sci 1994; 747: 162-71.

11 Choi DW. Calcium: still center-stage in hypoxic-ischemic neuronal death. Trends Neurosci 1995; 18: 58-60.

大蒜新素 脑细胞 细胞内液

过程

Fura-2

大蒜新素对分离大鼠脑细胞内游离  $Ca^{2+}$  的影响

R965.2

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**关键词** 脑; 培养的细胞; 大蒜新素; 细胞内液; 钙; Fura-2; 去甲肾上腺素; 谷氨酸钠

**目的:** 观察大蒜新素对不同刺激剂所致分离大鼠脑细胞内游离钙的影响. **方法:** 以 Fura 2-AM 为细胞内游离钙的荧光指示剂, 用 AR-CM-MIC 阳离子测定系统, 直接测定了分离新生大鼠脑细胞内游离钙 ( $[Ca^{2+}]_i$ ) 值, 观察了大蒜新素的影响. **结果:** 大蒜新素对脑细胞静息  $[Ca^{2+}]_i$  无明显影响, 大蒜新素  $1-100 \mu\text{mol}\cdot\text{L}^{-1}$  能剂量依赖性地抑制高  $K^+$  和谷氨酸引起的  $[Ca^{2+}]_i$  升高, 其中  $IC_{50}$  分别为  $59.7$  和  $69.9 \mu\text{mol}\cdot\text{L}^{-1}$ , 高剂量大蒜新素  $100 \mu\text{mol}\cdot\text{L}^{-1}$  能抑制去甲肾上腺素引起的  $[Ca^{2+}]_i$  升高. **结论:** 大蒜新素对高  $K^+$ 、去甲肾上腺素及谷氨酸引起的  $[Ca^{2+}]_i$  升高的抑制作用可能是其抗脑缺血作用机制之一.

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