

The expression and prognostic value of GLYATL1 and its potential role in hepatocellular carcinoma

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Background: Glycine-N-acyltransferase-like 1 (GLYATL1), which is involved in the detoxification of endogenous and exogenous acyl-CoA, promotes glutamine metabolism in xenobiotic metabolism. Recent evidence suggests an association between GLYATL1 and tumors. However, there are few comprehensive analyses of GLYATL1 in cancers. We evaluated the expression and prognostic value of GLYATL1 and explored the mechanism underlying the association between GLYATL1 and cancers.

Methods: GLYATL1 mRNA expression across cancers was investigated in the Oncomine database and confirmed in the UALCAN and Gene Expression Profiling Interactive Analysis (GEPIA) databases. Next, its prognostic value in different cancers was revealed by PrognoScan and Kaplan-Meier plotter. According to clinicopathologic features, we conducted a subgroup analysis of the prognosis of GLYATL1 in a cohort of hepatocellular carcinoma (HCC) patients from The Cancer Genome Atlas (TCGA) and the GSE116174 dataset. We further investigated the GLYATL1 promoter methylation profile in HCC. Next, a protein-protein interaction (PPI) network was constructed via the Search Tool for the Retrieval of Interacting Genes (STRING) database. Finally, we utilized gene set enrichment analysis (GSEA) to identify significantly enriched pathways and confirmed their associations using the Tumor Immune Estimation Resource (TIMER) and GEPIA databases.

Results: GLYATL1 is downregulated in many cancers and indicates a poor prognosis. Specifically, low GLYATL1 expression was associated with short overall survival (OS) in HCC patients. Interestingly, GLYATL1 expression was associated with poor OS in stage I-II HCC patients and was revealed as an independent prognostic factor. The promoter methylation level of GLYATL1 in HCC tissue was significantly higher than that in normal liver tissue. The PPI network suggested that GLYATL1 is co-expressed with ten genes, including *CNGA3* and *GNB5*. GSEA revealed that GLYATL1 is predominantly negatively enriched in xenobiotic metabolism, and the gene association analysis in TIMER and GEPIA showed a significantly negative association between the expression of GLYATL1 and the expression of most genes involved in mitochondrial glutamine metabolism, including SLC1A5 and SLC1A11.

Conclusions: Our study is the first to shed light on the expression and prognostic value of GLYATL1 in cancers and provide a potential regulatory mechanism underlying HCC development.

Keywords: GLYATL1; prognosis; biomarker; hepatocellular carcinoma (HCC); glutamine

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Introduction

Hepatocellular carcinoma (HCC) accounts for the majority of primary liver cancers and is one of the most malignant tumors and the third leading cause of cancer-related mortality worldwide (1). There are various treatments for HCC, including radical or palliative hepatectomy, intraoperative radiofrequency ablation, transarterial chemoembolization, and other targeted drugs. Among these treatments, radical or palliative hepatectomy is still the most effective strategy of HCC management (2). Though some HCC patients have the opportunity for surgical resection, long-term survival remains poor because of the high rates of recurrence and vascular tumor emboli (3,4). Moreover, the molecular mechanism underlying HCC recurrence remains unclear. Thus, it is significant for us to identify a molecular target that can be regarded as a prognostic indicator for HCC patients.

Glycine-N-acyltransferase (GLYAT) plays an important role in the detoxification of endogenous and exogenous acyl-CoA. In mammals, various carboxylic acid derivatives are combined with an amino acid, and the resulting polypeptide increases water solubility and can be excreted through urine, which occurs mainly in both the liver and kidney (5,6). This process involves mainly the following processes. First, carboxylic acid and coenzyme A activate ATP-dependent carboxylic acid, and then coenzyme A ligase forms Gatley, which is the intermediate product of acyl-CoA. Finally, under the action of GLYAT, the acyl group is transferred to the glycine amino group. Glycine-N-acyltransferase-like 1 (GLYATL1), one of the members of GLYAT that constitutes a gene cluster, is located in the cytoplasm (specifically, the mitochondrion). GLYATL1 was first identified at the mRNA and cellular level in 2007 (7) and encodes an enzyme involved in the catalysis of arylacetyl transfer (8). In contrast to GLYAT, GLYATL1 exerts phenylacetyl-CoA glutamine N-acyltransferase activities in xenobiotic metabolism (9). GLYATL1 is involved not only in normal physiological metabolism but also in tumorigenesis. A previous study demonstrated that the mRNA expression of GLYATL1 was unregulated by androgen and ETS variant transcription factor 1 (ETV1) in prostate cancer (10). Moreover, Kishimoto and colleagues showed that the protein expression of GLYATL1 in HCC tissues was lower than that in non-tumor tissues (9). However, few studies have performed comprehensive analyses of GLYATL1 in cancers. We aimed to evaluate the expression and prognostic value of GLYATL1 and

explore the mechanism underlying the connection between GLYATL1 and cancers.

In the present study, we took advantage of data available in public databases to investigate the expression level and prognostic significance of GLYATL1 in different cancers. To better understand the prognostic value and underlying mechanisms of GLYATL1 in HCC, we also investigated the association between its expression level and clinicopathologic features. Moreover, a protein network of predicted associations for GLYATL1 was constructed. Next, we conducted a functional analysis of GLYATL1 in cancers. Finally, we explored the correlation between GLYATL1 and genes involved in glutamine metabolism in HCC. Thus, we hypothesize that GLYATL1 catalyzes glutamine to affect mitochondrial glutamine metabolism, further inhibiting the rapid proliferation of HCC. In summary, our study sheds light on the expression and prognostic value of GLYATL1 across cancers and provides a potential regulatory mechanism underlying the development of HCC.

We present the following article in accordance with the Materials Design Analysis Reporting (MDAR) reporting checklist (available at: http://dx.doi.org/10.21037/jgo-20-186).

Methods

Oncomine database analysis

Oncomine (http://www.oncomine.org) is a web-based cancer microarray database and data-mining platform containing 715 databases and 86,733 samples (11). Thus, we can utilize it to compare differential expression between cancer and normal tissues. In our study, we used the Oncomine database to investigate mRNA levels in different cancer and corresponding non-tumor tissues. The parameters were set as follows: gene: GLYATL1; and data type: mRNA. Other cut-off values were as follows: P value: <1e-4; fold change: >2; and gene rank: top 10%.

UALCAN cancer database analysis

The UALCAN cancer database (http://ualcan.path. uab.edu/index.html) is a comprehensive and systematic web resource that provides publicly available cancer OMICS data, including data from The Cancer Genome Atlas (TCGA) (12). In our study, we first confirmed the expression of GLYATL1 across cancers in the TCGA. We further investigated the GLYATL1 promoter methylation

profile based on its clinicopathologic characteristics, including sample type, individual cancer stage, tumor grade, and nodal metastasis status. The level of DNA methylation was indicated by the β -values. The cut-off values of hypermethylation and hypomethylation were defined as 0.7 and 0.3, respectively (13).

PrognoScan database analysis

PrognoScan (http://dna00.bio.kyutech.ac.jp/PrognoScan/ index.html) was used to investigate associations between the expression level of GLYATL1 and survival time in patients with different cancers. PrognoScan is a new database that collects all cancer microarray datasets from public databases, including ArrayExpress, and provides researchers a convenient approach for confirming the prognostic value of genes. Overall survival (OS) was used as the study endpoint.

Kaplan-Meier plotter database analysis

We can readily extract RNA-seq data from multiple public databases via Kaplan-Meier plotter (14) (https://kmplot. com/analysis/). We conducted a survival analysis across 21 cancers by plotting Kaplan-Meier curves and compared their differences by the log-rank p-value based on the mRNA RNA-seq data of GLYATL1 from the TCGA. We further used the RNA-seq data of GLYATL1 and the corresponding clinical information from the TCGA to investigate the associations between the expression level and clinicopathologic features in HCC.

Gene Expression Omnibus (GEO) database analysis

We downloaded a gene microarray with survival data (GSE116174) from the GEO (https://www.ncbi.nlm.nih. gov/geo/) to confirm the prognostic value and clinical correlation of GLYATL1 in HCC. The GSE116174 cohort was divided into a high expression group and a low expression group according to the median GLYATL1 expression level. Survival analysis was conducted as described above to investigate the correlation between GLYATL1 expression and clinical features. Then, univariate Cox analysis was used to identify survival-associated variables (P<0.15), which were sequentially subjected to multivariate Cox analysis to investigate independent

prognostic factors (P<0.05).

Tumor Immune Estimation Resource (TIMER) database analysis

TIMER (https://cistrome.shinyapps.io/timer/) is a web resource that can help researchers investigate the associations between tumor cells and six immune cell types, including B cells, CD4+ T cells, CD8+ T cells, neutrophils, macrophages and dendritic cells, based on 10,897 samples from 32 cancer types in the TCGA (15). First, a gene module was used to investigate the correlation between GLYATL1 expression and the abundance of immune infiltrates. Then, a survival module was chosen to explore the associations between clinical characteristics and immune infiltration or GLYATL1 expression in a multivariable Cox proportional hazards model. Finally, we utilized a correlation module to identify the correlation between GLYATL1 and key genes involved in glutamine metabolism, including solute carrier family 1, member 5 (SLC1A5), solute carrier family 7, member 11 (SLC7A11), glutaminase (GLS), glutaminase 2 (GLS2), solute carrier family 38, member 5 (SLC38A5), glutamate dehydrogenase 1 (GLUD1), glutamic-oxaloacetic transaminase 1 (GOT1) and glutamic-oxaloacetic transaminase 2 (GOT2). Partial correlation was conditioned on tumor purity.

Search Tool for the Retrieval of Interacting Genes (STRING) database analysis

We constructed protein-protein interaction (PPI) networks of coregulated hub genes via the STRING database (version 11.0), which contains 5,090 total organisms, 24,584,628 proteins and 3,123,056,667 total interactions (https://string-db.org/) (16). The parameters were set as follows: protein name: GLYATL1; and organism: Homo sapiens. A combined score >0.4 suggested a statistically significant interaction, and the top 10 genes were screened as hub genes. Next, the interactions were exported to Cytoscape software (version 3.7.1) for visual presentation. To determine the potential biological functions of the hub genes, Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analyses of the 10 coregulated hub genes were performed. Only the top 15 GO enrichment pathways [including biological process (BP), cellular component (CC), and molecular function (MF)

terms] and the top five KEGG pathways were listed and visualized using a bubble diagram.

Gene set enrichment analysis (GSEA)

GSEA can be used to analyze and interpret genome-wide expression profiles based on biological knowledge (17). The RNA-seq data of HCC patients of the TCGA downloaded from Genomic Data Commons (https://portal.gdc.cancer. gov/) were used to investigate the potential mechanism underlying the development of HCC by GSEA (17). The patients were divided into two groups according to the median mRNA level of GLYATL1: the low expression group and the high expression group. The parameters were set as follows: gene sets database: h. All. V7.0 Symbols. gmt (Hallmarks); number of permutations: 1,000; and permutation type: phenotype. Significantly enriched pathways and genes were defined as those with a P value <0.05 and a false discovery rate (FDR) <0.25. These criteria were used to minimize the occurrence of false-positive results in multiple tests.

Gene Expression Profiling Interactive Analysis (GEPIA)

GEPIA (http://gepia.cancer-pku.cn/), which contains RNA sequencing expression data from the TCGA and Genotype-Tissue Expression (GTEx) project, is an enhanced web server for interactive analyses based on a standard processing pipeline (18). First, we utilized GEPIA to further confirm the mRNA expression of GLYATL1 in human cancers. Then, GEPIA was used to confirm the gene association between GLYATL1 and genes involved in glutamine metabolism. GLYATL1 was set as the X-axis, and genes related to glycolysis were set as the Y-axis. Then, we calculated the correlation coefficient in HCC and normal liver tissues from the TCGA.

Statistical analysis

We utilized SPSS software (version 20.0) and the GEO2R portal (https://www.ncbi.nlm.nih.gov/geo/info/geo2r. html) to conduct the analysis of GSE116174 cohort. The correlations between GLYATL1 and key genes involved in glutamine metabolism were indentified by Spearman's correlation analysis. Except for the Oncomine database analysis and GEO database analysis, a P value less than 0.05 was considered significant in the present study.

The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013).

Results

mRNA expression level of GLYATL1 across different cancer and corresponding non-tumor tissues

We first utilized the Oncomine database to investigate the mRNA expression level of GLYATL1 across different cancers. As shown in Figure 1A, the GLYATL1 mRNA levels were significant in four cancers: colorectal cancer, kidney cancer, liver cancer, and prostate cancer. Among these cancers, GLYATL1 was upregulated in colorectal cancer and prostate cancer and downregulated in kidney cancer and liver cancer. Next, the expression of GLYATL1 was verified in the UALCAN cancer database. The results revealed a significant difference in GLYATL1 expression between tumor and normal tissues in breast invasive carcinoma (P<1e-12), cholangiocarcinoma (P=3.53e-05), colon adenocarcinoma (P=1.11e-16), kidney chromophobe carcinoma (P=2.38e-04), clear cell renal cell carcinoma (P=5.67e-07), kidney renal papillary cell carcinoma (P=2.01e-06), HCC (P=1.62e-12), lung adenocarcinoma (P=1.7e-12), lung squamous cell carcinoma (P=1.84e-12), prostate adenocarcinoma (P=1.62e-12), rectal adenocarcinoma (P=5.55e-16), and uterine endometrial carcinoma (P=3.28e-8) (Figure 1B). Considering that there are few normal samples in the TCGA, we integrated the data on normal tissues from the GTEx database and the data on tumor tissues from the TCGA to analyze the expression differences between human tumor tissues and their matched normal tissues. As shown in Figure S1, further validation with the GEPIA database also indicated similar results.

Prognostic effects of GLYATL1 mRNA expression in patients with different cancers

As shown in Table S1, PrognoScan revealed that high GLYATL1 mRNA expression levels indicated poor OS of patients with breast cancer (HR =1.73, P=0.004) and skin melanoma (HR =2.51, P=0.036) but improved OS of patients with renal cell carcinoma (HR =0.20, P=0.040), lung adenocarcinoma (HR =0.56, P=0.034), and colorectal cancer (HR =0.47, P=0.016). To further confirm the association between GLYATL1 mRNA expression and the prognostic value in patients with different cancers, the mRNA RNA-seq data of GLYATL1 in 21 cancers from the

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Figure 2 Kaplan-Meier survival curves comparing high and low GLYATL1 mRNA expression in HCC patients from the TCGA database. (A) Overall survival (OS); (B) relapse-free survival (RFS); (C) progression-free survival (PFS); (D) disease-specific survival (DSS).

TCGA and survival information were used to plot Kaplan-Meier curves, and we compared differences in their log-rank P values using Kaplan-Meier plotter. As shown in *Figure 2A* and Figure S2, high GLYATL1 mRNA expression levels were associated with improved OS of patients with clear cell renal cell carcinoma (HR =0.49, P=3.1e-06), papillary renal cell carcinoma (HR =0.46, P=0.011), and HCC (HR =0.61, P=0.005). Of note, we found that the GLYATL1 expression level in HCC tissues was significantly lower than that in normal liver tissues, which indicated poor OS. Similar results were also obtained when we further investigated other survival outcomes, including progression-free survival (PFS) and disease-free survival (DSS), in HCC patients (Figure 2B,C,D).

Subanalysis of GLYATL1 mRNA expression and the clinicopathologic features of HCC patients

To determine how GLYATL1 mRNA expression affects the prognosis of HCC patients (based on results from Kaplan-Meier plotter), we further explored the correlation between GLYATL1 mRNA expression and the clinicopathologic features of HCC patients using the RNA-seq data of GLYATL1 and the corresponding clinical information from the TCGA. The results indicated that downregulated GLYATL1 mRNA expression was associated with poor

OS in Asian HCC patients, a subgroup of HCC patients from TCGA (HR =0.36, P=0.001) and male HCC patients (HR =0.53, P=0.0065). Regarding a history of alcohol consumption and hepatitis virus, we found a significant association between low GLYATL1 mRNA expression and poor OS in patients with a history of alcohol consumption (HR =0.42, P=0.0071). Surprisingly, unlike other cancer biomarkers, in our study, decreased GLYATL1 mRNA expression was associated with poor OS in HCC patients whose stages were I-II (HR =0.61, P=0.0489), whose grades were 3 (HR =0.46, P=0.0116) or with vascular invasion (HR =0.56, P=0.03). We also used data from the GSE116174 dataset to confirm GLYATL1 expression and clinical data. The results in the GSE116174 cohort indicated that high GLYATL1 mRNA expression indicated improved OS in patients in stages I-II (HR =0.42, P=0.047) and with no vascular invasion (HR =0.29, P=0.0016). Detailed information is listed in Table 1. High-throughput specimens may include immune cell infiltration, and immune cell infiltration could also impact the survival of HCC patients. We further investigated GLYATL1 expression and immune cell infiltration. As shown in Figure S3A, there was a significant association between immune cell infiltration and GLYATL1 expression. Thus, immune cell infiltration was regarded as a critical variable. We drew Kaplan-Meier plots for B cells, CD4+ T cells, CD8+ T cells, neutrophils, macrophages and dendritic cells to visualize the survival differences in the HCC patient cohort from the TCGA, and the results indicated that the expression of these six immune infiltrates did not lead to significant survival differences (Figure S3B). Univariate and multivariate Cox analyses indicated that GLYATL1 expression (HR =0.847, P<0.0001) was an independent prognostic factor for HCC patients (Table 2). We also analyzed immune cell infiltration in the GSE116174 dataset. As presented in Table S2, we obtained results that were similar to those of the TCGA cohort. Validation in the GSE116174 cohort also confirmed the role of GLYATL1 (HR =0.297, P<0.005) as an independent

role of GLYATL1 (HR =0.297, P<0.005) as an independent factor (*Table 2*). Taken together, these results suggest that GLYATL1 is an independent prognostic factor for HCC patients.

GLYATL1 promoter methylation profile based on the clinicopathologic characteristics of HCC patients

DNA methylation is a chemical modification of DNA that can alter genetic performance without changing the DNA sequence and control gene expression (19,20). Therefore, 1311

we further explored the GLYATL1 promoter methylation profile based on the clinicopathologic characteristics of HCC patients. As shown in *Figure 3*, the results indicated that the promoter methylation level of GLYATL1 in HCC tissue was significantly higher than that in normal liver tissue (P<1e-12). Next, we performed a subgroup analysis based on clinicopathologic characteristics. Of note, the GLYATL1 promoter methylation profile based on individual cancer stage, tumor grade and nodal metastasis status suggested significant differences (all P<0.0001). These results indicate that high GLYATL1 promoter methylation might suppress the expression level of GLYATL1, leading to the development of HCC.

Construction of the PPI network via the STRING database

The STRING database was used to construct PPI networks of GLYATL1 and its coregulated hub genes. The results indicated that GLYATL1 was co-expressed with cyclic nucleotide gated channel alpha 3 (CNGA3), guanine nucleotide-binding protein subunit beta-5 (GNB5), K acetyltransferase 2A (KAT2A), K acetyltransferase 2B (KAT2B), Opsin 4 (OPN4), polyamine oxidase (PAOX), the regulator of G protein signaling 9 (RGS9), RGS9binding protein (RGS9BP), spermidine/spermine N1acetyltransferase 1 (SAT1) and spermidine/spermine N1acetyltransferase 2 (SAT2) (Figure 4A). Among these genes, KAT2A had the maximum combined score (n=0.836), followed by RGS9BP, KAT2B, and OPN4 (n=0.8). Next, GO analysis with CC, BP, and MF terms and KEGG pathway analysis of these 10 coregulated hub genes were conducted. GO analysis showed that "photoreceptor outer segment", "ciliary membrane" and "Ada2/Gcn5/Ada3 transcription activator complex" in CC; "N-acyltransferase activity", "N-acetyltransferase activity" and "diamine N-acetyltransferase activity" in MF; and "putrescine catabolic process", "spermine metabolic process" and "spermidine metabolic process" in BP were the most significantly enriched pathways associated with the 10 coregulated hub genes (all P<0.0001). KEGG pathway analysis revealed that most genes were enriched in "arginine and proline metabolism", "ferroptosis" and "Notch signaling pathway" (all P<0.01) (*Figure 4B*).

Potential mechanisms of downregulated GLYATL1 that regulate the tumorigenicity of HCC

In our study, GLYATL1 expression was lower in HCC

Table 1 Prognostic value of GLYATL1 based on different clinicopathological factors in the LIHC cohort of the TCGA and the GSE116174 dataset

Variable		TCGA (n=364)			GSE116174 (n=64)	
variable	Ν	Hazard ratio	Р	Ν	Hazard ratio	Р
Gender						
Male	246	0.53 (0.34–0.85)	**	58	0.52 (0.24–1.13)	0.097
Female	118	0.58 (0.30–1.12)	0.1	6	-	-
Stage						
I–II	253	0.61 (0.38–1.00)	0.489	53	0.42 (0.18–0.99)	*
III–IV	87	0.67 (0.38–1.18)	0.16	11	2.87 (0.48–17.28)	0.249
Alcohol consump	tion					
Yes	115	0.42 (0.22–0.81)	**	13	0.66 (0.28–1.55)	0.338
No	202	0.79 (0.50–1.25)	0.31	51	0.33 (0.07–1.68)	0.333
Hepatitis virus						
Yes	150	0.53 (0.27–1.05)	0.064	47	0.59 (0.24–1.47)	0.257
No	167	0.68 (0.44–1.08)	0.099	17	0.61 (0.16–2.57)	0.502
Vascular invasion						
Yes	203	0.56 (0.33–0.95)	*	35	0.38 (0.10–1.53)	0.38
No	90	0.53 (0.24–1.18)	0.11	29	0.29 (0.10–0.79)	*
AJCC_T						
1	180	0.48 (0.26–0.88)	*			
2	90	0.68 (0.32–1.48)	0.305			
3	78	0.66 (0.36–1.20)	0.169			
Grade						
1	55	0.47 (0.18–1.22)	0.112			
2	174	0.59 (0.35–1.01)	0.052			
3	118	0.46 (0.25–0.85)	*			
Race						
White	181	0.85 (0.54–1.34)	0.47			
Asian	155	0.36 (0.19–0.68)	**			
Age						
>54				29	0.63 (0.19–2.05)	0.63
≤54				35	0.45 (0.15–1.34)	0.151
Smoking history						
Yes				31	0.80 (0.27–2.39)	0.688
No				32	0.32 (0.12–1.24)	0.392
Missing				1	-	_

*, P<0.05; **, P<0.01.

Parameters Universe Number learning State and set of ratio State and set o		TCGA				GSE116174							
Hazard ratio 95% Cl P Hazard ratio 95% Cl 95% Cl 95% Cl 958 Cl<	Parameters	Univariate analysis Multivariate analysis			ysis	Univariate analysis			Multivariate analysis		lysis		
Age 1.01 0.997- 1.024 0.130 0.26 1.01 0.25 0.164 0.162 0.142 0.949-1.034 0.663 0.533 0.154 0.239 Gender (male/ female) 0.82 0.573- 1.163 0.26 1.54 3.32 0.45-24.51 0.239 Race (White/ Black/Asian) 1.54 0.666- 3.622 0.21 1.54 1.54 1.54 1.54 1.54 1.54 Stage (IV/III/I/I) 1.42 0.827- 2.327 0.158 1.55 0.55 1.55		Hazard ratio	95% CI	Р	Hazard ratio	95% CI	Р	Hazard ratio	95% CI	Р	Hazard ratio	95% CI	Ρ
Gender (male/ female) 0.82 0.573 0.26 3.32 0.45-24.51 0.239 Face (White/ Black/Asian) 1.64 0.656- 0.321 5.55 <td>Age</td> <td>1.01</td> <td>0.997– 1.024</td> <td>0.139</td> <td>1.01</td> <td>0.996– 1.025</td> <td>0.164</td> <td>0.142</td> <td>0.949–1.034</td> <td>0.663</td> <td></td> <td></td> <td></td>	Age	1.01	0.997– 1.024	0.139	1.01	0.996– 1.025	0.164	0.142	0.949–1.034	0.663			
Bace (White/ Black/Asian) 1.54 0.656- 3.622 0.321 Stage (IV/II/I/I) 1.42 0.872- 2.323 0.158 Purity 2.07 0.901- 4.759 0.087 2.347 0.991- 5.555 0.052 GLYATL1 expression (righ/low) 0.866 0.79-0.94 0.001 0.847 0.774- 0.927 <001 HBV history (ves/no)	Gender (male/ female)	0.82	0.573– 1.163	0.26				3.32	0.45–24.51	0.239			
Stage (IV/III/I/I) 1.42 0.872- 2.323 0.158 Purity 2.07 0.901- 4.759 0.087 2.347 0.991- 5.555 0.052 GLYATL1 expression (righ/low) 0.86 0.79-0.94 0.001 0.847 0.774- 0.927 <0.001	Race (White/ Black/Asian)	1.54	0.656– 3.622	0.321									
Purity 2.07 0.901- 4.759 0.087 2.347 0.991- 5.55 0.052 GLYATL1 expression (righ/low) 0.86 0.79-0.94 0.001 0.847 0.774- 0.927 0.001 HBV history (res/no)	Stage (IV/III/II/I)	1.42	0.872– 2.323	0.158									
GLYATL1 expression (high/low) 0.86 0.79-0.94 0.001 0.847 0.774 0.927 <0.001	Purity	2.07	0.901– 4.759	0.087	2.347	0.991– 5.555	0.052						
HBV history 0.783 0.342-1.794 0.564 Alcohol history 1.099 0.443-2.727 0.839 Smoking history 0.673-3.163 0.339 Vascular invasion 0.258 0.115-0.58 0.001 0.162 0.066- 0.001 GLYATL1 0.266-1.209 0.142 0.297 0.128- 0.005 0.687 0.005 Stage (III/II/) 1.171 0.571-2.399 0.667 1.171 0.571-2.399 0.667 0.128- 0.005	GLYATL1 expression (high/low)	0.86	0.79–0.94	0.001	0.847	0.774– 0.927	<0.001						
Alcohol history (yes/no) 1.099 0.443-2.727 0.839 Smoking history (yes/no) 1.459 0.673-3.163 0.339 Vascular invasion (no/yes) 0.258 0.115-0.58 0.001 0.162 0.066- 0.399 <0.001	HBV history (yes/no)							0.783	0.342–1.794	0.564			
Smoking history (yes/no) 1.459 0.673–3.163 0.339 Vascular invasion (no/yes) 0.258 0.115–0.58 0.001 0.162 0.066– 0.399 GLYATL1 expression (high/low) 0.567 0.266–1.209 0.142 0.297 0.128– 0.005 Stage (III/II/) 1.171 0.571–2.399 0.667 0.005	Alcohol history (yes/no)							1.099	0.443–2.727	0.839			
Vascular invasion (no/yes) 0.258 0.115–0.58 0.001 0.066- <0.001	Smoking history (yes/no)							1.459	0.673–3.163	0.339			
GLYATL1 0.567 0.266–1.209 0.128– 0.005 expression (high/low) 0.567 0.267–1.209 0.128– 0.005 Stage (III/II/I) 1.171 0.571–2.399 0.667 5	Vascular invasion (no/yes)							0.258	0.115–0.58	0.001	0.162	0.066– 0.399	<0.001
Stage (III/II/I) 1.171 0.571–2.399 0.667	GLYATL1 expression (high/low)							0.567	0.266–1.209	0.142	0.297	0.128– 0.687	0.005
	Stage (III/II/I)							1.171	0.571–2.399	0.667			

Table 2 Univariate and multivariate Cox analyses of overall survival in patients with liver cancer from the TCGA and the GSE116174	dataset
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tissues than in normal tissues, and low GLYATL1 expression indicated a poor survival rate of HCC patients. Therefore, GLYATL1 might serve as a novel biomarker for patients with HCC. Compared with GO and KEGG pathway analyses, GSEA is more reliable because of its ability to distinguish expression changes in gene sets. Thus, GSEA was used to further explore the potential biological pathogenesis of GLYATL1 using the HCC patient cohort from the TCGA. As shown in *Figure 5A,B,C,D,E,F,G* and *Table 3*, the results indicated that low GLYATL1 expression is associated with bile acid metabolism, xenobiotic metabolism, fatty acid metabolism, coagulation, the peroxisome and adipogenesis.

Correlation analysis of GLYATL1 and genes involved in glutamine metabolism

We utilized TIMER to explore associations between GLYATL1 and the key genes involved in mitochondrial glutamine metabolism based on the expression datasets of HCC and normal liver tissue from the TCGA. As shown in *Figure 6*, the expression of GLYATL1 showed a significant



Figure 3 Promoter methylation level of GLYATL1 in LIHC based on clinicopathologic characteristics. (A) Sample type; (B) individual cancer stage; (C) tumor grade; (D) nodal metastasis status. *, P<0.05; ***, P<0.001.

association with the expression of most genes involved in mitochondrial glutamine metabolism, including SLC1A5 (partial.cor =-0.511, P=2.51e-24), SLC1A11 (partial. cor =-0.315, P=2.24e-09), GLS (partial.cor =-0.293, P=2.82e-08), GLS2 (partial.cor =0.453, P=7.17e-19), SLC38A5 (partial.cor =-0.399, P=1.21e-14), GOT1 (partial. cor =0.195, P=2.77e-04) and GOT2 (partial.cor =0.464, P=7.91e-20), in HCC tissues. As presented in *Table 4*, GEPIA also confirmed the association between GLYATL1 and genes involved in mitochondrial glutamine metabolism. Taken together, these results suggest that GLYATL1 facilitates the development of HCC by regulating mitochondrial glutamine metabolism. However, more experimental studies are needed to validate this conclusion.

Discussion

GLYATL1 is involved in both normal physiological metabolism and tumorigenesis. However, the prognostic value of GLYATL1 is still unknown. Here, to our knowledge, this is the first report on the expression level and prognostic value of GLYATL1 across different cancers. Notably, we found that the expression of GLYATL1 in HCC tissues was lower than that in normal liver tissues and indicated poor OS. Moreover, univariate and multivariate Cox analyses revealed that GLYATL1 could be an independent prognostic factor for patients with HCC. Interestingly, decreased GLYATL1 mRNA expression was associated with poor OS in HCC patients with stages I-II disease. Additionally, the promoter methylation level of GLYATL1 in HCC tissue was significantly higher than that in normal liver tissue. The PPI network was constructed with GLYATL1 and its coregulated hub genes. Furthermore, our study suggests that a low expression level of GLYATL1 in HCC is positively associated with xenobiotic metabolism and significantly associated with the expression of most genes involved in mitochondrial glutamine metabolism, such as SLC1A5, SLC1A11, GLS and SLC38A5. We further hypothesize that GLYATL1 catalyzes glutamine to affect mitochondrial glutamine metabolism, further inhibiting the rapid proliferation of HCC. In summary, we conducted a systematic assessment of the potential role of GLYATL1 in HCC and determined that GLYATL1 can be regarded as a novel and potential prognostic biomarker for HCC patients.

In the present study, the mRNA expression level of



Figure 4 Construction of the PPI network and gene enrichment of GLYATL1 and its coregulated hub genes. (A) The PPI network was constructed with the STRING database and visualized with Cytoscape software. The size and color of map nodes were determined by degree values. The width of the map edges was determined by combined scores. The lighter and smaller map nodes are, the smaller the degree values. The smaller the map edges are, the lower the combined scores. (B) Pathways identified by GO and KEGG analyses. Only the top 15 GO enrichment pathways (including BP, CC, and MF terms) are listed. PPI, protein-protein interaction; GO, Gene Ontology; KEGG, Kyoto Encyclopedia of Genes and Genomes; BP, biological process; CC, cellular component; MF, molecular function.

GLYATL1 across different cancers was explored by the Oncomine database and confirmed by the UALCAN cancer database. The results of the two databases indicated that consistent with prior studies (9,10), the expression of GLYATL1 was significantly downregulated in HCC, while it was significantly upregulated in prostate adenocarcinoma. Our findings regarding its expression in BRCA were in line with those from a previous study (21). Interestingly, compared with normal tissues, GLYATL1 expression in kidney cancer tissues, including kidney chromophobe, clear cell renal cell carcinoma and kidney renal papillary cell carcinoma, was lower. We further plotted Kaplan-Meier curves to evaluate the prognostic value of GLYATL1 in 21 cancer types using Kaplan-Meier plotter. The results demonstrated that high GLYATL1 mRNA expression levels were associated with improved OS in many cancers, including clear cell renal cell carcinoma, liver HCC, bladder carcinoma, cervical squamous cell carcinoma, lung adenocarcinoma, ovarian cancer, pancreatic ductal adenocarcinoma, pheochromocytoma, and paraganglioma. Surprisingly, the expression of the GLYATL1 gene was not significantly associated with OS (P>0.05), though it exhibited significantly different expression between prostate adenocarcinoma and adjacent normal samples. Notably,

we found that in HCC, both GLYATL1 expression and its prognostic value (including OS) were statistically significant. Moreover, univariate and multivariate Cox analyses revealed that GLYATL1 could be an independent prognostic factor for patients with HCC. We further conducted a subgroup analysis based on clinicopathologic characteristics. Surprisingly, unlike other prognostic biomarkers, decreased GLYATL1 mRNA expression was associated with poor OS in HCC patients with stages I-II. In the HCC cohort from the TCGA, decreased GLYATL1 mRNA indicated poor survival for patients with an Edmondson grade of 3. An Edmondson grade of 3 indicates that HCC cells are poorly differentiated. A previous study suggested that GLYAT may be repressed in dedifferentiated HCC cells (9). Thus, an Edmondson grade of 3 may lower the expression of GLYATL1, resulting in a poor survival rate. However, the mechanism underlying the association between stages I-II and the prognostic value of GLYATL1 is still unclear. These results suggest that the prognostic value of GLYATL1 might be associated with clinicopathologic characteristics.

DNA methylation is a common and critical epigenetic event that can lead to gene silencing at the level of pretranscription (22). Abnormal DNA methylation in the promoter might cause silencing of tumor suppressor genes



Figure 5 Enrichment plots from gene set enrichment analysis (GSEA). The identification of related signaling pathways associated with low GLYATL1 expression in HCC was conducted by GSEA. (A) Heat map of the top 50 features for each phenotype in single GLYATL1 gene enrichment analysis in the HCC patient cohort of the TCGA; (B) bile acid metabolism; (C) xenobiotic metabolism; (D) fatty acid metabolism; (E) coagulation; (F) peroxisome; (G) adipogenesis.

Table 3 Enrichment results of GSEA in HCC with low GLYATL1 exp	oression	phenotype
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Name	NES	NOM P value/ FDR q-value
HALLMARK_BILE_ACID_METABOLISM	-2.36	0
HALLMARK_XENOBIOTIC_METABOLISM	-2.31	0
HALLMARK_FATTY_ACID_METABOLISM	-1.97	0.5
HALLMARK_COAGULATION	-1.92	0.33
HALLMARK_PEROXISOME	-1.86	0.4
HALLMARK_ADIPOGENESIS	-1.61	0.43
HALLMARK_ADIPOGENESIS	-1.61	0.43

NES, normalized enrichment score; NOM P value, normalized P value; FDR q-value, false discovery rate q-value.



Figure 6 Correlations between GLYATL1 expression and genes involved in glutamine metabolism in the TIMER database. SLC1A5, solute carrier family 1, member 5; SLC7A11, solute carrier family 7, member 11; GLS, glutaminase; GLS2, glutaminase 2; SLC38A5, solute carrier family 38, member 5; GLUD1, glutamate dehydrogenase 1; GOT1, glutamic-oxaloacetic transaminase 1; GOT2, glutamic-oxaloacetic transaminase 2.

or aberrant activation of oncogenes. As a result, abnormal DNA methylation is considered a hallmark of human tumors (23). We also explored the GLYATL1 promoter methylation profile in HCC patients. The results suggested that the level of GLYATL1 promoter methylation was significantly increased in HCC tissues compared with that in normal tissues. Similar results were also found in the

subgroup analysis based on clinicopathologic characteristics, including individual cancer stage, tumor grade, and nodal metastasis status. Therefore, we postulate that high GLYATL1 promoter methylation might suppress the expression level of GLYATL1, leading to the development of HCC.

STRING was used to construct the PPI network

 Table 4 Correlation analysis between GLYATL1 and genes involved in glutamine metabolism in the GEPIA database

	Hepatocellular carcinoma					
Gene	Tumo	or	Normal			
	R	Р	R	Р		
SLC1A5	-0.48	***	-0.043	0.59		
SLC7A11	-0.27	***	-0.26	**		
GLS	-0.26	***	-0.25	**		
GLS2	0.45	***	0.42	***		
SLC38A5	-0.15	**	-0.07	0.63		
GLUD1	0.15	**	0.75	***		
GOT1	0.26	***	-0.0082	0.92		
GOT2	0.53	***	0.7	***		

, P<0.01; *, P<0.001. SLC1A5, solute carrier family 1, member 5; SLC7A11, solute carrier family 7, member 11; GLS, glutaminase; GLS2, glutaminase 2; SLC38A5, solute carrier family 38, member 5; GLUD1, glutamate dehydrogenase 1; GOT1, glutamic-oxaloacetic transaminase 1; GOT2, glutamicoxaloacetic transaminase 2.

based on the coexpression of GLYATL1 with CNGA3, GNB5, KAT2A, KAT2B, OPN4, PAOX, RGS9, RGS9BP, SAT1, and SAT2. A mutation in CNGA3 can lead to achromatopsia (24). Most investigations on the role of GNB5 have concentrated on neuronal signaling (25). OPN4 is a G protein-coupled receptor. In the latest study, scientists found that targeting OPN4 could suppress PKC/RAF/ MEK/ERK signaling and cell growth and lead to apoptosis in lung cancer cells (26). KAT2A and KAT2B participate in lysine acetylation, a common posttranslational modification that regulates multiple BPs (27). Moreover, accumulating evidence has revealed that the abnormal expression of KAT2A and KAT2B occurs in various cancers, including colon cancer and HCC, and contributes to resistance to cancer therapeutics (28,29). A previous study indicated that RGS9 was downregulated in lung adenocarcinoma and that there was an indirect association between RGS9, Kras and p53 (30). RGS9BP, the binding protein of RGS9, plays a role in bladder cancer (31). SAT1 and SAT2 are members of SAT, a regulator of polyamine catabolism that is involved in the inflammatory response and associated with the growth of several cancers, such as non-small cell lung cancer (32,33). Recently, Ou et al. demonstrated that p53 regulates the reactive oxygen species stress response and ferroptosis via the activation of SAT1 (34). These results indicate the roles

of these co-expressed genes in the development of various cancers, but the synergistic pathways of GLYATL1 and its co-expressed genes in cancers remain elusive. Thus, we conducted GO and KEGG pathway analyses to explore the potential mechanism. The results indicated that these genes are enriched mainly in amine substance metabolic processes. However, the detailed mechanism by which GLYATL1, along with its co-expressed genes, engages in the process of oncogenesis via amine substance metabolic processes is still unknown.

Another important aspect of this study is that the expression of GLYATL1 in HCC was associated with xenobiotic metabolism. By analyzing the mRNA level of GLYATL1 from the TCGA HCC cohort using GSEA, we found that low GLYATL1 expression was predominantly negatively enriched in xenobiotic metabolism and other metabolism pathways, including bile acid metabolism, fatty acid metabolism, coagulation, peroxisome and adipogenesis. Since GLYAT plays a role in the conjugation of carboxylic acids to glycine (6), which is a key metabolite in the rapid proliferation of tumor cells, inhibiting glycine uptake or biosynthesis may impair the growth of cancer cells by slowing the synthesis of nucleic acids (35). GLYATL1 is an acyltransferase that is involved in xenobiotic metabolism in the liver. It can transfer an acyl group to the N-terminus of glutamine and produce N²-acyl-L-glutamine, which is excreted in the urine (9). Thus, we should understand the role of glutamine in material metabolism, especially xenobiotic metabolism and mitochondrial glutamine metabolism, if we want to determine the association between GLYATL1 and mitochondrial glutamine metabolism in HCC. Glutamine, the most abundant circulating amino acid in human plasma, is essential for mitochondrial metabolism, supporting the rapid proliferation of tumor cells (36-38). Previous studies have shown that inhibiting mitochondrial glutamine metabolism could result in a strong cancer-fighting effect (39), such as apoptosis (40). Under certain conditions, the more glutamine GLYATL1 conjugates in xenobiotic metabolism, the less efficient glutamine is for mitochondrial glutamine metabolism. Thus, we conjecture that GLYATL1 catalyzes glutamine and affects mitochondrial glutamine metabolism, subsequently suppressing the inexorable growth of HCC cells. This ratiocination helps us illuminate the reason why GLYATL1 expression in HCC tissues was downregulated and indicated a poor survival rate of HCC patients.

We must acknowledge potential limitations to our analysis. First, we analyzed the impact of immune cell

infiltration, but the number of related variables was limited, and we did not use our HCC samples to verify the expression and prognostic value of GLYATL1. Second, although we found that the prognostic value of GLYATL1 might be associated with clinicopathologic characteristics, it was not corrected by multiple hypothesis testing. Third, we did not compare the predictive power of GLYATL1 with other classifications. Finally, the association between the expression of GLYATL1 and xenobiotic metabolism in HCC was explored by GSEA and confirmed with the TIMER and GEPIA databases. Our hypothesis should be further verified by *in vitro* and animal experiments.

In summary, we performed a comprehensive analysis of the role of GLYATL1 in different cancers. In particular, low GLYATL1 expression is associated with a poor prognosis and is an independent prognostic factor for patients with HCC. Moreover, GLYATL1 is associated with glutamine in xenobiotic metabolism in HCC. Therefore, GLYATL1 can be regarded as a potential prognostic indicator for HCC patients.

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Footnote

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interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. In our study, the statement of ethics approval and the informed consent were not required because all the liver sample information was downloaded from public databases, like The Cancer Genome Atlas (TCGA) and Gene Expression Omnibus (GEO). The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013).

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Supplementary

Dataset	Cancer type	Cut-point	Ν	Cox P value	HR (95% CI)
GSE17537	Colorectal cancer	0.109	55	0.001	0.01 (0.00–0.15)
GSE1456-GPL97	Breast cancer	0.629	159	0.004	1.73 (1.19–2.52)
GSE17536	Colorectal cancer	0.345	177	0.016	0.47 (0.25–0.87)
GSE17536	Colorectal cancer	0.429	177	0.023	0.26 (0.08–0.83)
GSE31210	Lung adenocarcinoma	0.152	204	0.034	0.56 (0.32–0.96)
GSE19234	Skin melanoma	0.737	38	0.036	2.51 (1.06–5.94)
E-DKFZ-1	Renal cell carcinoma	0.475	59	0.040	0.20 (0.04–0.93)

Table S1 Significant associations between GLYATL1 expression and overall survival across different cancers in PrognoScan

Table S2 Survival differences based on immune cell infiltration in patients with liver cancer from the GSE116174 dataset

Parameters	Hazard ratio	95% CI	P value
CD8A (high/low)	1.298	0.607–2.773	0.501
CD19 (high/low)	1.414	0.655–3.049	0.377
IRF5 (high/low)	0.642	0.298-1.384	0.258
CD163 (high/low)	1.541	0.715–3.323	0.27
NRP1 (high/low)	1.341	0.627-2.865	0.449
ITGAM (high/low)	1.318	0.535-2.421	0.78

Markers include CD8A (CD8+ T cells), CD19 (B cells), IRF5 (M1 macrophages), CD163 (M2 macrophages), NRP1 (dendritic cells), and ITGAM (neutrophils). IRF5, interferon regulatory factor 5; NRP1, neuropilin 1; ITGAM, integrin, alpha M.



Figure S1 mRNA level of GLYATL1 in different tumor types from the TCGA and GTEx project in the GEPIA cancer database. The red boxes represent cancer tissues, and the gray boxes represent normal tissues. *, P<0.05. BRCA, breast invasive carcinoma; CHOL, cholangiocarcinoma; COAD, colon adenocarcinoma; KICH, kidney chromophobe carcinoma; KIRP, kidney renal papillary cell carcinoma; LIHC, liver hepatocellular carcinoma; PRAD, prostate adenocarcinoma; READ, rectal adenocarcinoma; SARC, sarcoma.



Figure S2 Kaplan-Meier survival curves comparing high and low GLYATL1 mRNA expression across 21 cancers in the TCGA database. (A) Bladder carcinoma; (B) breast cancer; (C) cervical squamous cell carcinoma; (D) esophageal adenocarcinoma; (E) esophageal squamous cell carcinoma; (F) head-neck squamous cell carcinoma; (G) clear cell renal cell carcinoma; (H) kidney renal papillary cell carcinoma; (I) lung adenocarcinoma; (J) lung squamous cell carcinoma; (K) ovarian cancer; (L) pancreatic ductal adenocarcinoma; (M) pheochromocytoma and paraganglioma; (N) rectum adenocarcinoma; (O) sarcoma; (P) stomach adenocarcinoma; (Q) testicular germ cell tumor; (R) thymoma; (S) Thyroid carcinoma; (T) uterine endometrial carcinoma.



Figure S3 The correlations among immune cell infiltration, GLYATL1 expression and cumulative survival. (A) Correlations between GLYATL1 expression and six immune cell types: B cells, CD4+ T cells, CD8+ T cells, neutrophils, macrophages and dendritic cells. (B) Kaplan-Meier plots comparing high and low immune infiltrates in liver cancer tissues from the TCGA database.