



# The analysis of a ceRNA network and the correlation between lncRNA, miRNA, and mRNA in bladder cancer

Ranran Dai<sup>#</sup>, You Zhou<sup>#</sup>, Zhishan Chen, Zihao Zou, Ping Liu, Xingcheng Gao

Department of Urology Surgery, The Third Affiliated Hospital of Guangzhou Medical University, Guangzhou 510150, China

**Contributions:** (I) Conception and design: R Dai, Y Zhou, X Gao; (II) Administrative support: R Dai, Y Zhou, X Gao; (III) Provision of study materials or patients: All authors; (IV) Collection and assembly of data: All authors; (V) Data analysis and interpretation: All authors; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

<sup>#</sup>These authors contributed equally to this study.

**Correspondence to:** Dr. Xingcheng Gao. Department of Urology Surgery, The Third Affiliated Hospital of Guangzhou Medical University, No. 63 Duobao Road, Guangzhou 510150, China. Email: gaoxch008@126.com.

**Background:** To explore the correlation between the lncRNA-miRNA-mRNA and ceRNA network through the differential expression analysis of lncRNAs, miRNAs and mRNAs in bladder cancer based on The Cancer Genome Atlas (TCGA) database combined with Gene Ontology (GO) and Kyoto Encyclopedia of Genes Genomes (KEGG) enrichment analysis.

**Methods:** Firstly, the expression profile data and corresponding clinical data of RNAs in bladder cancer were searched and downloaded from TCGA database, and aberrantly expressed long non-coding RNA (lncRNA), microRNA (miRNA), and messenger RNA (mRNA) were screened and found by using TCGA database. The relationship between lncRNA-miRNA-mRNA was established by comparing these lncRNAs, miRNAs, and mRNAs, while the ceRNA network was constructed. Combined with the analysis of the GO annotation and KEGG pathway, the effects of lncRNA-miRNA-mRNA interaction on the development of bladder cancer were explored.

**Results:** A total of 1,742 differentially expressed lncRNA, 511 differentially expressed miRNAs, and 4,373 differentially expressed mRNAs were identified, and 328 lncRNAs, 73 miRNAs, and 677 mRNAs were screened by survival analysis. With the lncRNA-miRNA-mRNA correlation analysis, a ceRNA network consisting of 45 lncRNAs, 14 miRNAs, and 29 mRNAs was successfully constructed. The GO annotation and functional enrichment of target gene mRNAs in the network are mainly concentrated in the signal pathways and include fatty acid biosynthesis, gap junction, insulin signaling pathway, and the MAPK signaling pathway biological processes such as positive regulation of cellular process and system development.

**Conclusions:** We successfully identified the target gene correlating lncRNA, miRNA, and mRNA, and constructed a ceRNA network. Our findings can provide a potential target for the study of the occurrence, development, diagnosis, treatment, and prognosis of bladder cancer.

**Keywords:** Bladder cancer; long non-coding RNA (lncRNA); microRNA (miRNA); messenger RNA (mRNA); The Cancer Genome Atlas (TCGA)

Submitted Jul 08, 2019. Accepted for publication Nov 29, 2019.

doi: 10.21037/tcr.2019.12.27

View this article at: <http://dx.doi.org/10.21037/tcr.2019.12.27>

## Introduction

Bladder cancer is a malignant tumor originating from the bladder uroepithelium. It is one of the most common malignant tumors in the urinary system and seriously

endangers human health. The incidence of bladder cancer in the world ranks 11th among all malignant tumors, among which the incidence rate in males is 9.0/100,000, ranking 7th among male malignant tumors. In China, the incidence

of bladder cancer in men has exceeded that of prostate cancer, ranking first among malignant tumors in the male urinary system (1) (Standard for Diagnosis and Treatment of Bladder Cancer, 2018 Edition).

In recent years, great progress has been made in the study of bladder cancer, but its mechanisms are still unclear. Studies have shown that gene mutation and abnormal regulation of gene expression play an important role in the process of tumorigenesis and progression, and also play an important role in the occurrence and development of bladder cancer (2,3). MicroRNA (miRNA), for example, has been widely shown to be involved in the development of bladder cancer (4). Compared with miRNAs, there are few studies on newly discovered long non-coding RNA (lncRNA). lncRNA is a non-coding RNA transcript with a length of more than 200 bp, which does not encode proteins. lncRNA was once thought to have no biological functions and to be a “noise” in transcription. However, recent studies have shown that lncRNA plays an important role in epigenetic regulation, transcriptional regulation, and post-transcriptional regulation by acting as a signaling molecule, bait molecule, scaffold molecule, or guide molecule in the form of RNA, and participates in nearly all physiological and pathological processes of organisms. The abnormal expression of lncRNAs is closely related to the occurrence, development, and metastasis of tumors (5,6). The effect of abnormal expression of lncRNA in bladder cancer tissues on the proliferation, invasion, and migration of bladder cancer cells, and its guiding significance for clinical diagnosis and prognosis have been reported (7,8). miRNA is a non-coding single-stranded RNA encoded by endogenous genes. Its length is 22 bp. It can regulate the expression of mRNA through the microRNA response element (MRE) on the mRNA (9). Each miRNA can regulate the transcription and translation of multiple RNA, and each MRE can interact with multiple miRNAs (10,11).

Salmena *et al.* (12) proposed the endogenous competitive non-coding ceRNA hypothesis, indicating that RNA transcripts can communicate with each other through microRNA response elements (MRES), thus affecting the expression level of various RNAs, which plays a crucial role in tumorigenesis. Although progress has been made in the study of bladder cancer, the detection of specific lncRNAs associated with bladder cancer, survival analysis, and other clinically significant lncRNAs have not been reported. Therefore, we attempted to extract sample data from the Cancer Genome Atlas (TCGA) database, conduct differential expression, GO, KEGG, and survival analysis.

The aim of our study was to analyze the molecular functions of the integrated lncRNA itself and the related signaling pathways, and to find the related regulatory molecules. We hope our findings can provide a reliable theoretical basis for exploring the molecular mechanism of bladder cancer and finding molecular targets for clinical diagnosis or treatment.

## Methods

### *Data acquisition of bladder cancer and its adjacent samples*

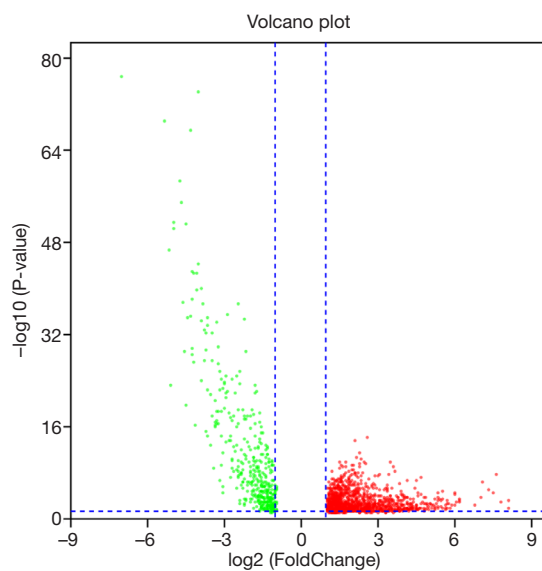
All the data in this study were collected from the TCGA database until January 28, 2016, and included 408 cases of expression of mRNA, 407 cases of expression of miRNAs, and 412 cases of clinical samples, along with 19 matched samples of cancer tissues and adjacent normal tissues. After eliminating the duplicate data and ultra-low expression data, only part of the data information and incomplete clinical information were recorded in the follow-up research.

### *Screening of lncRNAs, miRNAs, and mRNAs differentially expressed and significantly surviving in bladder cancer tissues*

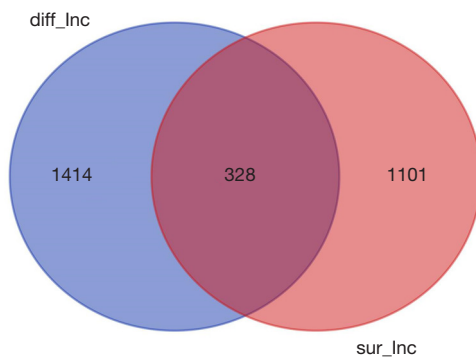
The data of RNA expression profiles were collected, and the differentially expressed lncRNAs, miRNAs, and RNAs were analyzed by R language edgeR package. The differentially expressed lncRNAs, miRNAs and mRNAs whose absolute value of fold change (FC) was more than 2, and whose false discovery rate (FDR) was less than 0.05, were screened. At the same time, the volcano maps were drawn by ggplot2 package, and the differentially expressed lncRNAs, miRNAs, and mRNA were obtained. Survival analysis was then conducted by using R language Survival package, and lncRNAs, miRNAs, and mRNAs with significant differences in expression and survival analysis were screened out.

### *Correlation analysis between lncRNA and miRNA-RNA, and construction of a ceRNA network in bladder cancer*

The expression data of differentially expressed and surviving significant RNA, lncRNA, and miRNAs were correlated with corresponding expression data (i.e., differentially expressed and surviving significant RNA corresponding to lncRNA and miRNA), and the correlation coefficient between them was calculated. The absolute value of the correlation coefficient equal to or more than 0.5 represented a significant correlation.



**Figure 1** Differential lncRNA volcano map of bladder cancer (red for up-regulation, green for down-regulation). lncRNA, long non-coding RNA.



**Figure 2** Venn diagram of differentially expressed lncRNAs and survival analysis of significant lncRNAs in bladder cancer. lncRNA, long non-coding RNA.

The interacting miRNAs were matched according to the miR-code database (<http://www.mircode.org/>) using the differentially expressed and surviving significant lncRNA and miRNAs from the above analysis (13). Target Scan ([http://www.targetscan.org/vert\\_72/](http://www.targetscan.org/vert_72/)), miRDB (<http://mirdb.org/>), and miRTarBase (<http://mirtarbase.mbc.nctu.edu.tw/php/index.php>) databases were used to predict the target genes of miRNAs (14-16), and the target genes present in all three databases were screened. Finally, a subset of sections was obtained from the differentially expressed and surviving target genes

screened, and the target genes differentially expressed and significantly surviving from earlier analysis. Based on the above-obtained lncRNA, miRNAs, and RNA, a ceRNA network between lncRNA and miRNAs was constructed using Cytoscape 3.7.1, and Gene Ontology (GO) annotation and Kyoto Encyclopedia of Genes Genomes (KEGG) enrichment analysis of the target genes in the network were carried out.

#### *Enrichment analysis and survival analysis*

The differentially expressed and surviving significant mRNAs, lncRNAs, significantly correlated mRNAs and miRNAs, along with significantly correlated mRNAs were analyzed by GO functional annotation and KEGG pathway enrichment, respectively. Combined with the survival data of patients with bladder cancer in TCGA, the K-M curve method was used to analyze the survival of patients with bladder cancer using the R language Survival package. The relationship between lncRNAs, miRNAs, and mRNAs and the prognosis of bladder cancer was analyzed.  $P < 0.05$  was considered to have statistical significance.

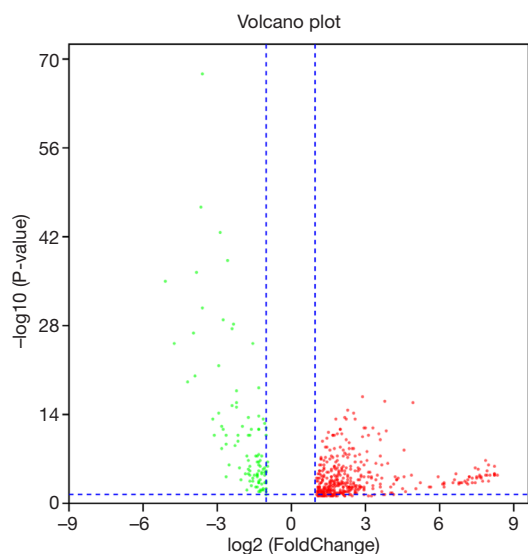
## **Results**

### *Differentially expressed and significantly surviving lncRNAs in bladder cancer*

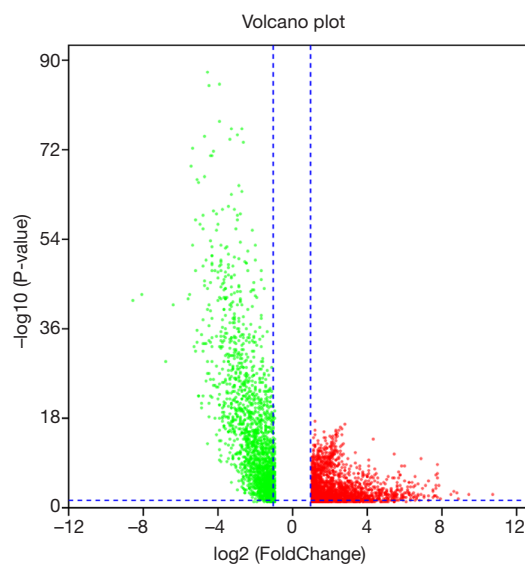
lncRNA was extracted from the expression matrix file downloaded from the TCGA database, and 1,742 differentially expressed lncRNAs were screened according to the criteria of  $|\log FC| (>2)$ ,  $FDR < 0.05$  using R language edgeR package. As shown in the volcano map (Figure 1), there were 1,292 up-regulated lncRNAs and 450 down-regulated lncRNAs. Survival analysis of differentially expressed lncRNAs showed that 328 differentially expressed and significantly analyzed lncRNAs were screened. As shown in the Venn diagram (Figure 2), 260 of them were up-regulated, and 68 were down-regulated.

### *Differentially expressed and significantly surviving miRNAs in bladder cancer*

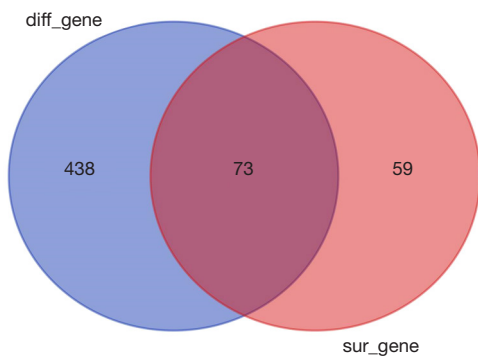
To study the direct relationship between lncRNA and miRNAs, the differentially expressed miRNAs of bladder cancer in the TCGA database were analyzed. In total, 511 differentially expressed miRNAs were identified according to the screening criteria of  $|\log FC| (>2)$  and  $FDR < 0.05$ , of which 406 were differentially up-regulated, and 105 were



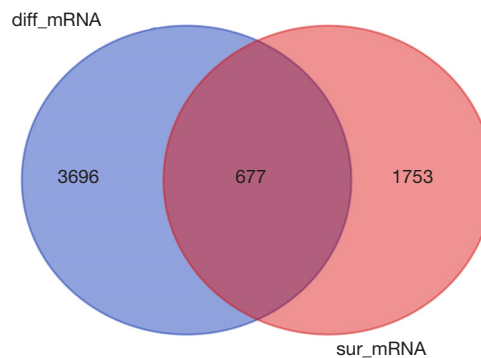
**Figure 3** Volcano map of differential miRNAs in bladder cancer (red for up-regulation and green for down-regulation). miRNA, microRNA.



**Figure 5** Volcano map of differential mRNAs in bladder cancer (red for up-regulation and green for down-regulation). mRNA, messenger RNA.



**Figure 4** Differentially expressed miRNAs in bladder cancer and significantly surviving miRNAs in the Venn diagram. miRNA, microRNA.



**Figure 6** Venn diagram of differentially expressed mRNAs in bladder cancer and significantly expressed mRNAs in survival analysis. mRNA, messenger RNA.

differentially down-regulated (*Figure 3*). By performing K-M survival analysis on differentially expressed miRNAs, a total of 73 differentially expressed and surviving miRNAs were obtained (*Figure 4*), of which 41 were up-regulated and 32 were down-regulated.

#### *mRNA differentially expressed and significantly survived in bladder cancer*

Similarly, according to the above method, the mRNA expression profiles of bladder cancer tissues and adjacent

tissues in the TCGA database were compared, and differential mRNAs with  $|\log FC| \geq 2$  and  $FDR < 0.05$  were screened. The results showed that there were 4,373 differentially expressed mRNAs in bladder cancer tumor tissues compared with normal tissues (*Figure 5*), of which 2,254 were up-regulated mRNA and 2,119 were down-regulated mRNA. The differentially expressed mRNA was subjected to survival analysis, and 677 mRNAs with significant differences and survival were screened (*Figure 6*), of which 246 were up-regulated and 431 were down-regulated.

### *Construction of ceRNA network of lncRNA-miRNA-mRNA in bladder cancer*

To further understand the role of differentially expressed lncRNAs in bladder cancer, we performed a correlation analysis between lncRNAs, miRNAs and mRNAs with variability of expression and significant survival. With 328 differentially expressed and significantly surviving lncRNAs and 73 miRNAs, the lncRNA-miRNA-mRNA ceRNA network was constructed. The results showed that 45 lncRNAs were involved in the regulation of 14 miRNAs (Table 1). After a subset of sections was obtained from 14 miRNA-regulated target genes and differentially expressed and surviving mRNAs, 29 target genes regulated by miRNAs were identified. Thus, 45 lncRNAs, 14 miRNAs, and 29 mRNAs constitute a direct regulatory relationship of lncRNA-miRNA-mRNA in bladder cancer (Figure 7).

### *GO and KEGG enrichment analysis of mRNA in the lncRNA-miRNA-mRNA ceRNA network of bladder cancer*

To further investigate the effect of mRNA in the relationship of lncRNA, miRN, and mRNA in bladder cancer on the cause and development of bladder cancer, GO and KEGG enrichment analysis was performed on lncRNA (differentially expressed and significantly surviving)-correlated mRNAs, miRNA (differentially expressed and significantly surviving)-correlated mRNAs, and differentially expressed and significantly surviving mRNAs. The results showed that some target genes were enriched in the signaling pathways, including cell adhesion molecules (CAMs), MAPK signaling pathway, and PI3K Akt signaling pathway. MAPK signaling pathway has been associated with promoting cell proliferation (17,18), while PI3K-Akt signaling pathway has been associated with tumor invasion and metastasis (19). Other pathways have been associated with cancer, adhesion, and abnormality in gene transcription (Figures 8-10).

Similarly, we performed GO and KEGG enrichment analysis on mRNAs of the lncRNA-miRNA-mRNA ceRNA network, and the GO annotation results are shown in Figure 11 (only the top 10 are visible). The results of KEGG enrichment analysis showed that 29 mRNAs were involved in cancer-related signaling pathways such as fatty acid biosynthesis, gap junction, insulin signaling pathway, prostate cancer, regulation of actin cytoskeleton, and MAPK signaling pathway (Table 2).

## **Discussion**

Bladder cancer is the most common malignant tumor in the urinary system in China, and its incidence and mortality are the first in urinary tract tumors. Advances in medical technology have made great progress in the treatment of bladder cancer. However, the recurrence rate of bladder cancer is high, and the 5-year survival rate is low. Therefore, patients with bladder cancer need to endure long-term detection and treatment. The difficulties of this disease may be due to an insufficient understanding of the potential mechanism and the lack of effective biomarkers. Earlier studies have shown that improper gene regulation may play an important role in bladder cancer (20) and has enormous potential as a biomarker. This article studies the mechanism of action in bladder cancer to better understand and identify biomarkers of bladder cancer. With the development of high-throughput sequencing technology, more and more studies have shown that lncRNA plays a crucial role in the development of cancer (2,3), and that the expression of lncRNA is closely related to the progress of different cancers. Studies have shown that lncRNA TSLNC8 can interact with TKT and STAT3, thereby regulating the phosphorylation levels of STAT3-Tyr705 and STAT3-Ser727 and the transcriptional activity of STAT3, leading to the inactivation of the IL-6/STAT3 signaling pathway and suppressing the development of tumors (21). At present, the research focus of related lncRNA in urinary tract tumors at home and abroad is focused on prostate cancer. The related lncRNAs include prostate cancer gene expression marker 1 (PCGEM1), prostate-specific antigen 3 (DD3/PCA3), prostate non-coding RNA1 (PRNCR1), and others. In bladder tumors, there are more studies on urothelial carcinoma-associated gene 1 (UCA1) and imprinted gene H19, and more lncRNA studies are still in their infancy. In this study, a series of lncRNAs were screened using public data of RNA expression profiles from TCGA database. Target gene enrichment analysis revealed correlations with pathways such as the cGMP-PKG signaling pathway.

As an area of intense research, miRNA has been studied in relation to various tumors, including bladder cancer. It has been proven that miR-141, miR-200c, miR-30b, miR-370, miR-1236, and miRNAs like miR-144, miR-223, and miR-let-7, etc. (22-25) play a key role in the proliferation, invasion, metastasis, apoptosis, and prognosis evaluation of bladder cancer. Yonemori *et al.* (26) found that by down-regulating miR-139-5p or miR-139-3p to enhance the migration and invasion of bladder cancer cells, and directly

**Table 1** Results of the correlation between lncRNA and miRNA in bladder cancer

miRNA	lncRNA	Q-value	R
hsa-miR-217-5p	SMC2-AS1	8.71E-262	0.974328
hsa-miR-216a-5p	SMC2-AS1	6.01E-261	0.974076
hsa-miR-204-5p	AC021242.3	3.32E-199	0.946584
hsa-miR-192-5p	HNF1A-AS1	2.59E-141	0.893255
hsa-miR-99a-5p	MIR99AHG	5.49E-130	0.877411
hsa-miR-99a-3p	MIR99AHG	7.44E-106	0.834522
hsa-let-7c-5p	MIR99AHG	4.28E-101	0.824251
hsa-let-7c-3p	MIR99AHG	2.50E-88	0.792994
hsa-miR-125b-2-3p	MIR99AHG	2.18E-81	0.773362
hsa-miR-504-5p	AC008537.3	2.26E-73	0.747935
hsa-miR-217-5p	AC117489.1	2.48E-72	0.744398
hsa-miR-216a-5p	AC117489.1	5.67E-72	0.74316
hsa-miR-125b-5p	MIR99AHG	1.25E-69	0.734916
hsa-miR-100-5p	MIR100HG	1.42E-53	0.668764
hsa-miR-133b	ADAMTS9-AS2	1.70E-48	0.643551
hsa-miR-133a-3p	ADAMTS9-AS2	3.79E-44	0.619897
hsa-miR-143-3p	ADAMTS9-AS2	2.28E-42	0.609598
hsa-miR-200a-3p	AL390719.2	2.13E-41	0.603821
hsa-miR-145-3p	ADAMTS9-AS2	1.52E-40	0.598635
hsa-miR-192-5p	SATB2-AS1	1.83E-39	0.591936
hsa-miR-504-5p	LINC01152	3.75E-37	0.577061
hsa-miR-133a-3p	CARMN	3.00E-36	0.571033
hsa-miR-323a-3p	VLDLR-AS1	5.78E-36	0.569108
hsa-miR-431-5p	VLDLR-AS1	1.48E-35	0.566324
hsa-miR-504-5p	AP001107.5	6.03E-35	0.562112
hsa-miR-758-5p	VLDLR-AS1	6.35E-35	0.561957
hsa-miR-377-5p	VLDLR-AS1	7.11E-35	0.561616
hsa-miR-133b	AP000892.3	2.96E-34	0.557273
hsa-miR-133a-3p	ADAMTS9-AS1	5.03E-34	0.555639
hsa-miR-654-3p	VLDLR-AS1	6.21E-34	0.55499
hsa-miR-133a-3p	AP000892.3	2.25E-33	0.550976
hsa-miR-379-3p	VLDLR-AS1	2.73E-33	0.550372
hsa-miR-133b	CARMN	7.19E-33	0.547319
hsa-miR-217-5p	AC120049.1	1.92E-32	0.544188
hsa-miR-216a-5p	AC120049.1	4.07E-32	0.541762

**Table 1** (continued)

Table 1 (continued)

miRNA	lncRNA	Q-value	R
hsa-miR-145-3p	CARMN	9.15E-32	0.539135
hsa-miR-145-3p	AP000892.3	1.94E-31	0.536667
hsa-miR-337-3p	VLDLR-AS1	3.14E-30	0.527373
hsa-miR-192-5p	PTPRD-AS1	6.65E-30	0.524818
hsa-miR-411-5p	VLDLR-AS1	1.29E-29	0.52254
hsa-miR-143-3p	AP000892.3	2.22E-29	0.52066
hsa-miR-504-5p	MAGI2-AS3	5.43E-29	0.517546
hsa-miR-379-5p	VLDLR-AS1	1.64E-28	0.51365
hsa-miR-133b	ADAMTS9-AS1	1.91E-28	0.513102
hsa-miR-5579-3p	HNF1A-AS1	5.75E-28	0.509164
hsa-miR-133b	LINC00702	8.72E-28	0.507662
hsa-miR-143-3p	CARMN	2.83E-27	0.50337

lncRNA, long non-coding RNA; miRNA, microRNA.

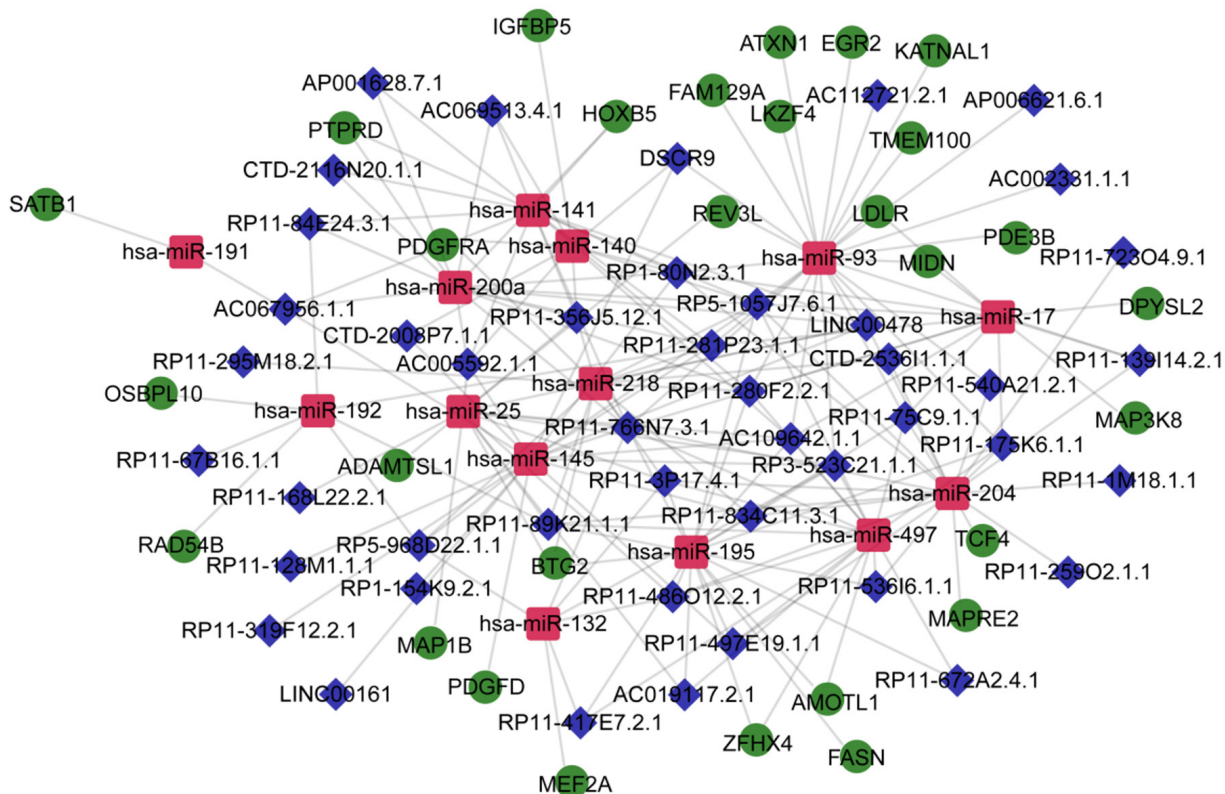
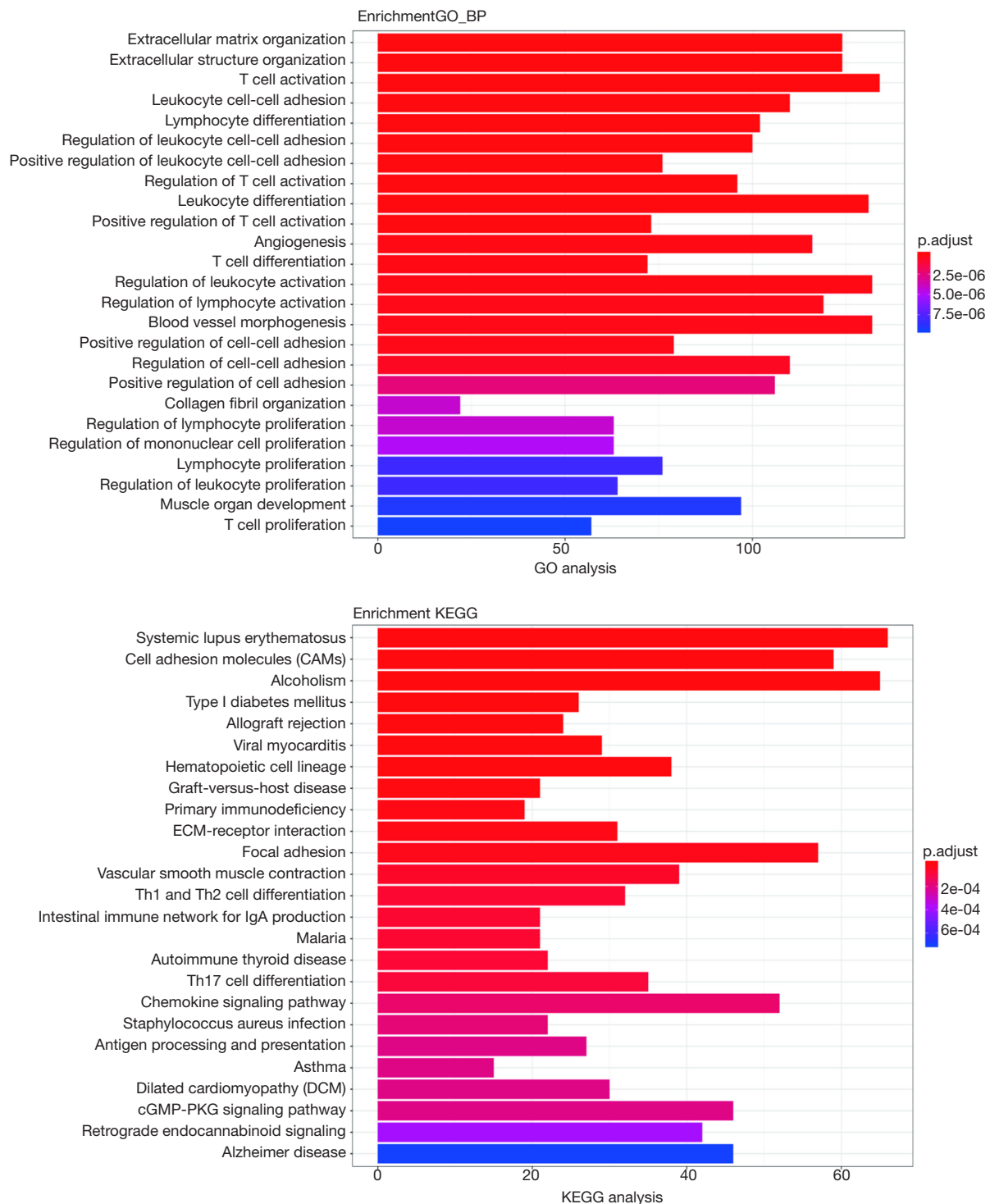
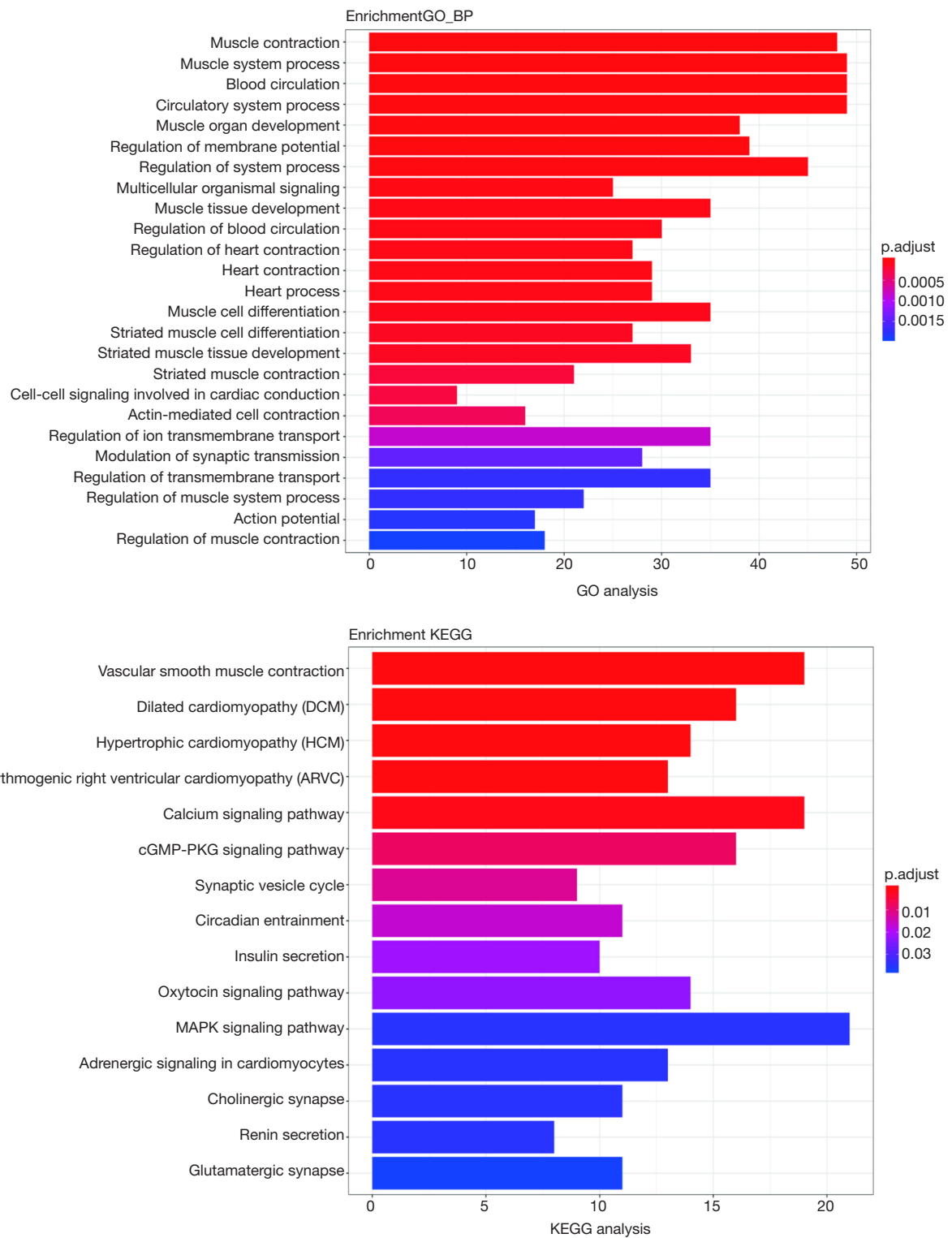


Figure 7 ceRNA regulatory network of lncRNA, miRNA, and mRNA in bladder cancer. Note: the red represents miRNA, the blue represents lncRNA, and the yellow represents mRNA. lncRNA, long non-coding RNA; miRNA, microRNA; mRNA, messenger RNA.

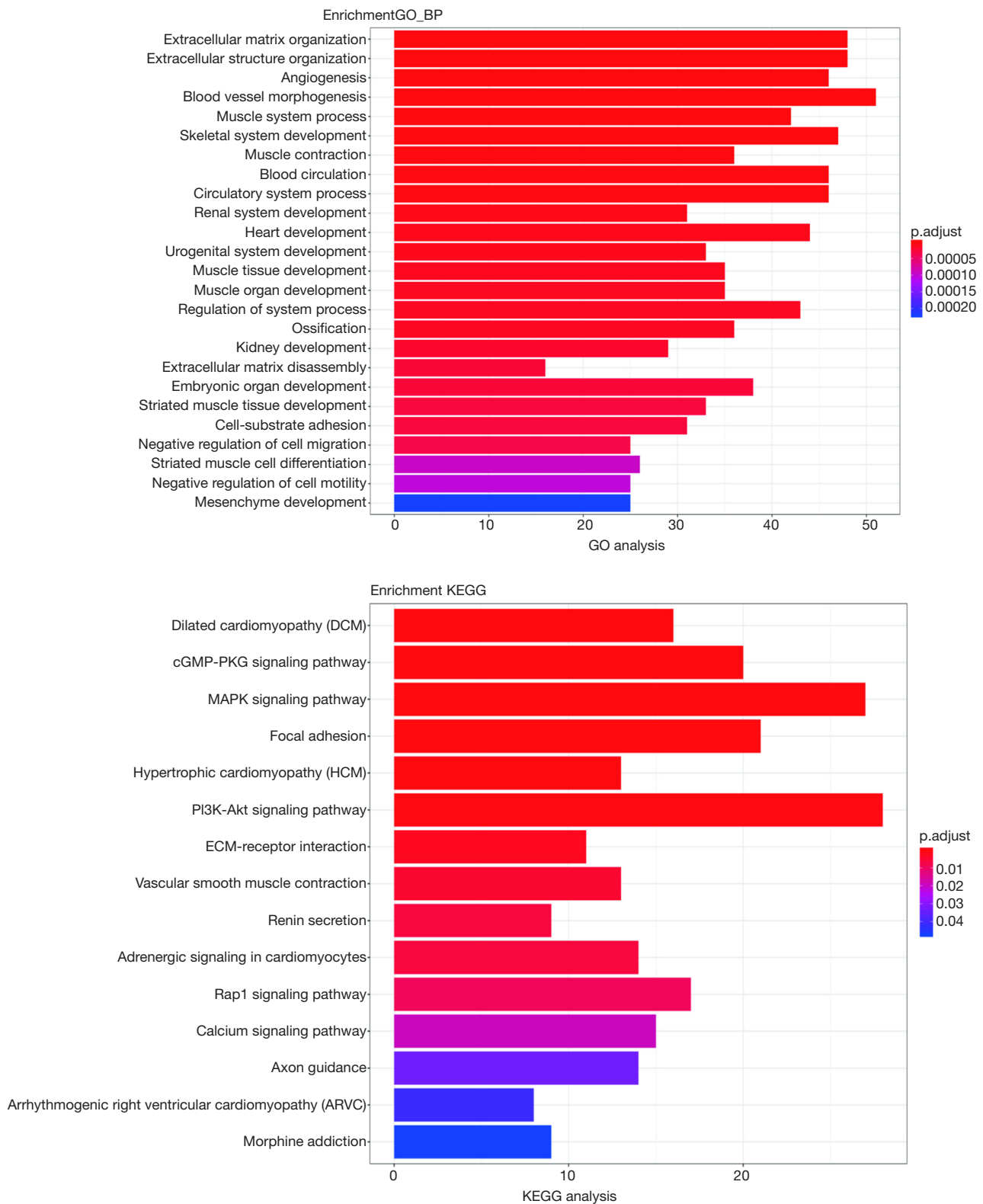


**Figure 8** GO and KEGG enrichment analysis results of lncRNA-related mRNA. GO, Gene Ontology; KEGG, Kyoto Encyclopedia of Genes Genomes; lncRNA, long non-coding RNA; mRNA, messenger RNA.

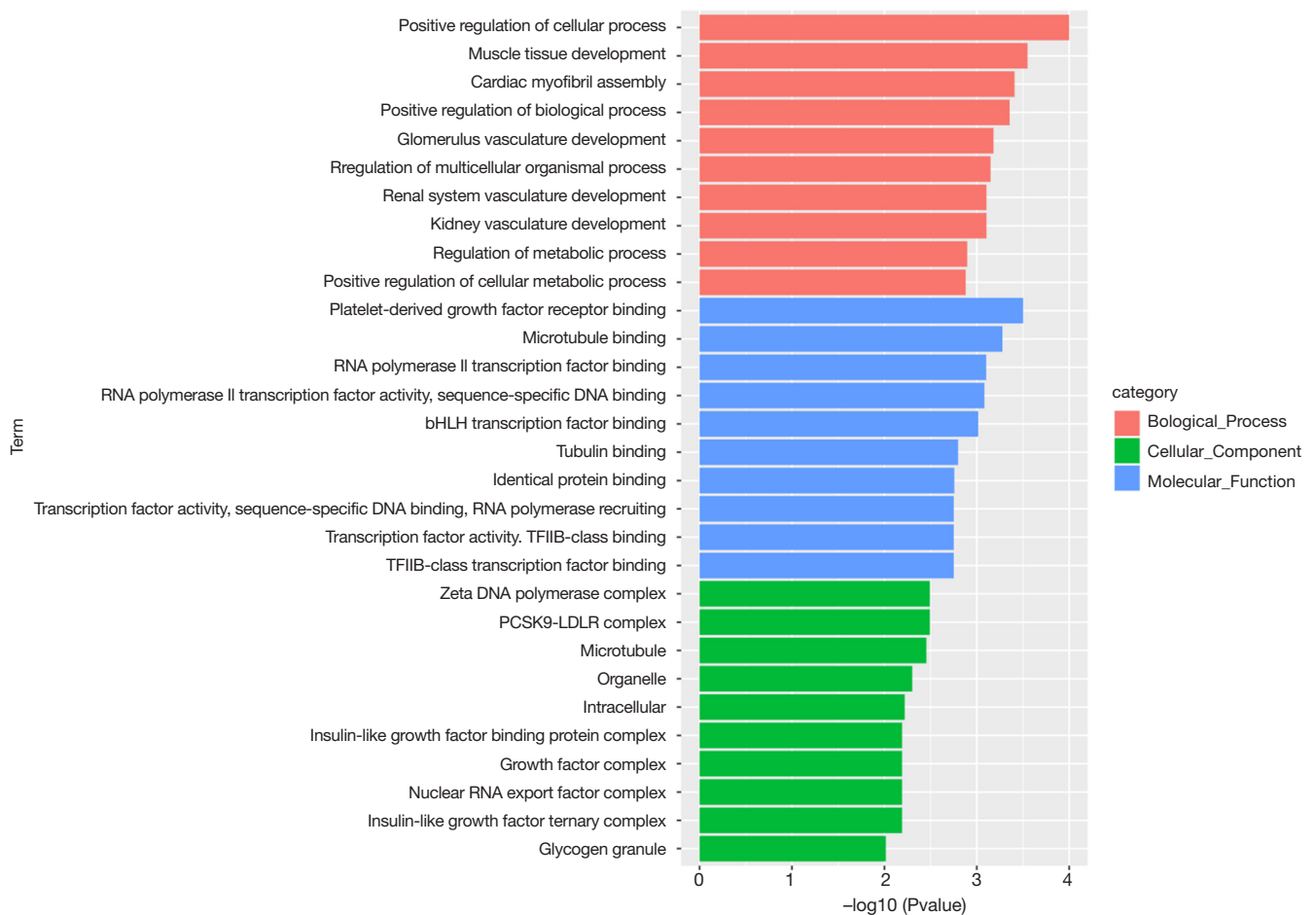




**Figure 9** GO and KEGG enrichment analysis of miRNA-related mRNA. GO, Gene Ontology; KEGG, Kyoto Encyclopedia of Genes Genomes; lncRNA, long non-coding RNA; mRNA, messenger RNA.



**Figure 10** GO and KEGG enrichment analysis of differentially expressed and significant mRNAs. GO, Gene Ontology; KEGG, Kyoto Encyclopedia of Genes Genomes; mRNA, messenger RNA.



**Figure 11** GO enrichment analysis on mRNAs of the lncRNA-miRNA-mRNA ceRNA network (top 10).GO, Gene Ontology; lncRNA, long non-coding RNA; miRNA, microRNA; mRNA, messenger RNA.

**Table 2** Results of the KEGG enrichment analysis of 29 mRNAs in a bladder cancer ceRNA network

KEGGID	Pathway	Gene symbol	P value
5218	Melanoma	<i>PDGFRA, PDGFD</i>	0.007544
61	Fatty acid biosynthesis	<i>FASN</i>	0.011313
4540	Gap junction	<i>PDGFRA, PDGFD</i>	0.011669
5215	Prostate cancer	<i>PDGFRA, PDGFD</i>	0.011669
4910	Insulin signaling pathway	<i>FASN, PDE3B</i>	0.026779
3440	Homologous recombination	<i>RAD54B</i>	0.051805
4510	Focal adhesion	<i>PDGFRA, PDGFD</i>	0.052877
4144	Endocytosis	<i>LDLR, PDGFRA</i>	0.053353
4810	Regulation of actin cytoskeleton	<i>PDGFRA, PDGFD</i>	0.058698
4010	MAPK signaling pathway	<i>MAP3K8, PDGFRA</i>	0.088056

KEGG, Kyoto Encyclopedia of Genes Genomes.

target MMP11, which can be used as a prognostic marker for bladder cancer patients. In this study, bioinformatics methods were used to identify several miRNAs with differential expression and significant survival, such as *MAP3K8* and *PDGFRA*, using the public resources of bladder cancer in TCGA database. Further enrichment analysis showed that these miRNA-related mRNAs were enriched in signaling pathways such as the MAPK signaling pathway, and this finding can provide a certain data foundation for further research.

The proposed ceRNA hypothesis provides an important guiding direction for the pathogenesis of tumors and also offers a new theoretical basis for the diagnosis and treatment of tumors. The interaction of RNA molecules like lncRNA, miRNA, and mRNA has important biological significance. Studies have shown that lncRNA, miRNA, and mRNA are in a state of equilibrium in the ceRNA network. Once this balanced microenvironment state is broken, cancer disease ensues (27). Studies have confirmed that lncRNA-regulated ceRNA networks have important regulatory effects in cancers such as gastric cancer, liver cancer, and prostate cancer (28-31). In this study, a ceRNA network consisting of 45 lncRNAs, 14 miRNAs, and 29 mRNAs was constructed by RNA omics data from TCGA database, which provided a basis for further experimental verification.

The inadequacy of this study is that all the data consist of changes in RNA levels provided by TCGA, with no data from the proteome, and thus we were unable to analyze mRNA at the protein level in ceRNA networks. In the next study, we will conduct quantitative analysis and experiments such as immunohistochemistry or Western blotting to further explore and verify the results of this study.

## Conclusions

In summary, we performed a differential analysis of lncRNA, miRNA, and mRNA data in the TCGA database, and combined them with survival analysis to screen out those lncRNAs, miRNAs, and mRNAs that were differentially expressed and significantly proliferated. The correlation between lncRNA, miRNA, and mRNA was also analyzed in pairs, and a ceRNA network was constructed to provide a potential target for the diagnosis, treatment, and prognosis evaluation of bladder cancer.

## Acknowledgments

*Funding:* None.

## Footnote

*Conflicts of Interest:* The authors have completed the ICMJE uniform disclosure form (available at <http://dx.doi.org/10.21037/tcr.2019.12.27>). The authors have no conflicts of interest to declare.

*Ethical Statement:* The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

*Open Access Statement:* This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

## References

1. Chen W, Zheng R, Baade PD, et al. Cancer statistics in China, 2015. *CA Cancer J Clin* 2016;66:115-32.
2. Goebell, PJ, Knowles MA. Bladder cancer or bladder cancers? Genetically distinct malignant conditions of the urothelium. *Urol Oncol* 2010;28:409-28.
3. Rentsch CA, Müller DC, Ruiz C, et al. Comprehensive Molecular Characterization of Urothelial Bladder Carcinoma: A Step Closer to Clinical Translation? *Eur Urol* 2017;72:960-1.
4. He L, Hannon GJ. MicroRNAs: small RNAs with a big role in gene regulation. *Nat Rev Genet* 2004;5:522-31.
5. Qiu MT, Hu JW, Yin R, et al. Long noncoding RNA: an emerging paradigm of cancer research. *Tumour Biol* 2013;34:613-20.
6. Kung JT, Colognori D, Lee JT. Long noncoding RNAs: past, present, and future. *Genetics* 2013;193:651-69.
7. Martens-Uzunova ES, Böttcher R, Croce CM, et al. Long Noncoding RNA in Prostate, Bladder, and Kidney Cancer. *Eur Urol* 2014;65:1140-51.
8. Zhang Q, Su M, Lu G, et al. The complexity of bladder cancer: long noncoding RNAs are on the stage. *Mol Cancer* 2013;12:101.
9. Ye W, Lv Q, Wong CK, et al. The effect of central loops in miRNA: MRE duplexes on the efficiency of miRNA-

- mediated gene regulation. *Plos One* 2008;3:e1719.
10. Amirkhah R, Schmitz U, Linnebacher M, et al. MicroRNA-mRNA interactions in colorectal cancer and their role in tumor progression. *Genes Chromosomes Cancer* 2015;54:129-41.
  11. Wang C, Hu DZ, Liu JZ. Identification of critical TF-miRNA-mRNA regulation loops for colorectal cancer metastasis. *Genet Mol Res* 2015;14:5485-95.
  12. Salmena L, Poliseno L, Tay Y, et al. ceRNA hypothesis: The Rosetta Stone of a hidden RNA language? *Cell* 2011;146:353-8.
  13. Jeggari A, Marks DS, Larsson E. Larsson, miRcode: a map of putative microRNA target sites in the long non-coding transcriptome. *Bioinformatics* 2012;28:2062-3.
  14. Hsu SD, Lin FM, Wu WY, et al. miRTarBase: a database curates experimentally validated microRNA-target interactions. *Nucleic Acids Res* 2011;39:D163-169.
  15. Zhang C, Zhang CD, Ma MH, et al. Three-microRNA signature identified by bioinformatics analysis predicts prognosis of gastric cancer patients. *World J Gastroenterol* 2018;24:1206-15.
  16. Riffo-Campos ÁL, Riquelme I, Brebi-Mieville P. Tools for Sequence-Based miRNA Target Prediction: What to Choose? *Int J Mol Sci* 2016;17:E1987.
  17. Hao YQ, Zheng HX. Study on the Mechanism of MAPK/ERK Signaling Pathway in Tumor Therapy. *China Journal of Leprosy and Skin Diseases* 2012;28:490-3.
  18. Chrzanowska-Wodnicka M. Regulation of angiogenesis by a small GTPase Rap1. *Vascul Pharmacol* 2010;53:1-10.
  19. Park JH, Kim JJ, Bae YS. Involvement of PI3K-AKT-mTOR pathway in protein kinase CKII inhibition-mediated senescence in human colon cancer cells. *Biochem Biophys Res Commun* 2013;433:420-5.
  20. Knowles MA, Hurst CD. Molecular biology of bladder cancer: new insights into pathogenesis and clinical diversity. *Nat Rev Cancer* 2015;15:25.
  21. Zhang J, Li Z, Liu L, et al. Long noncoding RNA TSLNC8 is a tumor suppressor that inactivates the interleukin-6/STAT3 signaling pathway. *Hepatology* 2018;67:171-87.
  22. Mahdavinezhad A, Mousavi-Bahar SH, Poorolajal J, et al. Evaluation of miR-141, miR-200c, miR-30b Expression and Clinicopathological Features of Bladder Cancer. *Int J Mol Cell Med* 2015;4:32-9.
  23. Wang C, Chen Z, Ge Q, et al. Up-regulation of p21WAF1/CIP1 by miRNAs and its implications in bladder cancer cells. *FEBS Lett* 2014;588:4654-64.
  24. Aygeris M, Mavridis K, Tokas T, et al. Uncovering the clinical utility of miR-143, miR-145 and miR-224 for predicting the survival of bladder cancer patients following treatment. *Carcinogenesis* 2015;36:528-37.
  25. Ichimi T, Enokida H, Okuno Y, et al. Identification of novel microRNA targets based on microRNA signatures in bladder cancer. *Int J Cancer* 2009;125:345-52.
  26. Yonemori M, Seki N, Yoshino H, et al. Dual tumor-suppressors miR-139-5p and miR-139-3p targeting matrix metalloprotease 11 in bladder cancer. *Cancer Sci* 2016;107:1233-42.
  27. Jeggari A, Marks DS, Larsson E. miRcode: a map of putative microRNA target sites in the long non-coding transcriptome. *Bioinformatics* 2012;28:2062-3.
  28. Poliseno L, Salmena L, Zhang J, et al. A coding-independent function of gene and pseudogene mRNAs regulates tumour biology. *Nature* 2010;465:1033-8.
  29. Xia T, Liao Q, Jiang X, et al. Long noncoding RNA associated-competing endogenous RNAs in gastric cancer. *Sci Rep* 2014;4:6088.
  30. Zhang R, Guo Y, Ma Z, et al. Long non-coding RNA PTENP1 functions as a ceRNA to modulate PTEN level by decoying miR-106b and miR-93 in gastric cancer. *Oncotarget* 2017;8:26079-89.
  31. Liu XH, Sun M, Nie FQ, et al. Lnc RNA HOTAIR functions as a competing endogenous RNA to regulate HER2 expression by sponging miR-331-3p in gastric cancer. *Mol Cancer* 2014;13:92.

**Cite this article as:** Dai R, Zhou Y, Chen Z, Zou Z, Liu P, Gao X. The analysis of a ceRNA network and the correlation between lncRNA, miRNA, and mRNA in bladder cancer. *Transl Cancer Res* 2020;9(2):869-881. doi: 10.21037/tcr.2019.12.27