

# First-line treatment selection with organoids of an *EGFR*m + *TP53*m stage IA1 patient with early metastatic recurrence after radical surgery and follow-up

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# Introduction

Stage I lung adenocarcinoma (LADC) has a 5-year overall survival (OS) of 68%, with stage IA1 reaching as high as 91% (1-3). Surgery is the conventional treatment and currently preferred local treatment that can achieve complete relief in early-stage non-small cell lung cancer (NSCLC) (4). However, the relapse of some early-stage NSCLC has brought attention to the post-operative management of resectable tumors (5). According to phase III trials, adjuvant chemotherapy after R0 resection may achieve an improvement of 5% in 5-year survival, but the result is limited among patients with NSCLC staged higher than IB. In stage IA patients, the use of adjuvant chemotherapy can be even detrimental (6). Moreover, the current clinical practice guideline does not recommend adjuvant chemotherapy in stage IA patients after R0 resection (7). With a low genetic testing rate in earlystage LADC, no evidence from phase III clinical trials has demonstrated that epidermal growth factor receptormutated (*EGFR*m) stage IA patients can benefit from adjuvant *EGFR*-tyrosine kinase inhibitors (TKIs).

Without adjuvant therapy, 5–10% patients will suffer from early recurrence 2-3 years after their curative surgery (8). It remains unclear how to identify the high-risk cohort in stage IA patients. The evaluation on the prognosis of stage IA patients is mainly based on pathological findings: the size of invasive component, lymphatic or vascular invasion, or visceral pleural invasion (9,10). Because genetic testing is mainly performed in advanced stage NSCLC (11,12), the prognostic and predictive values of genomic profiling has not been widely recognized in early-stage NSCLC (13,14). However, it has been reported that advanced NSCLC patients harboring co-mutation of EGFR and TP53 have a poorer prognosis (15-17). Relapse in resected NSCLC patients is often reported as local or regional recurrence and metastasis. For these advanced stage recurrent patients, there still lacks high-level evidence as to whether these patients should be treated in the same way as advanced stage patients for the first-line treatment, or if they should

be treated with a more aggressive regiment based on their formal relapsed history. It is also unclear whether these patients can benefit from first-line osimertinib in case of activating *EGFR* mutation. Yet, according to a multicenter phase III trial of osimertinib used as first-line treatment in *EGFR*m advanced stage NSCLC (FLAURA), though osimertinib established a substantial improvement in median progression-free survival (PFS) and median OS, which converged down toward the comparing arm of firstgeneration TKI (18). Based on the current evidence, a preclinical model could be applied in treatment selection in personalized precision therapy.

Patient-derived organoids (PDO) are 3D tumor cultures that are grown from tissue-specific stem cells derived from the primary tumor (19-21). They are cultured in a 3D environment with essential growth factors, collagen, and other extracellular matrix (ECM) components structured by Matrigel. PDO can recapitulate the histological, structural (20), and genetic characteristics (21-23) of the primary tumor. As a preclinical tumor model, PDO has higher similarity than cell line cultures and saves both time and cost compared to a patient-derived xenograft (24). Organoid drug sensitivity assay uses IC<sub>50</sub> as an indicator in individual drug screening for a primary tumor. Randomized clinical trials of colon-rectal cancer, breast cancer, and NSCLC are ongoing in Europe to evaluate the recapitulation of primary tumor drug sensitivity (TUMOROID, NL49002.031.14) and the efficacy of last-line drug selection (SENSOR, NL50400.031.14).

Herein, we report a case of a 66-year-old Chinese male who is a former smoker (40 pack-years) with a case of early metastatic recurrence of stage IA1 LADC harboring  $EGFR^{L858R}$ and  $TP53^{R110L}$  after R<sub>0</sub> resection and the response to firstline therapy osimertinib. An international multidisciplinary team (iMDT) discussion is presented that discusses the future treatment strategies. We present the following case in accordance with the CARE reporting checklist (available at http://dx.doi.org/10.21037/jtd-20-1882). Written informed consent was obtained from the patient for publication of this manuscript and any accompanying images.

## **Case presentation**

A 66-year-old male was admitted to our hospital for suspicious metastatic recurrence in his post-operative physical examination with no clinical manifestation. He received video-assisted thoracoscopic surgery (VATS) left upper lobectomy and associated hilar and mediastinal

lymph nodes dissections in other hospital 3 years ago. His primary tumor was pathologically diagnosed as LADC with immunohistochemistry (IHC) showing CK7(+), TTF(+), Napsin A(-), staged as  $T_{1a}N_0M_0$  (stage IA1), with a microscopically negative margin, no invasion of visceral pleura, lymphatic or venous vessels was observed. The diagnosis of metastatic recurrence was based on chest computed tomography (CT), which showed multiple nodules in the right lung (the largest one was sized 1.1 cm  $\times$  0.9 cm) and an enhanced mass with soft tissue density at the left hilum. <sup>18</sup>F-fluorodeoxyglucose positron emission tomography/computed tomography (<sup>18</sup>FDG-PET/CT) scan showed a 1-cm nodule in his right supraclavicular fossa with increased avidity and a maximum standard uptake value (SUV<sub>max</sub>) of 4.2 (Figure 1). The routine laboratory test [including carcinoembryonic antigen (CEA), CYFRA21-1, CA125, alpha-fetoprotein (AFP), CA19-9, CA15-3, CA72-4, total prostate-specific antigen (tPSA), free PSA (fPSA), squamous cell carcinoma antigen (SCC-Ag)] results showed elevations in multiple tumor markers, including tPSA, fPSA, SCC-Ag, and CYFRA21-1. Pelvic CT ruled out the suspicion of prostate cancer or metastasis. The patient presented with no complaint of discomfort or disability, and was subsequently assessed as ECOG-PS 0. The final pathological diagnosis was lymph node metastatic adenocarcinoma by surgical removal of the right supraclavicular lymph nodes (Figure 2). IHC staining was not repeated.

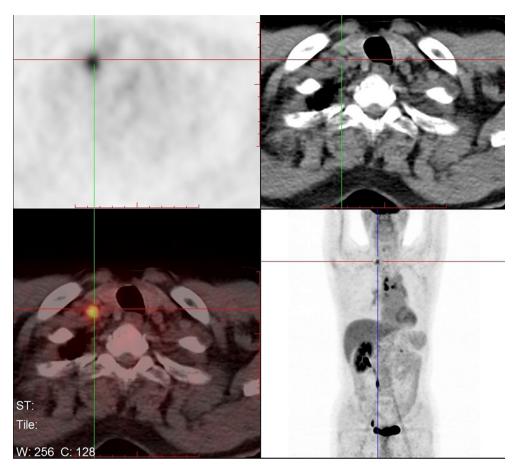
Molecular diagnosis of the patient's formalin-fixed paraffin-embedded (FFPE) tissue specimens based on next-generation sequencing (NGS) showed  $EGFR^{L858R}$  +  $TP53^{R110L}$  mutations in his primary tumor, and  $EGFR^{L858R}$  +  $TP53^{R110L}$  +  $CDKN2A^{H83Y}$  in the lymph nodes. Circulating tumor DNA (ctDNA) of the plasma was also tested, but no mutation was detected.

PDO cultures were successfully established using culture methods described in the supplemental materials (*Figure 3A*). Efficacy of *EGFR*-TKI was demonstrated by carrying out cell viability and proliferation assays using the established organoid cultures. Cell viability curves of organoids treated with gefitinib, erlotinib, afatinib, icotinib, and osimertinib are shown in *Figure 3B*. The results showed that the IC50 of osimertinib was the lowest among the *EGFR*-TKIs that were tested.

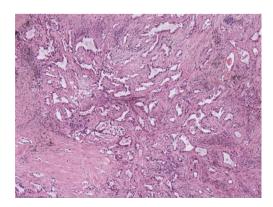
## Treatment

Based on the evidence listed above, the recurrent metastatic

Jia et al. Managing early recurrence in EGFR-positive NSCLC



**Figure 1** <sup>18</sup>FDG-PET/CT scan showing a 1-cm nodule in the right supraclavicular fossa with increased avidity of SUV<sub>max</sub> =4.2. <sup>18</sup>FDG-PET/CT, <sup>18</sup>F-fluorodeoxyglucose positron emission tomography/computed tomography; SUV<sub>max</sub>, maximum standard uptake value.



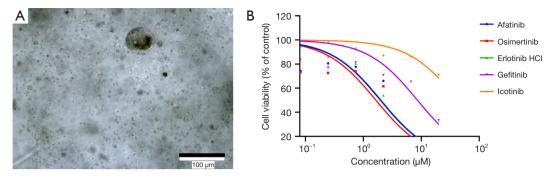
**Figure 2** Pathological diagnosis of right supraclavicular lymph nodes as lymph node metastatic adenocarcinoma. HE staining, original magnification ×40.

lymph node adenocarcinoma of this patient harbored both  $EGFR^{L858R}$  and  $TP53^{R110L}$ , and a novel lymph node specific mutation-l  $CDKN24^{H83Y}$ . Additionally, based on the results of the drug response curves of the 5 EGFR-targeted TKIs, osimertinib was suggested to be the most efficacious drug. Thus, the patient began receiving oral osimertinib, 80 mg, once daily and achieved a PFS of 9 months (*Figure 4*).

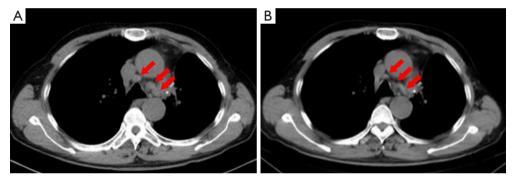
## **iMDT** discussion

#### Department of Oncology

In this case, osimertinib was chosen as first-line therapy based on the NGS result of the primary tumor and the



**Figure 3** Drug responses of patient-derived lymph node metastatic adenocarcinoma organoids. (A) Organoid culture of lymph node metastatic adenocarcinoma; (B) dose-response curves of patient-derived lymph node metastatic adenocarcinoma organoids treated with *EGFR*-TKI afatinib, osimertinib, erlotinib HCl, gefitinib, and icotinib. Cell viability was measured by an ATP-based assay after 4 days of drug incubation. TKI, tyrosine kinase inhibitor.



**Figure 4** Effect of osimertinib in this patient with multiple mediastinal lymph node metastasis. Computed tomography scan of the chest before (A) and after (B) treatment with osimertinib. Shrinkage of multiple mediastinal lymph nodes (arrows) was observed.

metachronous second primary tumor (MST). According to FLAURA (18), osimertinib used as first-line TKI in EGFRm NSCLC can achieve a median PFS of 18.9 months (8.7 months longer than standard EGFR-TKI) and a median OS of 38.6 months (6.8 months longer than standard EGFR-TKI). Although the OS converged at 48 months, nearly crossing-over with the standard EGFR-TKI, osimertinib still demonstrated a better prognosis for advanced NSCLC patients. However, FLAURA did not provide information regarding concomitant mutation, and whether patients with EGFR + TP53 would benefit more from osimertinib still lacks evidence. Thus, we performed organoid culture to examine the sensitivity of multiple EGFR-TKIs. Data analysis showed that osimertinib has the highest sensitivity, followed by first-generation EGFR-TKIs, and second-generation EGFR-TKIs show the lowest sensitivity. In conclusion, it is reasonable to use osimertinib as first-line treatment in this patient. However, extending the application of organoid drug-sensitive tests to all clinical circumstances requires randomized clinical trial validation.

# Department of Thoracic Surgery

Early recurrence in a stage IA LADC patient is a rare condition characterized by newly reported lung nodules or metastases in lymph nodes or other organs with the same genetic characteristics. A recurrence rate of 12.5–12.6% was reported in both retrospective and prospective studies (10,25). Among those who had a recurrent disease, median time to recurrence was 15 months (25). Median disease-free survival (DFS) was 47.2 months, and 2-year DFS rate was 93% (10). Due to its low recurrence rate and the treatment side effects, some surgeons insist on treat-when-recur, but others support the early application of adjuvant therapy due to the poor prognosis of these stage

IA patients after recurrence. Therefore, it is crucial to identify those patients with a high recurrence risk and to apply the most suitable adjuvant therapy as early as possible (10). Besides pathological features and serum tumor biomarkers (9), early genetic testing may aid earlystage LADC risk evaluation. Small panels covering classic driver genes tend to be cost-effective and an increasing trend of applying small panel in early-stage LADC has been observed. However, the predictive value of driver mutations still needs investigation (13,14). The coexistence of TP53 has been discussed as a poor prognostic predictor in LADCs harboring EGFR mutations and might be one of the factors that contributes to the early recurrence of the tumor (15-17). Despite this, TP53 is not included in the National Comprehensive Cancer Network (NCCN) clinical practice guideline of recommended molecular biomarkers. Thus, a panel of NGS including driver genes and TP53 might be needed in the predictive evaluation of early-stage NSCLC.

The application of adjuvant therapy in the overall management is a complement to curative resection, but the value of adjuvant therapy in early-stage NSCLC has not been widely recognized. For stage I patients with high-risk pathological features of recurrence, adjuvant chemotherapy can be beneficial and monotherapy is enough for these patients (10). Very few phase III clinical trials of adjuvant chemotherapy have included stage IA NSCLC. This may result from patients' fear toward adverse events brought by chemotherapy and clinical ethical challenges, as these subgroups of NSCLC patients show a 5-year OS of greater than 90% (1). To improve the risk-benefit balance, more attention has been drawn to target adjuvant therapy. However, it is still controversial whether adjuvant therapy should be applied to stage IA patients. In EGFRpositive NSCLC patients, the single-arm phase II trial of post-operative adjuvant erlotinib (SELECT) showed that 2-year DFS was improved compared to historical data. SELECT included 13 patients with stage IA disease and 32 with stage IB disease. The stage I patients achieved a 2-year DFS greater than 96%, but DFS converged after 3 years (26). None of the recent phase III randomized clinical trials on EGFR-TKI have included stage IA patients (27-30). Whether TKI adjuvant therapy would lead to resistance or early recurrence is still being validated through preclinical and real-world data.

The selection of the treatment regimen for this patient included 2 complementary strategies, the first was genedrug correlation, and the second was black box testing based on a 3D *in vitro* model of PDO. Due to the existence of *EGFR*<sup>L858R</sup>, the patient was responsive to *EGFR*-TKIs. The PDO strategy helped to evaluate which *EGFR*-TKI could exert better efficacy.

## Three questions for further discussion

Question 1: As indicated by the above, LADC that harbors EGFR + TP53 mutations has worse prognosis than single-EGFR-mutated LADC. However, it still remains controversial to recommend adjuvant therapy for stage I patients, especially in those whose lesions are smaller than 4 cm in diameter. Ultimately, the question is if adjuvant therapy should be used in stage I LADC patients with high risk molecular diagnosis, and which of EGFR-TKI or chemotherapy should be chosen as adjuvant therapy based on their durability and toxicity

## Expert opinion 1: Dr. Daniel C. Christoph

In case of proven pleural invasion and/or lymphatic or venous invasion adjuvant treatment should be discussed with the patient and the results of a large Japanese retrospective multicenter study presented at the last ASCO annual meeting should be explained. If the patient has no serious comorbidities (Charlson comorbidity index), I recommend adjuvant chemotherapy within 2 months after the initial resection, because targeted agents have not yet proven any OS benefit in the adjuvant setting in phase 3 prospective clinical trials. But if the postoperatively tested serum levels of tumors markers were within the regular range and no pathological signs of invasion of the primary tumor could be demonstrated, I would perform surveillance only.

## Expert opinion 2: Dr. Antonio Passaro

The role of adjuvant chemotherapy in NSCLC is established for resected disease with nodal involvement (either N1 or N2 levels), thought the achieved benefit is limited, accounting of about 4–5% in absolute increase in 5 years of survival. Cisplatin-based chemotherapy is worldwide approved as the standard adjuvant therapy, with the combination with cisplatin plus vinorelbine, that is the recommended standard worldwide.

For patients with N0, adjuvant treatment might be considered, with a lower level of evidence, for tumors larger than 4 cm in diameter.

At the present time, 8 clinical trials evaluated the role of *EGFR*-TKI in adjuvant setting, including patients

with resected stage IA–IIIA harbouring *EGFR* mutations, without OS benefit (31).

#### Expert opinion 3: Dr. Cesare Gridelli

In my opinion there is no indication to adjuvant chemotherapy or however to any adjuvant treatment for patients with stage I and T less than 4 cm. In particular, in lung adjuvant cisplatin evaluation (LACE) metaanalysis the use of adjuvant chemotherapy in stage I was even detrimental with worse survival as compared to the observational control arm. The role of adjuvant chemotherapy for stage I with T more or equal 4 cm is controversial and optional.

In case of  $EGFR^+$  patients, even with concurrent TP53positive mutation, adjuvant targeted therapy is not yet recommended because we are waiting for the results of ongoing phase III randomized trials with the best hope for the osimertinib study.

#### Expert opinion 4: Dr. Tomoyuki Hishida

My current answer is no. As the authors mention, no positive results about the survival benefit of platinum-based adjuvant chemotherapy have been demonstrated for p-stage IA disease. Although previous CALGB 9633 and JBR-10 trials indicated potential survival benefit in patients with p-stage IB tumors that are 4 cm or larger (p-stage IIA in the present TNM 8<sup>th</sup> edition), in a subset of analyses, no OS benefit was shown for patients with p-stage IB tumors less than 4 cm, and p-IA disease patients were excluded from both trials. The LACE meta-analysis showed adverse effects of chemotherapy for the p-stage IA group with a hazard ratio (HR) of 1.40 (95% CI: 0.95-2.06) (6). Adjuvant trials using EGFR-TKIs targeted more advanced p-stage IB, II, and IIIA, but did not demonstrate any OS benefit in these populations (27,32). Tegafur-uracil (UFT) has shown mild survival benefits for p-stage IA NSCLC (>2 cm) with a HR of 0.62 (95% CI: 0.42-0.90) in a meta-analysis of 6 Japanese trials (33). However, I think the benefits of UFT are very limited for high-risk patients. We feel tempted to use adjuvant therapy for p-stage IA patients with high-risk biological signatures; however, there is still no rationale for administering adjuvant therapy. Further investigations are needed to identify reasonable biomarkers to select good candidates for adjuvant treatment that will contribute towards prolonging patient survival.

Question 2: Considering the toxicity of adjuvant chemotherapy and economic burden of adjuvant target therapy, how should early-stage LADC patients be identified who have a high risk of relapse for administration of adjuvant therapy with a target? Also, is it rational to use NGS to test the genetic mutation in early-stage NSCLC?

## Expert opinion 1: Dr. Daniel C. Christoph

For the identification of these patients clinical/pathological factors of the patients need to be known: smoking status (EGFR-diagnostics should be performed in never smokers and light smokers) and proof of signs of invasion *resp.* aggressiveness of the primary tumor (please see above). There is no rationale to use NGS for every patient, but for patients with a higher suspicion of suffering from an *EGFR*-mutated lung cancer I recommend mutational diagnostics.

## Expert opinion 2: Dr. Antonio Passaro

The identification of high-risk factors (e.g., visceral pleural invasion, as lymphatic permeation or vascular invasion), for resected NSCLC, should be highly useful for a better patient's selection to improve survival outcome (34).

The use of NGS in early stages it is highly debated, considering cost and international guidelines, that confirmed no target agents is approved in early stage. Nowadays, the role *EGFR* mutations in adjuvant setting it is under evaluation and the survival results of the phase III randomized trial, ADAURA [NCT02511106], will be released very soon.

In this trial, patients with resected NSCLC in stage IB, II and IIIA harboring common *EGFR* mutations, received osimertinib or placebo for a treatment duration of up to three years, with the primary point of DFS. Based on the preliminary data, not yet presented, the trial results will be unblinded early following a recommendation from an Independent Data Monitoring Committee (IDMC) based on its determination of overwhelming efficacy.

# Expert opinion 3: Dr. Cesare Gridelli

To date is no evidence-based to do in clinical practice NGS for identify early-stage high risk patients. First, because we have no evidence that the presence of any tumor genetic alteration can drive adjuvant therapy. Second, the presence of any mutation or gene fusion or however genetic alteration can drive an adjuvant targeted therapy in clinical practice.

## Expert opinion 4: Dr. Tomoyuki Hishida

I think the use of NGS is useful for detecting potential druggable driver gene abnormalities for targeted therapy. However, targeted therapy is usually cytostatic and not cytotoxic. Previous adjuvant *EGFR*-TKI trials have shown prolonged recurrence-free survival, but no OS benefit in patients with p-stage IB–IIIA NSCLC with activated *EGFR* mutations (27,32). In my opinion, it is not too late to use targeted therapy, even if we start to use it at the time of recurrence. It remains unknown whether early use of targeted therapy as an adjuvant treatment is beneficial for prolonging patient survival and enhancing a rate of cure.

Question 3: In our case, organoids were cultured as a preclinical model for drug-sensitivity screening tests, the results of which met our expectations and facilitated the selection of treatment. However, more evidence needs to be collected from large-scale clinical trials focusing on the concordance of organoid drug sensitivity and clinical response. Despite the current evidence summarized in the introduction, it still remains unclear if organoids will play a more important role in future personalized treatment selection

# Expert opinion 1: Dr. Daniel C. Christoph

There might be a role of organoids if the primary tumor shows concomitant genetic mutations with different treatment options. During the period of postoperative convalescence, drug sensitivity might be tested and the results might guide the selection of the most effective systemic treatment.

## Expert opinion 2: Dr. Antonio Passaro

The role of preclinical model to test drug-sensitivity is highly potential, especially in the era of precision medicine. Lung cancer showed high genetic and phenotypic heterogeneity. Organoid evaluation should be useful and interesting to improve our knowledge on biological alteration (genomic and genetic) in early setting of lung cancer.

The next step of precision cancer medicine it will be to predict patient-specific drug responses through in vitro patient-specific drug trials.

# Expert opinion 3: Dr. Cesare Gridelli

Drug-sensitivity tests should be prospectively validated and

object of future phase III randomized trials. To date there is no recommendation even for the chemotherapeutic drugsensitivity test already tested in the past. In conclusion, to date there is no role for any kind of drug-sensitivity test in clinical practice.

# Expert opinion 4: Dr. Tomoyuki Hishida

It is an exciting avenue to utilize organoids as a personalized in vitro model to predict response to anticancer therapy. In this case, osimertinib was identified as the treatment of choice based on the results from organoids. However, osimertinib has been known to be a key drug and the 1stline therapy for LADCs with EGFR mutations (Ex19 del or L858R) (35), in spite of the lacking data for LADCs with EGFR + TP53 co-mutations, and whether it is a good candidate, can be decided without the use of an organoid model. In my opinion, the clinical benefit of organoids may be limited for LADC with driver gene abnormalities and the corresponding targeted therapies. Organoid models may pose potential benefits for LADC without driver gene abnormalities for predicting response toward conventional cytotoxic regimens and immune checkpoint inhibitors. Further studies are required to identify good targets of organoid models for the treatment of advanced NSCLC.

# Conclusions

For stage I NSCLC patients with nodules less than 4 cm, no current evidence supports the use of adjuvant therapy nor genetic testing is recommended. However, organoid, an *in vitro* personalized pre-clinical model, is highly potential for drug sensitive test in precision medicine. Further prospective investigations and clinical validations are needed to establish the feasibility of organoids.

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## Jia et al. Managing early recurrence in EGFR-positive NSCLC

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# 3772

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## Methods

#### Patient-derived tumor organoid culture

The primary organoids were established using previously described culture methods with modification (1). Briefly, solid lung cancer tissue was rinsed by 10 mL HBSS containing antibiotics, then minced by sterilized surgical scissors, and digested in 5 mL of 5 mg/mL collagenase type II (Invitrogen) in DMEM/F12 for about 4 h at 37 °C with gentle shaking and intermittent pipetting. The digested tissue suspension was strained over a 70 µm filter. The strained suspension was centrifuged at 300 rcf, and red blood cells were lysed using lysis buffer (00443357, Invitrogen eBioscience) for 5 min. Dissociated cells were washed, counted, and resuspended in Matrigel basement membrane Matrix (Corning, 356235). Next, 30 µL drops of Matrigel-cell suspension were allowed to solidify on pre-warmed 60mm culture plates in a 37 °C and 5% CO<sub>2</sub> cell culture incubator for 30 min. Organoids were cultured in Advanced DMEM/F12 (Thermo Fisher Scientific), supplemented with the additives listed in *Table S1*. Upon completed gelation, culture medium was added to each plate. Medium was refreshed every 4 days (21).

#### Organoid drug response assay

Organoids were harvested using  $1 \times$  TrypLe (Gibco) and dissociated to small clusters. Between 100 and 500 clusters were plated in 40-µL 5% Matrigel/organoid culture medium in a 384-well plate (Corning) in triplicate. Clear bottom 384-well plates were coated with 10-µL collagen (Thermo Fisher) prior to plating cells. Then, 48 h after plating, a concentration dilution series of each compound (50, 10, 2, 0.4, 0.08, 0.016 µM) was dispensed using liquid-handling robotics, and cell viability was assayed using CellTiter-Glo (Promega) after 4 d of drug incubation. Results were normalized to vehicle controls. Data analyses were performed using GraphPad Prism 7.0 software, and the values of IC50 were calculated by applying nonlinear regression (curve fit).

Additive	Supplier	Catalogue #	Concentration
N-Acetylcysteine	Sigma	A9165	1.25 mM
EGF	PeproTech	AF-100-15	50 ng/mL
FGF-10	PeproTech	100-26	20 ng/mL
FGF-basic	PeproTech	100-10B	1 ng/mL
Y-27632	Selleck	S1049	10 µM
A-83-01	Sigma	SML0788	500 nM
SB202190	Selleck	S1077	10 µM
Nicotinamide	Sigma	72340	10 mM
PGE2	Tocris	2296	1 µM
Noggin (conditional medium)	In-house	_	50 mL
R-Spondin (conditional medium)	In-house	_	50 mL
HEPES	Sigma	H6147	10 mM
GlutaMAX	Gibco	35050061	1×
B27	Gibco	17504044	1×
N2	Gibco	17502048	1×

Table S1 Organoid culture additives

R-Spondin 1 and Noggin conditioned media were prepared in house from the culture media of Rspo1 cell and the noggin cDNA transfected HEK-293 cell following the methods described before (36,37).

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