

Ca²⁺ sensitivity of contractile system and Ca²⁺ release from sarcoplasmic reticulum in skinned myocardium from rats with pressure overload LV hypertrophy and heart failure¹

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KEY WORDS congestive heart failure; left ventricular hypertrophy; calcium; sarcoplasmic reticulum; contractile proteins; trabecular meshwork; caffeine

ABSTRACT

AIM: To explore the possible subcellular mechanisms underlying the decreased contractility of myocardium in left ventricle failed hearts caused by pressure overload. **METHODS:** Left ventricle pressure overload hypertrophy (LVH) and congestive heart failure (CHF) models were created in rats by ascending aortic banding. Left ventricle trabeculae skinned fibers were prepared by treatment with saponin 500 or 50 mg·L⁻¹. Relative Ca²⁺-activated tensions (*T*_{6.0%} and *T*_{5.6%}) of saponin (500 mg·L⁻¹)-skinned fibers were taken as the indices of Ca²⁺ sensitivity of contractile protein. Caffeine-induced contracture of saponin 50 mg·L⁻¹-skinned fibers was an index of Ca²⁺ release from sarcoplasmic reticulum (SR). **RESULTS:** 1) There was no significant difference in relative Ca²⁺ activated tensions among CHF, LVH, and Sham-operated group in basic situation (*P* > 0.05). After treatment with caffeine 10 mmol·L⁻¹, the increase in value of δT % were significantly higher in LVH and CHF compared with that in Sham-operated control (*P* < 0.01). 2) The amplitudes of caffeine (5 and 10 mmol·L⁻¹)-induced contracture were (0.66 ± 0.14) and (1.20 ±

0.27) g/mm² in control group, 19.8 % and 25.8 % lower in LVH (*P* < 0.05), 78.8 % and 80.9 % much more lower in CHF (*P* < 0.01). **CONCLUSION:** Decreased SR Ca²⁺ release was the main factor responsible for depressed contractility in failed myocardium while the Ca²⁺ sensitivity of contractile protein might not be involved.

INTRODUCTION

Contractility of myocardium is determined by 1) availability of intracellular calcium to myofilaments, mainly from calcium release of sarcoplasmic reticulum (SR) via ryanodine receptor (RyR); 2) responsiveness of contractile system to calcium. Cardiac hypertrophy induced by long term pressure overload is known as an early compensated stage with maintenance of myocardium contractile force, which is followed by later decompensated stage of heart failure characterized by depressed myocardial contractility. Abnormality of Ca²⁺ release from SR and Ca²⁺ sensitivity of contractile protein are supposed to be 2 subcellular mechanisms underlying the suppressed contractility of failed myocardium.

Recently, investigations on the SR Ca²⁺ release channel-RyR protein function and gene encoding RyR expression have been well documented^[1,2,3,4]. It has been shown that the activity and density of Ca²⁺ release channel in SR are abnormal in ischemic failed human heart^[1] and different animal pathological models^[2]. But little is known about the abnormal change of SR Ca²⁺ release function and contractile protein Ca²⁺ sensitivity on the level of skinned myocardium.

In this paper we used saponin-skinned left ventricle trabeculae fibers from rats with left ventricular hypertrophy (LVH) and heart failure caused by

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experimental ascending aortic stenosis, to explore how the above functions change during cardiac hypertrophy and heart failure, which is in charge of the decreased contractility of failed heart.

MATERIALS AND METHODS

Preparation of pathological animal model

Heart failure and hypertrophy rat models were made according to the method of Feldman with small modification^[5]. Wistar ♂ rats of 4-wk-old (weighing 50–70 g) (Grade II, Certificate No 022) were chosen for operation. Silver clip (0.9 mm internal diameter) was used to create ascending aorta stenosis, animals of age-matched controls underwent a same left thoracotomy without placement of the clip. All rats were fed normal rat chow and water *ad lib*. Eighteen weeks after operation, rats with aortic stenosis showing clinical signs associated with heart failure (eg, despnæa, cyanopathy) were denoted as congestive failure group (CHF), while those without clinical changes were denoted as LVH group.

Preparation of skinned left ventricular trabeculae CHF, LVH, and Sham-operated rats were killed at the end of period and hearts were quickly removed to oxygenated Krebs-Henseleit buffer. Left ventricular trabeculae (< 1 mm in diameter) were isolated carefully and transferred to the perfusion chamber with one end of the fiber attached to isometric force transducer (Statham UC2, Gould, USA). The fibers were subjected to skinning procedure previously described in detail^[6]. After skinning procedure, the permeability of sarcolemma and membrane of SR of the trabeculae were checked by MgATP removal and caffeine-induced contracture, respectively^[7].

Solutions Three basic solutions were used as previously described^[6]: 1) Ca^{2+} free relaxing solution, 2) skinning solution containing saponin 50 or 500 $\text{mg} \cdot \text{L}^{-1}$, 3) activating solution containing different concentration of free Ca^{2+} was expressed as pCa (negative lg of the Ca^{2+} concentration). pCa 6.0, pCa 5.6, and pCa 4.5 were used in this experiment. All chemicals were purchased from Sigma Chemical Co. Solution pH was adjusted to 7.0 with KOH 1 $\text{mol} \cdot \text{L}^{-1}$ and checked by a digital pH meter (Model 5986-62, Cole Parmer, Germany).

Measurement of Ca^{2+} sensitivity of

contractile protein After incubation in saponin 500 $\text{mg} \cdot \text{L}^{-1}$ skinning solution, both sarcolemma and the membrane of SR were perforated. Fibers were exposed to activating solutions of pCa 6.0, pCa 5.6, and pCa 4.5 sequentially and the Ca^{2+} -activated forces were recorded and revised by muscle cross-area (mm^2). The Ca^{2+} -activated tension ($T_{6.0}$ and $T_{5.6}$) expressed as the % of maximal tension (T_{max}) obtained at pCa 4.5 was taken as the index of Ca^{2+} sensitivity of contractile protein.

Measurement of Ca^{2+} release from SR

After incubation in saponin (50 $\text{mg} \cdot \text{L}^{-1}$) skinning solution, the sarcolemma of trabeculae became perforated but the membrane of SR remained intact. Fibers were exposed to caffeine 25 $\text{mmol} \cdot \text{L}^{-1}$ to empty the Ca^{2+} store in SR^[8]. After reloading SR by pCa 7.0 solution for 3 min, fibers were exposed to caffeine 5 or 10 $\text{mmol} \cdot \text{L}^{-1}$ and contracture force was recorded and revised by CSA. The amplitude of caffeine-induced contracture tension was taken as the index of the function of Ca^{2+} release from SR.

Statistical analysis All values were presented as $\bar{x} \pm s$. Differences between groups were compared by ANOVA and Newman-Keuls *q* test.

RESULTS

Ca^{2+} sensitivity of contractile protein In preparations skinned by saponin 500 $\text{mg} \cdot \text{L}^{-1}$, Ca^{2+} -activated forces induced by pCa 6.0, pCa 5.6, and pCa 4.5 were recorded in basic situation and under the treatment of caffeine 10 $\text{mmol} \cdot \text{L}^{-1}$. The data of T_{max} were similar in three groups: (0.84 ± 0.23) g/mm^2 in CHF, (0.91 ± 0.28) g/mm^2 in LVH and (0.97 ± 0.31) g/mm^2 in Sham control group ($P > 0.05$) (Fig 1).

The relative Ca^{2+} -activated tension induced by pCa 6.0 ($T_{6.0}\%$) and pCa 5.6 ($T_{5.6}\%$) in CHF, LVH, and Sham groups were calculated. No significant difference was observed among these groups in basic situation ($P > 0.05$) (Tab 1).

Caffeine 10 $\text{mmol} \cdot \text{L}^{-1}$ increased Ca^{2+} -activated forces in three groups at different extents (Fig 1). Under the treatment with caffeine 10 $\text{mmol} \cdot \text{L}^{-1}$, the increase in value of relative Ca^{2+} -activated tension ($\delta T_{6.0}\%$) and ($\delta T_{5.6}\%$) were significantly higher in CHF and LVH groups compared with that in Sham-

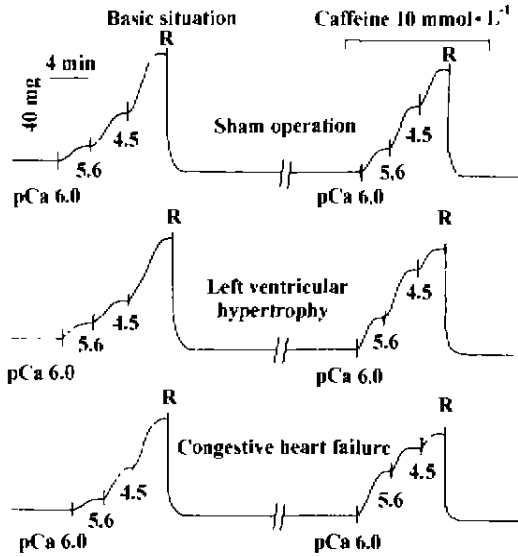


Fig 1. Ca^{2+} activated forces in saponin ($500 \text{ mg} \cdot \text{L}^{-1}$)-skinned ventricular trabeculae fibers from age-matched rat hearts (R: relaxing solution, pCa: $-\lg [\text{Ca}^{2+}]$). $n = 8$.

Tab 1. Relative tension activated by pCa 6.0 ($T_{6.0}\%$) and pCa 5.6 ($T_{5.6}\%$) in saponin ($500 \text{ mg} \cdot \text{L}^{-1}$)-skinned trabeculae and increase in value ($\delta T\%$) treated with caffeine $10 \text{ mmol} \cdot \text{L}^{-1}$. $n = 8$. $^{\ast}P < 0.01$ vs Sham.

	$T_{6.0}\%$ -	$\delta T_{6.0}\%$ + Caffeine	$T_{5.6}\%$ -	$\delta T_{5.6}\%$ + Caffeine
Sham	21.2 ± 5.3	11.2 ± 3.3	45.6 ± 9.9	21.0 ± 2.9
LVH	19.3 ± 3.8	$21.8 \pm 2.6^{\ast}$	43.8 ± 6.2	$31.6 \pm 3.6^{\ast}$
CHF	18.1 ± 4.6	$21.5 \pm 3.9^{\ast}$	39.2 ± 7.1	$35.5 \pm 3.3^{\ast}$

Sham; sham-operated control, LVH; left ventricular hypertrophy, CHF; congestive heart failure.

operated control ($P < 0.01$) (Tab 1).

Ca^{2+} release function of SR After reloading SR with pCa 7.0 solution, typical caffeine-induced contractures were produced by the treatment of caffeine 5 and $10 \text{ mmol} \cdot \text{L}^{-1}$ in LVH and Sham control groups, but not in CHF group. The amplitudes of contracture induced by caffeine 5 and $10 \text{ mmol} \cdot \text{L}^{-1}$ in Sham group were (0.66 ± 0.14) and $(1.20 \pm 0.27) \text{ g}/\text{mm}^2$. The values in LVH group were (0.53 ± 0.10) and $(0.89 \pm 0.15) \text{ g}/\text{mm}^2$, which were lower than that in Sham control ($P < 0.05$). The values in CHF group were (0.14 ± 0.07) and $(0.23 \pm 0.08) \text{ g}/\text{mm}^2$, which were much more decreased compared with that in Sham control and in LVH ($P < 0.01$) (Fig 2).

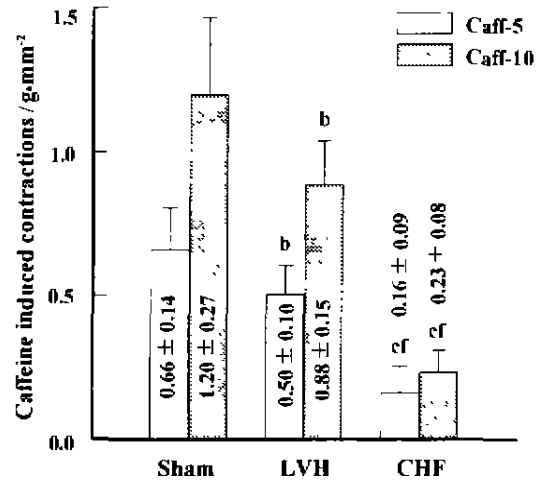


Fig 2. Comparison of the amplitude of caffeine-induced contractures in saponin $50 \text{ mg} \cdot \text{L}^{-1}$ skinned left ventricular trabeculae from sham operated control (Sham), hypertrophy (LVH) and heart failure (CHF) groups. $n = 8$. $\bar{x} \pm s$. $^bP < 0.05$, $^cP < 0.01$ vs Sham; $^{\ast}P < 0.01$ vs LVH. (Caff-5: caffeine $5 \text{ mmol} \cdot \text{L}^{-1}$, Caff-10: caffeine $10 \text{ mmol} \cdot \text{L}^{-1}$)

DISCUSSION

The myocytes Ca^{2+} responsiveness was decreased apparently in the end stage of right ventricular hypertrophy in rat hearts^[9]. In the present paper, we observed that in basic situation, there was no change on myocardial Ca^{2+} sensitivity in left ventricular hypertrophic and failed hearts compared with that in age-matched control, this result was similar to those of hypertrophy in guinea pig^[10] and ferret hearts^[11]. From the results, we can infer that abnormality of Ca^{2+} sensitivity of contractile protein was not involved in the mechanisms underlying the depressed contractility in left ventricular failed hearts. But interestingly, we found that the Ca^{2+} -activated tension in pathological myocardium was more sensitive to caffeine. Further study is needed to explain the difference of responsiveness to caffeine between pathological and normal preparations.

In the preparations with intact SR membrane, caffeine-induced contractures deeply decreased in failed hearts compared with those in hypertrophic or age-matched control group, indicating that the abnormality of SR Ca^{2+} release function existed in the end stage of failed hearts in rats. This abnormal

change of SR Ca^{2+} release might be the main factor responsible for the depressed contractility in heart failure. There are 2 possible explanations on the reduced SR Ca^{2+} release function; a down-regulated number and function of RyR protein, or reduced available stored Ca^{2+} in SR due to decreased SR Ca^{2+} uptake^[12]. Plenty of evidences showed that the gene expression of SR Ca^{2+} -ATPase decreased in failed hearts^[3,5,13,14] which supported the possibility of the reduced stored Ca^{2+} in SR due to decreased SR Ca^{2+} uptake. Whether the gene expression of SR Ca^{2+} release channel changed in this situation still remained controversy^[3,4]. Further researches on the molecular mechanisms of decreased SR Ca^{2+} release are necessary.

REFERENCES

- 1 Brillantes AM, Allen P, Takahashi T, Izumo S, Marks AR. Differences in cardiac calcium release channel (ryanodine receptor) expression in myocardium from patients with end-stage heart failure caused by ischemic versus dilated cardiomyopathy. *Circ Res* 1992; 71: 18-26.
- 2 Rannou F, Sainte-beuve C, Oliviero P, Do E, Trouve P, Charlemagne D. The effects of compensated cardiac hypertrophy on dihydropyridine and ryanodine receptors in rat, ferret and guinea-pig hearts. *J Mol Cell Cardiol* 1995; 27: 1225-34.
- 3 Schillinger W, Meyer M, Kuwajima G, Mikoshiba K, Just H, Hasenfuss G. Unaltered ryanodine receptor protein levels in ischemic cardiomyopathy. *Mol Cell Biochem* 1996; 160/161: 297-302.
- 4 Arai M, Suzuki T, Nagai R. Sarcoplasmic reticulum genes are upregulated in mild cardiac hypertrophy but downregulated in severe cardiac hypertrophy induced by pressure overload. *J Mol Cell Cardiol* 1996; 28: 1583-90.
- 5 Feldman AM, Weinberg EO, Ray PE, Iorell BH. Selective changes in cardiac gene expression during compensated hypertrophy and the transition to cardiac decompensation in rats with chronic aortic banding. *Circ Res* 1993; 73: 184-92.
- 6 Huang Y, He ZH, Li YX. Effects of MCI-154 on calcium sensitivity of contractile system and calcium release from sarcoplasmic reticulum in saponin-skinned rat myocardium. *Acta Pharmacol Sin* 1997; 18: 234-7.
- 7 Huang Y, He ZH, Li YX. Effect of caffeine and theophylline on Ca^{2+} sensitivity of troponin and Ca^{2+} release from SR in saponin-skinned rat cardiac muscle. *Acta Physiol Sin* 1997; 49: 267-72.
- 8 D'Agnoles A, Luciani GB, Mazzucco A, Gallucci V, Salvati G. Contractile properties and Ca^{2+} release activity of the sarcoplasmic reticulum in dilated cardiomyopathy. *Circulation* 1992; 85: 518-25.
- 9 Fan D, Wannenburg T, Tombe PP. Decreased myocyte tension development and calcium responsiveness in rat right

- ventricular pressure overload. *Circulation* 1997; 95: 2312-7.
- 10 Ventura-Clapier R, Mekhfi H, Oliviero P, Swynghedauw B. Pressure overload changes cardiac skinned-fiber mechanics in rats, not in guinea pigs. *Am J Physiol* 1988; 254: H517-24.
- 11 Baudet S, Ventura-Clapier R. Differential effects of caffeine on skinned fibers from control and hypertrophied ferret hearts. *Am J Physiol* 1990; 259: H1803-8.
- 12 Cory CR, McCutcheon LJ, O'Grady M, Pang AW, Geiger JD, O'Brien PJ. Compensatory downregulation of myocardial Ca^{2+} channel in SR from dogs with heart failure. *Am J Physiol* 1993; 264: H926-37.
- 13 Boluyt MO, O'Neill L, Meredith AL, Bing OHL, Brooks WW, Conrad CH, et al. Alterations in cardiac gene expression during the transition from stable hypertrophy to heart failure. *Circ Res* 1994; 75: 23-32.
- 14 Peters DG, Mitchell HL, McCune SA, Park S, Williams JH, Kandarian SC. Skeletal muscle sarcoplasmic reticulum Ca^{2+} -ATPase gene expression in congestive heart failure. *Circ Res* 1997; 81: 703-10.

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16

压力超负荷左心室肥大和心衰大鼠心肌收缩系统 Ca^{2+} 敏感性和肌浆网 Ca^{2+} 释放¹

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关键词 充血性心力衰竭; 左心室肥大; 钙; 肌浆网; 收缩蛋白质类; 小梁网; 咖啡因

目的: 探讨压力超负荷左心衰心肌收缩能力下降的亚细胞机制。 **方法:** 采用升主动脉缩窄法制备左心室压力超负荷心肌肥大(LVH)和充血性心衰(CHF)大鼠病理模型; 采用皂苷 500 或 50 $\text{mg} \cdot \text{L}^{-1}$ 制备左室肌小梁蜕膜标本, 皂苷 500 $\text{mg} \cdot \text{L}^{-1}$ 蜕膜标本的相对 Ca^{2+} 激活张力($T_{6.0\%}$ 和 $T_{5.6\%}$)和皂苷 50 $\text{mg} \cdot \text{L}^{-1}$ 蜕膜标本的咖啡因挛缩幅值分别是收缩系统 Ca^{2+} 敏感性和肌浆网 Ca^{2+} 释放的指标。 **结果:** 1)基础状态下, LVH、CHF 和同龄假手术对照大鼠的相对 Ca^{2+} 激活张力不存在显著差异($P > 0.05$); 但变力因素咖啡因 10 $\text{mmol} \cdot \text{L}^{-1}$ 干预后 Ca^{2+} 激活张力的增量 $\delta T\%$ 在 LVH 组和 CHF 组明显高于对照组($P < 0.01$); 2)咖啡因 5 和 10 $\text{mmol} \cdot \text{L}^{-1}$ 引发的挛缩幅值在对照组为 (0.66 ± 0.14) 和 (1.20 ± 0.27) g/mm^2 , LVH 组略低于对照组 19.8% 和 25.8% ($P < 0.05$), CHF 组大幅低于对照组 78.8% 和 80.9% ($P < 0.01$)。 **结论:** 导致衰竭心肌收缩能力低下, 可能主要源于肌浆网 Ca^{2+} 释放功能的显著下降而非收缩系统的 Ca^{2+} 敏感性改变。
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