



Antitumor activity of integrin $\alpha_v\beta_3$ antibody conjugated-cationic microbubbles in liver cancer

Jiale Li¹, Ping Zhou¹, Hongbo Xu², Shuangming Tian¹, Wengang Liu¹, Yongfeng Zhao¹, Zheyu Hu³

¹Department of Ultrasound, ²Department of General Surgery, the Third Xiangya Hospital, Central South University, Changsha 410013, China;

³Department of Breast Medical Oncology and Central Laboratory, the Affiliated Cancer Hospital of Xiangya School of Medicine, Central South University, Changsha 410013, China

Contributions: (I) Conception and design: J Li, P Zhou; (II) Administrative support: P Zhou; (III) Provision of study materials or patients: J Li, H Xu, S Tian, W Liu, Y Zhao, Z Hu; (IV) Collection and assembly of data: J Li; (V) Data analysis and interpretation: J Li; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

Correspondence to: Ping Zhou. Department of Ultrasound, the Third Xiangya Hospital, Central South University, No. 138, Tongzipo Road, Yuelu District, Changsha 410013, China. Email: zhouping_xythird@163.com.

Background: The overexpression of integrin $\alpha_v\beta_3$ in hepatocarcinoma (HCC) promotes tumor progression, metastasis, and clinical staging. Thus, the inhibition of integrin $\alpha_v\beta_3$ might be potentially effective as an anti-cancer agent in HCC.

Methods: In this study, we aimed to investigate the antitumor effect of integrin $\alpha_v\beta_3$ antibody conjugated cationic microbubbles (CMBs) in HCC model. By conjugating with integrin $\alpha_v\beta_3$ antibody with non-targeting CMBs, CMBs $_{\alpha_v\beta_3}$ was constructed. The antitumor effect of CMBs $_{\alpha_v\beta_3}$ was evaluated in HepG2 cells *in vitro* and in HepG2 xenograft mice models. Bcl-2, p53 and CD31 mRNA level, and caspase-3 activity were examined in xenograft tumors. Cell proliferation assay and scratch test were performed to evaluate the anti-migrant effect of CMBs $_{\alpha_v\beta_3}$ *in vitro*.

Results: CMBs $_{\alpha_v\beta_3}$ could specifically target to HCC HepG2 cells and improve pEGFP-KDRP-CD/TK plasmid transfection efficiency. In HepG2 xenograft mice models, CMBs $_{\alpha_v\beta_3}$ treatment significantly suppressed tumor weights and volumes. CMBs $_{\alpha_v\beta_3}$ treatment suppressed Bcl-2 and p53 mRNA level in tumors. In HepG2 cells, CMBs $_{\alpha_v\beta_3}$ significantly impaired wound healing and inhibited cell proliferation. Moreover, when combined with CD/TK double suicide gene transfection and 5-FC/GCV treatment, caspase-3 was activated and the cell proliferation was tremendously inhibited.

Conclusions: CMBs $_{\alpha_v\beta_3}$ not only suppresses cell migration and proliferation, but also facilitates 5-FC/GCV plus CD/TK double suicide gene-induced apoptotic cell death. CMBs $_{\alpha_v\beta_3}$ is a promising gene delivery agent with potential anti-tumor activity itself.

Keywords: Integrin $\alpha_v\beta_3$; antibody; conjugated-cationic microbubbles (CMBs); anti-tumor activity; HepG2 xenografts; migration

Submitted Dec 20, 2018. Accepted for publication May 15, 2019.

doi: 10.21037/tcr.2019.05.29

View this article at: <http://dx.doi.org/10.21037/tcr.2019.05.29>

Introduction

Integrins are cell-surface glycoproteins that trigger a diversity of signaling pathways, including cell adhesion and angiogenesis (1,2), and are involved in various human cancers. Integrin expression is hypothesized to

render cancer cells to proliferate and migrate (3). Thus, integrins represent attractive targets for the prevention of cancer spread and tumor progression (4). Integrin $\alpha_v\beta_3$ is constitutively expressed in quiescent endothelial cells at low level (5), but it is highly expressed in tumors, such as prostate cancer (6), breast cancer (7), and melanoma (4).

Integrin α_vβ₃ overexpression in hepatocarcinoma (HCC) is associated PI3K/Akt and TGF-β/ERK signaling pathways and promotes tumor progression, metastasis, and clinical staging (8-10).

The inhibition of integrin α_vβ₃ might suppress tumor proliferation. A series of integrin α_vβ₃ inhibitors have been developed to target tumors. RGD (Arg-Gly-Asp) peptides, a group of canonical antagonist of integrin α_vβ₃, show inhibitory activity in cell mobility and cell attachment in breast cancer (11). By enhancing internalization rate of these micelles in melanoma, the integrin α_vβ₃ targeting peptide (RGDfK) show synergistic cytotoxicity with docetaxel/cisplatin-co-loaded micelles (12). Besides RGD peptides, by binding to NC1 domain of collagen XIX, integrin α_vβ₃ could inhibit FAK/PI3K/Akt/mTOR pathway (13).

Our previous study also has demonstrated that by conjugating with α_vβ₃ integrin antibody, non-targeting cationic microbubbles (CMBs) can specifically target to HepG2 cells (14), and substantially increase pEGFP-KDRP-CD/TK plasmid transfection efficiency. The development of ultrasound contrast agent opens up a new idea of carrying target-delivery genes or drugs for chemotherapy in liver tumor patients (15). Ultrasound-targeted microbubble destruction (UTMD) provides a non-invasive, safe, and repeatable method for gene delivery (16-18). To further improve gene delivery efficiency, researchers design conjugated MBs to specifically bind to the membrane proteins on the surface of tumor cells. In the present study, CMBs_{α_vβ₃} could significantly suppress tumor growth in HepG2 xenografts mice model. *In vitro*, CMBs_{α_vβ₃} significantly impaired the wound healing and inhibited cell proliferation of HepG2 cells. CMBs_{α_vβ₃} also facilitated 5-FC/GCV + CD/TK double suicide gene-induced anti-tumor activity. These findings suggested CMBs_{α_vβ₃} as a promising gene delivery agent with potential anti-tumor activity itself.

Methods

Cell line and preparation of non-targeting CMBs

As previously described (14), human liver cancer HepG2 cells were purchased from Xiangya Cell Bank, Central South University (Changsha, China). 1,2-distearoyl-sn-glycero-3-phosphocholine (DSPC), dipalmitoylphosphatidylcholine (DPPC), biotinylated dipalmitoylphosphatidylethanolamine (DSPE-PEG2000-Biotin) and 1,2-distearoyl-3-trimethyl-ammonium-propane (DTAP; Avanti, Alabaster, AL, USA) were mixed in a 5 mL plastic tube

to form a suspension at a molar ratio of 46:36:8:2 (19). Perfluorinated propane (C₃F₈) was purchased from the Special gas Co., Ltd. Factory (Nanjing, China). Following lyophilization, 1 mL of phosphate-buffered saline (PBS) was added to the samples to rehydrate them and then C₃F₈ gas was slowly injected into the container to replace the air. Samples were then agitated using an ultrasonic mechanical vibrator with high speed shearing method for 90 s to form a milky white solution.

Preparation of CMBs_{α_vβ₃}

Integrin α_vβ₃ antibody was conjugated to the distal end of the DSPE-PEG2000-Biotin molecules through biotin-streptavidin coupling chemical method (20). Briefly, 500 μL CMBs (1×10⁹/mL) were mixed with 100 μg biotinylated anti-α_vβ₃ antibody in an ultrasonic agitating reaction for 30 min. Then, after centrifugation at 50 g for 5 min, the upper layer (CMBs_{α_vβ₃}) was washed with PBS three times and then collected. The morphology and particle distribution of CMBs_{α_vβ₃} was observed by optical microscopy. The particle size and surface potential were measured by a Zetasizer 3000HS (Malvern, Worcestershire, UK). All experiments were performed for five times.

Plasmids

As described in our previous study (14), the restructured plasmid pEGFP-KDRP-CD/TK coding for green fluorescent protein (GFP) contained CD/TK double suicide gene and was driven by KDR promoter (21). The molecular weight of this plasmid is 2,300 kDa with 3,486 bp. The plasmid was amplified and then isolated and purified using QIAGEN plasmid giga kit (Qiagen, Valencia, CA, USA) following the manufacturer's protocol.

Fluorescent assay

To confirm binding of CMBs_{α_vβ₃} to HepG2 cells, rhodamine mouse anti-human immunoglobulin (Ig) G was adopted to detect α_vβ₃ antibody. After treated with CMBs_{α_vβ₃} alone or plus pEGFP-KDRP-CD/TK plasmid, HepG2 cells were incubated with rhodamine mouse anti-human Ig G for 1 h at room temperature. Unbound (Ig) G was removed by two rounds of centrifugal washing. The binding was observed under a fluorescence microscope (CKX41; Olympus, Tokyo, Japan). EGFP expression levels were also evaluated by fluorescence microscope.

Animal experiments

As described previously (14), nude mice bearing HepG2 liver cancer were randomly divided into 3 groups: normal saline (NS) group, CMBs treatment group and CMBs_{αvβ3} treatment group. All treatments were terminated at 10 days. The tumor growth was observed and tumor volumes were calculated. Five days after treatment, HepG2 xenografts were removed and weighted. Tumor tissues RNA were extracted for quantitative RT-PCR (qRT-PCR) and caspase-3 activity assay.

qRT-PCR screening

Tumor RNA was isolated and converted into cDNA. qRT-PCR was performed using ABI-7500 (Applied Biosystems, Foster City, CA, USA) by mixing equal amounts of cDNAs, iQ SYBR Green Supermix (Bio-Rad, Hercules, CA, USA) and specific primers. All real-time data were normalized to β-actin.

Caspase-3 activity assay

Tumor tissue protein was extracted as previously described (22). Caspase-3 activity was tested instantly using the colorimetric substrate Ac-DEVD-pNA (Beyotime, Nanjing, China) according to the manufacturer's instructions. After incubation overnight at 37 °C, caspase-3 activity was measured on an enzyme-linked immunosorbent assay instrument (Bio-Rad, USA) at 490 nm.

Scratch closure test

The migration of HepG2 cells exposed to CMBs_{αvβ3} was evaluated by scratch closure test. We took images of the scratched area at 0, 12, and 24 h after scratch and measured widths of the scratched by the Image-Pro Plus 6.0 software.

Anti-tumor effect evaluation

To determine the effects of CMBs_{αvβ3} on the cell cycle, propidium iodide (PI) staining after 75% alcohol fixation was used, followed by flow cytometry analysis. MTT assay was performed to evaluate the cell proliferation inhibition after CMBs_{αvβ3} treatment.

Statistical analysis

All data are expressed as mean ± standard deviation (SD),

unless otherwise noted. Differences among groups were analyzed by one-way ANOVA and LSD method for multiple comparisons among groups using the SPSS software package (Version 19.0 for windows, SPSS, Chicago, Illinois, USA) with P<0.05 considered statistically significant.

Results

CMBs_{αvβ3} treatment suppresses HepG2 tumor growth

To investigate the anti-tumor effect of CMBs_{αvβ3} *in vivo*, HepG2 xenograft mice models were used. The nude mice bearing HepG2 liver cancers were randomly divided into 3 groups: NS group, CMBs treatment group and CMBs_{αvβ3} treatment group. The whole treatment period was 10 days. Tumor volumes were evaluated at day 0 to day 17 from the treatment initial day. As shown in *Figure 1*, mice in NS (*Figure 1A*), CMBs (*Figure 1B*), CMBs_{αvβ3} (*Figure 1C*) groups were sacrificed and tumors were removed and weighed at 17 days after treatment. Compared to control (NS) group, CMBs_{αvβ3} treatment significantly suppressed tumor growth (P<0.05; *Figure 2A*). But CMBs treatment had no such effect. The xenograft tumors were weighted at the experiment endpoint. Compared to control group and CMBs group, CMBs_{αvβ3} treatment significantly suppressed tumor weight (P<0.05, *Figure 2B*). Mice body weights of all three treatment groups did not change significantly during the experiment period (*Figure 2C*). These findings implied an anti-tumor effect of CMBs_{αvβ3}.

CMBs_{αvβ3} binds to tumor cells in xenografts and suppresses Bcl-2 and p53 mRNA level

CMBs_{αvβ3} is supposed to specifically bind to integrin α_vβ₃ on HepG2 cell surface through the conjugated integrin α_vβ₃ antibody. To confirm this binding, we adopted rhodamine mouse anti-human immunoglobulin (Ig) G and fluorescence microscope to detect the localization of CMBs_{αvβ3} on HepG2 cells. As demonstrated in *Figure S1*, the binding of CMBs_{αvβ3} to the surface of HepG2 cells was detected under fluorescence microscope. The red fluorescent signals suggested a tight binding of CMBs_{αvβ3} to HepG2 xenograft cells.

In previous reports, integrin α_vβ₃ inhibitor has shown effect to inhibit Bcl-2 expression (23) and modify p53 level (24). Consistent with these reports, qRT-PCR results in *Figure 3A,B* showed that CMBs_{αvβ3} treatment significantly suppressed the Bcl-2 and p53 mRNA levels, compared to control. CD31 mRNA level was not affected by CMBs_{αvβ3}

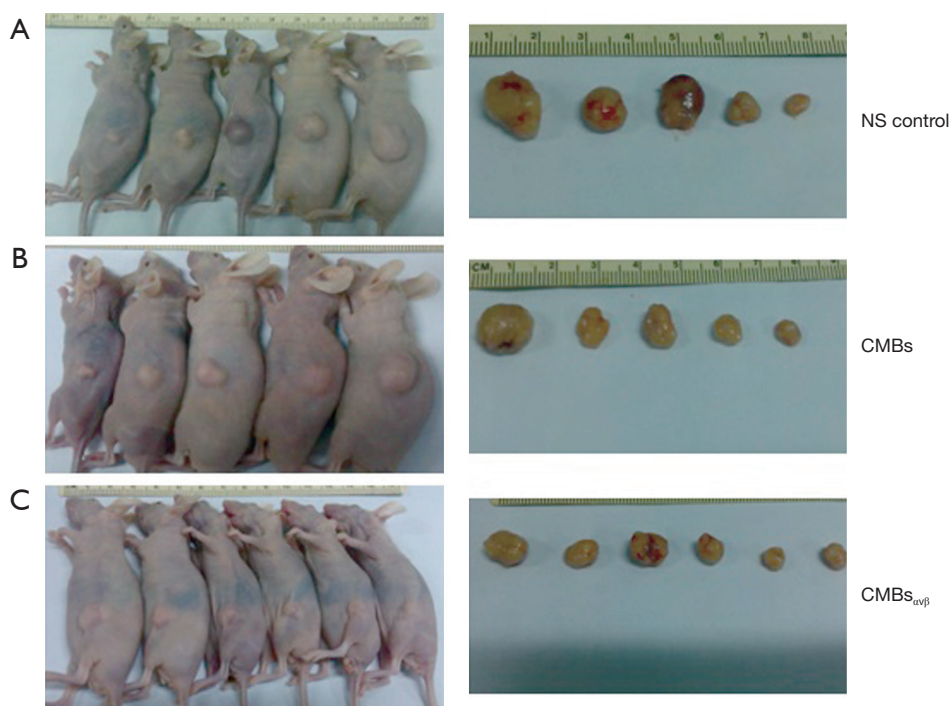


Figure 1 HepG2-bearing nude mice were treated with NS (A); CMBs (B); CMBs_{αvβ3} (C). Seventeen days after treatment, mice were sacrificed and HepG2 xenograft tumors were removed for further evaluation. All data are expressed as mean ± SD. Differences among groups were analyzed by one-way ANOVA and LSD method for multiple comparisons among groups using the SPSS software package (Version 19.0 for windows, SPSS, Chicago, Illinois, USA) with $P < 0.05$ considered statistically significant. NS, normal saline; CMBs, cationic microbubbles; SD, standard deviation.

treatment (Figure 3C). No obvious caspase-3 activation was detected in CMBs_{αvβ3} treatment group (Figure 3D). These data implied that the suppression of Bcl-2 and p53 might be responsible for CMBs_{αvβ3}-mediated anti-tumor effect. But caspase-3-mediated apoptosis pathway was not involved in CMBs_{αvβ3}-mediated anti-tumor effect.

CMBs_{αvβ3} inhibits HepG2 cells proliferation and migration in vitro

The anti-tumor CMBs_{αvβ3} effect of was also evaluated by cell cycle and MTT assays. As shown in Figure 4A, cell cycle distribution was not modified by CMBs_{αvβ3} treatment. MTT assay showed that HepG2 cell proliferation was obviously inhibited by CMBs_{αvβ3} treatment (Figure 4B). Integrin $\alpha_v\beta_3$ triggers signaling pathway to promote cell adhesion and migration (3). To examine the anti-migrant effect of CMBs_{αvβ3}, HepG2 cells were exposed to CMBs_{αvβ3} for 24 h, and then evaluated by scratch closure test. At 24 h after scratch, areas of the scratched wounds were measured

by Image J software (Figure 5), Compared to NS treatment (Figure 5A), at 24 h after scratch, CMBs_{αvβ3} treatment significantly inhibited the healing of scratched wounds (Figure 5B).

CMBs_{αvβ3} facilitates CD/TK transfection and promotes antitumor activity

In our previous publication (14), we had demonstrated that CMBs_{αvβ3} combined with CD/TK transfection + 5-FC/GCV had a higher suppressing effect in HepG2 xenograft mice model than CD/TK transfection + 5-FC/GCV alone. Moreover, CMBs_{αvβ3} plus CD/TK + GCV/5-FC treatment induced more TUNEL-positive cells than CD/TK transfection + 5-FC/GCV treatment alone (14). Here, as demonstrated in Figure S1, the green fluorescence signals indicated a successful CD/TK transfection in HepG2 cells. qRT-PCR assay further confirmed the expression of CD and TK mRNA in HepG2 cells (Figure S2). When coupled with CMBs_{αvβ3}, CD/TK gene expression levels were higher

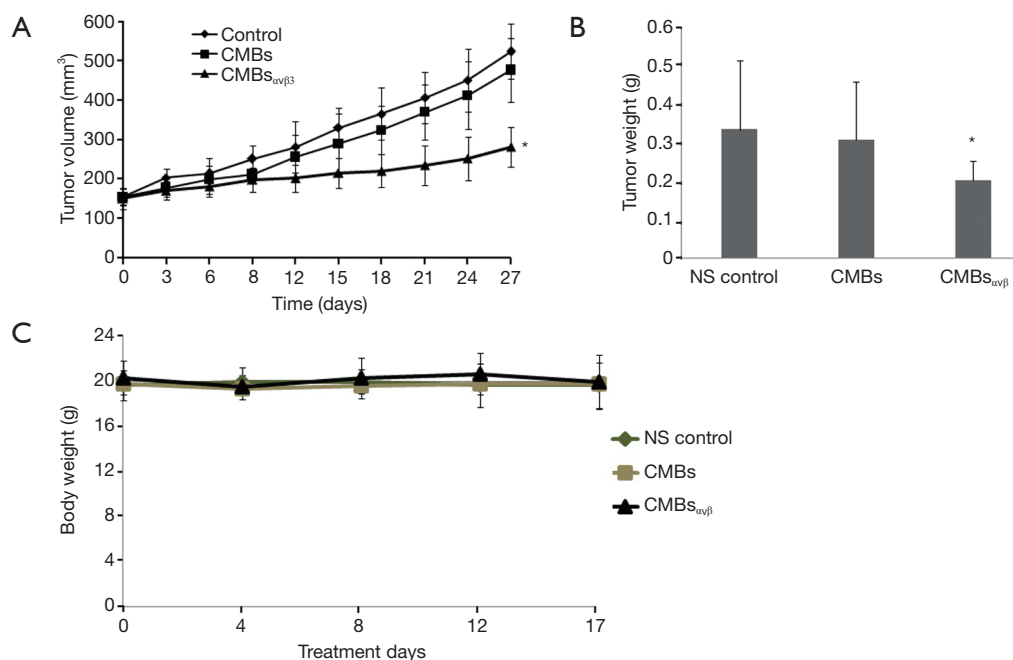


Figure 2 CMBs_{αvβ3} inhibited tumor growth in HepG2-bearing nude mice. Differences among groups were analyzed by one-way ANOVA and LSD method for multiple comparisons among groups with P<0.05 considered statistically significant. (A) Tumor volumes were measured on day 0 to day 27 for every three days. The tumor volume in CMBs_{αvβ3} group was significantly lower compared with NS group; (B) the tumor weight in CMBs_{αvβ3} group was significantly lower compared with CMBs group and NS group; (C) mice body weights kept unchanged among all treatment groups. *, P<0.05. NS, normal saline; CMBs, cationic microbubbles.

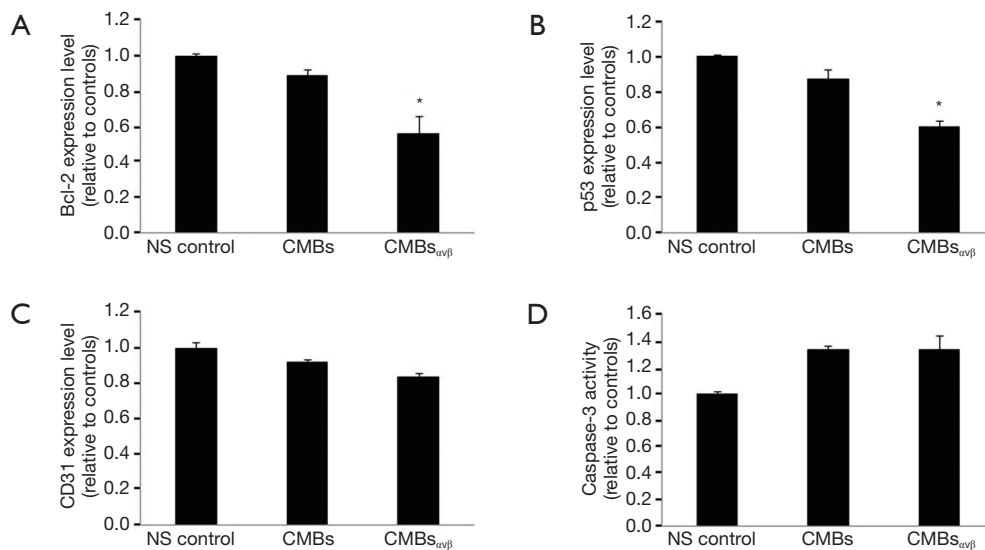


Figure 3 CMBs_{αvβ3} suppresses Bcl-2 and p53 mRNA level. Differences among groups were analyzed by one-way ANOVA and LSD method for multiple comparisons among groups with P<0.05 considered statistically significant. mRNA expression levels of Bcl-2 (A), p53 (B) and CD31 (C) were determined by qRT-PCR. β-Actin was used as a control to confirm equal loading of cDNAs. Data are shown as means ± SD of three experiments; (D) caspase-3 activity was evaluated colorimetric substrate and measured by Bio-Rad reader. Data are shown as means ± SD of three experiments. *, P<0.05. NS, normal saline; CMBs, cationic microbubbles; qRT-PCR, quantitative RT-PCR; SD, standard deviation.

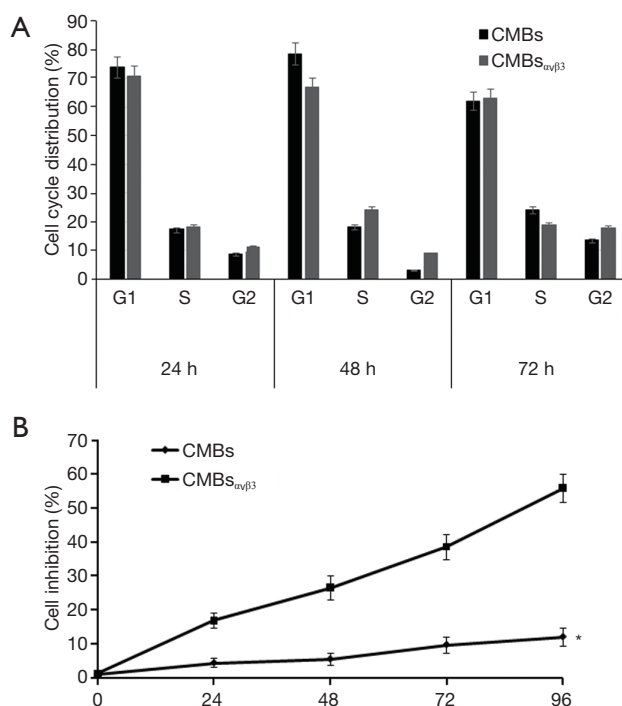


Figure 4 CMBs_{αvβ3} inhibits cell proliferation *in vitro*. Differences among groups were analyzed by one-way ANOVA and LSD method for multiple comparisons among groups with $P < 0.05$ considered statistically significant. (A) The effects of CMBs_{αvβ3} treatment on the cell cycle distribution of HepG2 cells were analyzed using Multicycle software program (Phoenix Flow System) (mean \pm SD of three experiments); (B) the anti-proliferation effect of CMBs_{αvβ3} in HepG2 cells was measured by MTT assay (mean \pm SD of three experiments). Compared to CMBs group, CMBs_{αvβ3} treatment significantly inhibited the cell proliferation. *, $P < 0.05$. CMBs, cationic microbubbles; SD, standard deviation.

than CD/TK plasmid alone in HepG2 cells ($P < 0.05$; Figure S2B,C).

The effect of CMBs_{αvβ3} in GCV/5-FC + CD/TK activity was also evaluated. As demonstrated in Figure 5C, CD/TK suicide gene transfection alone did not prevent the wound healing. When combined with CMBs_{αvβ3}, CD/TK + GCV/5-FC could significantly suppress the healing of the scratched wounds (Figure 5E). Figure 5F compared the percentages of wound area relative to the initial wound area among treatment groups. Compared to NS control, both CMBs_{αvβ3} treatment alone and CD/TK + GCV/5-FC treatment alone could significantly suppress the wounds healing ($P < 0.05$). Moreover, compared to CD/TK +

GCV/5-FC treatment alone group, CMBs_{αvβ3} plus CD/TK + GCV/5-FC treatment had even lower wound healing rate ($P < 0.05$). These results implied that CMBs_{αvβ3} not only inhibited HCC cell migration itself, but also increased the anti-migrant effect of CD/TK + GCV/5-FC treatment.

While no obvious caspase-3 activation was detected in CMBs_{αvβ3} treatment group, CD/TK + 5-FC/GCV treatment and CD/TK + 5-FC/GCV plus CMBs_{αvβ3} treatment could induce significant caspase-3 activation in HepG2 cells (Figure 6A). These data implied that caspase-3-mediated apoptosis pathway was involved in CD/TK + 5-FC/GCV-mediated anti-tumor effect, but not in CMBs_{αvβ3} treatment. MTT assay showed a significant higher suppression in cell proliferation in CD/TK + 5-FC/GCV plus CMBs_{αvβ3} treatment group, compared to CMBs_{αvβ3} alone or CD/TK + 5-FC/GCV treatment alone group (Figure 6B). Based on these findings, we suggested that CMBs_{αvβ3} not only had potential anti-tumor activity itself, but also facilitated CD/TK gene transfection and promoted the anti-tumor effects (Figure 7).

Discussion

In the era of precision medicine, targeting and accuracy become more and more important and practicable. Application of CMBs to treat cancers (25), brain disease (26,27), hepatic fibrosis (21), and heart diseases (28) are non-invasive, targeted and safe. Our designed CMBs_{αvβ3} carries CD/TK double suicide gene, and focused UTMD helps local gene delivery.

In this study, we reported a novel system CMBs_{αvβ3}, with high affinity to integrin $\alpha_v\beta_3$ on HepG2 surface, and showed potent anti-tumor activity in HepG2 mice models. This integrin $\alpha_v\beta_3$ targeting system not only suppressed tumor growth alone, but also promoted CD/TK gene expression and 5-FC/GCV killing effect. Unlike other integrin ligands/analogues/peptides or integrin-based drug-load systems, CMBs_{αvβ3} initiated a completely new research field for the combination of multiple anti-tumor effects in one system. We believe the major anti-tumor actions of CMBs_{αvβ3} includes following two aspects: (I) inhibiting migration of integrin $\alpha_v\beta_3$ -overexpressed tumor cells; (II) facilitating CD/TK suicide gene transfection and promoting 5-FC/GCV-induced apoptotic cell death. This proposed model was described in Figure 7.

Biotinylated specific antibodies coupled to streptavidin-containing microbubbles through the biotin-streptavidin linkage have been used for molecular ultrasound imaging

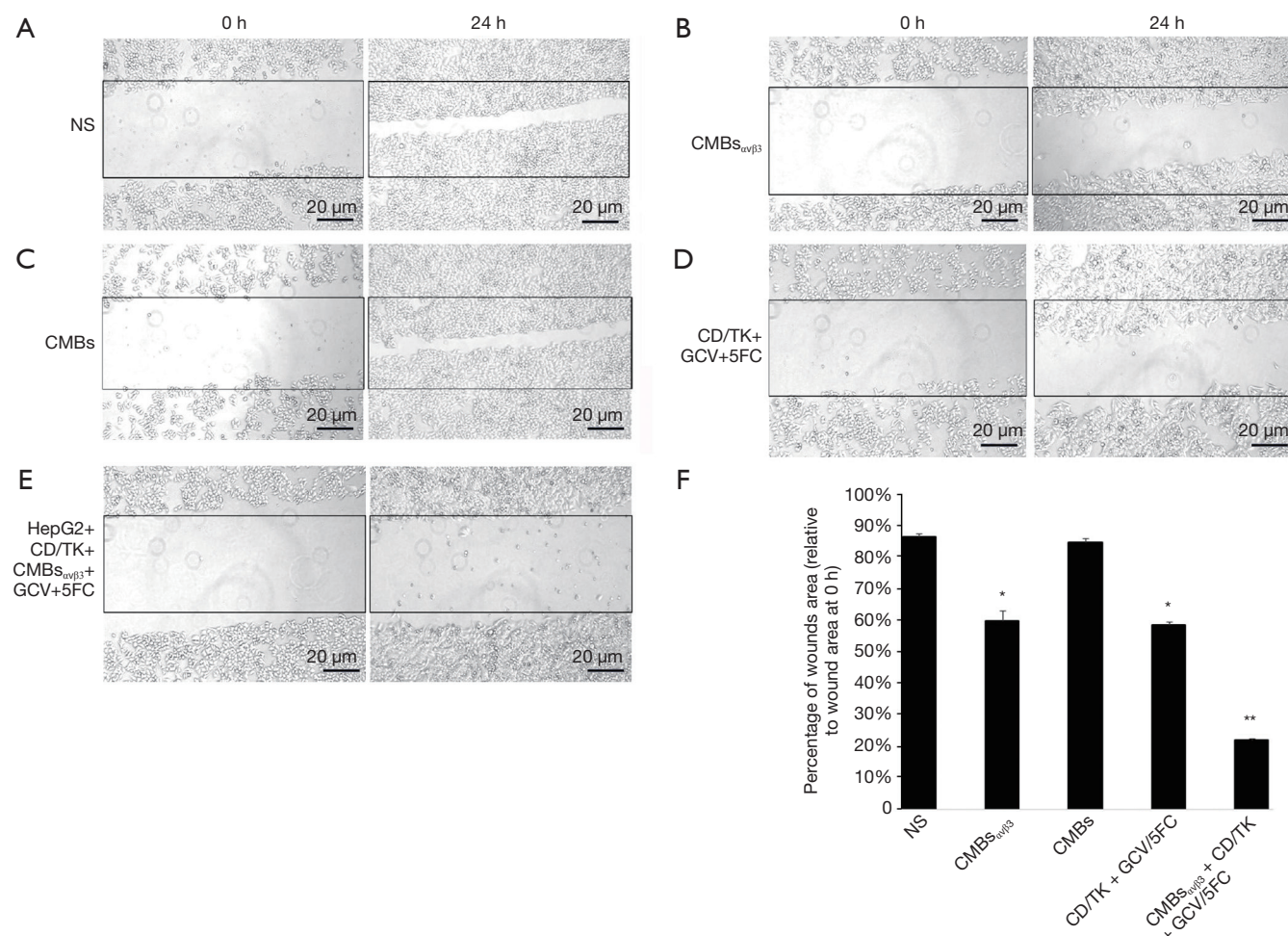


Figure 5 CMBs_{αvβ3} inhibits migration. Differences among groups were analyzed by one-way ANOVA and LSD method for multiple comparisons among groups with $P < 0.05$ considered statistically significant. Effects of NS (A), CMBs_{αvβ3} (B), CMBs (C), CD/TK transfection + GCV/5FC (D) and CMBs_{αvβ3} + CD/TK transfection + GCV/5FC (E) on wound closure in HepG2 cells. 10 visions were chosen at random and each experiment was repeated for three times (mean \pm SD of three experiments); (F) wounds healing areas were compared among five treatment groups. CMBs_{αvβ3} and CD/TK transfection + GCV/5FC treatment significantly suppressed wounds healing. Compared to CD/TK transfection + GCV/5FC treatment, CMBs_{αvβ3} + CD/TK transfection + GCV/5FC further inhibited wounds healing. Data are shown as means \pm SD of three experiments. *, $P < 0.05$, **, $P < 0.01$. NS, normal saline; CMBs, cationic microbubbles; SD, standard deviation.

to monitor the receptor expression (29), such as VEGF receptor (30,31). Like Cyclic RGD peptides, biotin-streptavidin linkage is commonly used to specifically couple quantum dots to integrins (32). Cilengitide, a cyclic pentapeptide, is efficient to treat glioblastoma by targeting integrin $\alpha_v\beta_3$ and $\alpha_v\beta_5$ (33,34). Combination of UTMD with cilengitide-nanotherapy increases the tumor Cilengitide level over 3-fold and significantly reduce renal clearance, which help reduce Cilengitide dose level and increase killing effect (35). From our animal experiment results,

focused UTMD with CMBs_{αvβ3} showed significantly anti-tumor effect in HepG2 xenograft tumors (Figure 1).

Integrins $\alpha_v\beta_3$ and $\alpha_v\beta_5$ ligands contain the RGD sequence. Cyclic RGD peptides are canonical integrin inhibitors, such as cilengitide (36). Humanized monoclonal integrin antibodies abrituzumab and vedolizumab have demonstrated anti-tumor activity preclinically (37). Other inhibitors including salvicine and borrelidin are non-specific (38). CMBs_{αvβ3} specifically targets to integrin $\alpha_v\beta_3$ and showed high affinity to HepG2 cells in our models

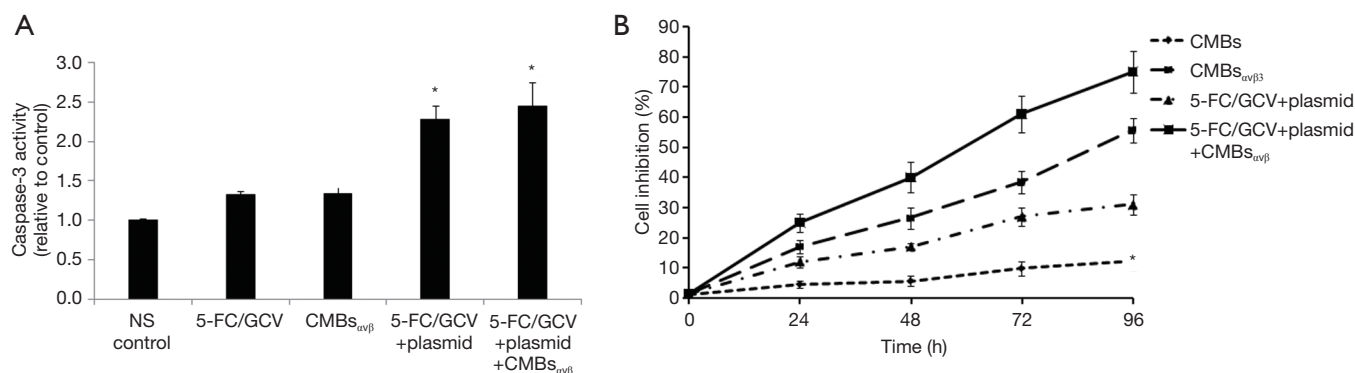


Figure 6 $\text{CMBs}_{\alpha_v\beta_3}$ facilitates the anti-tumor effect of CD/TK + GCV/5FC *in vitro*. Differences among groups were analyzed by one-way ANOVA and LSD method for multiple comparisons among groups with $P < 0.05$ considered statistically significant. (A) The caspase-3 activity was evaluated in NS, 5-FC/GCV, $\text{CMBs}_{\alpha_v\beta_3}$, 5-FC/GCV + CD/TK plasmid and 5-FC/GCV + CD/TK plasmid + $\text{CMBs}_{\alpha_v\beta_3}$ group. Both 5-FC/GCV + CD/TK plasmid treatment and 5-FC/GCV + CD/TK plasmid + $\text{CMBs}_{\alpha_v\beta_3}$ treatment induced significantly higher caspase-3 activity; (B) the anti-proliferation effect of CMBs, $\text{CMBs}_{\alpha_v\beta_3}$, 5-FC/GCV + CD/TK plasmid and 5-FC/GCV + CD/TK plasmid + $\text{CMBs}_{\alpha_v\beta_3}$ in HepG2 cells was measured by MTT assay (mean \pm SD of three experiments). Compared to CMBs alone or CD/TK + 5-FC/GCV treatment alone group, CD/TK + 5-FC/GCV plus $\text{CMBs}_{\alpha_v\beta_3}$ treatment showed a significant higher suppression in cell proliferation. *, $P < 0.05$. NS, normal saline; CMBs, cationic microbubbles; SD, standard deviation.

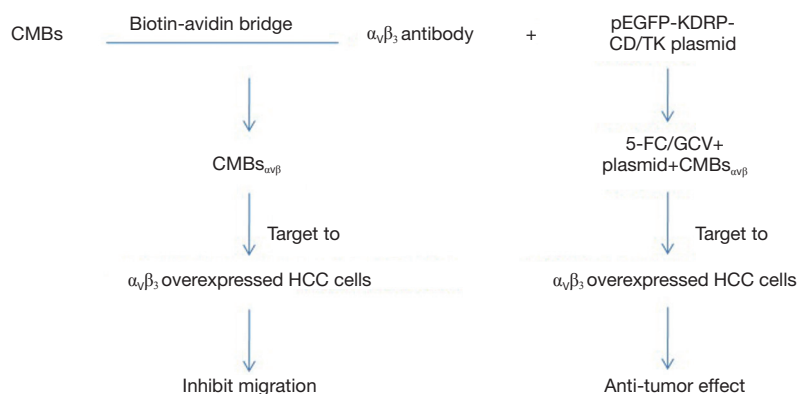


Figure 7 Hypothesis diagram of the anti-tumor effect of $\text{CMBs}_{\alpha_v\beta_3}$. HCC, hepatocarcinoma; CMBs, cationic microbubbles.

(Figure S1).

Integrin inhibitors always demonstrate anti-angiogenic (39) and anti-metastasis (40) functions. By targeting to integrin $\alpha_v\beta_3$ on neovasculature, cilengitide increased systemic radio-immunotherapy efficacy of therapy in p53 mutant and bcl-2 overexpressing breast cancer cells (41). In this study, $\text{CMBs}_{\alpha_v\beta_3}$ significantly suppressed Bcl-2 and p53 levels, while CD31 level was moderately suppressed in mice models (Figure 3). *In vitro* experiment and inhibited HepG2 cell migration (Figure 5). Caspase-3 activity was not influenced by $\text{CMBs}_{\alpha_v\beta_3}$, which is consistent with our previous study that $\text{CMBs}_{\alpha_v\beta_3}$ alone did not induce apoptotic death in HepG2 animal model (14).

Conclusions

$\text{CMBs}_{\alpha_v\beta_3}$ exerted the anti-tumor activities by inhibiting migration of integrin $\alpha_v\beta_3$ -overexpressed tumor cells and facilitating CD/TK suicide gene transfection and promoting 5-FC/GCV-induced apoptotic cell death. Application of $\text{CMBs}_{\alpha_v\beta_3}$ could initiate an accurate-targeting field for the combination of multiple anti-tumor effects in one system.

Acknowledgments

We thank Dr. Yu Sun from Nanjing Origin Biosciences Incorporation for animal model and CMBs construction.

Funding: This work was funded by a grant from National Natural Science Foundation of China (81271680) and Ultrasound Department of the Third Xiangya Hospital in Changsha, Hunan Province, China.

Footnote

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <http://dx.doi.org/10.21037/tcr.2019.05.29>). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The animal experiment in this study complied with the Third Xiangya Hospital guidelines and approved by the Third Xiangya Hospital Ethics Committee (2012-S119).

Open Access Statement: This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

References

- Kampen KR. Membrane proteins: the key players of a cancer cell. *J Membr Biol* 2011;242:69-74.
- Sayed-yahosseini S, Dagnino L. Integrins and small GTPases as modulators of phagocytosis. *Int Rev Cell Mol Biol* 2013;302:321-54.
- Weber MR, Zuka M, Lorger M, et al. Activated tumor cell integrin alphavbeta3 cooperates with platelets to promote extravasation and metastasis from the blood stream. *Thromb Res* 2016;140 Suppl 1:S27-36.
- Yacobovich S, Tuchinsky L, Kirby M, et al. Novel synthetic cyclic integrin alphavbeta3 binding peptide ALOS4: Antitumor activity in mouse melanoma models. *Oncotarget* 2016;7:63549-60.
- Brooks PC, Clark RA, Cheresh DA. Requirement of vascular integrin alpha v beta 3 for angiogenesis. *Science* 1994;264:569-71.
- McCabe NP, De S, Vasanji A, et al. Prostate cancer specific integrin alphavbeta3 modulates bone metastatic growth and tissue remodeling. *Oncogene* 2007;26:6238-43.
- Chakravarty R, Chakraborty S, Dash A. Molecular Imaging of Breast Cancer: Role of RGD Peptides. *Mini Rev Med Chem* 2015;15:1073-94.
- Jin Y, Chen JN, Feng ZY, et al. OPN and alphavbeta3 expression are predictors of disease severity and worse prognosis in hepatocellular carcinoma. *PLoS One* 2014;9:e87930.
- Xia H, Chen J, Shi M, et al. EDIL3 is a novel regulator of epithelial-mesenchymal transition controlling early recurrence of hepatocellular carcinoma. *J Hepatol* 2015;63:863-73.
- Xu ZZ, Xiu P, Lv JW, et al. Integrin alphavbeta3 is required for cathepsin B-induced hepatocellular carcinoma progression. *Mol Med Rep* 2015;11:3499-504.
- Georgoulis A, Havaki S, Drosos Y, et al. RGD binding to integrin alphavbeta3 affects cell motility and adhesion in primary human breast cancer cultures. *Ultrastruct Pathol* 2012;36:387-99.
- Song W, Tang Z, Zhang D, et al. Anti-tumor efficacy of c(RGDfK)-decorated polypeptide-based micelles co-loaded with docetaxel and cisplatin. *Biomaterials* 2014;35:3005-14.
- Oudart JB, Doue M, Vautrin A, et al. The anti-tumor NC1 domain of collagen XIX inhibits the FAK/ PI3K/Akt/ mTOR signaling pathway through alphavbeta3 integrin interaction. *Oncotarget* 2016;7:1516-28.
- Xue Q, Liu Y, He R, et al. Lyophilized Powder of Catalpol and Puerarin Protects Neurovascular Unit from Stroke. *Int J Biol Sci* 2016;12:367-80.
- Jing H, Cheng W, Zhang JW, et al. Galactosylated poly-L-lysine targeted microbubbles for ultrasound mediated antisense c-myc gene transfection in hepatocellular carcinoma cells. *Arch Med Sci* 2015;11:292-300.
- Cui K, Yan T, Luo Q, et al. Ultrasound microbubble-mediated delivery of integrin-linked kinase gene improves endothelial progenitor cells dysfunction in pre-eclampsia. *DNA Cell Biol* 2014;33:301-10.
- Fujii H, Li SH, Wu J, et al. Repeated and targeted transfer of angiogenic plasmids into the infarcted rat heart via ultrasound targeted microbubble destruction enhances cardiac repair. *Eur Heart J* 2011;32:2075-84.
- Suzuki J, Ogawa M, Takayama K, et al. Ultrasound-microbubble-mediated intercellular adhesion molecule-1 small interfering ribonucleic acid transfection attenuates neointimal formation after arterial injury in mice. *J Am Coll Cardiol* 2010;55:904-13.

19. Hou L, Zhao X, Wang P, et al. Antitumor activity of antimicrobial peptides containing CisoDGRC in CD13 negative breast cancer cells. *PLoS One* 2013;8:e53491.
20. Kaur S, Kenny HA, Jagadeeswaran S, et al. β 3-integrin expression on tumor cells inhibits tumor progression, reduces metastasis, and is associated with a favorable prognosis in patients with ovarian cancer. *Am J Pathol* 2009;175:2184-96.
21. Zhao YZ, Lin Q, Wong HL, et al. Glioma-targeted therapy using Cilengitide nanoparticles combined with UTMD enhanced delivery. *J Control Release* 2016;224:112-25.
22. Saini R, Sorace AG, Warram JM, et al. An animal model allowing controlled receptor expression for molecular ultrasound imaging. *Ultrasound Med Biol* 2013;39:172-80.
23. Pochon S, Tardy I, Bussat P, et al. BR55: a lipopeptide-based VEGFR2-targeted ultrasound contrast agent for molecular imaging of angiogenesis. *Invest Radiol* 2010;45:89-95.
24. Nunn A, Pochon S, Tardy I, et al. Microbubble-conjugated vascular endothelial growth factor receptor 2 binding peptide. In: Bethesda: National Center for Biotechnology Information. Molecular Imaging and Contrast Agent Database (MICAD), 2004.
25. Lieleg O, Lopez-Garcia M, Semmrich C, et al. Specific integrin labeling in living cells using functionalized nanocrystals. *Small* 2007;3:1560-5.
26. Ishida J, Onishi M, Kurozumi K, et al. Integrin inhibitor suppresses bevacizumab-induced glioma invasion. *Transl Oncol* 2014;7:292-302.e1.
27. Kurozumi K, Ichikawa T, Onishi M, et al. Cilengitide treatment for malignant glioma: current status and future direction. *Neurol Med Chir (Tokyo)* 2012;52:539-47.
28. Ruffini F, Graziani G, Levati L, et al. Cilengitide downmodulates invasiveness and vasculogenic mimicry of neuropilin 1 expressing melanoma cells through the inhibition of α v β 5 integrin. *Int J Cancer* 2015;136:E545-58.
29. Élez E, Kocakova I, Hohler T, et al. Abituzumab combined with cetuximab plus irinotecan versus cetuximab plus irinotecan alone for patients with KRAS wild-type metastatic colorectal cancer: the randomised phase I/II POSEIDON trial. *Ann Oncol* 2015;26:132-40.
30. Hariharan S, Gustafson D, Holden S, et al. Assessment of the biological and pharmacological effects of the α nu β 3 and α nu β 5 integrin receptor antagonist, cilengitide (EMD 121974), in patients with advanced solid tumors. *Ann Oncol* 2007;18:1400-7.
31. Harisi R, Jeney A. Extracellular matrix as target for antitumor therapy. *Onco Targets Ther* 2015;8:1387-98.
32. Oliveira-Ferrer L, Hauschild J, Fiedler W, et al. Cilengitide induces cellular detachment and apoptosis in endothelial and glioma cells mediated by inhibition of FAK/src/AKT pathway. *J Exp Clin Cancer Res* 2008;27:86.
33. Burke PA, DeNardo SJ, Miers LA, et al. Cilengitide targeting of α (v) β (3) integrin receptor synergizes with radioimmunotherapy to increase efficacy and apoptosis in breast cancer xenografts. *Cancer Res* 2002;62:4263-72.
34. Xu J, Zeng X, Liu Y, et al. A novel dual-targeted ultrasound contrast agent provides improvement of gene delivery efficiency in vitro. *Tumour Biol* 2016;37:8609-19.
35. Fan CH, Ting CY, Lin CY, et al. Noninvasive, Targeted, and Non-Viral Ultrasound-Mediated GDNF-Plasmid Delivery for Treatment of Parkinson's Disease. *Sci Rep* 2016;6:19579.
36. Tan JK, Pham B, Zong Y, et al. Microbubbles and ultrasound increase intraventricular polyplex gene transfer to the brain. *J Control Release* 2016;231:86-93.
37. Zhang SH, Wen KM, Wu W, et al. Efficacy of HGF carried by ultrasound microbubble-cationic nanoliposomes complex for treating hepatic fibrosis in a bile duct ligation rat model, and its relationship with the diffusion-weighted MRI parameters. *Clin Res Hepatol Gastroenterol* 2013;37:602-7.
38. Sun L, Huang CW, Wu J, et al. The use of cationic microbubbles to improve ultrasound-targeted gene delivery to the ischemic myocardium. *Biomaterials* 2013;34:2107-16.
39. Christiansen JP, French BA, Klibanov AL, et al. Targeted tissue transfection with ultrasound destruction of plasmid-bearing cationic microbubbles. *Ultrasound Med Biol* 2003;29:1759-67.
40. Lindner JR, Song J, Xu F, et al. Noninvasive ultrasound imaging of inflammation using microbubbles targeted to activated leukocytes. *Circulation* 2000;102:2745-50.
41. Martenson RE, Deibler GE, Kies MW. Extraction of rat myelin basic protein free of other basic proteins of whole central nervous system tissue. An analysis of its electrophoretic heterogeneity. *J Biol Chem* 1969;244:4268-72.

Cite this article as: Li J, Zhou P, Xu H, Tian S, Liu W, Zhao Y, Hu Z. Antitumor activity of integrin α v β 3 antibody conjugated-cationic microbubbles in liver cancer. *Transl Cancer Res* 2019;8(3):899-908. doi: 10.21037/tcr.2019.05.29

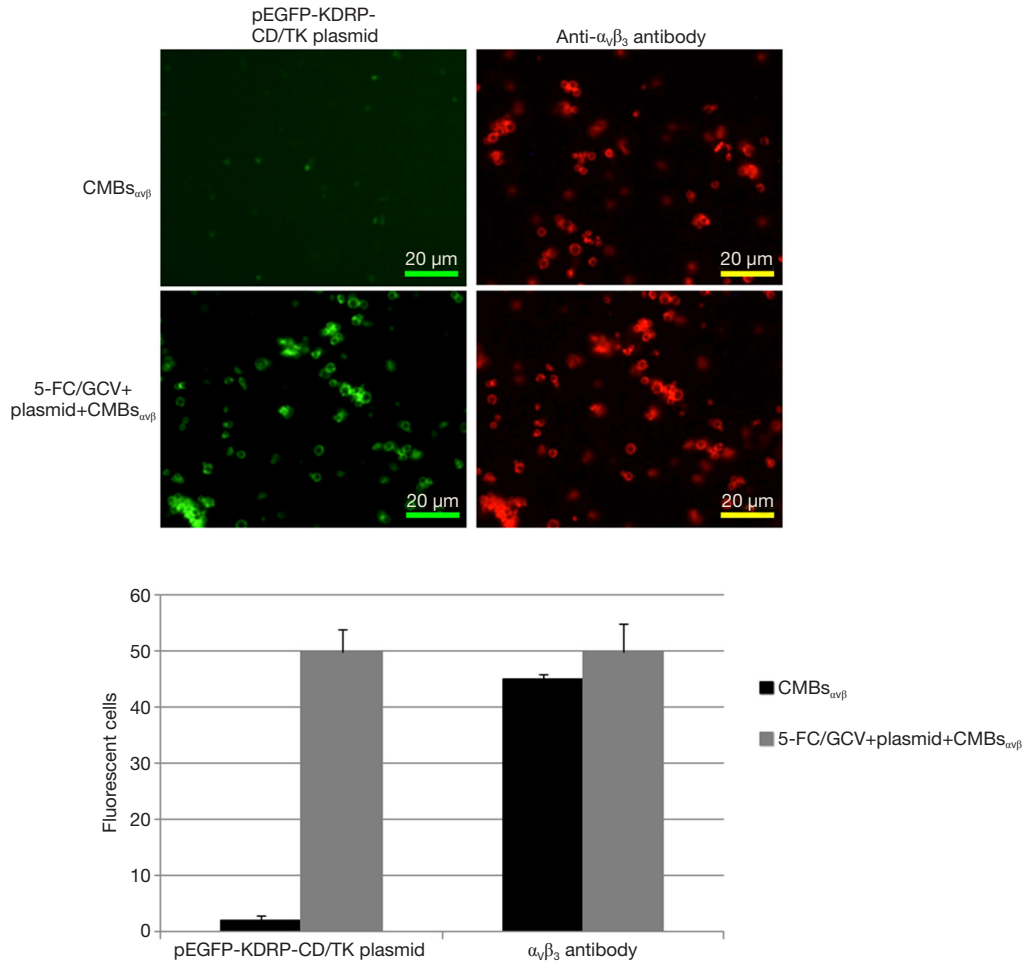


Figure S1 Binding of CMBs_{αvβ3} to HepG2 cells in xenograft tumors. (A) Expression of CD/TK in tumor cells was detected by EGFP green fluorescence. Red fluorescence was observed on the surface of CMBs_{αvβ3} using immunofluorescent microscopy. Ten visions were chosen at random and each experiment was repeated for three times (mean ± SD of three experiments); (B) CMBs_{αvβ3} specifically targets to integrin α_vβ₃ and showed high affinity to HepG2 cells in our models. CMBs, cationic microbubbles; SD, standard deviation.

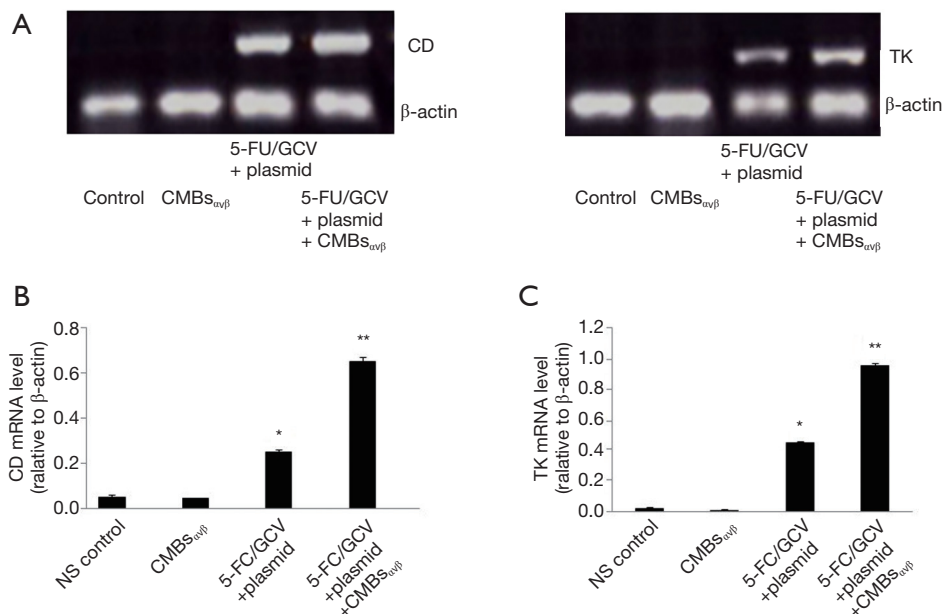


Figure S2 CMBs_{αvβ3} facilitates CD/TK gene transfection in HepG2 cells. (A) RT-PCR showed the expression level of CD and TK mRNA in CMBs_{αvβ3} + CD/TK group was higher than that in CD/TK group; mRNA expression levels of CD (B) and TK (C) were determined by qRT-PCR. β-Actin was used as a control to confirm equal loading of cDNAs. Data are shown as means ± SD of three experiments. *, P<0.05; **, P<0.01. NS, normal saline; CMBs, cationic microbubbles; qRT-PCR, quantitative RT-PCR.