

# Tetrahydroprotoberberines inhibit lipid peroxidation and scavenge hydroxyl free radicals<sup>1</sup>

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**KEY WORDS** stepholidine; lipid peroxidation; malondialdehyde; mitochondria; free radicals

## ABSTRACT

**AIM:** To study the effects of tetrahydroprotoberberines (THPB) on rat liver and brain lipid peroxidation (LPO) and oxygen free radicals generation. **METHODS:** The malondialdehyde (MDA) levels in rat brain and liver homogenates, induction of MDA by Fe<sup>2+</sup>-Vit C in mitochondria, OH<sup>·</sup> generation by Fenton reaction, and O<sub>2</sub><sup>-·</sup> generation by pyrogallol oxidation were observed *in vitro*. **RESULTS:** (1) THPB lowered the MDA contents in the liver homogenate and mitochondria, and the IC<sub>50</sub> values of *l*-THPB-18 and *l*-stepholidine (SPD) in the liver mitochondria were 3.1 and 12.7 μmol·L<sup>-1</sup> respectively. SPD decreased the MDA contents in the brain homogenate and mitochondria with IC<sub>50</sub> values of 102 and 35.0 μmol·L<sup>-1</sup> respectively. (2) THPB scavenged OH<sup>·</sup>, and the IC<sub>50</sub> values of *l*-THPB-18 and SPD were 0.21 and 3.8 μmol·L<sup>-1</sup> respectively, but no effect on O<sub>2</sub><sup>-·</sup> was observed. **CONCLUSION:** THPB could reduce the MDA contents and scavenge OH<sup>·</sup> and THPB-18 was the most potent amongst them.

## INTRODUCTION

We have found that some dihydroxy-tetrahydroprotoberberine analogues (dihydroxy-THPB), such as *l*-stepholidine (SPD), THPB-18, and scoulerine, possess an intrinsic activity to D<sub>1</sub> receptors<sup>[1]</sup>, and have a D<sub>1</sub> agonistic action in the 6-OHDA-lesioned rats, a Parkinson

disease (PD) animal model<sup>[2-4]</sup>. Furthermore, SPD has been used to treat tardive dyskinesia and alleviate PD syndrome in preliminary clinical trails<sup>[5,6]</sup>.

According to the current insight, PD is a neurodegenerative disease of DA neurons in the substantia nigra based on the oxygen free radicals and lipid peroxidation (LPO) induced by the oxidative stress<sup>[7-9]</sup>. The new effective remedy that could stop or slow down this degeneration can be a strategical drug, which both directly activates DA receptors (particularly D<sub>1</sub>) and protects the surviving DA neurons in the substantia nigra<sup>[10,11]</sup>. Although this viewpoint has been well understood, the perfect drug remains elusive. The present work studied the anti-oxidative effects of THPB, particularly SPD and THPB-18, on LPO and oxygen free radicals generation.

## MATERIAL AND METHODS

**Animals and Materials** Sprague-Dawley rats (male, Grade II, certificate No 005, weighing 200-300 g) were purchased from Shanghai Experimental Animal Center, Chinese Academy of Sciences. Eleven THPB and 2 protoberberines (PB) (Fig 1), supplied by Shanghai Institute of Materia Medica, were dissolved in sulphuric acid 0.1 mol·L<sup>-1</sup>, and adjusted with NaOH 0.1 mol·L<sup>-1</sup> to pH 4-5. Other reagents were AR.

**Measurement of MDA in rat liver and brain homogenate** The contents of MDA in the liver or brain homogenate were measured with the thiobarbituric acid (TBA) reaction<sup>[12]</sup>. In brief, the liver and brain were homogenized with Tris-KCl buffer (Tris 0.1 mol·L<sup>-1</sup>, KCl 1 mol·L<sup>-1</sup>, pH 7.4). The supernatants were incubated with THPB at 37 °C, and then with TBA and kept in boiling water bath. After cooling, the optical density (OD) value was measured at 535 nm (752 C spectrophotometer). MDA content was expressed as nmol·g<sup>-1</sup> wet tissue.

**Measurement of MDA initiated by Fe<sup>2+</sup>-Vit**

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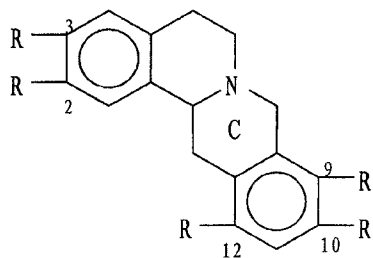
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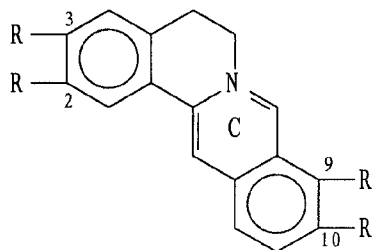
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THPBs	R <sub>2</sub>	R <sub>3</sub>	R <sub>9</sub>	R <sub>10</sub>	R <sub>12</sub>
<i>l</i> -THPB-18	OH	OCH <sub>3</sub>	OH	OCH <sub>3</sub>	Cl
<i>d</i> -THPB-18	OH	OCH <sub>3</sub>	OH	OCH <sub>3</sub>	Cl
<i>l</i> -THPB-19	OH	OCH <sub>3</sub>	OH	OCH <sub>3</sub>	Br
scoulerine	OH	OCH <sub>3</sub>	OH	OCH <sub>3</sub>	H
SPD	OH	OCH <sub>3</sub>	OCH <sub>3</sub>	OH	H
THPB-1	OH	OH	OCH <sub>3</sub>	OCH <sub>3</sub>	H
H-143	OH	OCH <sub>3</sub>	H	H	H
H-25	OH	OCH <sub>3</sub>	H	Cl	H
H-149	OH	OCH <sub>3</sub>	C <sub>2</sub> H <sub>5</sub> O	H	OCH <sub>3</sub>
<i>l</i> -THP	OCH <sub>3</sub>	OCH <sub>3</sub>	OCH <sub>3</sub>	OCH <sub>3</sub>	H
THB	—OCH <sub>2</sub> O—	OCH <sub>3</sub>	OCH <sub>3</sub>	OCH <sub>3</sub>	H



PBs	R <sub>2</sub>	R <sub>3</sub>	R <sub>9</sub>	R <sub>10</sub>
palmatine	OCH <sub>3</sub>	OCH <sub>3</sub>	OCH <sub>3</sub>	OCH <sub>3</sub>
berberine	—OCH <sub>2</sub> O—	OCH <sub>3</sub>	OCH <sub>3</sub>	OCH <sub>3</sub>

Fig 1. The chemical structures of tetrahydro-protoberberines (THPBs) and proto-berberines (PBs).

**C in liver and brain mitochondria** Rat liver and brain were homogenized with 10% (w/v) ice-cold Tris-KCl buffer, then the supernatant was centrifuged at  $18\,000 \times g$ ,  $4\text{ }^\circ\text{C}$  for 15 min (HITACHI 25-PR520). The mitochondrial suspension 0.5 mL (containing 1 mg protein) was incubated at  $37\text{ }^\circ\text{C}$  for 60 min, in the presence of  $\text{FeSO}_4$   $5\ \mu\text{mol}\cdot\text{L}^{-1}$ , Vit C  $10\ \mu\text{mol}\cdot\text{L}^{-1}$ , and the test compounds, then 2 mL  $0.1\ \text{mol}\cdot\text{L}^{-1}$  HCl and 1 mL 0.67% TBA were added, suspension mixture was then heated for 15 min in a boiling water bath, and extracted with 4 mL BuOH after cooling. The OD value of the BuOH phase was measured at  $535\ \text{nm}$ <sup>[13]</sup>. The MDA content was expressed as  $\mu\text{mol}\cdot\text{g}^{-1}$  protein.

#### Measurement of hydroxyl radical ( $\text{OH}^\cdot$ ) gen-

#### erated in $\text{Fe}^{2+}$ -EDTA and $\text{H}_2\text{O}_2$ system *in vitro*

The suspension mixture 3.5 mL containing benzoic acid  $2.187\ \text{mmol}\cdot\text{L}^{-1}$ ,  $\text{FeSO}_4$   $21\ \mu\text{mol}\cdot\text{L}^{-1}$ , edetic acid-2Na  $64\ \mu\text{mol}\cdot\text{L}^{-1}$ ,  $\text{K}_2\text{HPO}_4$ - $\text{KH}_2\text{PO}_4$  buffer  $109\ \text{mmol}\cdot\text{L}^{-1}$  (pH 7.4),  $\text{H}_2\text{O}_2$   $32\ \mu\text{mol}\cdot\text{L}^{-1}$ , and the test compound was incubated for 5 h at  $25\text{ }^\circ\text{C}$ . The fluorescence intensity of  $\text{OH}^\cdot$  radicals generated as a result was measured ( $300/408\ \text{nm}$ , HITACHI 650-10S fluorospectrophotometer)<sup>[14]</sup>.

**Measurement of superoxide anion ( $\text{O}_2^{\cdot-}$ ) resulting from pyrogallol autoxidation** The effects of THPBs on scavenging  $\text{O}_2^{\cdot-}$  was measured by determining of rate of pyrogallol autoxidation<sup>[15]</sup>.

**Statistical analysis** The inhibitory concentration 50% ( $\text{IC}_{50}$ ) and its 95% confidence limits were calculated by logit's method. Statistical analysis was performed using unpaired *t* test.

## RESULTS

**Effect of SPD on MDA content** SPD decreased the MDA contents in rat liver and brain homogenate in a dose-dependent manner (Fig 2). At the concentrations from  $1.875 - 300\ \mu\text{mol}\cdot\text{L}^{-1}$ , the inhibitory rates (IR) in the liver and brain homogenate were  $(4 \pm 7)\%$  to  $(83 \pm 4)\%$  and  $(18 \pm 5)\%$  to  $(80 \pm 6)\%$  respectively, and its  $\text{IC}_{50}$  values (95% confidence limits) were 18.5 ( $15.5 - 22.0$ ) and 102 ( $94 - 110$ )  $\mu\text{mol}\cdot\text{L}^{-1}$ , respectively.

SPD could also inhibit the MDA induced by  $\text{Fe}^{2+}$ -Vit C in rat brain mitochondria (Fig 3). At the concentrations from  $7.5 - 240\ \mu\text{mol}\cdot\text{L}^{-1}$ , IR were  $(24 \pm 5)\%$  to  $(92 \pm 14)\%$  with  $\text{IC}_{50}$  value of 35.0 ( $21.7 - 56.7$ )  $\mu\text{mol}\cdot\text{L}^{-1}$ . These results indicate that SPD possesses anti-LPO activity.

**Effect of THPB on MDA content of rat liver homogenate** Among the tested 11 THPB and 2 PB, *l*-THPB-18, *d*-THPB-18, *l*-THPB-19, THPB-1, H-143, and SPD all lowered the MDA level at  $5\ \mu\text{mol}\cdot\text{L}^{-1}$  ( $n = 5$ ,  $P < 0.05$  vs control) with IR of 68%, 56%, 56%, 56%, 32%, and 28% respectively. The anti-LPO effect of *l*-THPB-18 was more potent than that of SPD and other THPB. At  $10\ \mu\text{mol}\cdot\text{L}^{-1}$ , *l*-THP could lower the MDA content ( $P < 0.01$  vs control) with an IR of 89%, but palmatine (dehydrogenated *l*-THP) had no obvious effect on it.

**Effect of THPB on  $\text{Fe}^{2+}$ -Vit C initiated MDA content in rat liver mitochondria**  $\text{Fe}^{2+}$ -Vit C

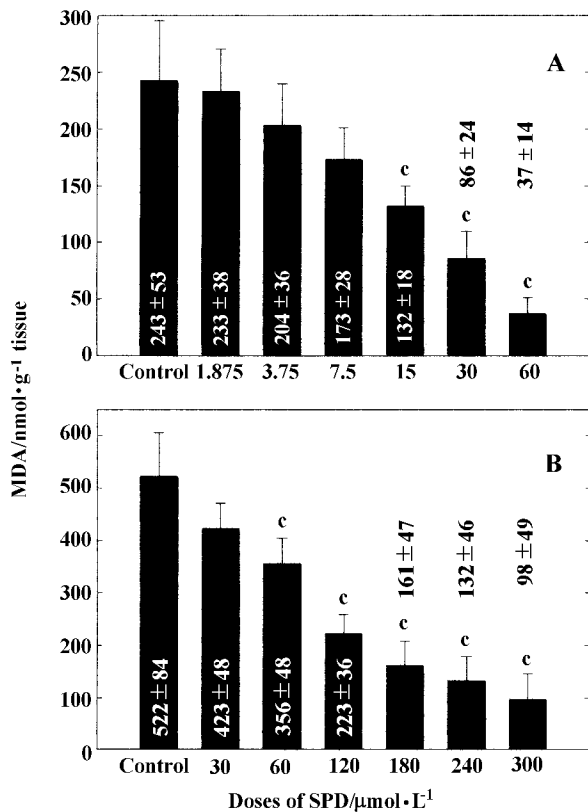


Fig 2. Effect of SPD on the MDA levels in rat liver (A) and brain (B) homogenate.  $n = 4$  experiments in triplicate.  $\bar{x} \pm s$ .  $^c P < 0.01$  vs control.

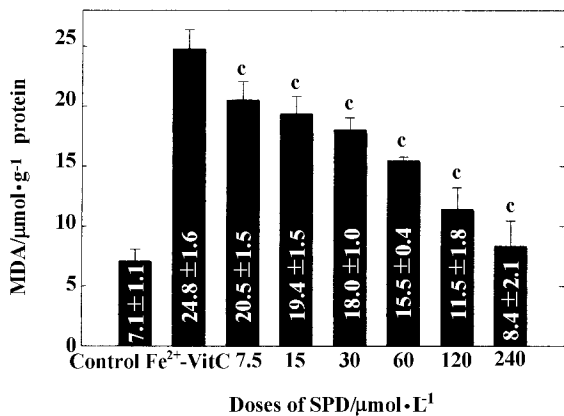


Fig 3. Effect of SPD on Fe<sup>2+</sup>-Vit C induced increase in MDA levels in rat brain mitochondria.  $n = 4$  experiments in duplicate.  $\bar{x} \pm s$ .  $^c P < 0.01$  vs Fe<sup>2+</sup>-Vit C group.

significantly increased the MDA levels in rat liver mitochondria ( $P < 0.01$  vs control). *l*-THPB-18 (0.5 – 10 μmol·L<sup>-1</sup>), *d*-THPB-18 (0.5 – 10 μmol·L<sup>-1</sup>), THPB-1 (1 – 50 μmol·L<sup>-1</sup>), H-143 (0.3 – 30 μmol·L<sup>-1</sup>), H-25 (1 – 10 μmol·L<sup>-1</sup>), scoulerine (1 – 45 μmol·L<sup>-1</sup>), SPD (3.75 – 60 μmol·L<sup>-1</sup>), and tea

polyphenols (0.625 – 5 μg·mL<sup>-1</sup>) all inhibited the MDA levels dose-dependently ( $r = 0.9186 - 0.9973$ ,  $P < 0.01$  or  $P < 0.05$ ). Among them, *l*-THPB-18 was most potent, while SPD was the least (Tab 1). The IC<sub>50</sub> of tea polyphenols was 1.49 ± 0.20 μg·mL<sup>-1</sup> ( $n = 4$ ).

Tab 1. IC<sub>50</sub> values of THPB on MDA induced with Fe<sup>2+</sup>-Vit C in rat liver mitochondria and fluorescence intensity of the hydroxyl radical (OH<sup>·</sup>).  $\bar{x} \pm s$ .  $n = 4$  experiments in duplicate.  $^c P < 0.01$  vs *l*-THPB-18.

Compounds	IC <sub>50</sub> /μmol·L <sup>-1</sup>	
	MDA	OH <sup>·</sup>
<i>l</i> -THPB-18	3.1 ± 0.3	0.21 ± 0.04
<i>d</i> -THPB-18	3.9 ± 0.6	0.24 ± 0.19
THPB-1	4.1 ± 3.1	8.3 ± 2.9 <sup>c</sup>
H-25	4.2 ± 0.2 <sup>c</sup>	8.0 ± 1.9 <sup>c</sup>
H-143	4.9 ± 1.2 <sup>c</sup>	9.0 ± 0.9 <sup>c</sup>
Scoulerine	7.7 ± 2.0 <sup>c</sup>	0.59 ± 0.15 <sup>c</sup>
SPD	12.7 ± 2.5 <sup>c</sup>	3.8 ± 0.7 <sup>c</sup>

### Effects of THPB on generation of hydroxyl radical (OH<sup>·</sup>) and superoxide anion (O<sub>2</sub><sup>·-</sup>)

*l*-THPB-18 (0.01 – 3 μmol·L<sup>-1</sup>), *d*-THPB-18 (0.1 – 3 μmol·L<sup>-1</sup>), *l*-THPB-19 (0.1 – 2 μmol·L<sup>-1</sup>), THPB-1 (1 – 20 μmol·L<sup>-1</sup>), scoulerine (0.03 – 30 μmol·L<sup>-1</sup>), H-25 (0.1 – 30 μmol·L<sup>-1</sup>), H-143 (0.3 – 60 μmol·L<sup>-1</sup>), and SPD (0.3 – 60 μmol·L<sup>-1</sup>) all could significantly scavenge OH<sup>·</sup> in a dose-dependent manner. The rank order of the inhibitory potency of THPB was *l*-THPB-18, *d*-THPB-18, *l*-THPB-19 > scoulerine > SPD > H-25, H-143, THPB-1 (Tab 1). The IC<sub>50</sub> of *l*-THPB-19 was 0.23 ± 0.09 μmol·L<sup>-1</sup> ( $n = 3$ ). However, none of them had any significant effect on pyrogallol autoxidation rates even with the higher doses (100 – 640 μmol·L<sup>-1</sup>), which indicated that THPB could not scavenge O<sub>2</sub><sup>·-</sup>.

### DISCUSSION

The MDA level, used as an autoxidation index of LPO in tissues, is used to screen the antioxidants. The present results show that some THPB can remarkably lower the MDA levels, which indicate that they possess the antioxidant effects. In addition, THPB can scavenge OH<sup>·</sup> radicals *in vitro*. Among them, the most potent one is THPB-18 which has a halogen substituted group at C<sub>12</sub> position, suggesting that the halogen group is impor-

tant for enhancing the pharmacological effects of THPB analogs<sup>[2,16]</sup>.

THPB can scavenge OH<sup>·</sup> radical, but has no effect on O<sub>2</sub><sup>-</sup>. This property of THPB is to that of bromocriptine (D<sub>2</sub> agonist)<sup>[11]</sup>. Whether THPB has the scavenging effects in other O<sub>2</sub><sup>-</sup> generating systems such as the xanthine-xanthine oxidase system remains to be further studied.

The oxygen free radicals, especially OH<sup>·</sup>, induced in DA oxidative stress is very important in the neurodegeneration of DA neurons of the substantia nigra in PD<sup>[8,9]</sup>. The present results show that THPB, such as THPB-18 and SPD possess anti-oxidative pharmacological properties. Furthermore, recent *in vitro* and *in vivo* results have demonstrated that SPD and THPB-18 could protect DA neurons from the damage induced by selective DA neuron toxin MPTP and 1-methyl-4-phenyl pyridinium ion (MPP<sup>+</sup>) (unpublished data). Recently, SPD has been preliminarily used to treat the PD patients in the later stage and has alleviated the syndrome by combination with the lowest dose of bromocriptine<sup>[6]</sup>. Thus it can be inferred that THPB can be potentially beneficial by preventing further DA neuro degeneration in PD.

## REFERENCES

- 1 Dong ZJ, Chen LJ, Jin GZ. GTP regulation of (-)-stepholidine binding to R<sub>H</sub> of D<sub>1</sub> dopamine receptors in calf striatum. *Biochem Pharmacol* 1997; 54: 227-32.
- 2 Guo X, Wang LM, Liu J, Jin GZ. Characteristics of tetrahydroprotoberberines on dopamine D<sub>1</sub> and D<sub>2</sub> receptors in calf striatum. *Acta Pharmacol Sin* 1997; 18: 225-30.
- 3 Liu J, Guo X, Wang BC, Jin GZ. Increased phosphorylation of DARPP-32 by D<sub>1</sub> agonistic action of *l*-stepholidine in the 6-OHDA-lesioned rat striatum. *Acta Physiol Sin* 1999; 51: 65-72.
- 4 Zou LL, Liu J, Jin GZ. Involvement of receptor reserve in D<sub>1</sub> agonistic action of stepholidine in lesioned rats. *Biochem Pharmacol* 1997; 54: 233-40.
- 5 Cai N, Jin GZ, Zhang ZD. A controlled study on treatment of tartive dyskinesia by *l*-stepholidine. *Chin J Neurol Psychiatry* 1988; 21: 281-3.
- 6 Li RK, Chen LJ, Zhao H, Jin GZ. Treatment of Parkinson disease with *l*-stepholidine (SPD) plus bromocriptine. *Chin J Integrated Tradit West Med* 1999; 19: 428-9.
- 7 Ogawa N, Edamatsu R, Mizukawa K, Asanuma M, Kohno M, Mori A. Degeneration of dopaminergic neurons and free radicals: possible participation of levodopa. *Adv Neurol* 1993; 60: 242-50.
- 8 Adams JD Jr, Odunze IN. Oxygen free radicals and Parkinson's disease. *Free Radic Biol Med* 1991; 10: 161-9.

- 9 Fahn S, Cohen G. The oxidant stress hypothesis in Parkinson's disease: evidence supporting it. *Ann Neurol* 1992; 32: 804-12.
- 10 Grondin R, Bedard PJ, Britton DR, Shiosaki K. Potential therapeutic use of the selective dopamine D<sub>1</sub> receptor agonist, A-86929: an acute study in parkinsonian levedopa-primed monkeys. *Neurology* 1997; 49: 421-6.
- 11 Ogawa N, Tanaka K, Asanuma M, Kawai M, Masumizu T, Kohno M, *et al*. Bromocriptine protects mice against 6-hydroxydopamine and scavenges hydroxyl free radicals *in vitro*. *Brain Res* 1994; 657: 207-13.
- 12 Xu CF, Sun LY. The effect of extract form Equisetum Hymale on forming of mouse' brain heart and lung lipid peroxide. *Chin J Mod Appl Pharm* 1998; 15: 5-7.
- 13 Lu XY. Injury in rat liver mitochondria induced by ascorbic acid and ferrous sulfate. *Prog Biochem Biophys* 1989; 16: 372-4, 382.
- 14 Wang J, Wu JF, Zhang JT. Studies on the anti-cerebral ischemia effect of total salvianolic acid. *Chin Pharmacol Bull* 1999; 15: 164-6.
- 15 Xie WH, Yao JF, Yuan QS. Modification of pyrogallol autoxidation method for assay of superoxide dismutase. *Pharm Ind* 1988; 19: 217-20.
- 16 Chen LJ, Xi Y, Pang DW, Zhou QT, Jin GZ. Effect of (±)12-chloroscoulerine on brain dopamine receptors. *Acta Pharmacol Sin* 1996; 17: 185-9.

## 四氢原小檗碱同类物抑制脂质过氧化和清除羟自由基<sup>1</sup>

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关键词 千金藤立定; 脂质过氧化; 丙二醛; 线粒体; 自由基

目的: 研究 THPB 对大鼠肝、脑脂质过氧化(LPO)和氧自由基的影响。方法: 体外测定大鼠肝、脑匀浆 LPO 和 Fe<sup>2+</sup>-Vit C 诱导肝、脑线粒体 LPO 产生的 MDA, Fenton 反应产生 OH<sup>·</sup> 和邻苯三酚氧化产生 O<sub>2</sub><sup>-</sup>。结果: (1) THPB 减少肝匀浆和线粒体 MDA, 其中 *l*-THPB-18 和 SPD 的 IC<sub>50</sub> 为 3.1 和 12.7 μmol·L<sup>-1</sup>。SPD 减少脑匀浆和脑线粒体 MDA, IC<sub>50</sub> 分别为 102 和 35.0 μmol·L<sup>-1</sup>。(2) THPB 能清除 OH<sup>·</sup>, 其中 *l*-THPB-18 和 SPD 的 IC<sub>50</sub> 为 0.21 和 3.8 μmol·L<sup>-1</sup>; 但对 O<sub>2</sub><sup>-</sup> 无抑制作用。结论: THPB 能减少 MDA 和清除 OH<sup>·</sup>, 其中 THPB-18 作用最强。

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