

Endothelin-1 releases endothelium-derived endoperoxides and thromboxane A₂ in porcine coronary arteries with regenerated endothelium¹

Seung-Jung PARK, John J LEE, Paul M VANHOUTTE² (Center for Experimental Therapeutics, Department of Medicine, Baylor College of Medicine, One Baylor Plaza, Houston, TX 77030, USA)

KEY WORDS coronary vessels; endothelin-1; indomethacin; prostaglandin-endoperoxide synthase; nitric oxide; thromboxane A₂; vascular endothelium

ABSTRACT

AIM: To determine the role of endothelium-derived contracting factor (EDCF) in the response to endothelin-1 in arteries with regenerated endothelium. **METHODS:** Rings of porcine coronary arteries, with and without endothelium of previously deendothelialized left anterior descending coronary arteries and native left circumflex coronary arteries, were suspended in conventional organ chambers for the measurement of isometric force. **RESULTS:** In quiescent rings of the previously deendothelialized left anterior descending coronary artery treated with the NO-synthase inhibitor nitro-L-arginine, endothelin-1 caused contractions which were larger in rings with than that in those without endothelium. Under the same experimental conditions, in the left circumflex coronary artery, the contractions to endothelin-1 were augmented markedly by the removal of the endothelium. In rings with endothelium of the previously deendothelialized left anterior descending coronary artery, indometacin (inhibitor of cyclooxygenase) and ridogrel (thromboxane A₂ receptor antagonist and inhibitor of thromboxane

synthase) inhibited contractions to endothelin-1. Dazoxiben (inhibitor of thromboxane synthase) inhibited, to the same extent as indometacin and ridogrel, the response to higher concentrations of endothelin-1. The endothelium-dependent component of the response to lower concentrations of endothelin-1 was inhibited by indometacin and ridogrel, but not by dazoxiben. In rings without endothelium of both previously deendothelialized left anterior descending and native left circumflex coronary arteries, indometacin and ridogrel did not affect the contractions to endothelin-1. **CONCLUSION:** These findings suggest that in regenerated endothelium, high concentrations of endothelin-1 stimulate the release of thromboxane A₂. Endoperoxides generated by activation of endothelial cyclooxygenase may be the endothelium-derived contracting factor(s) released in regenerated endothelium by lower concentrations of the peptide.

INTRODUCTION

Endothelin-1 is a potent vasoconstrictor substance that causes slow and long-lasting contractions of isolated blood vessels^[1,2]. The aorta of the spontaneously hypertensive rat is characterized by the occurrence of endothelium-dependent contractions^[3-7]. In this preparation, endothelin-1 stimulates the release of a cyclooxygenase-dependent, endothelium-derived contracting factor (EDCF), presumably thromboxane A₂^[3,4,8]. Similar conclusions have been reached in the pulmonary artery of the rat^[9] human placental vessels^[10] and afferent arterioles of the SHR^[11].

Following balloon deendothelialization, porcine

¹ Project supported in part by NIH grant HL 31547.

² Correspondence to Prof Paul M Vanhoutte, MD, PhD. IRIS, 6, Place des Pléiades, 92415 Courbevoie Cédex, France.

Phn 33-1-5572-6123. Fax 33-1-5572-7276.

E-mail vanhoutt@servier.fr

Received 1999-04-23

Accepted 1999-06-18

coronary arteries with regenerated endothelial cells exhibit not only selectively impaired pertussis-toxin sensitive endothelium-dependent relaxations, but also augmented endothelium-dependent contractions to serotonin, norepinephrine and platelets^[12-16]. This dysfunction of regenerated endothelium may play an important role in the pathogenesis of vasospasm^[15-16]. The present study was designed to determine whether or not the response to endothelin-1 is modulated by regeneration of the endothelium in the porcine coronary artery.

MATERIALS AND METHODS

Male Yorkshire pigs [8 wk of age ($n = 5$; 20–25 kg)] were used. The animals were anesthetized with Telazol (a mixture of tiletamine hydrochloride, arylaminocycloalkane, and zolazepam hydrochloride, 100 mg/animal, im) and atropine (0.4 mg, im) followed by inhalation of halothane (2 L/min). Using aseptic surgical technique, the left carotid artery was dissected free and a 7F guiding catheter (hockey stick or multipurpose) was introduced into the left coronary ostium under fluoroscopic guidance. Before denudation, heparin (100 $\mu\text{g}\cdot\text{kg}^{-1}$) and lidocaine HCl (20 mg) were administered via the arterial sheath. During the procedure, the arterial blood pressure and the electrocardiogram (ECG, lead II, avL) were monitored continuously. A 2.5 or 3 mm sized balloon catheter (USCI, over-the guide wire system) was advanced through the guiding catheter into the left anterior descending coronary artery. The balloon was then gently rubbed against the proximal 3 to 4 cm of the arterial endothelium. Successful denudation of the coronary endothelium was confirmed by ischemic ECG changes (0.1 mV of ST segment depression or elevation) and/or decreases in luminal diameter changes upon intracoronary injection of serotonin (10 $\mu\text{g}\cdot\text{kg}^{-1}$)^[8-15,17]. The animals were then housed individually in temperature-controlled animal quarters and fed regular chow. The daily food intake was limited to an amount equal to 3% of the body weight to prevent excessive weight gain. The organ chamber experiments were performed after 4 wk of feeding. All procedures were in accordance with institutional guidelines.

Organ chamber experiments After 4 wk, the

pigs were anesthetized with Telazol (100 mg im) and sodium pentobarbital (12.5 $\text{mg}\cdot\text{kg}^{-1}$ iv). The hearts were then removed. Both left coronary [anterior descending coronary artery (LAD) and circumflex (LCX)] arteries were dissected free, immersed in cold modified Krebs-Ringer bicarbonate solution [consisting of ($\text{mmol}\cdot\text{L}^{-1}$) NaCl 118.3, KCl 4.7, CaCl_2 2.5, MgSO_4 1.2, KH_2PO_4 1.2, NaHCO_3 25, glucose 11.1, and calcium disodium edetate 0.026 at pH 7.4 (control solution)], and cleaned of connective tissue. They were cut into rings (3 to 4 mm length). The proximal 3 to 4 cm portions of the LAD were used for the organ chamber experiments, and the same anatomic portions of the LCX were used as controls. In some rings, the endothelium was removed mechanically by inserting the tip of a watchmaker's forceps into the lumen and gently rolling the preparation back and forth over a paper tissue wetted with cold control solution. The rings were suspended horizontally in organ chambers filled with 25 mL of control solution (37 °C), gassed with 95% O_2 + 5% CO_2 (pH 7.4), and stretched to the optimal point of their length-active tension relation as determined by the contractile response to KCl (60 $\text{mmol}\cdot\text{L}^{-1}$) at progressive levels of stretch. The tissues were allowed to equilibrate for 60 min before beginning the experiments. The presence of the endothelium was confirmed by the occurrence of relaxations to bradykinin (0.01 $\mu\text{mol}\cdot\text{L}^{-1}$) in rings contracted with prostaglandin $\text{F}_{2\alpha}$ (2 $\mu\text{mol}\cdot\text{L}^{-1}$). Rings with and without endothelium of the same coronary arteries were studied in parallel. After one hour of equilibration concentration-response curves to endothelin-1 (0.0001 to 0.1 $\mu\text{mol}\cdot\text{L}^{-1}$) were obtained by cumulative addition of the peptide either in control solution or after incubation (45 min) with indometacin (10 $\mu\text{mol}\cdot\text{L}^{-1}$; inhibitor of cyclooxygenase), dazoxiben (100 $\mu\text{mol}\cdot\text{L}^{-1}$; inhibitor of thromboxane synthase), or ridogrel (1 $\mu\text{mol}\cdot\text{L}^{-1}$; antagonist of thromboxane A_2 receptors and inhibitor of thromboxane synthase)^[19] where indicated. Certain experiments were performed in the presence of nitro-*L*-arginine (100 $\mu\text{mol}\cdot\text{L}^{-1}$) to prevent the production of nitric oxide (NO).

Drugs The following drugs were used: bradykinin, endothelin (ET)-1, indometacin, potassium chloride, prostaglandin $\text{F}_{2\alpha}$ ($\text{PGF}_{2\alpha}$) (all from Sigma Chemical Company, St Louis Mo. USA); dazoxiben (Pfizer, Groton, Conn); N^G -nitro-*L*-arginine

(Aldrich, Milwaukee, WIS); and ridogrel (Janssen Pharmaceutica, Beerse, Belgium). All drugs were prepared with distilled water on the day of the study, except indometacin which was dissolved in water and Na_2CO_3 , and sonicated. Na_2CO_3 had no effect at the concentration of $5 \mu\text{mol} \cdot \text{L}^{-1}$. The concentrations of the drugs are expressed as final molar (M) concentration in the bath solution.

Statistical analysis The results are expressed as $\bar{x} \pm s_x$. Unless otherwise specified, n refers to the number of animals studied. Statistical evaluation of the data was performed with t -test for either paired or unpaired observations (two tailed). Values were considered to be statistically different when P was less than 0.05.

RESULTS

In the experiments which were performed in the presence of nitro- L -arginine, nitro- L -arginine induced an increase in tension, which averaged 0.88 ± 0.12 g in both left anterior descending and circumflex coronary arteries with endothelium.

Native endothelium In quiescent rings of left circumflex coronary arteries (control arteries), endothelin-1 caused concentration-dependent contractions, which were markedly less in rings than that in those without endothelium (Fig 1, left panel).

Tab 1. Effect of indometacin, dazoxiben and ridogrel on contractions evoked by endothelin-1 in porcine coronary arteries with and without endothelium.

	$\text{ED}_{50} / -\lg \text{mmol} \cdot \text{L}^{-1}$		Maximal contraction / %	
	Denuded LAD	LCX	Denuded LAD	LCX
Rings with endothelium				
Control	8.68 ± 0.05	8.73 ± 0.20	145 ± 5^c	119 ± 8
Indometacin, $10 \mu\text{mol} \cdot \text{L}^{-1}$	8.28 ± 0.03^b	8.46 ± 0.15	114 ± 2.7^b	119 ± 6
Dazoxiben, $100 \mu\text{mol} \cdot \text{L}^{-1}$	9.32 ± 0.09^b	8.96 ± 0.14	109 ± 7^b	118 ± 4
Ridogrel, $1 \mu\text{mol} \cdot \text{L}^{-1}$	8.31 ± 0.07^b	8.49 ± 0.10	115 ± 4^b	118 ± 7
Rings without endothelium				
Control	8.48 ± 0.11	8.57 ± 0.17	117 ± 3	126 ± 8
Indometacin $10 \mu\text{mol} \cdot \text{L}^{-1}$	8.34 ± 0.15	8.52 ± 0.17	128 ± 5	122 ± 6
Dazoxiben, $100 \mu\text{mol} \cdot \text{L}^{-1}$	8.79 ± 0.23	8.86 ± 0.22	103 ± 6	116 ± 5
Ridogrel, $1 \mu\text{mol} \cdot \text{L}^{-1}$	8.35 ± 0.09	8.55 ± 0.11	130 ± 7	123 ± 6

Data are expressed as $\bar{x} \pm s_x$; $n = 5$ pigs. ED_{50} : effective concentration producing 50 % of the maximal response to KCl $60 \text{mmol} \cdot \text{L}^{-1}$. Maximal contraction; maximal contraction to KCl $60 \text{mmol} \cdot \text{L}^{-1}$. Denuded; previously balloon endothelial denuded; LAD; left anterior descending coronary artery; LCX; left circumflex coronary artery. $^bP < 0.05$ compared with controls. $^cP < 0.05$ compared with LCX. All experiments were performed in the presence of nitro- L -arginine $100 \text{mmol} \cdot \text{L}^{-1}$.

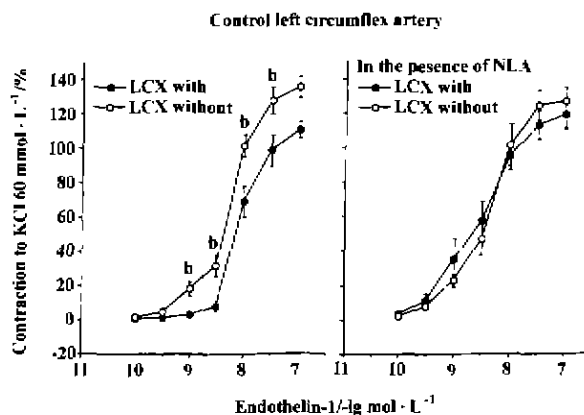


Fig 1. Cumulative concentration-response curves to endothelin-1 ($0.0001 - 0.1 \mu\text{mol} \cdot \text{L}^{-1}$) in quiescent rings, with and without endothelium, of control left circumflex porcine coronary arteries (LCX) in control solutions (left panel), and in the presence nitro- L -arginine (NLA $100 \mu\text{mol} \cdot \text{L}^{-1}$) right panel). Data shown as $\bar{x} \pm s_x$ ($n = 5$ pigs), and expressed as percent of a reference contraction to KCl ($60 \text{mmol} \cdot \text{L}^{-1}$). $^bP < 0.05$ vs with endothelium group.

However, in the presence of nitro- L -arginine, the contractions in rings with and without endothelium were comparable (Fig 1, right panel).

In the presence of nitro- L -arginine, inhibitors of the arachidonic acid cascade did not affect the contractions to endothelin-1 in rings with and without endothelium (Tab 1).

Regenerated endothelium In quiescent rings of previously deendothelialized left anterior descending coronary arteries, endothelin-1 ($0.0001 - 0.1 \mu\text{mol} \cdot \text{L}^{-1}$) caused concentration-dependent, comparable contractions in rings with and without endothelium (Fig 2, left panel).

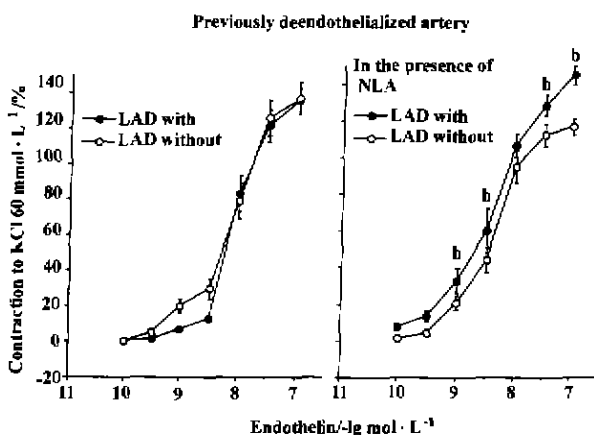


Fig 2. Cumulative concentration-response curves to endothelin-1 ($0.0001 - 0.1 \mu\text{mol} \cdot \text{L}^{-1}$) in quiescent rings with and without endothelium of previously deendothelialized porcine left anterior descending coronary arteries (LAD) in control solution (left panel), and in the presence of nitro-*L*-arginine (NLA $100 \mu\text{mol} \cdot \text{L}^{-1}$) (right panel). Data shown as $\bar{x} \pm s_x$ ($n = 5$ pigs), and expressed as percent of a reference contraction to KCl ($60 \text{ mmol} \cdot \text{L}^{-1}$). ^b $P < 0.05$ vs rings without endothelium.

Contractions to endothelin-1 ($0.3 - 0.1 \mu\text{mol} \cdot \text{L}^{-1}$) in rings of LAD with regenerated endothelium were markedly larger than those in LCX with native endothelium (Fig 1, 2, Tab 1). In the presence of nitro-*L*-arginine, endothelin-1 caused markedly larger contractions in rings than that in those without endothelium (Fig 2, right panel). In rings with endothelium of the previously deendothelialized LAD, indometacin and ridogrel caused a markedly reduced contraction, the ED_{50} was increased (Fig 3, Tab 1).

Dazoxiben did, markedly augmented the response to low, but caused a significant decrease in the response to high concentrations of endothelin-1 ($0.3 - 0.1 \mu\text{mol} \cdot \text{L}^{-1}$) (Fig 3, Tab 1). In rings without endothelium, inhibitors of the arachidonic acid cascade did not affect the contractions to endothelin-1 (Tab 1).

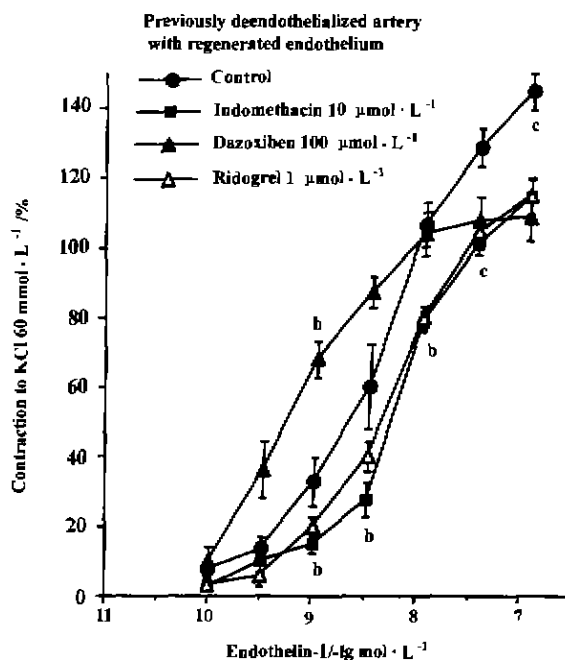


Fig 3. Effect of indometacin, dazoxiben, BQ-123 and ridogrel on contractions evoked by endothelin-1 in the presence of nitro-*L*-arginine ($100 \mu\text{mol} \cdot \text{L}^{-1}$) in quiescent rings with endothelium of previously deendothelialized left anterior descending coronary arteries. Data shown as $\bar{x} \pm s_x$ ($n = 5$ pigs), and expressed as percent of a reference contraction to KCl ($60 \text{ mmol} \cdot \text{L}^{-1}$). ^b $P < 0.05$, ^c $P < 0.01$ vs control.

DISCUSSION

The present study suggests that endothelin-1 releases endothelium-dependent contracting factors in porcine coronary arteries with regenerated endothelium. Two endothelin receptors have been cloned; one shows high specificity for endothelin-1 and is expressed mainly in vascular smooth muscle (ET_A receptor)^[18], and the other binds equally to all isoforms of the peptide and is found preferentially on the endothelium (ET_B receptor). The latter mediates the release of prostacyclin and endothelium-derived relaxing factor (EDRF)^[19]. The removal of the endothelium and treatment with an inhibitor of nitric oxide synthase augments contractions evoked by endothelins^[20-22].

In the present study, in rings with previously deendothelialized regenerated endothelium, contractions to endothelin-1 were greater than those in rings with native endothelium. This can be explained by the

depressed release of endothelium-derived relaxing factor(s), or by the release of endothelium-derived contracting factors from the regenerated endothelium, or both. In control left circumflex arteries, the contractions to endothelin in rings with endothelium were less than that in those without endothelium, and the inhibitor of NO synthase (nitro-*L*-arginine) resulted in greater contractions in rings with endothelium. These findings indicate that endothelin-1 stimulates the release of EDRF in the porcine coronary artery or, alternatively, that the basal release of EDRF attenuates the contractile response to endothelin-1. However, there was no evidence of release of cyclooxygenase-dependent contracting factors from native endothelium and vascular smooth muscle.

In chronic regenerated endothelium from previously deendothelialized LAD, the inhibitor of nitric oxide synthase (nitro-*L*-arginine) also results in greater contractions in rings with regenerated endothelium. This suggests that either the basal release of endothelium-derived relaxing factor is preserved in regenerated endothelium, or that the regenerated endothelium releases EDRF in response to endothelin-1. The effects of indometacin (inhibitor of cyclooxygenase) and ridogrel (antagonist of thromboxane A₂ receptors and inhibitor of thromboxane synthase) indicate that endothelin-1 induces the production of a cyclooxygenase-dependent, endothelium-derived contracting factor(s) by the regenerated endothelium; the action of which is mediated by a endoperoxide/thromboxane receptor on the vascular smooth muscle.

In response to lower concentrations of endothelin-1, the augmented contractions were inhibited by indometacin and ridogrel, but not by dazoxiben (inhibitor of thromboxane synthase). These observations suggest that in response to lower concentrations of endothelin-1, endoperoxides rather than thromboxane A₂ may be the cyclooxygenase-dependent, endothelium-derived contracting factor(s), as is the case in the SHR aorta [for acetylcholine and serotonin^[5-7,23]]. In fact, dazoxiben augmented the endothelium-dependent contraction in response to endothelin-1 1 nmol·L⁻¹. This may be due to blockade of thromboxane synthesis resulting in accumulation of endoperoxides in the vascular smooth muscle. Endoperoxides can cause contraction of vascular smooth muscle by activation of endoperoxide/thromboxane receptors^[24,25]. However,

in response to higher concentrations of endothelin-1, dazoxiben had a comparable inhibitory effect to those of indomethacin and ridogrel. Thus, it is logical to conclude that higher concentrations of endothelin-1 stimulate the release of cyclooxygenase-dependent, endothelium-derived contracting factor(s) which most likely is thromboxane A₂. The present findings suggest that cyclooxygenase-dependent, endothelium-derived contracting factors, thromboxane A₂ and endoperoxides, contribute to the augmented contractions to endothelin-1 in porcine coronary arteries with regenerated endothelium.

Pathophysiological implications Circulating endothelin-1 levels are increased in various ischemic heart diseases including acute myocardial infarction^[26-28]. Endothelins are potent activators of most vascular smooth muscle. However, they by themselves are not likely to contribute to acute endothelium-dependent changes in tension as they are not stored in endothelial cells. Any augmented release would require de novo protein synthesis. Thus, it is likely that endothelin-1 may be involved in long-term regulation of vascular tone^[29]. However, threshold concentrations of endothelin-1 potentiate the contractile response to norepinephrine and serotonin^[30], which may be important mediators of coronary vasospasm. These observations taken in conjunction with the present findings imply that locally increased levels of endothelin-1 may indirectly contribute to the enhanced vasoconstrictor responses through the release of EDCF.

ACKNOWLEDGMENT To Mr Bamabas DESTA, Mr Dewayne O CONEY, and Mr Daryl SCHULZ for outstanding technical help as well as Ms Marie PALUMBO for great editorial assistance.

REFERENCES

- 1 Yanagisawa M, Kurihara H, Kimura S, Tomobe Y, Kobayashi M, Mitsui Y, *et al.* A novel potent vasoconstrictor peptide produced by vascular endothelial cell. *Nature* 1988; 322: 411-5.
- 2 Schini-Kerth VB, Vanhoutte PM. Endothelin-1; a potent vasoactive peptide. *Pharmacol Toxicol* 1991; 69: 1-7.
- 3 Auch-Schwelk W, Vanhoutte PM. Contractions to endothelin in normotensive and spontaneously hypertensive rats; role of endothelium and prostaglandins. *Blood Pressure* 1992; 1: 45-9.

- 4 Taddei S, Vanhoutte PM. Role of endothelium in endothelin-evoked contractions in the rat aorta. *Hypertension* 1993; 21: 9-15.
- 5 Luscher TF, Vanhoutte PM. Endothelium-dependent contractions to acetylcholine in the aorta of the spontaneously hypertensive rat. *Hypertension* 1986; 8: 344-8.
- 6 Auch-Schwelk W, Katusic ZS, Vanhoutte PM. Contractions to oxygen-derived free radicals are augmented in the aorta of the spontaneously hypertensive rats. *Hypertension* 1989; 13: 859-64.
- 7 Auch-Schwelk W, Vanhoutte PM. Endothelium-derived contracting factor released by serotonin in the aorta of the spontaneously hypertensive rat. *Am J Hypertens* 1991; 4: 767-72.
- 8 Zerrouk A, Champeroux P, Safar M, Brisac AM. Role of endothelium in the endothelin-1-mediated potentiation of the norepinephrine response in the aorta of hypertensive rats. *J Hypertens* 1997; 15: 1101-11.
- 9 Curzen N, Griffiths MJD, Evans TW. Contraction to endothelin-1 in pulmonary arteries from endotoxin-treated rats is modulated by endothelium. *Am J Physiol* 1995; 37: H2260-H2266.
- 10 Howarth SR, Vallance P, Wilson CA. Role of thromboxane A₂ in the vasoconstrictor response to endothelin-1, angiotensin II and 5-hydroxytryptamine in human placental vessels. *Placenta* 1995; 16: 679-89.
- 11 Gonzalez MR, Villa E, Garcia-Robles R, Angulo J, Peiro C, Marin J, *et al.* Effects of indomethacin and iloprost on contraction of the afferent arterioles by endothelin-1 in juxtamedullary nephron preparations from normotensive Wistar-Kyoto and spontaneously hypertensive rats. *J Cardiovasc Pharmacol* 1996; 28: 809-16.
- 12 Flavahan NA, Shimokawa H, Vanhoutte PM. Pertussis toxin inhibits endothelium-dependent relaxations to certain agonists in porcine coronary arteries. *J Physiol (Lond)* 1989; 408: 549-60.
- 13 Shimokawa H, Flavahan NA, Vanhoutte PM. Natural course of the impairment of endothelium-dependent relaxations after balloon endothelium removal on porcine coronary arteries. Possible dysfunction of a pertussis toxin-sensitive G protein. *Circ Res* 1989; 65: 740-53.
- 14 Shimokawa H, Flavahan NA, Vanhoutte PM. Loss of endothelial pertussis toxin-sensitive G protein function in atherosclerotic porcine coronary arteries. *Circulation* 1991; 83: 652-60.
- 15 Shimokawa H, Aarhus LL, Vanhoutte PM. Porcine coronary arteries with regenerated endothelium have a reduced endothelium-dependent responsiveness to aggregating platelets and serotonin. *Circ Res* 1987; 61: 256-70.
- 16 Vanhoutte PM, Shimokawa H. Endothelium-derived relaxing factor and coronary vasospasm. *Circulation* 1989; 80: 1-9.
- 17 Shimokawa H, Vanhoutte PM. Angiographic demonstration of hyperconstriction induced by serotonin and aggregating platelets in porcine coronary arteries with regenerated endothelium. *J Am Coll Cardiol* 1991; 17: 197-202.
- 18 Arai H, Hori S, Aramori I, Ohkubo H, Nakanishi S. Cloning and expression of a cDNA encoding an endothelin receptor. *Nature* 1990; 348: 730-2.
- 19 Sakurai T, Yanagisawa M, Takawa Y, Miyazaki H, Kimura S, Goto K, *et al.* Cloning of a cDNA encoding a non-isopeptide-selective subtype of the endothelin receptor. *Nature* 1990; 348: 732-5.
- 20 Schini VB, Kim ND, Vanhoutte PM. The basal and stimulated release of EDRF inhibits the contractions evoked by endothelin-1 and endothelin-3 in aorta of normotensive and spontaneously hypertensive rats. *J Cardiovasc Pharmacol* 1991; 17 (Suppl 7): S267-S271.
- 21 Wright CE, Fozard JR. Differences in regional vascular sensitivity to endothelin-1 between spontaneously hypertensive and normotensive Wistar-Kyoto rats. *Br J Pharmacol* 1990; 100: 107-13.
- 22 Namiki A, Hirata Y, Ishikawa M, Moroi M, Aikawa J, Machii K. Endothelin-1 and endothelin-3 induced vasorelaxation via common generation of endothelium-derived nitric oxide. *Life Sci* 1992; 50: 677-82.
- 23 Ge T, Hughes H, Junquero DC, Wu KK, Vanhoutte PM, Boulanger CM. Endothelium-dependent contractions are associated with both augmented expression of prostaglandin H synthase-1 and hypersensitivity to prostaglandin H₂ in the SHR aorta. *Circ Res* 1995; 76: 1003-10.
- 24 Coleman RA, Humphrey PPA, Kennedy I, Levy GP, Lumley P. Comparison of the actions of U-46619, a prostaglandin H₂-analog, with those of prostaglandin H₂ and thromboxane A₂ on some isolated smooth muscle preparations. *Br J Pharmacol* 1981; 73: 773-8.
- 25 Moncada S, Vane JR. Pharmacology and endogenous roles of prostaglandin endoperoxides, thromboxane A₂, and prostacyclin. *Pharmacol Rev* 1979; 30: 293-331.
- 26 Emori T, Hirata Y, Aizawa T, Ando T, Ando K, Shichiri M, *et al.* Plasma endothelin levels in patients with coronary artery disease undergoing percutaneous coronary angioplasty. *Circulation* 1989; 80 Suppl II: II 2327.
- 27 Yasuda M, Kohno M, Tahara A, Itagane H, Toda I, Akioka K, *et al.* Circulating immunoreactive endothelin in ischaemic heart disease. *Am Heart J* 1990; 119: 801-6.
- 28 Salminen K, Tikkanen I, Saijonmaa O, Nieminen M, Fyhrquist F, Frick MH. Modulation of coronary tone in acute myocardial infarction by endothelin. *Lancet* 1989; 2: 747.
- 29 Vanhoutte PM. The other endothelium-derived vasoactive factors. *Circulation* 1993; 87 (Suppl V): V-9-V-17.
- 30 Yang ZH, Richard V, von Segesser L, Bauer E, Stulz P, Turina M, *et al.* Threshold concentration of endothelin-1 potentiates contractions to norepinephrine and serotonin in human arteries. A new mechanism of vasospasm? *Circulation* 1990; 82: 188-95.

内皮素-1 促进猪冠状动脉再生内皮释放
内皮衍生内过氧化物和血栓素 A₂¹

Seung-Jung PARK, John J LEE,
Paul M VANHOUTTE²
(Center for Experimental Therapeutics, Department of
Medicine, Baylor College of Medicine, One Baylor

Plaza, Houston TX 77030, USA)

关键词 冠状血管; 内皮素-1; 吲哚美辛;
前列腺素内过氧化物合酶; 一氧化氮;
血栓素 A₂; 血管内皮

吴子健

(责任编辑 朱倩蓉)

R977.6

We are pleased to announce that the
Third Workshop on
ENDOTHELIUM-DERIVED HYPERPOLARIZING FACTOR
will be held at the **Abbaye des Vaux de Cernay (France)**
between **June 14 – 16, 2000**

This Workshop will be Chaired by Prof P M VANHOUTTE (France),
and we will discuss the progress in the identification of EDHF
and the understanding of its role in physiology and pathology.

The scientific programme of the Workshop
will be constructed based on submitted Abstracts.

In order to obtain abstract and registration forms for this Workshop, please write
to **Mrs Denise MAGGI (Organization Secretariat)** at the following address,
clearly stating the number of forms required:

Servier International
22, rue Garnier-92200 NEUILLY SUR SEINE (FRANCE)
Fax (33.Ø1)01.55.72.68.77/(33.Ø1)01.55.72.72.95

Participation is limited to **75 people**,
and will be based on the quality of the submitted abstracts.

**Deadline for receipt of abstracts, registration
and hotel accommodation forms:**

JANUARY 31, 2000