

©2003, Acta Pharmacologica Sinica  
Chinese Pharmacological Society  
Shanghai Institute of Materia Medica  
Chinese Academy of Sciences  
<http://www.ChinaPhar.com>

# Pharmacokinetics and tissue distribution of iv injection of polyphase liposome-encapsulated cisplatin (KM-1) in rats<sup>1</sup>

WANG Shan, MI Jie-Bo, LI Yuan-Zong, CHANG Wen-Bao<sup>2</sup>, CI Yun-Xiang  
*College of Chemistry and Molecular Engineering, Peking University, Beijing 100871, China*

ZHAO Min-Zheng, ZHAO Yun-Kun, ZHU Li-Ya, XU Guang  
*Research Institute of Precious Metals, Kunming 650221, China*

**KEY WORDS** pharmacokinetics; liposomes; cisplatin; induced coupled plasma atomic emission spectrometry

## ABSTRACT

**AIM:** The pharmacokinetics and biodistribution of cisplatin encapsulated in polyphase liposome (KM-1) were compared with those of free drug in rats. **METHODS:** The platinum levels in serum and normal organs, after a single dose of iv injection of free or encapsulated cisplatin to rats, were determined by induced coupled plasma atomic emission spectrometry. **RESULTS:** Serum platinum concentration-time curve after a single iv dose of KM-1 4.5 mg/kg in rats was fitted with an open three-compartment model. The pharmacokinetic parameters were as follows:  $V_c=0.10$  L/kg,  $T_{1/2\alpha}=0.3$  h,  $T_{1/2\alpha'}=3.5$  h,  $T_{1/2\beta}=2.7$  h,  $AUC=265$  mg·h·L<sup>-1</sup>, and  $CL(s)=0.02$  g·L<sup>-1</sup>·h<sup>-1</sup>. KM-1 was cleared from the circulation much more slowly than free cisplatin. Liver and spleen had the highest concentration of platinum after KM-1 treatment. **CONCLUSION:** KM-1 remained in the bloodstream longer than its free drug, and was taken mainly by the reticuloendothelial system.

## INTRODUCTION

Cisplatin (*cis*-dichlorodiamine platinum II) is an anti-neoplastic platinum agent commonly used alone or in combination with other agents in a variety of chemotherapeutic regimens<sup>[1-3]</sup>. However, expansion of the clinical utility of cisplatin has been limited by its toxicity, as well as the emergence of intrinsic and acquired resistance in many common tumor types. These make the improvements of the drug necessary<sup>[4-6]</sup>.

Liposomal encapsulation reduces the toxicity as-

sociated with many antitumor agents by altering their pharmacokinetics and distribution<sup>[7-11]</sup>. But the development of liposome containing cisplatin has been hampered by poor water solubility and low lipophilicity of free drug, which results in unstable liposome formulations that have a low drug encapsulation rate and tend to leak their content rapidly during storage. With the help of water-solubility enhancement reagents, we have developed a new formulation of cisplatin, polyphase liposome KM-1, which has a high total drug encapsulation rate (85 %) and is stable at 4 °C for at least 30 months. Preliminary studies have shown that, compared with cisplatin, KM-1 showed a lower cisplatin-associated toxicity and was superior in the treatment of mouse tumor in liver. In this report, the pharmacokinetics and biodistribution of KM-1 and free cisplatin in rats at a dose of 4.5 mg/kg were comparatively studied.

<sup>1</sup> Projects supported by Yunnan Science and Technology Committee.

<sup>2</sup> Correspondence to Prof CHANG Wen-Bao.  
Phn 86-10-6275-4905. Fax 86-10-6275-1708.  
E-mail wangshan420@163.net

Received 2002-05-16

Accepted 2002-12-16

**MATERIALS AND METHODS**

**Animals** Wistar rats (Grade II,  $n=110$ ,  $170 \text{ g} \pm 20 \text{ g}$ , Male, Certificate No SCXK11-00-0006) were supplied by Breeding Center of the Institute of Experimental Animals, Chinese Academy of Medical Sciences.

**Chemicals and drugs** Cisplatin was purchased from Qilu Pharmaceutical Manufactory. Platinum standard solution ( $1.000 \text{ g/L}$ ) was purchased from National Test Center of Steel Material, Ministry of Metallurgical Industry, China. All other chemicals were of AR grade.

**Instrument** Leeman abs, INC Profile induced coupled plasma atomic emission spectrometer was used.

**Medication and sampling** On the day of dosing, rats received a  $4.5 \text{ mg/kg}$  dose of either KM-1 or cisplatin by tail vein injection. Doses were based on individual animal body weight. Animals were sacrificed by decapitation (five per time-point). Blood was collected immediately and centrifuged at  $1500 \times g$  for 10 min to isolate serum. The liver, kidneys, spleen, lung, heart, and testicles of rats sacrificed at 2, 8, 16, 24, and 48 h were removed, blotted dry, and weighed prior to freezing. Serum and tissues were stored frozen ( $-20^\circ \text{C}$ ) for later analysis.

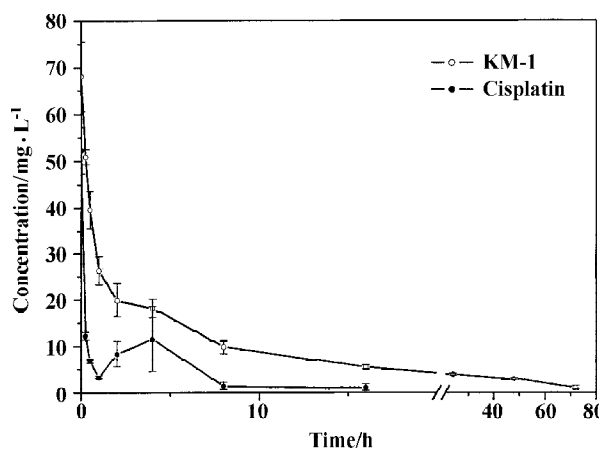
**Sample preparations** Serum  $1.0 \text{ mL}$  was diluted with  $0.5 \text{ mL}$  deionized water and analyzed directly. Tissue samples were digested in concentrated acid prior to analysis. After overnight incubation in nitric acid, the tissue slurry was gently heated to achieve partial digestion, nitric acid and perchloric acid mixture in the ratio of 1:1 was added followed by heating to complete the digestion. Evaporated to near dryness, the residue was resolved in a 3 % nitric acid solution to a final volume of  $2.5 \text{ mL}$ .

**Platinum determination** The total platinum levels in all samples were determined using a sensitive and specific ICP-AES based method. The assay does not differentiate liposomal and nonliposomal platinum or free and protein-bound platinum. The sensitivity of the assay is  $12.8 \text{ mg/L}$ . The calibration curve for Pt was linear over the range of  $0\text{-}100 \text{ mg/L}$ . Linear equation:  $C=0.2268I-123.73$  ( $r=0.998$ ). The within and between day variabilities were within 5 % and 10 %, respectively. The recoveries of Pt from tissues were above 70 %. All concentrations are expressed as  $\text{mg cisplatin equivalent/L}$  or  $\mu\text{g cisplatin equivalent/g}$ .

**Data analysis** The platinum concentration-time curves were analyzed with 3P87 program to determine the pharmacokinetic parameters.

**RESULTS**

**Pharmacokinetics** KM-1 was cleared from the circulation much more slowly than cisplatin (Fig 1). The serum platinum concentration with cisplatin treatment was  $12 \text{ mg/L}$  at 15 min, while that with KM-1 treatment was  $51 \text{ mg/L}$  at the same time point, showing a 4-fold increase of platinum in serum. At 8 h, the serum platinum concentration with KM-1 treatment was still  $10 \text{ mg/L}$ , while only  $1 \text{ mg/L}$  of platinum was left in that with cisplatin treatment. The serum platinum concentration-time data in KM-1-treated group were well characterized by an open three-compartment model. Some of the parameters were listed in Tab 1. Due to cisplatin fast elimination from the circulation in free cisplatin-treated group, serum platinum concentration dropped quickly with time. There were not enough concentration-time data available in free cisplatin group for the calculation of pharmacokinetics parameters in this study.



**Fig 1.** Serum concentrations of cisplatin following KM-1 or cisplatin administration ( $4.5 \text{ mg/kg}$  of body weight each).  $n=5$ . Mean $\pm$ SD.

**Tab 1.** Pharmacokinetic parameters of KM-1 following a single iv dose of  $4.5 \text{ mg/kg}$ .

Parameter	Value
$V_d/\text{L} \cdot \text{kg}^{-1}$	0.10
$T_{1/2\pi}/\text{h}$	0.3
$T_{1/2\alpha}/\text{h}$	3.5
$T_{1/2\beta}/\text{h}$	2.7
$\text{AUC}/\text{mg} \cdot \text{h} \cdot \text{L}^{-1}$	265
$\text{CL}(\text{s})/\text{g} \cdot \text{L}^{-1} \cdot \text{h}^{-1}$	0.02

**Tissue distribution** After a single iv dose of KM-1, both the liver and spleen showed the highest platinum concentration at 24 h (Fig 2), 4-, and 3-fold higher, respectively, than those after free cisplatin treatment ( $P<0.01$ ). In the kidney, the primary target organ of cisplatin toxicity, both KM-1 and cisplatin treatment had similar platinum levels ( $P>0.05$ ). Concentration of KM-1 in kidney reached its highest level at 16 h and then began to decline with time, while in cisplatin-treated group, the platinum concentration in kidney fluctuated with time during the whole observation period. The platinum levels in heart, lung, and testicle were slightly higher following treatment with KM-1 than with cisplatin.

## DISCUSSION

It has been demonstrated that, due to the altering

of pharmacokinetics and disposition of the drugs, liposomal encapsulation of many antitumor agents attenuates their toxicity or changes their target organs of chemotherapy. And this is also the case in cisplatin. Gondal *et al* has reported a formulation of cisplatin encapsulated in multilamellar liposomes<sup>[11]</sup>. They found that the liposome had a lower platinum level in the kidney and maintained reduced nephrotoxicity compared with free cisplatin. Newman *et al* had cisplatin encapsulated in long-circulating, pegylated liposomes (SPI-077). They observed an increasing tumor Pt disposition and improved antitumor effect in murine colon and lung cancer models<sup>[10]</sup>. Our present study showed that the platinum levels in the spleen and liver following KM-1 treatment were several times higher than those achieved with cisplatin, which was caused by the greater uptake of liposomal formulation by the reticuloendothelial system. This offers a greater potential for the treat-

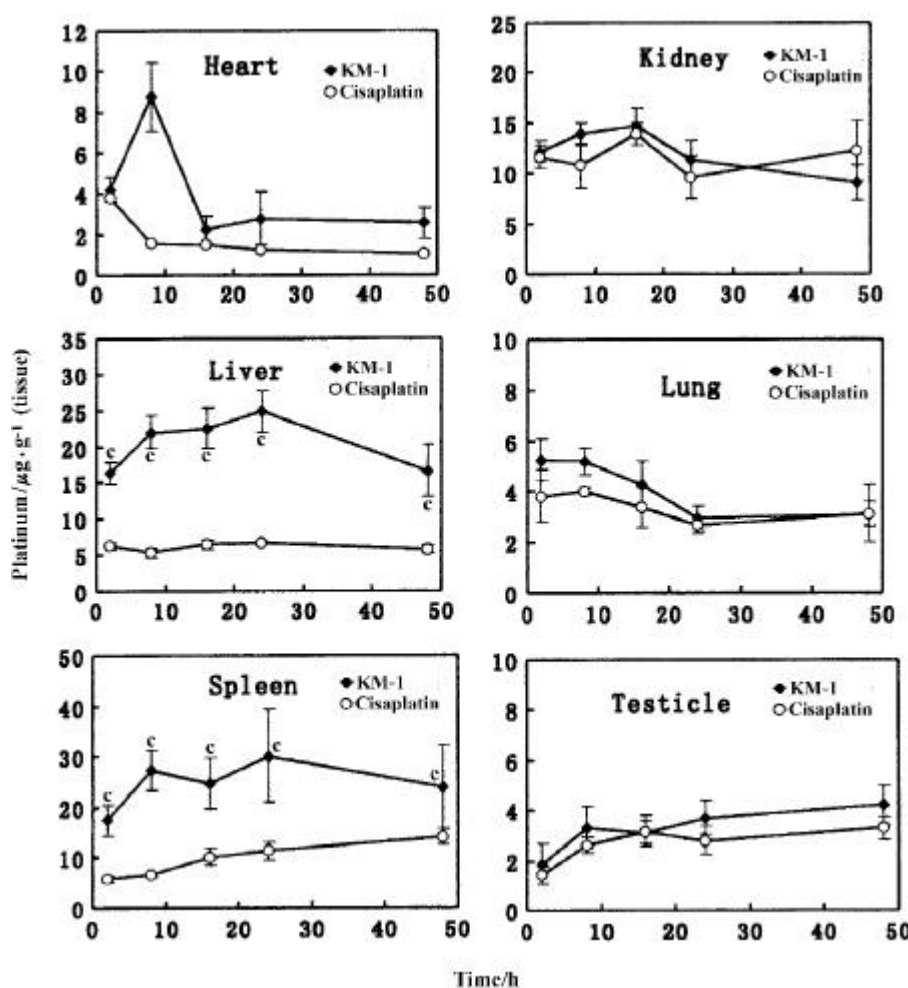


Fig 2. Concentration-time profiles of platinum in tissues following a single iv dose of KM-1 or cisplatin 4.5mg/kg.  $n=5$ . Mean $\pm$ SD. \* $P<0.01$  vs cisplatin group.

ment of tumors in these organs. And it agrees with our preliminary pharmacodynamic study with KM-1. The study showed that KM-1 had the equivalent antitumor efficacy with cisplatin in liver tumor bearing mice, but was superior to cisplatin in that it could alleviate the damage of liver caused by the tumor, while cisplatin would aggravate the damage.

We did not discern lower platinum concentrations in the kidney with KM-1 than with free cisplatin. To outward seeming, this is not consistent with our preliminary study showing that nephrotoxicity in KM-1-treated mice was less than that in cisplatin-treated ones. The explanation may lie in the different reactive mechanism between free drug with tissues and liposomal-entrapped drug with tissues. Since most of the drug is retained in liposome, the total amount of platinum measured in KM-1-treated group is not equivalent to that of platinum that can react with biological substances and has a pharmacological or toxicological efficacy. Cisplatin encapsulated in liposome was released gradually with time and tissues were continuously exposed to a low-dose drug environment. This could avoid a high peak free cisplatin level that would produce a severe toxicological effect. Similar situation occurred in liver with KM-1 treatment, which had high platinum levels and showed no hepatotoxicity. The result agrees with the studies of Gondal *et al*<sup>[11]</sup>.

No significant eliminations of platinum in tissues were observed during the first 48 h after dosing. This is due to the characteristics of cisplatin which would bind to protein or other tissue compositions firmly through covalent bonds. Excretion studies had also shown that only 25 % of total KM-1 taken by rats was excreted during (0-48) h period of time.

## REFERENCES

- 1 Rosenberg b, VanCamp L, Trosko JE, Monsour VH. Platinum compounds: a new class of potent antitumor agents. *Nature* 1969; 222: 385-6.
- 2 Einhorn LH, Donohue J. *cis*-Diaminedichloroplatinum, vinblastine, and bleomycin chemotherapy in disseminated testicular cancer. *Ann Intern Med* 1977; 87: 293-8.
- 3 Pearson ADJ, Craft AW, Pinkerton CR, Meller ST, Reid MM. High-dose rapid schedule chemotherapy for disseminated neuroblastoma. *Eur J Cancer* 1992; 28A: 1654-9.
- 4 Raymond BW, Michaele CC. New cisplatin analogues in development. *Drugs* 1993; 46: 360-77.
- 5 Kelland LR, Clarke SJ, Mckeage MJ. Advances in platinum complex cancer chemotherapy. *Platinum Metals Rev* 1992; 36: 178-84.
- 6 Yang YK, Xiong HZ, Pu SP, Liu WP, Chen XZ, He J. Study on the third generation platinum anticancer drugs. *Precious Metals* 1996; 17: 50-7.
- 7 Van Hoesel QGCM, Steenberg PA, Crommelin DJ, Vandijk A, Vanoort W, Klein S, *et al*. Reduced cardiotoxicity and nephrotoxicity with preservation of antitumor-activity of doxorubicin entrapped in stable liposomes in the LOU/M Wsl rats. *Cancer Res* 1984; 44: 3698-705.
- 8 Herman EH, Rahman A, Ferrans VJ, Vick JA, Schein PS. Prevention of chronic doxorubicin cardiotoxicity in beagles by liposomal encapsulation. *Cancer Res* 1983; 43: 5427-32.
- 9 Gabizon A, Catane R, Uziely B. Prolonged circulation time and enhanced accumulation in malignant exudates of doxorubicin encapsulated in polyethylene-glycol coated liposomes. *Cancer Res* 1994; 54: 987-92.
- 10 Newman MS, Colbern GT, Working PK, Engbers C, Amantea MA. Comparative pharmacokinetics, tissue distribution, and therapeutic effectiveness of cisplatin encapsulated in long-circulating, pegylated liposomes (SPI-077) in tumor-bearing mice. *Cancer Chemother Pharmacol* 1999; 43: 1-7.
- 11 Gondal JA, Preuss HG, Swartz R, Rahman A. Comparative pharmacological, toxicological and antitumoral evaluation of free and liposome-encapsulated cisplatin in rodents. *Eur J Cancer* 1993; 29A: 1536-42.

©2003, Acta Pharmacologica Sinica  
Chinese Pharmacological Society  
Shanghai Institute of Materia Medica  
Chinese Academy of Sciences  
<http://www.ChinaPhar.com>

# Pharmacokinetics and tissue distribution of iv injection of polyphase liposome-encapsulated cisplatin (KM-1) in rats<sup>1</sup>

WANG Shan, MI Jie-Bo, LI Yuan-Zong, CHANG Wen-Bao<sup>2</sup>, CI Yun-Xiang  
*College of Chemistry and Molecular Engineering, Peking University, Beijing 100871, China*

ZHAO Min-Zheng, ZHAO Yun-Kun, ZHU Li-Ya, XU Guang  
*Research Institute of Precious Metals, Kunming 650221, China*

**KEY WORDS** pharmacokinetics; liposomes; cisplatin; induced coupled plasma atomic emission spectrometry

## ABSTRACT

**AIM:** The pharmacokinetics and biodistribution of cisplatin encapsulated in polyphase liposome (KM-1) were compared with those of free drug in rats. **METHODS:** The platinum levels in serum and normal organs, after a single dose of iv injection of free or encapsulated cisplatin to rats, were determined by induced coupled plasma atomic emission spectrometry. **RESULTS:** Serum platinum concentration-time curve after a single iv dose of KM-1 4.5 mg/kg in rats was fitted with an open three-compartment model. The pharmacokinetic parameters were as follows:  $V_c=0.10$  L/kg,  $T_{1/2\alpha}=0.3$  h,  $T_{1/2\alpha'}=3.5$  h,  $T_{1/2\beta}=2.7$  h,  $AUC=265$  mg·h·L<sup>-1</sup>, and  $CL(s)=0.02$  g·L<sup>-1</sup>·h<sup>-1</sup>. KM-1 was cleared from the circulation much more slowly than free cisplatin. Liver and spleen had the highest concentration of platinum after KM-1 treatment. **CONCLUSION:** KM-1 remained in the bloodstream longer than its free drug, and was taken mainly by the reticuloendothelial system.

## INTRODUCTION

Cisplatin (*cis*-dichlorodiamine platinum II) is an anti-neoplastic platinum agent commonly used alone or in combination with other agents in a variety of chemotherapeutic regimens<sup>[1-3]</sup>. However, expansion of the clinical utility of cisplatin has been limited by its toxicity, as well as the emergence of intrinsic and acquired resistance in many common tumor types. These make the improvements of the drug necessary<sup>[4-6]</sup>.

Liposomal encapsulation reduces the toxicity as-

sociated with many antitumor agents by altering their pharmacokinetics and distribution<sup>[7-11]</sup>. But the development of liposome containing cisplatin has been hampered by poor water solubility and low lipophilicity of free drug, which results in unstable liposome formulations that have a low drug encapsulation rate and tend to leak their content rapidly during storage. With the help of water-solubility enhancement reagents, we have developed a new formulation of cisplatin, polyphase liposome KM-1, which has a high total drug encapsulation rate (85 %) and is stable at 4 °C for at least 30 months. Preliminary studies have shown that, compared with cisplatin, KM-1 showed a lower cisplatin-associated toxicity and was superior in the treatment of mouse tumor in liver. In this report, the pharmacokinetics and biodistribution of KM-1 and free cisplatin in rats at a dose of 4.5 mg/kg were comparatively studied.

<sup>1</sup> Projects supported by Yunnan Science and Technology Committee.

<sup>2</sup> Correspondence to Prof CHANG Wen-Bao.  
Phn 86-10-6275-4905. Fax 86-10-6275-1708.  
E-mail wangshan420@163.net

Received 2002-05-16

Accepted 2002-12-16

**MATERIALS AND METHODS**

**Animals** Wistar rats (Grade II,  $n=110$ ,  $170 \text{ g} \pm 20 \text{ g}$ , Male, Certificate No SCXK11-00-0006) were supplied by Breeding Center of the Institute of Experimental Animals, Chinese Academy of Medical Sciences.

**Chemicals and drugs** Cisplatin was purchased from Qilu Pharmaceutical Manufactory. Platinum standard solution ( $1.000 \text{ g/L}$ ) was purchased from National Test Center of Steel Material, Ministry of Metallurgical Industry, China. All other chemicals were of AR grade.

**Instrument** Leeman abs, INC Profile induced coupled plasma atomic emission spectrometer was used.

**Medication and sampling** On the day of dosing, rats received a  $4.5 \text{ mg/kg}$  dose of either KM-1 or cisplatin by tail vein injection. Doses were based on individual animal body weight. Animals were sacrificed by decapitation (five per time-point). Blood was collected immediately and centrifuged at  $1500 \times g$  for 10 min to isolate serum. The liver, kidneys, spleen, lung, heart, and testicles of rats sacrificed at 2, 8, 16, 24, and 48 h were removed, blotted dry, and weighed prior to freezing. Serum and tissues were stored frozen ( $-20^\circ \text{C}$ ) for later analysis.

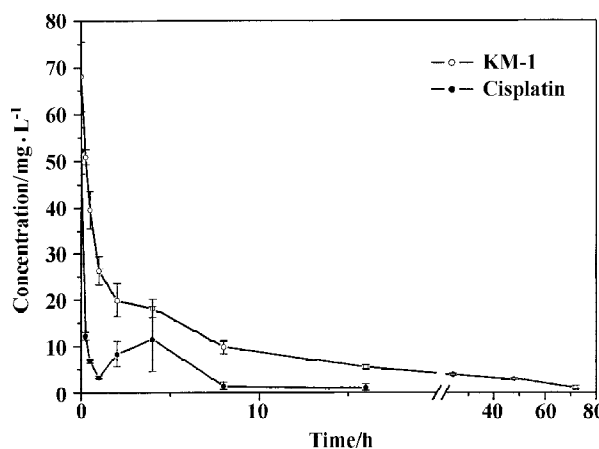
**Sample preparations** Serum  $1.0 \text{ mL}$  was diluted with  $0.5 \text{ mL}$  deionized water and analyzed directly. Tissue samples were digested in concentrated acid prior to analysis. After overnight incubation in nitric acid, the tissue slurry was gently heated to achieve partial digestion, nitric acid and perchloric acid mixture in the ratio of 1:1 was added followed by heating to complete the digestion. Evaporated to near dryness, the residue was resolved in a 3 % nitric acid solution to a final volume of  $2.5 \text{ mL}$ .

**Platinum determination** The total platinum levels in all samples were determined using a sensitive and specific ICP-AES based method. The assay does not differentiate liposomal and nonliposomal platinum or free and protein-bound platinum. The sensitivity of the assay is  $12.8 \text{ mg/L}$ . The calibration curve for Pt was linear over the range of  $0\text{-}100 \text{ mg/L}$ . Linear equation:  $C=0.2268I-123.73$  ( $r=0.998$ ). The within and between day variabilities were within 5 % and 10 %, respectively. The recoveries of Pt from tissues were above 70 %. All concentrations are expressed as  $\text{mg cisplatin equivalent/L}$  or  $\mu\text{g cisplatin equivalent/g}$ .

**Data analysis** The platinum concentration-time curves were analyzed with 3P87 program to determine the pharmacokinetic parameters.

**RESULTS**

**Pharmacokinetics** KM-1 was cleared from the circulation much more slowly than cisplatin (Fig 1). The serum platinum concentration with cisplatin treatment was  $12 \text{ mg/L}$  at 15 min, while that with KM-1 treatment was  $51 \text{ mg/L}$  at the same time point, showing a 4-fold increase of platinum in serum. At 8 h, the serum platinum concentration with KM-1 treatment was still  $10 \text{ mg/L}$ , while only  $1 \text{ mg/L}$  of platinum was left in that with cisplatin treatment. The serum platinum concentration-time data in KM-1-treated group were well characterized by an open three-compartment model. Some of the parameters were listed in Tab 1. Due to cisplatin fast elimination from the circulation in free cisplatin-treated group, serum platinum concentration dropped quickly with time. There were not enough concentration-time data available in free cisplatin group for the calculation of pharmacokinetics parameters in this study.



**Fig 1.** Serum concentrations of cisplatin following KM-1 or cisplatin administration ( $4.5 \text{ mg/kg}$  of body weight each).  $n=5$ . Mean $\pm$ SD.

**Tab 1.** Pharmacokinetic parameters of KM-1 following a single iv dose of  $4.5 \text{ mg/kg}$ .

Parameter	Value
$V_d/\text{L} \cdot \text{kg}^{-1}$	0.10
$T_{1/2\pi}/\text{h}$	0.3
$T_{1/2\alpha}/\text{h}$	3.5
$T_{1/2\beta}/\text{h}$	2.7
$\text{AUC}/\text{mg} \cdot \text{h} \cdot \text{L}^{-1}$	265
$\text{CL}(\text{s})/\text{g} \cdot \text{L}^{-1} \cdot \text{h}^{-1}$	0.02

**Tissue distribution** After a single iv dose of KM-1, both the liver and spleen showed the highest platinum concentration at 24 h (Fig 2), 4-, and 3-fold higher, respectively, than those after free cisplatin treatment ( $P<0.01$ ). In the kidney, the primary target organ of cisplatin toxicity, both KM-1 and cisplatin treatment had similar platinum levels ( $P>0.05$ ). Concentration of KM-1 in kidney reached its highest level at 16 h and then began to decline with time, while in cisplatin-treated group, the platinum concentration in kidney fluctuated with time during the whole observation period. The platinum levels in heart, lung, and testicle were slightly higher following treatment with KM-1 than with cisplatin.

## DISCUSSION

It has been demonstrated that, due to the altering

of pharmacokinetics and disposition of the drugs, liposomal encapsulation of many antitumor agents attenuates their toxicity or changes their target organs of chemotherapy. And this is also the case in cisplatin. Gondal *et al* has reported a formulation of cisplatin encapsulated in multilamellar liposomes<sup>[11]</sup>. They found that the liposome had a lower platinum level in the kidney and maintained reduced nephrotoxicity compared with free cisplatin. Newman *et al* had cisplatin encapsulated in long-circulating, pegylated liposomes (SPI-077). They observed an increasing tumor Pt disposition and improved antitumor effect in murine colon and lung cancer models<sup>[10]</sup>. Our present study showed that the platinum levels in the spleen and liver following KM-1 treatment were several times higher than those achieved with cisplatin, which was caused by the greater uptake of liposomal formulation by the reticuloendothelial system. This offers a greater potential for the treat-

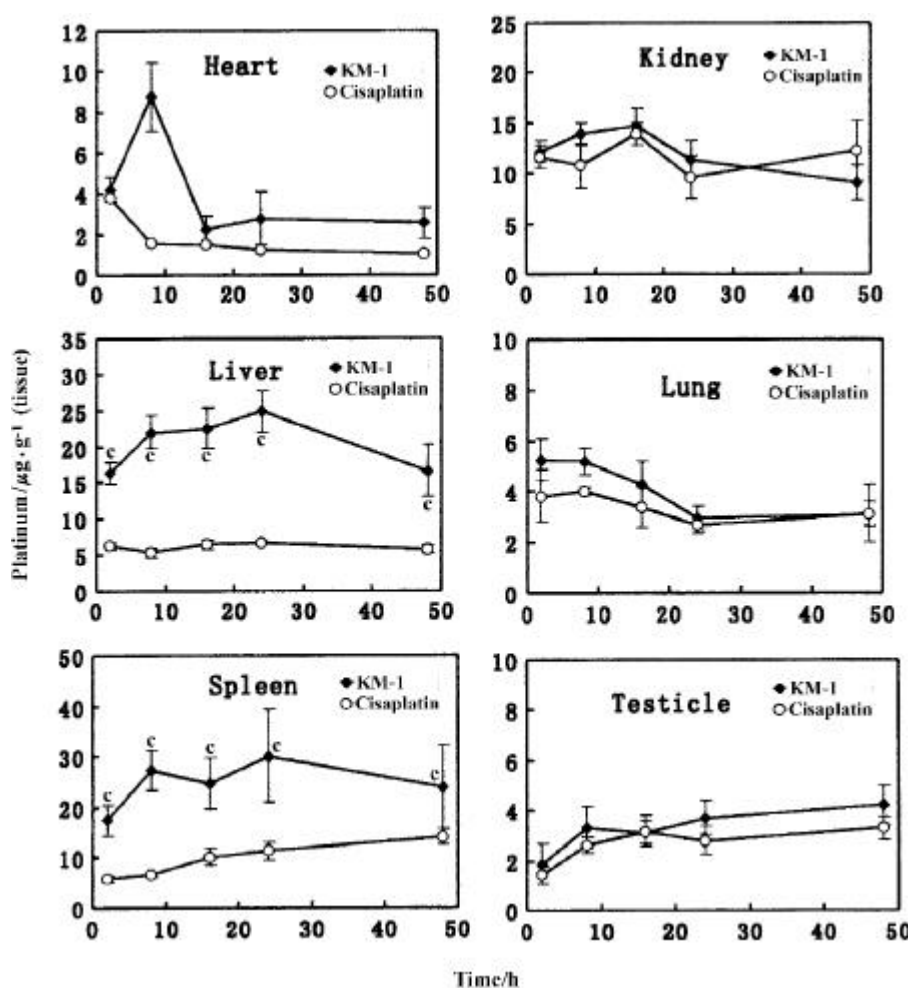


Fig 2. Concentration-time profiles of platinum in tissues following a single iv dose of KM-1 or cisplatin 4.5mg/kg.  $n=5$ . Mean $\pm$ SD. \* $P<0.01$  vs cisplatin group.

ment of tumors in these organs. And it agrees with our preliminary pharmacodynamic study with KM-1. The study showed that KM-1 had the equivalent antitumor efficacy with cisplatin in liver tumor bearing mice, but was superior to cisplatin in that it could alleviate the damage of liver caused by the tumor, while cisplatin would aggravate the damage.

We did not discern lower platinum concentrations in the kidney with KM-1 than with free cisplatin. To outward seeming, this is not consistent with our preliminary study showing that nephrotoxicity in KM-1-treated mice was less than that in cisplatin-treated ones. The explanation may lie in the different reactive mechanism between free drug with tissues and liposomal-entrapped drug with tissues. Since most of the drug is retained in liposome, the total amount of platinum measured in KM-1-treated group is not equivalent to that of platinum that can react with biological substances and has a pharmacological or toxicological efficacy. Cisplatin encapsulated in liposome was released gradually with time and tissues were continuously exposed to a low-dose drug environment. This could avoid a high peak free cisplatin level that would produce a severe toxicological effect. Similar situation occurred in liver with KM-1 treatment, which had high platinum levels and showed no hepatotoxicity. The result agrees with the studies of Gondal *et al*<sup>[11]</sup>.

No significant eliminations of platinum in tissues were observed during the first 48 h after dosing. This is due to the characteristics of cisplatin which would bind to protein or other tissue compositions firmly through covalent bonds. Excretion studies had also shown that only 25 % of total KM-1 taken by rats was excreted during (0-48) h period of time.

## REFERENCES

- 1 Rosenberg b, VanCamp L, Trosko JE, Monsour VH. Platinum compounds: a new class of potent antitumor agents. *Nature* 1969; 222: 385-6.
- 2 Einhorn LH, Donohue J. *cis*-Diaminedichloroplatinum, vinblastine, and bleomycin chemotherapy in disseminated testicular cancer. *Ann Intern Med* 1977; 87: 293-8.
- 3 Pearson ADJ, Craft AW, Pinkerton CR, Meller ST, Reid MM. High-dose rapid schedule chemotherapy for disseminated neuroblastoma. *Eur J Cancer* 1992; 28A: 1654-9.
- 4 Raymond BW, Michaele CC. New cisplatin analogues in development. *Drugs* 1993; 46: 360-77.
- 5 Kelland LR, Clarke SJ, Mckeage MJ. Advances in platinum complex cancer chemotherapy. *Platinum Metals Rev* 1992; 36: 178-84.
- 6 Yang YK, Xiong HZ, Pu SP, Liu WP, Chen XZ, He J. Study on the third generation platinum anticancer drugs. *Precious Metals* 1996; 17: 50-7.
- 7 Van Hoesel QGCM, Steenberg PA, Crommelin DJ, Vandijk A, Vanoort W, Klein S, *et al*. Reduced cardiotoxicity and nephrotoxicity with preservation of antitumor-activity of doxorubicin entrapped in stable liposomes in the LOU/M Wsl rats. *Cancer Res* 1984; 44: 3698-705.
- 8 Herman EH, Rahman A, Ferrans VJ, Vick JA, Schein PS. Prevention of chronic doxorubicin cardiotoxicity in beagles by liposomal encapsulation. *Cancer Res* 1983; 43: 5427-32.
- 9 Gabizon A, Catane R, Uziely B. Prolonged circulation time and enhanced accumulation in malignant exudates of doxorubicin encapsulated in polyethylene-glycol coated liposomes. *Cancer Res* 1994; 54: 987-92.
- 10 Newman MS, Colbern GT, Working PK, Engbers C, Amantea MA. Comparative pharmacokinetics, tissue distribution, and therapeutic effectiveness of cisplatin encapsulated in long-circulating, pegylated liposomes (SPI-077) in tumor-bearing mice. *Cancer Chemother Pharmacol* 1999; 43: 1-7.
- 11 Gondal JA, Preuss HG, Swartz R, Rahman A. Comparative pharmacological, toxicological and antitumoral evaluation of free and liposome-encapsulated cisplatin in rodents. *Eur J Cancer* 1993; 29A: 1536-42.