Pharmacokinetics of flutamide and its metabolite 2-hydroxyflutamide in normal and hepatic injury rats

XU Chang-Jiang, LI Duan¹ (Department of Pharmacology, School of Pharmacy, Shanghai Medical University, Shanghai 200032, China)

KEY WORDS flutamide; 2-hydroxyflutamide; pharmacokinetics; liver; carbon tetrachloride poisoning; high pressure liquid chromatography

AIM: To develop a new HPLC assay to study the pharmacokinetics of flutamide (Flu) and its active metabolite 2-hydroxyflutamide (HF) in rats. METHODS: Normal or hepatic injury rats were given ig Flu 50 mg kg 1. Reverse phase HPLC was developed with a μ -Bondapak C 18 column. Internal standard was methyltestosterone. The mobile phase was a mixture of methanol: acetonitrile: water: diethyl ether = 40:20:35:1 (vol). Absorbance was measured RESULTS: The pharmacokinetic at λ_{234} nm. parameters of Flu were as follows: in normal rats, K $=0.62 \pm 0.16 \text{ h}^{-1}$, $Cl = 6.0 \pm 1.0 \text{ L} \cdot \text{kg}^{-1} \cdot \text{h}^{-1}$, AUC = $8.6 \pm 1.3 \text{ mg} \cdot \text{L}^{-1} \cdot \text{h}$, $C_{\text{max}} = 2.4 \pm 0.7$ mg·L⁻¹; in hepatic injury rats, $K = 0.16 \pm 0.03$ h^{-1} , $Cl = 0.63 \pm 0.29 \text{ L} \cdot \text{kg}^{-1} \cdot h^{-1}$, $AUC = 100 \pm$ 44 mg · L⁻¹ · h, $C_{\text{max}} = 6.7 \pm 2.8 \text{ mg} \cdot \text{L}^{-1}$. The pharmacokinetic parameters of HF were as follows: in normal rats, $K_{(m)} = 0.07 \pm 0.01 \text{ h}^{-1}$, AUC = 219 ± 22 mg·L⁻¹·h, $C_{\text{max}} = 8.6 \pm 0.6 \text{ mg·L}^{-1}$; in hepatic injury rats, $K_{(m)} = 0.05 \pm 0.01 \text{ h}^{-1}$, AUC = 170 ± 42 $mg \cdot L^{-1} \cdot h$, $C_{max} = 3.8 \pm 0.8 \text{ mg} \cdot L^{-1}$. There were significant differences between the parameters of normal and hepatic injury rats (P < 0.01) except AUC of HF (P > 0.05). CONCLUSION: This HPLC assay was sensitive and precise, and the elimination of Flu and HF was inhibited significantly due to hepatic injury.

Flutamide (Flu) is a nonsteroid antiandrogen drug employed in the treatment of prostate cancer, and it is rapidly and completely absorbed and excreted mainly through the kidney. At least 11 metabolites are present. The major metabolite 2-hydroxyflutamide

(HF) blocks androgen recepors greater than Flu. The elimination half-time of HF is greater than Flu, too^[1]. The therapeutic effect of Flu is largely due to HF *in vivo*, so it is important to mornitor the plasma levels of Flu and HF simultaneously.

Several methods for the determination of Flu and HF have been reported. They have certain limitations, such as complex detection equipment employing radioactivity and electron capture detection $(2 \text{ mL})^{(4)}$, lack of an internal standard (3-6), or only for determination of $(3 \text{ HF}^{(4.5)})$. Accordingly, we intended to develop a simple and sensitive HPLC method to determine Flu and HF simultaneously.

Flu is primarily metabolized via hepatic metabolism. Hepatic injury is associated with decreases in metabolizing enzymes and cytochrome P450 contents^[7]. The clearance of Flu might be prolonged due to hepatic injury. This study was designed to compare the pharmacokinetics of Flu and its active metabolite HF, and study the excretion in urine during hepatic injury.

$$(CH_3)_2C$$
 $\stackrel{R}{\longrightarrow}$ $\stackrel{O}{\longrightarrow}$ $\stackrel{H}{\longrightarrow}$ $\stackrel{CF_3}{\longrightarrow}$ NO_2

R = H Flutamide OH 2-Hydroxyflutamide

MATERIALS AND METHODS

Reagents Flu (yellow crystal, purity > 99 %, mp 113 – 114.5 °C) was made by Hongqi Pharmaceutical Factory of Shanghai Medical University. Suspension of Flu was prepared in 0.5 % sodium carboxymethyl cellulose at the concentration of 5.0 g $^{\circ}$ L⁻¹. HF was synthesized by Prof XIA Peng (Department of Organic Chemistry, School of Pharmacy, Shanghai Medical University). Methanol and acetonitrile were HPLC reagents. Cyclohexane, diethyl ether, and CCl₄ were of AR. Methyltestosterone was used as an internal standard, and purchased from Sigma.

¹ Correspondence to Prof LI Duan. Phn 86-21-6404-1900, ext 2558. Fax 86-21-6403-7260. E-mail dli@shmu.edu.cn Received 1997-03-14 Accepted 1997-07-02

Standard solution Stock solution of Flu or HF was prepared in methanol at the concentration of 1.0 g $^{\circ}$ L⁻¹, and stored below 4 $^{\circ}$ C.

Tab 1. Serum biochemistry after CCl₄ poisoning. n = 5 rats, $x \pm s$. ${}^{a}P > 0.05$, ${}^{c}P < 0.01$ vs normal.

Time/h	AlaAT/U	Bil/mg·L-1	Alb/g·L-1
0	28 ± 11	7.4 ± 2.5	23 4±1.7
24	$440 \pm 248^{\circ}$	$168 \pm 121^{\circ}$	23.0 ± 2.0^{a}
48	$1.000 \pm 600^{\circ}$	$78 \pm 49^{\circ}$	25.2 ± 1.9^{a}
72	455 ± 145°	19.2 ± 0.2^{c}	23.4 ± 2.1^{a}

Medication and sampling Rats were given ig Flu 50 mg · kg⁻¹. Blood samples were collected at 0.5, 1, 1.5, 2, 3, 4, 6, 8, 10, 12, 24, and 48 h via caudal vein heparinized, plasma was separated. Urine samples were collected by individual metabolic cage. Plasma 0.1 mL or urine 0.5 mL with internal standard methyltestosterone (15 mg · L⁻¹, 0.75 μ g in methanol 0.05 mL) and 1 mL pH 7.6 phosphate buffer solution (0.01 mol·L⁻¹) was extracted with 3 mL of organic solution (cyclohexane : diethyl ether = 9 : 1, vol/vol) twice, after centrifugation (2000 × g, 15 min), the solvent was evaporated at 50 °C under a nitrogen flow. The residue was dissolved in 50 μ L of methanol, and 10 μ L was injected for HPLC.

HPLC Waters HPLC system consisted of 510 pump, 486 UV detector, U6k injector (Millipore Corporation, USA). HPLC column: μ -Bondapak C 18, 10 μ m, 3,9 mm × 300 mm (USA). An equilibrating model recorder XWT-104 was the product of Shanghai Dahua Instrument Factory. Mobile phase was a mixture of methanol acetonitrile: water diethyl ether = 40: 20: 35:1 (vol), prepared and degassed daily. Chromatography assay was performed at room temperature (20 °C) using a flow rate of 1 mL·min⁻¹ which produced a back pressure of 2000 PSI. Absorbance was measured at 234 nm with 0.03 absorbance units of full scale (AUFS).

Quantitation and linearity The standard curves showed a good linearity over a range of $0.1 - 20 \text{ mg} \cdot \text{L}^{-1}$ for Flu (plasma $\hat{Y} = -0.054 + 1.41 \text{ } X$, r = 0.9989; urine $\hat{Y} = -0.0078 + 1.11 \text{ } X$, r = 0.9993) and HF (plasma $\hat{Y} = -0.0065 + 1.11 \text{ } X$, r = 0.9998; urine $\hat{Y} = -0.019 + 1.20 \text{ } X$, r = 0.9994).

Recovery Extracting recovery for the method was determined by evaluating the peak height ratio of extracted plasma to unextracted methanol standards by adding 0.1, 0.5, or $2.0~\mu g$ Flu and HF to 0.1~mL blank plasma or 0.5~mL blank urine (Tab 2).

Tab 2. Extracting recovery of Flu and HF in plasma or urine. n = 4 experiments, $x \pm s$.

Added/	Plasma\`%		Urine %	
mg·L-1	Flu	HF	Ðμ	HF
1	94 ± 7	93 ± 9	89±8	88 ± 7
5	95 ± 4	96.0 ± 2.0	90 ± 7	89 ± 8
20	98 ± 5	96 ± 10	94±5	92 ± 6

The accuracy and precision of the method were determined by calculating the coefficiency of variation (CV) at each control concentration of either Flu or HF (Tab 3).

Tab 3. Combined intra and inter-day accuracy and precision. n=5 experiments, $x \pm s$.

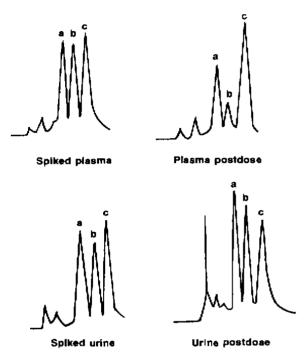
Dru /mg	g*L-1	Intra-day /mg·L ⁻¹	CV %	lmer-day ≠mg·L ⁻¹	CV %
flu	1.0 5.0 20.0	1.04 ± 0.01 5.03 ± 0.07 20.1 ± 0.8	6.46 1.39 1.09	1.04 ± 0.05 5.01 ± 0.11 20.1 ± 1.0	5.02 2.15 5.14
HF	1.0 5.0 2 0.0	0.96 ± 0.06 5.03 ± 0.11 19.9 ± 1.2	5.71 2 22 6.14	$ 1.00 \pm 0.07 5.01 \pm 0.12 20.5 \pm 0.8 $	6,70 2,21 3,82

Pharmacokinetics The pharmacokinetic parameters of Flu and its metabolite HF were calculated according to the method^[9]. The data were analyzed with a PK-GRAPH software (provided by Department of Pharmacology, Shanghai Second Medical University) on a 586 personal computer to determine the compartment model and the pharmacokinetic parameters. Comparison of pharmacokinetic parameters between groups was carried out with *F*-test.

- 41 -

Chromatography There were no endogenous interferences at the retention time (T_r) of Flu, HF, and methyltestosterone from the biological drug-free control (Fig. 1). The T_r of HF, Flu, and methyltestosterone were 4.5, 5.5, and 6.6 min, respectively. The minimal detection concentrations of Flu and HF were 0.05 and $0.1~\text{mg} \cdot L^{-1}$, respectively, at signal-to-noise ratio of 2. The detection limits of

Flu and HF were 1 and 2 ng, respectively.

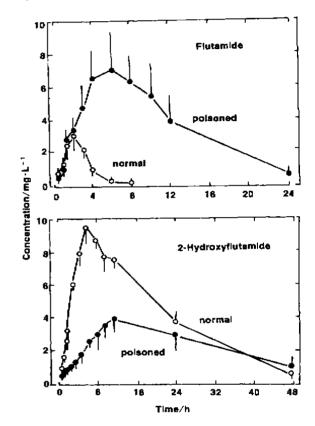


Chromatograms of HF (a), Flu (b), and Fig 1, methyltestosterone (c).

Pharmacokinetics The curve for Flu after ig in rats was fitted with 1-compartment model. elimination was inhibited due to hepatic injury in rats, K decreased from 0.62 ± 0.16 to 0.16 ± 0.03 h⁻¹(P < 0.01), $T_{\frac{1}{2}}$ was prolonged from 1.19 ± 0.29 to 4.4 $\pm\,0.8$ h ($P\,{<}\,0.01$), clearance decreased from $6.0\,\pm\,$ 1.0 to $0.63 \pm 0.29 \text{ L} \cdot \text{kg}^{-1} \cdot \text{h}^{-1} (P < 0.01)$. The AUC of Flu was higher in rats with hepatic injury (100 $\pm 44 \text{ mg} \cdot \text{L}^{-1} \cdot \text{h}$) than that in normal rats (8.6 ± 1.3) $\text{mg} \cdot \text{L}^{-1} \cdot \text{h}$) (P < 0.01). The V_d for Flu was lower in rats with hepatic injury $(3.9 \pm 1.8 \text{ L} \cdot \text{kg}^{-1})$ than that in normal rats $(10.2 \pm 2.4 \text{ L} \cdot \text{kg}^{-1})$ (P < 0.01).

 $K_{(m)}$ is the constant of elimination rate of

 $K_{(m)}$ of HF were 0.07 ± 0.01 h⁻¹ in metabolite. normal rats and $0.05 \pm 0.01 \text{ h}^{-1}$ in hepatic injury rats $T_{\frac{1}{2}}$ was prolonged from 9.9 ± 0.7 to (P < 0.01). 14.8 ± 1.9 h due to hepatic injury (P < 0.01). AUC of HF tended to be lower in rats with hepatic injury (P >0.05). The elimination of HF was inhibited, too. The elimination of HF was fitted with elimination rate limited (ERL) model, and K was greater than $K_{(m)}$ (Fig 2, Tab 4).



Flu and HF concentrations of plasma in normal and CCl₄-poisoned rats. n = 5 rats, $x \pm s$.

Excretion in urine Percentage of cumulative excretion of Flu in urine in normal and hepatic injury rats at the dosage of 50 mg \cdot kg⁻¹ were 0.078 % \pm 0.023~% and $0.078~\% \pm 0.027~\%$, respectively (n =5 rats, P > 0.05); but there was difference in % of cumulative urinary excretion of HF between normal and hepatic injury rats; $0.067 \% \pm 0.024 \%$ vs 0.11 % ± 0.023 % (n = 5 rats, P < 0.05).

DISCUSSION

We developed a simple and sensitive procedure which could be employed for the analysis of Flu and

Tab 4. Pharmacokinetics of Flu and HF in normal and CCl₄-poisoned rats. n = 5 rats, $x \pm s$. P < 0.01 vs normal of Flu, P > 0.05, P < 0.01 vs normal of HF.

D	Flutamide		2-Hydroxyflutamide	
Parameter	Normal	Poisoned	Normal	Poisoned
$K_{\rm s}/{\rm h}^{-1}$	1.03 ± 0.24	$0.24 \pm 0.08^{\circ}$		
K/h ⁻¹	0.62 ± 0.16	$0.16 \pm 0.03^{\circ}$		
$K_{(m)}/h^{-1}$			0.07 ± 0.01	$0.05 \pm 0.01^{\circ}$
$T_{\frac{1}{2}}/h$	1.19 ± 0.29	$4.4 \pm 0.8^{\circ}$	9.9 ± 0.7	$14.8 \pm 1.9^{\circ}$
$V_{\rm d}/{\rm L}\cdot{\rm kg}^{-1}$	10.2 ± 2.4	$3.9 \pm 1.8^{\circ}$		
$Cl \cdot F^{-1} / L \cdot kg^{-1} \cdot h^{-1}$	6.0 ± 1.0	$0.63 \pm 0.29^{\circ}$		
AUC∕mg·L ^{−1} ·h	8.6 ± 1.3	$100 \pm 44^{\circ}$	219 ± 22	170 ± 42^{d}
$T_{ m max}/{ m h}$	1.61 ± 0.27	$5.8 \pm 1.5^{\circ}$	8.4 ± 0.4	$15.8 \pm 2.0^{\circ}$
$C_{\rm max}/{ m mg}\cdot { m L}^{-1}$	2.4 ± 0.7	$6.7 \pm 2.8^{\circ}$	8.6 ± 0.6	$3.8 \pm 0.8^{\circ}$

HF simultaneously. The three-dimentional structures of Flu and HF were similar to testosterone, so we used methyltestosterone as the internal standard for HPLC assay. The maximum absorbance wavelengths of Flu, HF, and methyltestosterone were 227, 226, and 241 nm, respectively. They all had evident absorbance at 234 nm. The detection limits of Flu and HF were 1 and 2 ng, respectively. The mobile phase had no inorganic salt, and its pH approximated to 5.6. There were three HPLC methods for analysis of Flu or HF^[4-6]. They were all lack of internal standard. There was only one method for analysis of Flu and HF simultaneously^[6]. Its detection limits of Flu and HF were 1.5 and 4.5 ng, respectively. The pH of mobile phase was 2.9, it would threat the column life. extraction procedure was simpler than ours. The other two methods were only used to determine HF. There was phosphate in the second method, and its detection limit was 6 $ng^{(5)}$. The plasma sample volume was 2 mL in the third method, and its detection limit was 2 ng⁽⁴⁾. There was one gas - liquid chromatographic method⁽²⁾ using an internal standard to determine Flu and HF, but it was limited by employing 63Ni electron capture detector. The internal standard was 4-nitro-3trifluoromethyl-hexananilide, which was difficult to obtain.

$$A \xrightarrow{K_{m} = f_{m} \cdot K} A_{(m)} \xrightarrow{K_{(m)}} A_{e(m)}$$

 $K_{\rm m}$ is the constant of formation rate of metabolite, and $f_{\rm m}$ is fraction of metabolite formation. $f_{\rm m}=1$ is the prerequisite for the following equation:

$$C_{(m)} = \frac{K \cdot c}{K_{(m)} - K} (e^{-K_1} - e^{-K_{(m)}t})$$

While K is greater than $K_{(m)}$, after enough time, e^{-Kt} tends to zero, so we can calculate $K_{(m)}^{(9)}$.

Our data showed the elimination of active metabolite HF was fitted with ERL model, $K > K_{(m)}$ both in normal and hepatic injury rats. $K_{(m)}$ and $T_{\frac{1}{2}}$ of HF were $0.07 \pm 0.01 \ h^{-1}$ and 9.9 ± 0.7 h in normal rats, respectively. If rats were given ig HF 50 mg·kg⁻¹ directly, its K and $T_{\frac{1}{2}}$ were $0.14 \pm 0.02 \ h^{-1}$ and 4.9 ± 0.7 h, respectively (unpublished data). Our results suggested there might be feedback inhibition of HF elimination due to other metabolites while the rats were given Flu.

The decreases in $V_{\rm d}$ and clearance of Flu resulted in statistically significant changes in K and $T_{\frac{1}{2}}$ in rats with hepatic injury. These results reflected a slower metabolic clearance of Flu with hepatic injury in rats, resulting in a greatly reduced first-pass effect. The AUC of the metabolite HF showed a lower trend in rats with hepatic injury (P > 0.05).

REFERENCES

- Katchen B, Buxbaum S. Disposition of a new, nonsteroid, anfiandrogen, α, α, α-trifluoro-2-methyl-4'-nitro-m-propionotoluidide (Flutamide), in men following a single oral 200 mg dose.
 Clin Endocrinol Metab 1975; 41: 373 9.
- 2 Radwanski E, Perentesis G, Symchowicz S, Zampagtione N. Single and multiple dose pharmacokinetic evaluation of flutamide in normal genatric volunteers. J Clin Pharmacol 1989; 29: 554-8.
- 3 Schulz M, Schmoldt A, Donn F, Becker H. The pharmacolunetics of flutamide and its major metabolites after a single oral dose and during chronic treatment. Eur J Clin Pharmacol 1988; 34; 633 – 7.
- 4 Asade RH, Prizoni L, Muino JP, Tessler J.

Steady-state hydroxyflutamide plasma levels after the administration

Cancer Chemother Pharmacol 1991: 27: 401 - 5.

- 5 Belanger A, Giasson M, Couture J, Dupont A, Cusan L, Labrie F. Plasma levels of hydroxy-flutamide in patients with prostatic cancer receiving the combined hormonal therapy: an LHRH agonist and flutamide. Prostate 1988; 12: 79-84.
- 6 Farthing D., Sica D., Fakhry I., Walters DL., Cefali EA, Allan G. Determination of flutamide and hydroxytlutamide in dog plasma by a sensitive high performance liquid chromatography method utilizing mid-bore chromatography. Biomed Chromatogr 1994; 8: 251 4.
- 7 Howden CW, Birnie GG, Brodie MJ. Drug metabolism in liver disease. Pharmacol Ther 1989; 40; 439 – 74.

of two dosage forms of flutamide

- 8 Huang WY, Liu GT. Mechanism of the protective action of kopxinine against hepatotoxicity of carbon tetrachloride. Acta Pharmacol Sin 1989; 10: 461-4.
- 9 Li D. Drug active metabolite and clinical monitoring. New Drugs Clin Remedies 1987; 6: 102-6.

戴他胺及其代谢产物 2-羟基氟他胺 在正常及肝损伤大鼠的药物动力学

许长江,李 端

R979.1

(上海医科大学药学院药理学教研室,上海 200032,中国)

关键词 氟他胺; 2-羟基氟他胺; 药物动力学; 肝; 四氯化碳中毒; 高压液相色谱法 BF+16/15

目的:建立一新的高压液相色谱法用来研究氟他 胺(Flu)及其活性代谢产物 2-羟基氟他胺(HF)的药物动力学. 方法:正常及肝损伤大鼠灌胃 Flu 50 mg·kg⁻¹. 采用反相高压液相色谱法,以甲基睾丸素为内标,流动相为甲醇:乙腈:水:乙醚 \approx 40:20:35:1 (体积比),检测波长为 234 nm. 结果: Flu 的 K与Cl分别由 0.62 ± 0.16 h⁻¹及 6.0 ± 1.0 L·kg⁻¹·h⁻¹减小到 0.16 ± 0.03 h⁻¹及 0.63 ± 0.29 L·kg⁻¹·h⁻¹(P<0.01),AUC 与 C_{max} 分别由 8.6 ± 1.3 mg·L⁻¹·h 及 0.7 ± 0.01 h⁻¹减小到 0.05 ± 0.01 h⁻¹减小到 0.05 ± 0.01 h⁻¹(P<0.01). 结论:在肝损伤大鼠,Flu 与 HF 消除受到显著抑制.

5th International Chinese Peptide Symposium 1998

1998 Jul 14 -- 18

Lanzhou, China

Abstracts of researches in all areas of peptides are welcome. The official language of this symposium is English. The abstract must be received before April 1, 1998.

Please contact:

Dr WANG Rui

Department of Biology, Lanzhou University

Lanzhou 730000, China

Phn 86-931-891-3382 or 86-931-891-2567. Fax 86-931-861-0862. E-mail wangrui@lzu.edu.cn