

New players in the Bruton's tyrosine kinase vs. ibrutinib match

Stefania Fiorcari, Rossana Maffei, Roberto Marasca

Hematology Unit, Department of Medical and Surgical Sciences, University of Modena and Reggio Emilia, Modena, Italy *Correspondence to:* Prof. Roberto Marasca, MD. Hematology Unit, Department of Medical and Surgical Sciences, University of Modena and Reggio Emilia, Via Del Pozzo 71, 41124, Modena, Italy. Email: roberto.marasca@unimore.it.

Comment on: Bottoni A, Rizzotto L, Lai TH, et al. Targeting BTK through microRNA in chronic lymphocytic leukemia. Blood 2016;128:3101-12.

Submitted Mar 10, 2017. Accepted for publication Mar 24, 2017. doi: 10.21037/tcr.2017.03.75 View this article at: http://dx.doi.org/10.21037/tcr.2017.03.75

The Bruton's tyrosine kinase, also known as BTK, is a member of the Tec family kinases with a well-characterized role in B-cell receptor (BCR) signaling and B-cell activation. BTK is able to transmit and amplify several signals that are involved in the complex crosstalk between tumor cells and microenvironment. These signals include cytokines and growth factors, chemokine receptors, antigen receptors as BCR and integrins (1). Chronic lymphocytic leukemia (CLL) cells show an increased expression and activation of BTK that with a chain reaction modulates signaling pathways essential for CLL-cells survival, as the protein kinase AKT, extracellular signal-regulated kinase (ERK) and nuclear factor kappa light-chain enhancer of activated B cells (NF-KB) pathways (2). In addition, BTK is involved in chemokine-mediated migration, homing and adhesion of CLL cells (3). Given the known importance of BCR signaling in CLL and the central role of BTK in this pathway, in the recent years a new drug called ibrutinib has been used in clinical practice to induce inhibition of this kinase. Ibrutinib is a relatively selective, irreversible inhibitor of BTK binding covalently the Cys-481 in the ATP binding domain of the kinase. Inhibition of BTK in CLL cells leads to an impressive delocalization of leukemic cells from the protective tissue microenvironment to the periphery interfering with pathogenic mechanism of recirculation and homing. Several studies demonstrate the considerable clinical success of ibrutinib showing a good safety profile determining an improvement of the quality of life of CLL patients (4).

However, it has been immediately clear the limited capacity of this agent to induce a complete eradication of neoplastic clone, with the detection of persistent disease in blood and tissues of patients after years of singleagent therapy. Moreover, some patients lose the response and progress during treatment and, although infrequent, fail to respond, developing a significant resistance to treatment (5). These unwanted effects may be related, in part, to a wide spectrum of ibrutinib off-targets since its molecular effect is not completely restricted to the CLL clone but also regulates key functions in other cellular elements as NK cells, T cells, macrophages and osteoclasts (6-8). Moreover, ibrutinib resistance is known to be explained in the majority of cases by the acquired mutation in BTK at its binding site with a cysteine to serine substitution (C481S) and others involving a gainof-function mutation of the phospholipase C γ 2, an important molecule downstream of BTK. The C481S mutation switches the irreversible inhibition of ibrutinib to a reversible binding to BTK leading to an impairment of inhibition of BTK autophosphorylation and downstream signaling (9,10). The possibility to manage the onset of mutant BTK clone combining ibrutinib with other agents is a therapeutic need for CLL patients. In order to dissect the complex and essential role of BTK and ibrutinib in the pathobiology of CLL, Bottoni and colleagues (11) in this issue of Blood, described for the first time the possibility to regulate BTK expression through the modulation of miRNAs by the use of small molecule inhibitors of histone deacetylase (HDAC). miRNAs are small non-coding RNA molecules that are involved in the regulation of gene expression binding target mRNA to silence protein production. For this reason, dysregulation of miRNA expression is involved in tumor initiation and progression (12). The comparison of miRNA profile between normal B cells and malignant CLL lymphocytes shows substantial differences that are linked to initial progression and drug resistance (13). Bottoni et al. with their elegant experiments identified a peculiar signature

of miRNAs expressed in CLL cells that are able to target BTK. Of interest, the putative BTK-targeting miRNAs are expressed at lower levels in CLL cells compared to normal B cells, implying the existence of a regulation mechanism in leukemic cells. Since in CLL it has been shown that overexpression of HDAC and chromatin-modulating enzymes mediates the epigenetic silencing of miRNA-15A, miRNA-16 and miRNA-29d (14), the authors proposed and validated a causal mechanism in which the recruitment of HDAC to the promoters for the BTK-targeting miRNAs is able to silence their expression. On this scenario, the possibility to revert this repression may lead to target BTK indirectly by inhibiting HDAC activity which in turn may overexpress BTK-targeting miRNAs with the consequent downregulation of its target genes. For this reason, the authors decided to use first in an in vitro model and then in vivo, two different HDAC inhibitors abexinostat and panobinostat. As expected, the HDAC inhibitors induced a significant increased expression of BTK-targeting miRNAs and the consequent reduction of BTK expression. Functionally, this result influences the BTK downstream pathways with reduction of pPLC₂, pERK, pAKT and the consequent induction of CLL cells death. At this point, the possibility to combine the reduction of BTK expression by HDAC inhibitors and inhibition of its kinase activity by ibrutinib is appealing. In vitro and in vivo Eu-TCL-1 mouse model, abexinostat and ibrutinib showed an interesting synergistic activity with cytotoxicity in CLL cells and a reduced lymphocytosis. These important data put forward the basis for the possibility to use HDAC inhibitors to target mutated BTK cells since in vitro a reduction of BTK signaling pathway and CLL cell death has seen. In conclusion, the study of Bottoni et al. provides new interesting data on the potential targeting of BTK by using HDAC inhibitors that are able to silence BTK-targeting miRNAs leading to the reduction of BTK expression. Although the possibility to influence BTK expression by modulating an epigenetic silencing is potentially relevant, what is not known is how these HDAC inhibitors, alone or in combination with ibrutinib, could be use in the clinical practice. All HDAC inhibitors evaluated in phase I-II trials for the treatment of CLL patients showed the development of significant unwanted adverse effects limiting the possibility to continue therapy (15,16). A further concern is mainly related to the CLL patients with a mutant BTK, because is not yet completely known if mutations are present pre-treatment or if are acquired during treatment for induction of drug pressure. These open questions are

to be addressed to optimize the use of ibrutinib with other agents, as HDAC inhibitors.

Acknowledgments

Funding: This work was supported by Associazione Italiana per la Ricerca sul Cancro (TRIDEO 16923 to R Maffei; AIRC IG14376 to R Marasca and FIRC/AIRC Triennal Fellowship 16430 to S Fiorcari), Milan, Italy; Ricerca Finalizzata Giovani Ricercatori 2011-2012, Ministero della Salute (GR-2011-02349282 to R Maffei), Rome, Italy.

Footnote

Provenance and Peer Review: This article was commissioned and reviewed by the Section Editor Peipei Xu (Department of Hematology, The Affiliated Drum Tower Hospital of Nanjing University Medical School, Nanjing, China).

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at http://dx.doi. org/10.21037/tcr.2017.03.75). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Open Access Statement: This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: https://creativecommons.org/licenses/by-nc-nd/4.0/.

References

- Ponader S, Chen SS, Buggy JJ, et al. The Bruton tyrosine kinase inhibitor PCI-32765 thwarts chronic lymphocytic leukemia cell survival and tissue homing in vitro and in vivo. Blood 2012;119:1182-9.
- 2. Herman SE, Gordon AL, Hertlein E, et al. Bruton tyrosine kinase represents a promising therapeutic target for treatment of chronic lymphocytic leukemia

Translational Cancer Research, Vol 6, Suppl 3 May 2017

and is effectively targeted by PCI-32765. Blood 2011;117:6287-96.

- Herman SE, Mustafa RZ, Jones J, et al. Treatment with Ibrutinib Inhibits BTK- and VLA-4-Dependent Adhesion of Chronic Lymphocytic Leukemia Cells In Vivo. Clin Cancer Res 2015;21:4642-51.
- Byrd JC, Furman RR, Coutre SE, et al. Three-year followup of treatment-naïve and previously treated patients with CLL and SLL receiving single-agent ibrutinib. Blood 2015;125:2497-506.
- Woyach JA. Patterns of resistance to B cell-receptor pathway antagonists in chronic lymphocytic leukemia and strategies for management. Hematology Am Soc Hematol Educ Program 2015;2015:355-60.
- Maffei R, Fiorcari S, Martinelli S, et al. Targeting neoplastic B cells and harnessing microenvironment: the "double face" of ibrutinib and idelalisib. J Hematol Oncol 2015;8:60.
- Fiorcari S, Maffei R, Audrito V, et al. Ibrutinib modifies the function of monocyte/macrophage population in chronic lymphocytic leukemia. Oncotarget 2016;7:65968-81.
- Boissard F, Fournié JJ, Quillet-Mary A, et al. Nurse-like cells mediate ibrutinib resistance in chronic lymphocytic leukemia patients. Blood Cancer J 2015;5:e355.
- 9. Cheng S, Guo A, Lu P, et al. Functional characterization of BTKC481S mutation that confers ibrutinib resistance:

Cite this article as: Fiorcari S, Maffei R, Marasca R. New players in the Bruton's tyrosine kinase *vs.* ibrutinib match. Transl Cancer Res 2017;6(Suppl 3):S469-S471. doi: 10.21037/tcr.2017.03.75

exploration of alternative kinase inhibitors. Leukemia 2015;29:895-900.

- Woyach JA, Furman RR, Liu TM, et al. Resistance mechanisms for the Bruton's tyrosine kinase inhibitor ibrutinib. N Engl J Med 2014;370:2286-94.
- Bottoni A, Rizzotto L, Lai TH, et al. Targeting BTK through microRNA in chronic lymphocytic leukemia. Blood 2016;128:3101-12.
- 12. Adams BD, Parsons C, Walker L, et al. Targeting noncoding RNAs in disease. J Clin Invest 2017;127:761-771.
- Balatti V, Pekarky Y, Croce CM. Role of microRNA in chronic lymphocytic leukemia onset and progression. J Hematol Oncol 2015;8:12.
- Sampath D, Liu C, Vasan K, et al. Histone deacetylases mediate the silencing of miR-15a, miR-16, and miR-29b in chronic lymphocytic leukemia. Blood 2012;119:1162-72.
- Byrd JC, Marcucci G, Parthun MR, et al. A phase 1 and pharmacodynamic study of depsipeptide (FK228) in chronic lymphocytic leukemia and acute myeloid leukemia. Blood 2005;105:959-67.
- Blum KA, Advani A, Fernandez L, et al. Phase II study of the histone deacetylase inhibitor MGCD0103 in patients with previously treated chronic lymphocytic leukaemia. Br J Haematol 2009;147:507-14.