©2003, Acta Pharmacologica Sinica Chinese Pharmacological Society Shanghai Institute of Materia Medica Chinese Academy of Sciences http://www.ChinaPhar.com

Relationship between cardiotonic effects and inhibition on cardiac sarcolemmal Na⁺, K⁺-ATPase of strophanthidin at low concentrations¹

SU Su-Wen, WANG Yong-Li², LI Jun-Xia, MEI He-Shan, YIN Jing-Xiang

Department of Pharmacology, School of Basic Medicine, Hebei Medical University, Shijiazhuang 050017, China

KEY WORDS strophanthidin; myocardial contraction; Na⁺-K⁺-exchanging ATPase; guinea pigs

ABSTRACT

AIM: To evaluate the effects of strophanthidin (Str) on cardiac contractile function and sarcolemmal Na⁺, K⁺-ATPase activities in isolated guinea-pig hearts. METHODS: Isolated guinea-pig hearts were perfused through aorta in a Langendorff mode. Heart rate (HR), left ventricular pressure (LVP), and first derivatives (\pm dp/dt_{max}) of LVP were recorded by eight-channel physiological instrument. Cardiac sarcolemmal Na⁺, K⁺-ATPase activities were determined with colorimetry. **RESULTS:** Str 0.1 nmol/L stimulated the Na⁺, K⁺-ATPase activities (P<0.05), but had no effect on HR, LVP, and \pm dp/dt_{max}. Str 1 nmol/L increased +dp/dt_{max} (P<0.05) and Na⁺, K⁺-ATPase activities (P<0.01). Str 10 and 100 nmol/L significantly increased both LVP (P<0.05) and +dp/dt_{max} (P<0.05 or P<0.01), and had no significant effects on Na⁺, K⁺-ATPase activities. However, Str 1-100 µmol/L at first enhanced the LVP and \pm dp/dt_{max} (P<0.01), then reduced them resulting from irregular contraction, and effects of Str on Na⁺, K⁺-ATPase activities revealed a concentration-dependent inhibition (P<0.01). **CONCLUSION:** The positive inotropic effects and irregular contraction produced by Str at higher concentrations result from the inhibition of Na⁺, K⁺-ATPase activities, and the positive inotropic effects of Str at lower concentrations are not related to the inhibition of the Na⁺, K⁺-ATPase activities.

INTRODUCTION

Cardiac glycosides have been thought to produce positive inotropic actions in failure heart via the inhibition of Na⁺, K⁺-ATPase activities^[1]. However, the therapeutic concentrations (1-10 nmol/L) of cardiac glycosides in patients with the heart failure are much lower than those shown to inhibit the Na⁺, K⁺-ATPase *in vitro*^[2,3]. Hougen *et al*^[4] and Gao *et al*^[5] have also demonstrated that cardiac glycosides 1 to 10 nmol/L stimulated Na⁺, K⁺-ATPase activities. Thus, the inhibition theory of Na⁺, K⁺-ATPase seems to be inadequate in explaining the inotropic action of glycosides at therapeutic levels. The reports about the effects of glycosides at low concentrations on cardiac performance were contradictory, which showed a positive inotropic effect in some cases^[6], a negative inotropic, or no effect in other experiments^[7,8]. So the relationship between the inotropic actions and effects on Na⁺, K⁺-ATPase activities of Str needs further to be confirmed. To assess the relationship, we measured the positive inotropic actions and the changes

¹ Project supported by Natural Science Foundation of Hebei Province, No 301360.

² Correspondence to Prof WANG Yong-Li.

Phn/Fax 86-311-605-7291.E-mail wangyl52@heinfo.netReceived 2002-08-20Accepted 2003-03-19

of Na⁺, K⁺-ATPase activities produced by strophanthidin (Str, a kind of cardiac glycosides) at the concentration range from 0.1 nmol/L-100 μ mol/L in Langendorff guinea-pig hearts.

MATERIALS AND METHODS

Drugs and chemicals Str (Sigma) was dissolved in dimethyl sulfoxide (Me₂SO) as stock solution. LiBr was from Beijing Chemical Co. Na⁺, K⁺-ATPase kit was purchased from Nanjing Jiancheng Study Co. All reagents were of AR grade.

Preparation of the isolated hearts Guinea pigs of either sex, weighing 250±20 g, were provided by Experimental Animal Center of Hebei Medical University (Grade II, Certificate No 04064). The hearts were rapidly excised, mounted through aorta, and perfused on a modified Langendorff apparatus at a constant perfusion pressure (10 kPa). A polyethylene cannula containing saline was introduced into the left ventricle cavity and was connected with a pressure transducer, by which heart rate (HR), left ventricular pressure (LVP), and its first derivatives $(\pm dp/dt_{max})$ were recorded on a polygraph system (RM 6000, Nihon kohden). The hearts were perfused with K-H buffer solution (37 °C, pH 7.4, saturated with 95 % O_2 and 5 % CO_2). The K-H buffer solution contained the following (in mmol/L): NaCl 118, NaHCO₃ 25.0, KCl 4.7, KH₂PO₄ 1.2, MgSO₄ 1.2, CaCl₂ 2.5, and glucose 11.0.

All the hearts were first perfused with K-H solution for 20-30 min for stabilization in a Langendorff apparatus and then were randomly divided into 8 groups (6-8 hearts each group). The hearts were perfused with K-H buffer solution in control group, and in seven Str groups with K-H solution containing different concentrations of Str (0.1, 1, 10, 100 nmol/L or 1, 10, 100 μ mol/L). HR, LVP and $\pm dp/dt_{max}$ were continuously recorded for 20 min.

Preparation of cardiac sarcolemma Guinea-pig hearts were taken out and placed in ice-cold 10 mmol/L Tris-HCl buffer solution (pH 7.4) containing 1 mmol/L ethylene-diaminetetracetate. The left ventricles were minced with scissors, and homogenized in a homogenizer. The resultant suspension was filtered through 4 layers of gauze and centrifuged for 10 min at $1000 \times g$. After the supernatant was discarded, the pellet was resuspended in 10 mmol/L Tris-HCl buffer solution and subjected to a sequence of Tris-HCl washing buffers and a further KCl extraction step^[9,10]. A part of the final pellets were used to observe the purity of sarcolemma with electron micrograph, the others were resuspended in 1 mmol/L Tris-HCl (pH 7.0) to determine the activities of Na⁺, K⁺-ATPase. All procedures were performed at 0-4 °C. The protein content was measured by Coomassie brilliant blue method.

Observation of sarcolemmal purity A part of the pellets were fixed overnight in 4 % glutaraldehydephosphate buffer (pH 7.4), diced into pieces of about 0.5 mm×0.5 mm×0.5 mm, post fixed for 1 h in 1 % osmium tetraoxide. Then the pellet was dehydrated in a graded acetone series (50 %-100 %) and embedded in 812-epoxy resin. Thin sections were made with LKB-V# ultramicrotome (Sweden) and double-stained with uranium acetate and lead citrate. The sections were examined and photographed using H-7500 electron microscope (Nihon kohden).

Determination of the cardiac sarcolemmal Na⁺, K⁺-ATPase activities The cardiac sarcolemmal Na⁺, K⁺-ATPase activities were determined by colorimetry. The suspension of cardiac sarcolemma (50-70 µg proteins per mL) was divided into 8 groups (8 samples each group) and incubated with saline or different concentrations (0.1, 1, 10, 100 nmol/L and 1, 10, 100 µmol/L) of Str at 37 °C for 10 min according to the method provided by manufacturer. Enzyme active unit was expressed as mmol inorganic phosphate per gram sarcolemmal protein per hour (mmol·h⁻¹·g⁻¹ protein).

Statistics Data were expressed as mean±SD. Statistical analysis was performed using paired *t*-test.

RESULTS

Effects of Str on cardiac contractility In the control group, perfused guinea-pig hearts were continuously observed for 50 min and showed no changes in HR, LVP, and $\pm dp/dt_{max}$. Str 0.1 nmol/L did not affect the cardiac contractile function; Str 1 nmol/L increased $+dp/dt_{max}$ (P<0.05), but had no effects on other parameters; Str 10 and 100 nmol/L increased both LVP and $+dp/dt_{max}$ (P<0.05 or P<0.01). The peak time of LVP and $+dp/dt_{max}$ was about 10-15 min, and after that the plateau of increase was sustained for whole observation. There was no obviously change in HR and $-dp/dt_{max}$; Str 1, 10, and 100 µmol/L caused the increase (P < 0.01) and subsequent decrease in LVP and $\pm dp/dt_{max}$ resulting from irregular contraction (the incident rate of irregular contraction were 57 %, 75 %, and 100 %, respectively). A peak effect was reached in 10 min. A concentration-dependent decrease in HR was observed (P<0.05 or P<0.01) (Fig 1).



Effects of Str on cardiac sarcolemmal Na⁺,K⁺-ATPase activities The electron microscopic examination showed that the membrane fraction was consisted of membranous sacs of varying shapes and sizes. No intact mitochondria, nuclei or myofibrils were found (Fig 2). Str 0.1 and 1 nmol/L increased (P<0.05) and 1-100 µmol/L inhibited the Na⁺, K⁺-ATPase activities (P<0.01). But Str 10 and 100 nmol/L did not affect

 \cdot 1106 \cdot



Fig 2. Electron micrograph of the guinea-pig heart sarcolemmal pellet (×10 000).

Na⁺, K⁺-ATPase activities (Tab 1).

Tab 1. Effects of Str on cardiac sarcolemmal Na⁺, K⁺-ATPase activity in guinea-pig hearts. n=8. Mean±SD. ^bP<0.05, ^cP<0.01 vs control.

Doses	Na ⁺ ,K ⁺ -ATPase activity /mmol·h ⁻¹ ·g ⁻¹ protein	Change rate/%
0	23.2±2.8	0
0.1 nmol/L	25.9±1.6 ^b	13
1 nmol/L	27.4±2.0°	20
10 nmol/L	22.9±2.4	2
100 nmol/L	22.6±2.5	-6
1 mmol/L	19.1±2.7°	-18
10 mmol/L	15.5±1.6°	-25
100 mmol/L	6.3±1.4°	-72

The relationship between the cardiac function parameters and the inhibition of Na⁺, K⁺-ATPase The correlation analysis between the maximum of cardiac contractile parameters or incident rate of irregular contraction and the percent inhibition of Na⁺, K⁺-ATPase of Str at higher concentrations (1-100 μ mol/L) were summarized in Tab 2. The results showed that there were a close correlation between LVP, +dp/dt_{max} or -dp/dt_{max}, percentage of irregular contraction and the percent inhibition of Na⁺, K⁺-ATPase activities. Tab 2. Correlation analysis between the cardiac function parameters and the inhibition of Na⁺, K⁺-ATPase.

	Slope(b) Intercept(a) r			Р
LVP (Y) vs I-Na (X)	0.19	7.64	0.92	< 0.01
$+dp/dt_{max}(Y) vs$ I-Na (X)	0.9	73.6	0.97	< 0.01
$-dp/dt_{max}(Y)$ vs I-Nz (X)	1.0	68.0	0.86	< 0.01
Percentage of irregular con	traction			
(Y) vs I-NA (X)	0.7	50.5	0.95	< 0.01

Y=a+b*X*, *r*: correlation coefficent.

DISCUSSION

Our results showed that Str 1-100 nmol/L increased the cardiac contractile function. Str 0.1, 1 nmol/L stimulated and 10,100 nmol/L did not affect the Na⁺, K⁺-ATPase activities. These results suggest that the positive inotropic actions of Str at lower concentrations are not related to the changes of Na⁺, K⁺-ATPase activities, either inhibition or stimulation, for Str 0.1 nmol/L increased the Na⁺, K⁺-ATPase activities without any effects on the cardiac contractile function. There was a close correlation between the increase of cardiac contractile function and the inhibition of Na⁺, K⁺-ATPase at higher concentrations of Str (1-100 µmol/L), suggesting that the inotropic actions result from inhibition of Na⁺, K⁺-ATPase activities. So there are at least two coexisted inotropic mechanisms of cardiac glycosides: one related to Na⁺, K⁺-ATPase inhibition, and the other, occurring at lower concentrations, involved a mechanism other than Na⁺,K⁺-ATPase inhibition. Our results is agreement with other researcher's work^[6,11,12]. The present experiment further confirmed the inotropic effects of Str at lower concentrations are not the result of Na⁺, K⁺-ATPase inhibition.

Since intracellular Ca^{2+} concentration ($[Ca^{2+}]_i$) plays a crucial role in the excitation-contraction coupling, and Na⁺, K⁺-ATPase is the receptor of cardiac glycosides, it is very interesting to know how cardiac glycosides at lower concentrations induced the $[Ca^{2+}]_i$ elevation. In general, inhibition of the Na⁺, K⁺-ATPase activities caused the increase of intracellular Na⁺ concentration ($[Na^+]_i$) and subsequent increase of $[Ca^{2+}]_i$ through Na⁺-Ca²⁺ exchange. Obviously, the positive inotropic effects of Str at lower concentrations is not related to increased $[Na^+]_i$ secondary to the inhibition of Na⁺, K⁺-ATPase. In fact, cardiac glycosides at lower concentrations did increase $[Ca^{2+}]_i$ without any change in $[Na^+]_i^{[13-15]}$. What is the mechanism of positive inotropic effects of Str at lower concentrations? Santana et al^[16] reported that nanomolar concentrations of cardiac glycosides activated the slip-mode conductance of the TTX-sensitive Na⁺-channels by which Ca²⁺ enters the cell, leading to an increase in $[Ca^{2+}]_i$. Sagawa *et al*^[17] also reported cardiac glycosides at lower concentrations activated the cardiac ryanodine receptors and resulted in Ca²⁺ release from sarcoplasmic reticulum. Mounting evidence suggests that there exists an endogenous ouabain-like factor which has hormone-like properties, and can relay message through various signal transducion pathways^[18-20]. Recently, Tian et al^[21] demonstrated that ouabain's effect on $[Ca^{2+}]_i$ in rat cardiac myocytes is throuth the signal transducing function of Na⁺, K⁺-ATPase activities. Yin et al^[22] proved that low concentration of dihydroouabain increased $[Ca^{2+}]_i$ by Ca^{2+} influx via L-type Ca^{2+} channels, TTXsensitive Na⁺ channels or/and by directly triggering intracellular calcium release. Thus, the effects of glycosides might be involved in more complex mechanisms. More studies are necessary to further understand the molecular basis of the action.

In conclusion, Our results suggest that Str at lower concentrations still has the positive inotropic effects, which involve a mechanism other than Na⁺, K⁺-ATPase inhibition.

REFERENCES

- Langer GA. The "sodium pump lag" revisited. J Mol Cell Cardiol 1983; 15: 647-51.
- 2 Noble D. Mechanism of action of therapeutic levels of cardiac glycosides. Cardiovas Res 1980; 14: 495-514.
- 3 Adams KF Jr, Gheorghiade M, Uretsky BF, Patterson JH, Schwartz TA, Young JB. Clinical benefits of low serum digoxin concentrations in heart failure. J Am Coll Cardiol 2002; 39: 946-53.
- 4 Hougen TJ, Spicer N, Smith TW. Stimulation of monovalent cation active transport by low concentrations of cardiac glycosides. J Clin Invest 1981; 68: 1207-14.
- 5 Gao J, Wymore RS, Wang Y, Gaudette GR, Krukenkamp IB, Cohen IS, *et al.* Isoform-specific stimulation of cardiac Na/K pumps by nanomolar concentrations of glycosides. J Gen Physiol 2002; 119: 297-312.
- 6 Godfraind T, Ghysel-Burton J. Independence of the positive inotropic effect of ouabain from the inhibition of the

heart Na⁺-K⁺ pump. Proc Natl Acad Sci USA 1980; 77: 3067-9.

- 7 Poole-Wilson PA, Galindez E, Fry CH. Effect of ouabain in therapeutic concentrations on K⁺ exchange and contractions of human and rabbit myocardium. Clin Sci 1979; 57: 415-20.
- 8 Hart G, Noble D, Shimoni Y. The effects of low concentrations of cardiotonic steroids on membrane currents and tension in sheep Purkinje fibres. J Physiol 1983; 334: 103-31.
- 9 Halla NS, Anand-Srivastava MB, Tuana BS, Khandelwal RL. Solubilization of a calcium dependent adenosine triphosphatase from rat heart sarcolemma. J Mol Cell Cardiol 1981; 13: 413-23.
- 10 Alto LE, Elimban V, Lukas A, Dhalla NS. Modification of heart sarcolemmal Na⁺, K⁺-ATPase activity during development of the calcium paradox. Mol Cell Biochem 2000; 207: 87-94.
- 11 Hougen TJ, Smith TW. Inhibition of myocardial monovalent cation active transport by subtoxic doses of ouabain in the dog. Circ Res 1978; 42: 856-63.
- 12 Roberto BMD, Mario VMD. The inotropic effects of strophanthidin in purkinje fibers and the sodium pump. Circulation 1984; 69: 618-31.
- 13 Arnon A, Hamlyn JM, Blaustein MP. Ouabain augments Ca²⁺ transients in arterial smooth muscle without raising cytosolic Na⁺. Am J Physiol Heart Circ Physiol 2000; 279: H679-91.
- 14 Jewell EA, Shamraj OL, Lingrel JB. Isoforms of the alpha subunit of Na⁺, K⁺-ATPase and their significance. Acta Physiol Scand 1992; 607: 161-9.
- 15 O'Brien WJ, Lingrel JB, Wallick ET. Ouabain binding kinetics of the rat alpha two and alpha three isoforms of the sodium-potassium adenosine triphosphate. Arch Biochem Biophys 1994; 310: 32-9.
- 16 Santana LF, Gomez AM, Lederer. Ca²⁺ flux through promiscuous cardiac Na⁺ channels: slip-mode conductance. Science 1998; 273: 1027-32.
- 17 Sagawa T, Sagawa K, Kelly JE, Tsushima RG, Wasserstrom JA. Activation of cardiac ryanodine receptors by cardiac glycosides. Am J Physiol Heart Circ Physiol 2002; 282: H1118-26.
- 18 Doris PA. Regulation of Na, K-ATPase by endogenous ouabain-like materials. Exp Biol Med 1994; 205: 202-12.
- Ferrandi M, Manunta P. Ouabain-like factor: is this the natriuretic hormone? Curr Opin Nephrol Hypertens 2000; 9: 165-71.
- 20 Ke YS. Endoxin: a major factor regulating cardiovascular system. Acta Pharmacol Sin 2001; 22: 201-9.
- 21 Tian J, Gong X, Xie Z. Signal transducing function of Na⁺/K⁺ ATPase is essential for ouabain's effect on [Ca²⁺]_i in rat cardiac myocytes. Am J Physiol Heart Circ Physiol 2001; 281: H1899-907.
- 22 Yin JX, Wang YL, Li Q, Shang ZL, SU SW. Effects of low concentration of dihydroouabain on intracellular calcium in guinea pig ventricular myocytes. Acta Physiol Sin 2002; 54: 359-62.