



New frontiers in fecal immunochemical and other fecal tests in colorectal cancer screening

Stefano Rapi¹, Filippo Cellai², Tiziana Rubeca²

¹Department of Service, Central Laboratory, Careggi Hospital, Florence, Italy; ²Regional Cancer Prevention Laboratory, Cancer Prevention and Research Institute (ISPO), Florence, Italy

Correspondence to: Stefano Rapi. Department of Service, Central Laboratory, Careggi Hospital, AOU Careggi, Largo Brambilla 3, 50139 Florence, Italy. Email: rapis@aou-careggi.toscana.it.

Comment on: Grobbee EJ, Schreuders EH, Hansen BE, *et al.* Association Between Concentrations of Hemoglobin Determined by Fecal Immunochemical Tests and Long-term Development of Advanced Colorectal Neoplasia. *Gastroenterology* 2017. [Epub ahead of print].

Received: 22 September 2017; Accepted: 10 October 2017; Published: 17 November 2017.

doi: 10.21037/jlpm.2017.10.03

View this article at: <http://dx.doi.org/10.21037/jlpm.2017.10.03>

Introduction: colorectal cancer (CRC) and fecal immunochemical test for hemoglobin

Since 1993, fecal immunochemical tests for hemoglobin (FIT-Hb) have been used as an initial test in CRC screening programs (1,2,3) and a large amount of data have supported the efficacy of this strategy to reduce CRC mortality (3,4). In addition, new evidence of a reduced CRC incidence in subjects enrolled in long term screening programs comes from about 2,000 reports (4,5). The ‘cost-effectiveness’ of CRC screening programs are related to cost, advantages and harms and overall compliance of the target population together to organizational and institutional considerations, therefore it is difficult to evaluate this cost in detail. Nevertheless, several factors indicate the FIT based strategies as the most effective non-invasive investigation for CRC screening (6,7). Importantly, after that the European Union Guidelines have suggested the adoption of FIT-Hb screening in the European Community in 2010 (8), a lot of countries have started population screening programs and a growing interest in both analytical and clinical aspects of FIT was observed worldwide. Today, there are two main fields of research regarding FIT-Hb based CRC population screening.

Gastrointestinal bleeding detectable by FIT-Hb

The first research topic is related to interesting biological aspects that consist in the transition from ‘pre-neoplastic’ to ‘malignant’ lesions and the detection of gastrointestinal

bleeding during ‘pre-neoplastic’ and/or inflammatory process. Indeed, detectable Hb concentrations can be found in the gastrointestinal tract, not only in neoplastic lesions, but also in ‘pre-neoplastic’ adenoma, hyperplastic polyps, diverticular disease, hemorrhoids, intestinal inflammatory process and in a significant number of participants with normal bowel on colonoscopy. Recent evidence indicates an increased individual CRC risk when detectable Hb levels with current FIT-Hb analyses are detectable.

The biological significances of low amounts of bleeding are not fully understood and Hb values below the cut-off used in FIT-Hb based screening programs (most frequently 20 µg Hb/g feces) are rarely associated to lesions detected by colonoscopy (9). But, the consequences of such gastrointestinal bleeding detectable by FIT-Hb are clearly reported in literature (10). The study of Grobbee *et al.* (11) should be considered in this context because it confirms the importance of fully understand the meaning of low fecal Hb concentrations and it addresses the specific related risk in individuals screened population in order to ‘customize’, for the single participant, the fecal Hb results obtained over sequential rounds of the screening. The value of finding a Hb value between the detection and decisional limits (reliable concentrations under selected the applied cut-off) consists in the identification of a sub-group of participants with an increased risk of developing CRC in subsequent years.

Epidemiological data have confirmed the close

associations of gastrointestinal bleeding with neoplastic risk, supporting to the concept of stratifying the risk of single individual participating in screening program. In particular, the study of Carrera *et al.* (12) found an increased risk of developing CRC in the years subsequent a negative colonoscopy into a cohort of participants with fecal Hb concentrations over the 'cut-off', applied in the screening program, and without detectable lesions at colonoscopy. It is known that a significant number of CRC are not detected by screening programs and during colonoscopy, hence, the investigation of interval cancers between subsequent screening rounds can offer an estimate of undetected neoplasms in a specific screening program. However, the number of undetected CRC in this kind of cohort seems to be too high to support epidemiological results on the bases of incomplete or incorrect colonoscopies.

Interestingly, a large multicenter case-control study on fecal DNA reports an increased sensitivity for 'pre-cancerous' lesions in proportion to lesion size from 57% for lesions >1 cm to 83% for those >3 cm, confirming the data on fecal Hb concentrations and severity of colorectal disease (13). Therefore, the increased risk to develop CRC seems to be related to the gastrointestinal 'bleeding' and the pre-malignant lesions more than to the neoplasms being undetected during colonoscopy. This significant epidemiological evidence must be considered to better address the frequency of FIT sampling in individuals. Medium and high-risk cohorts could have different intervals of FIT sampling or different control strategy, set overall by the screening programs in order to reduce the interval cancer proportion and optimize outcomes (14). As affirmed in 'Gut': development of a screening algorithm that combines FIT concentrations with other risk indicators (age, sex, screening history, deprivation) may enable improved clinical outcomes within the same colonoscopy resource and sets this concept as one of the main current topics in CRC screening (15).

Pre-analytical and analytical aspects of FIT-Hb

The second significant topic on FIT-Hb methods is strictly related to the method performance and characteristics of fecal tests and directly involves the sampled materials. The effectiveness of FIT based screening arises from two main advantages: the high compliance (high rate of uptake in the population invited to CRC screening programs), a major issue in population screening, and the possibility of 'determining a cut-off appropriate for the available colonoscopy resources' (15). High compliance with FIT

is directly related to the initial materials used for the investigation, including the requirement only to sample one fecal sample into a simple to use, hygienic specimen collection device. The possibility to select subjects at higher risk, starting with a non-invasive and self-sampled material is probably the main reason for the high uptake recorded as compared to other CRC screening strategies, such as flexible sigmoidoscopy, or other organized screening programs based on laboratory investigation operating in Italy (16). The use of postal delivery (17), under controlled conditions, can avoid the inconvenience of delivering materials to specific collecting sites and can assure an optimum uptake, together with a higher analytical quality of the estimation of the fecal Hb concentration. The positive effect of the use of a self-sampled biological material is 'unconventional' in the laboratory of medicine setting, but, can be somewhat negated by significant problems caused by lack of knowledge and consideration of the effects of the sampled material on the specimen collection, handling and the analytical technique.

The importance of the sampling phase and the effects of feces consistency were pointed out some years ago by Fraser *et al.* (18), generating the second important research topic for the FIT-Hb, mainly addressed to the laboratory and the analytical aspects. Pre-analytical effects on FIT-Hb measurements involve a number of different items. Indeed, the interaction of fecal materials with sample collection devices (design of the specimen collection probe) can affect the amounts of feces collected during self-sampling (19). The adoption of a correct metrological strategy ($\mu\text{g Hb/mL faces}$) (20) to indicate the amount of material collected by different sampling devices can also cause significant progress to be made in the understanding and addressing sampling variability and the overall pre-analytical sources of variation. Whereas, the stability of the Hb which in the specimen collection devices are strictly related to the buffer composition, presence of stabilizers and preservatives (21). In this context, it is relevant to remember that the compositions of buffers are covered by manufacturers' licenses. Finally, the ionic strength and the pH of the buffer in the device can change after addition of the sampled fecal materials. These changes can affect the final results because of factors related to the capability of buffers to reduce the changes and the overall amount of feces added to the device (22).

All these aspects of the pre-analytical and analytical phases of FIT-Hb should be fully addressed to control

the overall variability of results allowing comparison of the Hb concentrations across geography and methods eliminating the necessity to perform complex and expensive epidemiological investigation to determine the appropriate cut-offs for the available colonoscopy resources (17,23). This topic, together with other analytical aspects, is the subject of work, since January 2017 (23), by a Working Group of the Scientific Division of the International Federation of Clinical Chemistry and Laboratory Medicine and it has been extensively studied by the Italian SIBioC-GISCoR Working Group (Italian Society of Clinical Biochemistry and Clinical Molecular Biology—Italian Colorectal Screening Group) since 2014 (24).

Newer fecal tests

Although related to malignancy, the presence of Hb in gut cannot be considered ‘cancer specific’ and other new analytical methods should be developed to improve the performance of CRC screening programs. In 2014, the Food and Drug Administration (FDA) has approved a fecal DNA method as a screening test for CRC; a composite test that includes an immunochemical assay, similar to the one used in FIT, plus methylated markers and molecular mutations markers associated with CRC. Previous multicenter studies comparing fecal DNA test to FIT using colonoscopy as the gold standard showed that the fecal DNA test had a higher sensitivity than FIT for detecting CRC (92% *vs.* 74%), adenomas with high-grade dysplasia (69% *vs.* 46%), and serrated sessile polyps (42% *vs.* 5%) (24). Nevertheless, the high cost and the low specificity of this fecal DNA test (87–90%, compared to 95–96% of FIT-HB) (25) make the introduction of the test expensive in population screening, whereas DNA analysis can represent a most interesting strategy to improve the performances of the CRC screening.

Acknowledgments

Prof CG Fraser is gratefully acknowledged for manuscript revision.

Funding: None.

Footnote

Provenance and Peer Review: This article was commissioned and reviewed by the Section Editor Dr. Dao-Jun Hu (Department of Clinical Laboratory, Xin Hua Hospital Affiliated to Shanghai Jiaotong University School of

Medicine, Chongming Branch, Shanghai, China).

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <http://dx.doi.org/10.21037/jlpm.2017.10.03>). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Open Access Statement: This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

References

1. Mandel JS, Bond JH, Church TR, et al. Reducing mortality from colorectal cancer by screening for fecal occult blood. Minnesota Colon Cancer Control Study. *N Engl J Med* 1993;328:1365-71.
2. Saito H, Soma Y, Koeda J, et al. Reduction in risk of mortality from colorectal cancer by fecal occult blood screening with immunochemical hemagglutination test. A case-control study. *Int J Cancer* 1995;61:465-9.
3. Zappa M, Castiglione G, Grazzini G, et al. Effect of faecal occult blood testing on colorectal mortality: results of a population-based case-control study in the district of Florence, Italy. *Int J Cancer* 1997;73:208-10.
4. Mandel JS, Church TR, Bond JH, et al. The effect of fecal occult-blood screening on the incidence of colorectal cancer. *N Engl J Med* 2000;343:1603-7.
5. Ventura L, Zappa M, Carreras G, et al. What is the best screening strategy to detect advanced colorectal adenomas? Simulation from ongoing Italian screening experiences. *Tumori* 2011;97:547-50.
6. Heitman SJ, Hilsden RJ, Au F, et al. Colorectal cancer screening for average-risk North Americans: an economic evaluation. *PLoS Med* 2010;7:e1000370.
7. van Rossum LG, van Rijn AF, Verbeek AL, et al. Colorectal cancer screening comparing no screening,

- immunochemical and guaiac fecal occult blood tests: a cost-effectiveness analysis. *Int J Cancer* 2011;128:1908-17.
8. Von Karsa L, Patrick J, Segnan N. European guidelines for quality assurance in colorectal cancer screening and diagnosis. 1st edition. Luxembourg: Publication Office of the European Union, 2010.
 9. Grazzini G, Visioli CB, Zorzi M, et al. Immunochemical faecal occult blood test: number of samples and positivity cutoff. What is the best strategy for colorectal cancer screening? *Br J Cancer* 2009;100:259-65.
 10. Chen LS, Yen AM, Fraser CG, et al. Impact of faecal haemoglobin concentration on colorectal cancer mortality and all-cause death. *BMJ Open* 2013;3:e003740.
 11. Grobbee EJ, Schreuders EH, Hansen BE, et al. Association Between Concentrations of Hemoglobin Determined by Fecal Immunochemical Tests and Long-term Development of Advanced Colorectal Neoplasia. *Gastroenterology* 2017. [Epub ahead of print].
 12. Carrera A, McClements PL, Watling C, et al. Negative screening colonoscopy after a positive guaiac faecal occult blood test: not a contraindication to continued screening. *Colorectal Dis* 2012;14:943-6.
 13. Lidgard GP, Domanico MJ, Bruinsma JJ, et al. Clinical performance of an automated stool DNA assay for detection of colorectal neoplasia. *Clin Gastroenterol Hepatol* 2013;11:1313-8.
 14. Zappa M, Castiglione G, Paci E, et al. Measuring interval cancers in population-based screening using different assays of fecal occult blood testing: the District of Florence experience. *Int J Cancer* 2001;92:151-4.
 15. Moss S, Mathews C, Day TJ, et al. Increased uptake and improved outcomes of bowel cancer screening with a faecal immunochemical test: results from a pilot study within the national screening programme in England. *Gut* 2017;66:1631-44.
 16. I programmi di screening in Italia 2014. Available online: http://www.salute.gov.it/imgs/C_17_pubblicazioni_2305_allegato.pdf
 17. Passamonti B, Malaspina M, Fraser CG, et al. A comparative effectiveness trial of two faecal immunochemical tests for haemoglobin (FIT). Assessment of test performance and adherence in a single round of a population-based screening programme for colorectal cancer. *Gut* 2016. [Epub ahead of print].
 18. Fraser CG, Allison JE, Halloran SP, et al. A proposal to standardize reporting units for fecal immunochemical tests for hemoglobin. *J Natl Cancer Inst* 2012;104:810-4.
 19. Rapi S, Rubeca T, Fraser CG. How to improve the performances of Fecal Immunological Tests (FIT): Need for standardization of the sampling and pre-analytical phases and revision of the procedures for comparison of methods. *Int J Biol Markers* 2015;30:e127-31.
 20. Fraser CG, Rapi S, Rubeca T. RE: A Proposal to Standardize Reporting Units for Fecal Immunochemical Tests for Hemoglobin. *J Natl Cancer Inst* 2015;108. pii: djv312.
 21. Grazzini G, Ventura L, Rubeca T, et al. Impact of a new sampling buffer on faecal haemoglobin stability in a colorectal cancer screening programme by the faecal immunochemical test. *Eur J Cancer Prev* 2017;26:285-91.
 22. Rapi S, Berardi M, Cellai F, et al. Effects of fecal sampling on preanalytical and analytical phases in quantitative fecal immunochemical tests for hemoglobin. *Int J Biol Markers* 2017;32:e261-6.
 23. Rubeca T, Peruzzi B, Confortini M, et al. Overall evaluation of an immunological latex agglutination system for fecal occult blood testing in the colorectal cancer screening program of Florence. *Int J Biol Markers* 2012;27:e195-202.
 24. Imperiale TF, Ransohoff DF, Itzkowitz SH. Multitarget stool DNA testing for colorectal-cancer screening. *N Engl J Med* 2014;371:187-8.
 25. IFCC-FIT Working Group (FIT-WG). <http://www.ifcc.org/media/461890/IFCCeNewsJune2017.pdf> last access 24 august 2017

doi: 10.21037/jlpm.2017.10.03

Cite this article as: Rapi S, Cellai F, Rubeca T. New frontiers in fecal immunochemical and other fecal tests in colorectal cancer screening. *J Lab Precis Med* 2017;2:88.