Biochim Biophys Acta 1994; 1191; 384-8.

Bouchelouche P, Friche E, Sebested M, Jensen PB. Skovsgaard T Cytosolic free Ca²⁺ in daunorubicin and vincustine resistant Ehrlich ascites tumor cells. Drug accumulation is independent of intracellular Ca²⁺ changes.

Biochem Pharmacol 1991; 41: 243 - 53

71-80

2-苯-3-(3', 5'-双吗啉甲基-4'-羟基)-苯甲酰吲哚 (HWL-12)逆转肿瘤多药抗药性

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Fura-2; 钙; 培养的肿瘤细胞; 维拉帕米; 依他酸

目的: 採讨吲哚衍生物 HWL-12 逆转 MDR 的作用 及其作用机制。方法: 细胞毒的测定用 MTT 法; P-糖蛋白的测定用我们新近建立的 Fura 2-AM 法。细胞内阿霉素(Dox)的积累测定用荧光分光光度 计法。结果: HWL-12 能显著地增加 MDR 细胞内 Fura-2 的积累和逆转 MDR 17.2 倍。在 HWL-12 的作用下,MDR 细胞内阿霉素的积累亦明显增加,但在介质中高钙或低钙离子浓度对 Dox 积累均无影响。结论: HWL-12 具有较强的逆转作用,它的逆转机制可能与增加 MDR 细胞内阿霉素的积累有关,而与钙离子浓度无关。

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Effect of catecholamic acid on detoxication and distribution of NiCl₂ in mice and rats¹

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KEY WORDS chelating agents; catecholamic acid; nickel; lethal dose 50; autoradiography; tissue distribution

AIM: To study the effect of catecholamic acid (CBMIDA) on detoxication of NiCl₂. METHODS: Mice and rats were injected sc or im CBMIDA immediately after ip NiCl₂. Each mouse was injected ip CBMIDA after iv ⁶³NiCl₂ 185 kBq, and radioactivities of various tissues were measured with liquid scintillation counter at 24 h. The localization of ⁶³Ni was shown by the whole-body autoradiography. RESULTS: CBMIDA sc 0.5 – 1.5 g·kg⁻¹ markedly reduced the mortality from acute poisoning of ip NiCl₂ 500 mg·kg⁻¹. After ip NiCl₂ in mice, the LD₅₀ was 82.7 mg·kg⁻¹. Mice were injected sc CBMIDA 1.5 or 2.5 g·kg⁻¹ after Ni poisoning, the LD₅₀ of NiCl₂ were raised to 789 or 820 mg·kg⁻¹, respectively.

The LD₅₀ of NiCl₂ was 39 mg · kg⁻¹ in rat. If CBMIDA was injected im 0.5 g·kg⁻¹ after ip NiCl₂, the LD₅₀ was 332 mg \cdot kg⁻¹. CBMIDA 1.5 g \cdot kg⁻¹ im after iv 63 NiCl2, decreased the contents of 63 Ni in blood and lung of mice vs control mice at 24 h. The contents of 63 Ni in brain, heart, spleen, and kidney were similar to those of the control mice. The content of 63 Ni in bone was more than the control. excretions of 63 Ni through urine and feces were not increased by CBMIDA at 24 h. The whole-body autoradiograpy showed that the radioactivity was highly localized in the kidney, lung, and Harder's gland. There was a moderate level of 63 Ni in the liver, bone, skin, and blood. A pronounced accumulation occurred in the bone. There was a marked reduction of 63 Ni in the lung, skin, liver, and blood after ip CBMIDA. CONCLUSION: The CBMIDA markedly raised the survival rate of nickel-poisoned mice and rats, and decreased 63 Ni levels in lung and blood.

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Nickel compounds widely used in industry were absorbed into the body and deposited mainly in lung. so the lungs are the critical organ after exposure to A long period of exposure to nickel compounds may cause lung cancer[1], and most chelating agents are ineffective to treat nickel poisoning.

Catecholamic acid [catechol-3, 6-bis(methyleneiminodiacetic acid), CBMIDA] first synthesized by Prof XIE Yu-Yuan in our Institute was a chelating agent of a low toxicity[2], and increased excretion of uranium⁽³⁾ and plutonium⁽⁴⁾ in rats.

Catecholamic acid (CBMIDA)

The purpose of the present study was to explore the effects of CBMIDA on detoxication, removal, and tissue distribution of NiCl₂.

MATERIALS

Kunming mice (\frac{1}{2}) and Sprague-Dawley rats (\frac{1}{2}) were obtained from Shanghai Experimental Animal Center, Chinese Academy of Sciences, Certificate No 005.

Catecholamic acid was synthesized by Department of Synthetic Chemistry in our Institute, purity > 95 % and mp 184 °C. It was dissolved in distilled water with the aid of $NaHCO_3$, pH 6 - 7, and prepared before 30 min administration. NiCl₂(AR) was produced by Shanghai Qingong Chemical Plant. ⁶³ NiCl₂ was made by the Radiochemical Center Ltd., UK (radioactivity 247 GBq • L-1). The X-ray film was LKB Ultrafilm S-161 25 Bromma, Sweden. The whole-body section was made on Lipshaw Cryotome 1800-N freezing microtome, USA. Liquid scintillation counter (model YSJ-78) was made by Shanghai Institute of Nuclear Research, Chinese Academy of Sciences.

METHODS AND RESULTS

Detoxication of nickel by CBMIDA (weighing $20.0 \pm s + 1.6 \text{ g}$) injected ip NiCl₂ 500 mg·kg⁻¹ were divided into 5 groups, 20 mice/group. Then the control group mice were injected sc 0.9~%

saline, and other 4 groups were immediately injected sc CBMIDA 0.5, 1, 1.5, and 2 $g \cdot kg^{-1}$, respectively. The mortality was recorded within 3 d. diarrhoea, respiratory distress, and lethargy were noted in toxicity tests. All mice of control group died. In the CBMIDA-treated groups, $0.5 - 1.5 \text{ g} \cdot \text{kg}^{-1}$, exhibited a dose-response relationship (Tab 1).

Tab 1. Detoxificating effect of sc catecholamic acid (CBMIDA) after ip NiCl₂ 500 mg·kg⁻¹ within 3 d. P < 0.01 vs 0; P > 0.05, P < 0.05 vs 0.5; P > 0.05

CBMIDA/g·kg ⁻¹	Injected mice	Survived mice
0	20	0
0.5	20	13°
1.0	20	17 ^{od}
1.5	20	20^{ceg}
2.0	20	20 ^{oeg}

Various chelating agents on detoxication of **nickel** Mice (weighing $19.8 \pm s \cdot 1.3$ g, divided into 9 groups, 40 mice/group) were injected ip NiCl₂ 70. 150, 250, and 500 mg · kg⁻¹, (n = 10). After nickel poisoning, 7 chelating agents were injected sc: CBMIDA 1.5 or 2.5; sodium dimercaptosuccinate (Na-DMS) 1; qunamic acid (QA) 0.5; penicillamine (PCA) 0.3; 1,2-cyclohexylene dinitrilotetraacetic acid (CDTA) 0.15; edetic acid 0.15; dithiocarb (DDC) 0.5 g·kg⁻¹. CBMIDA remarkably raised the survival rate of mice, as compared with other chelating agents. The effect of CBMIDA 1.5 g kg -1 was similar to that of $2.5 \text{ g} \cdot \text{kg}^{-1}(\text{Tab } 2)$.

Tab 2. Survived mice after sc 7 chelating agents after ip NiCl₂ (n=10).

$NiCl_2/mg \cdot kg^{-1}$					
Chelator/g·kg	Ţ ⁻	70	140	250	500
		3	0	0	0
CBMIDA	1.5	10	10	10	10
	2.5	10	10	10	9
CDTA*	0.15	10	1	2	0
Dithiocarb	0.5	0	1	1	0
Edetic acid	0.15	10	8	2	0
Na-DMS	1	10	10	6	2
Penicillamine	0.3	10	10	0	0
Qunamic acid	0.5	10	7	0	0

^{*1,2-}cyclohexylene dinitrilotetraacetic acid

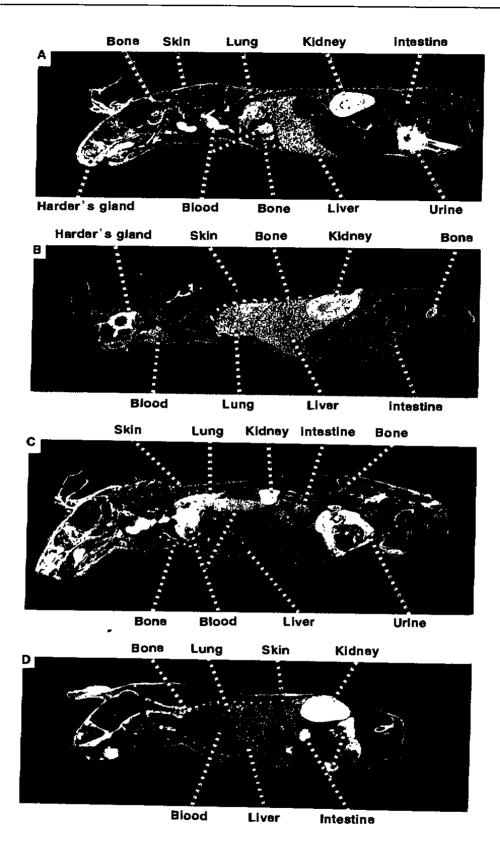


Fig 1. Autoradiograms from $\hat{\gamma}$ mice iv 63 NiCl₂ alone or followed by CBMIDA 3 h after Ni (A) or Ni + CBMIDA (B); 8 h after Ni (C) or Ni + CBMIDA (D).

Effects of chelating agents on LD₅₀ of NiCl₂ Mice (n = 260, weighing $20.4 \pm s \cdot 1.5 \text{ g}$) were divided into 5 groups. Control group was injected up only NiCl₂ at various doses (n = 10). Other 4 groups were respectively injected sc: CBMIDA 1.5 or 2.5, Na-DMS 1, edetic acid 0.15 g·kg⁻¹ after ip NiCl₂. The 3-d LD₅₀ (95 % confidence limits) were calculated by Finney's way.

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After ip a single dose to mice, the LD_{50} of nickel chloride was $82.7~\text{mg} \cdot \text{kg}^{-1}$, they were immediately injected so chelating agents, the LD_{50} of $NiCl_2$ was raised as the following order: CBMIDA > Na-DMS > edetic acid (Tab 3).

Tab 3. LD_{50} of ip NiCl₂ alone or followed by chelating agents.

Specie	s n	Chelator	Dose/g·kg ⁻¹	Route	LD ₅₀ /mg·kg ⁻¹ (95 % confidence limits)
Mice	70	_	_	_	82.7 (76.9 - 88.9)
	50	CBMIDA	1.5	sc	789 (693 – 899)
	60		2.5	SC	820 (693 – 970)
	40	Edetic acid	0.15	sc	187 (152 - 230)
	40	Na-DMS	I	sc	310 (245 - 392)
Rats	50	_	-	_	38.8 (36.7-41.0)
	50	CBMIDA	0.5	im	332 (305 – 362)

Rats (n = 100, weighing $182 \pm s 21$ g) were divided into 2 groups. The toxic group was injected ip various doses of NiCl₂. The detoxification group was injected im CBMIDA $0.5 \text{ g} \cdot \text{kg}^{-1}$ after ip NiCl₂. The LD₅₀ of NiCl₂ was elevated from 39 to 332 mg·kg⁻¹, about 8.6 times of that in toxic group (Tab 3).

Effect of CBMIDA on ⁶³Ni tissue distribution and elimination Mice (weighing $20.4 \pm s$ $0.5 \,\mathrm{g}$) were divided into 2 groups, 10 mice/group. The control group was injected iv ⁶³NiCl₂ 185 kBq/mouse; the treated group was injected ip CBMIDA 1.5 $\mathrm{g} \cdot \mathrm{kg}^{-1}$ after iv ⁶³NiCl₂. Each mouse was kept in a metabolism cage for 24 h, and then killed. The heart, lung, brain, liver, spleen, kidney, bone, and blood were weighed, and $7-15 \,\mathrm{mg}$, were digested in 0.1 mL formic acid and 0.1 mL $\mathrm{H_2O_2}$ at 80 °C for 30 min. Radioactivity was measured in 8 mL of 0.01 % POPOP and 0.4 % PPO with liquid scintillation counter for 1 min. In the CBMIDA-treated group, the contents of ⁶³Ni were reduced remarkably in blood and

lung, but increased in bone. ⁶³Ni in urine, feces, and other tissues were similar to those of control group at 24 h (Tab 4).

Tab 4. Tissue radioactivity at 24 h after iv 63 NiCl₂ 185 kBq and ip CBMIDA 1.5 g · kg⁻¹ in mice n = 10. 6 P > 0.05, 6 P < 0.01 vs control.

	Radioactivity (kBq/g wet tissue)		
	Control	CBMIDA	
Blood (L)	967 ± 367	433 ± 100°	
Bone	0.52 ± 0.13	$1.8 \pm 0.7^{\circ}$	
Brain	0.55 ± 0.22	0.38 ± 0.18^{a}	
Heart	0.60 ± 0.25	0.47 ± 0.22^a	
Kidney	2.9 ± 1.1	2.8 ± 1.2^{a}	
Liver	$\mathbf{0.8 \pm 0.4}$	0.7 ± 0.3^{a}	
Lung	1.9 ± 0.5	$0.55 \pm 0.22^{\circ}$	
Spleen	1.1 ± 0.6	1.1 ± 0.8^{a}	
Urine (24 h)	105 ± 33	$109 \pm 37^{\rm s}$	
Feces (24 h)	15 ± 14	17 ± 16^{4}	

Effect of CBMIDA on ⁶³Ni localization in mice Mice (n = 4, weighing 28 g) injected iv ⁶³NiCl₂ 370 kBq/mouse were divided into 2 groups, 2 mice served as control, the other 2 mice were immediately injected ip CBMIDA 1.5 g·kg⁻¹. At 3 and 8 h, respectively, 1 mouse in each group was anesthetized with ether, and frozen in mixture of dry ice and hexane (-75°C). Autoradiography was made^[5]. Whole-body sections were made 20 μ m in thickness, at -20°C, the X-film was exposed for 43 wk for the 3-h group, and exposed for 48 wk for the 8-h group.

At 3 h after the mice were injected iv ⁶³NiCl₂, the concentrations were high in kidney, lung, and Harder's gland; moderate in bone, liver, skin, and blood. There were trace radioactivities in intestines (Fig 1A). In the CBMIDA-treated mice, the radioactivities were high in kidney, bone, and Harder's gland; moderate in lung, liver, and blood. Remarkable accumulation of ⁶³Ni was found in bone, and reduced in lung and skin (Fig 1B).

At 8 h, in general, radioactivities were reduced in tissues after iv ⁶³NiCl₂. The radioactivities were moderate in kidney, lung, blood, and urine, and low in bone, intestine, skin, and liver (Fig 1C). In CBMIDA-treated mice, the levels were increased in kidney, bone, intestine, and skin, but remarkably decreased in lung and blood (Fig 1D).

DISCUSSION

Antidote principle of chelating agent was to combine with metal ion to form complex with a lower toxicity, it reduced the metal ion to the target organs poisoned or enhanced the urinary excretion of the ion. The present study showed that CBMIDA markedly raised the survival rate of NiCl2-poisoned mice and rats, but the excretion of 63Ni through urine and feces was not increased at 24 h. The detoxificating effect of CBMIDA on nickel might be that the chelate was combined with nickel, to form complex of lower toxicity in body.

The tissue distribution and localization of 63Ni were similar. The 63 Ni deposited in organs showed as the reports^[6], whereas given CBMIDA is more effective for removing nickel from lung, but to accumulate in bone. The metabolism of CBMIDA was mainly deposited in bone except the excretion⁽⁷⁾. It might be that CBMIDA formed the complex of CBMIDA-Ni, and enhanced the ⁶³Ni content in bone due to its action in affinity bone.

In conclusion, CBMIDA markedly raised survival rate of nickel-poisoned mice and rats and decreased content of ⁶³Ni in lung and blood.

REFERENCES

- 1 Nieboer E., Rossetto FE., Menon CR. Toxicology of nickel compounds. In: Sigel H, Sigel A, editors Metal ions in biological systems; vol 23. Nickel and its role in biology. New York; Marcel Dekker; 1988. p 359 - 402.
- 2 Tao ZQ, Xu XH, Yan XM, Chen ZJ, Zhang JS, Liang YY, et al. Pharmacological studies on catecholamic acid, an efficient chelating agent for metal mobilization.

Pizen lek Sborn 1988; 56 Suppl; 95 - 7.

3 Tao ZQ, Xu XH, Yan XM, Chen ZJ, Zhang JS, Liang YY, et al. Detoxication and mobilization of uranium by catecholamic acid. Acta Pharmacol Sin 1987; 8; 284 - 8.

- 4 Fukuda S, Inda H, Hseih YY, Chen WZ. Effect of CBMIDA [catechol-3, t-bis (methyleiminodiacetic acid)] on removal of plutonium in rats. Hoken Butsuri 1992; 27; 11-5.
- 5 Waddekk WJ, Marlowe C. Autoradiography. In: Garrett ER, Hirtz JL, editors. Drug fate and metabolism; methods and techniques; vol 1. 1st ed. New York: Marcel Dekker; 1977. pt-25.
- 6 Oskarsson A, Tjälve H. Autoradiography of nickel chloride and nickel carbonyl in mice. Acta Pharmacol Toxicol 1977; 41 Suppl 1: 158-9.
- Zhang JS, Liang YY, Yan XM. Whole-body autoradiographic localization of catecholamic acid in mice.

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双酚胺酸对小鼠和大鼠氯化镍解毒及分布的影响1

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螯合剂; 双酚胺酸; 镍; 半数致死量; 放射自显影术;组织分布

目的: 研究双酚胺酸(CBMIDA)对氯化镍的解毒作 用. 方法: NiCl。中毒后,立即给予 CBMIDA,记 录动物存活数;小鼠 iv 63NiCl。后给药,测定 24 h 组织中63镍; 用整体放射自显影术, 显示小鼠体 内ⁿ³镍分布. 结果; sc CBMIDA 0.5 - 1.5 g·kg⁻¹ 对 ip NiCl₉ 500 mg·kg⁻¹有解毒作用;小鼠 ip NiCl₉ LD₅₀为 82.8 mg·kg⁻¹, 给药 1.5 或 2.5 g·kg⁻¹, LD₅₀分别为 789 和 820 mg·kg⁻¹; 大鼠 im CBMIDA 500 mg·kg-1使 NiCl。的 LDso提高 8 倍;组织中的镍 测定和定位显示, CBMIDA 减少肺和血液中部镍, 增加了骨中的镍、24 h 尿、粪的镍排出与对照组无 明显差异。 结论: CBMIDA 有效地解除镍毒性, 提高动物存活率,降低镍在肺部的滞留.