A case of sinobronchial syndrome progressing to diffuse panbronchiolitis despite low-dose, long-term macrolide therapy

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Introduction

Sinobronchial syndrome (SBS) is a disease involving chronic upper and lower airway inflammation caused by a decline in bronchial mucociliary transport or mucosal immunity. This article describes a case of refractory SBS that progressed despite macrolide therapy, and discusses the possible causes of the intractable nature of the disease.

Case presentation

A 73-year-old man complained of productive cough and visited the authors' hospital. Although the patient was healthy in childhood, he had a history of operations for sinusitis in his sixth decade of life. He had been smoking one pack per day till the age of 30 years, after which he stopped smoking. At the initial visit, no rales were heard on auscultation and chest radiography revealed no abnormalities (Figure 1A). However, sinus radiography revealed a decrease in permeability of the maxillary sinus; consequently, he was treated with a low-dose macrolide [clarithromycin (200 mg/day)]. Nevertheless, his productive cough persisted. He also developed effort dyspnea and rhonchi were observed on auscultation. One year after treatment initiation, chest radiography revealed fine nodular shadows in the bilateral lower lung fields. Chest computed tomography (CT) revealed diffuse small nodules in the bilateral lower lobes (Figure 1B). His lung function had deteriorated since his first visit and demonstrated mixed ventilator impairment (%vital capacity =78.6%, forced expiratory volume% =65.8%). A blood examination revealed

signs of minor inflammation (white blood cells, 8,530/µL and neutrophils 81%; C-reactive protein level, 1.08 mg/dL). Cold agglutinin levels were in the normal range. A blood gas analysis revealed hypoxemia (PaO₂, 63.4 Torr). The disease progressed and developed into diffuse panbronchiolitis (DPB) despite treatment with low-dose macrolide. After obtaining informed written consent, a bronchoscopy was performed for work up. Bronchoalveolar lavage revealed an increased neutrophil ratio in the bronchial segment. P. aeruginosa was isolated from a bronchoalveolar lavage fluid culture. Bronchial epithelial specimens were obtained and the ultrastructure of the bronchial epithelial cilia was observed using transmission electron microscopy (Figure 2). The patient's inner dynein arms were defective on the large part of cilia; in contrast, the outer dynein arms were intact. The nasal clearance time according to saccharin test was prolonged [46 min (normal range ≤ 15 min)]. It was speculated that ciliary dysfunction could explain treatment resistance. Clarithromycin was replaced with another macrolide, erythromycin, and an inhaled bronchodilator (long-acting agonist and long-acting muscarinic antagonist combination drug) was added. Although the disease course did not improve, it did stabilize.

Discussion

SBS is a comorbid disease accompanied by chronic upper and lower airway inflammation (1). SBS includes not only mild cases of complicated sinusitis and bronchitis accompanied by productive cough, but also severe diseases



Figure 1 Chest computed tomography at the initial visit (A) revealed no abnormal shadow. Chest computed tomography one year later (B) revealed diffuse small nodular shadows on bilateral lower lobes.



Figure 2 Ultrastructure of the bronchial epithelial cilia viewed on transmission electron microscopy. Outer dynein arms were present (arrows); however, the inner dynein arms were defective on the large part of cilia (scale bar =100 nm).

such as primary ciliary dyskinesia (PCD), DPB, immune globulin deficiency or hypo-immunoglobulinemia, and cystic fibrosis (1,2). This particular case presented only with productive cough and coexisting sinusitis at the initial visit, which became progressively worse despite macrolide treatment, ultimately progressing to DPB. It was probable that inner dynein arm defects led to reduced bronchial clearance and caused refractory disease.

Ciliary activity without inner dynein arms is slightly decreased compared with that of normal cilia, but is increased compared with that of cilia without outer dynein arms, or without both outer and inner dynein arms (3). Cilia with an isolated inner dynein arm defect are observed to have a highly abnormal, stiff forward power stroke with markedly reduced amplitude (4). However, it is acknowledged that inner dynein arms are more difficult to observe due to decreased repeats along the ciliary axoneme compared with the outer dynein arms (5). Some studies (6,7) have emphasized the difficulty of demonstrating the absence of inner dynein arms due to the low contrast of these structures on electron microscopy (8). PCD is a congenital disorder of the respiratory cilia characterized by abnormal motility generally related to ultrastructural ciliary defects responsible for impaired mucociliary transport (8). However, patients with suspected PCD due to an inner dynein arm defect should have the diagnosis reassessed if it has been based on only one diagnostic sample (5). From these perspectives, our patient should undergo re-testing of his ciliary ultrastructure. Despite treatment, however, his symptoms and abnormalities on chest imaging did not improve; it was believed that his bronchial disorder had persisted. Conjecture based on details of this case suggests that a defect in the inner dynein arms caused a minimal disturbance of ciliary chronic bronchial inflammation developing at an older age compared with common PCD cases.

Low-dose, long-term macrolide therapy has been widely used for DPB since the study by Kudoh *et al.* (9), and has generally demonstrated satisfactory efficacy for DPB. Recently, it was reported to be effective against chronic sinusitis (10), cystic fibrosis (11), and non-cystic fibrosis bronchiectasis (12,13). agonists increase ciliary bend amplitude and ciliary beating frequency (14). Muscarinic antagonists have been reported to be effective in reducing bronchial secretions (15,16). In this case, the combination of erythromycin and long-acting agonist/long-acting muscarinic antagonist inhalation did not improve the disease symptoms, but did stop its progression.

Although the term "SBS" is rarely used today, it is

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more useful than "non-cystic fibrosis bronchiectasis" because it includes such conditions before progressing to bronchiectasis. Earlier intervention is possibly more effective than intervention initiated after bronchiectasis has run its course.

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Footnote

Conflicts of Interest: The authors have no conflicts of interest to declare.

Informed Consent: Written informed consent was obtained from the patient for publication of this manuscript and any accompanying images.

References

- Sugiyama Y. Sinobronchial syndrome. Nihon Rinsho 1999;57:2119-22.
- 2. McBride JT, Brooks J. Sinobronchial syndrome. Ear Nose Throat J 1984;63:177-9.
- Noone PG, Leigh MW, Sannuti A, et al. Primary Ciliary Dyskinesia. Am J Respir Crit Care Med 2004;169:459-67.
- Chilvers MA, Rutman A, O'Callaghan C. Ciliary beat pattern is associated with specific ultrastructural defects in primary ciliary dyskinesia. J Allergy Clin Immunol 2003;112:518-24.
- O'Callaghan C, Rutman A, Williams GM, et al. Inner dynein arm defects causing primary ciliary dyskinesia: Repeat testing required. Eur Respir J 2011;38:603-7.
- Chao J, Turner J, Sturgess J. Genetic heterogeneity of dynein-deficiency in cilia from patients with respiratory disease. Am Rev Respir Dis 1982;126:302-5.

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- van der Baan S, Veerman A, Bezemer P, et al. Primary ciliary dyskinesia: quantitative investigation of the ciliary ultrastructure with statistical analysis. Ann Otol Rhinol Laryngol 1987;96:264-72.
- 8. Escudier E, Couprie M, Duriez B, et al. Computer-assisted analysis helps detect inner dynein arm abnormalities. Am J Respir Crit Care Med 2002;166:1257-62.
- Kudoh S, Azuma A, Yamamoto M, et al. Improvement of survival in patients with diffuse panbronchiolitis treated with low-dose erythromycin. Am J Respir Crit Care Med 1998;157:1829-32.
- Shimizu T, Suzaki H. Past, present and future of macrolide therapy for chronic rhinosinusitis in Japan. Auris Nasus Larynx 2016;43:131-6.
- Saiman L, Marshall BC, Mayer-Hamblett N, et al. Azithromycin in patients with cystic fibrosis chronically infected with pseudomonas aeruginosa. JAMA 2003;290:1749-56.
- Altenburg J, de Graaff CS, Stienstra Y, et al. Effect of azithromycin maintenance treatment on infectious exacerbations among patients with non – cystic fibrosis bronchiectasis: the BAT randomized controlled trial. JAMA 2013;309:1251-9.
- Anwar GA, Bourke SC, Afolabi G, et al. Effects of longterm low-dose azithromycin in patients with non-CF bronchiectasis. Respir Med 2008;102:1494-6.
- Komatani-Tamiya N, Daikoku E, Takemura Y, et al. Procaterol-stimulated increases in ciliary bend amplitude and ciliary beat frequency in mouse bronchioles. Cell Physiol Biochem 2012;29:511-22.
- Pera T, Zuidhof A, Valadas J, et al. Tiotropium inhibits pulmonary inflammation and remodelling in a guinea pig model of COPD. Eur Respir J 2011;38:789-96.
- Cortijo J, Mata M, Milara J, et al. Aclidinium inhibits cholinergic and tobacco smoke-induced MUC5AC in human airways. Eur Respir J 2011;37:244-54.