



High-mobility group A1 proteins may be involved in estrogen receptor status of breast cancer

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The three types of breast cancer

Breast cancer is one of the most common of cancers in woman. About 316,700 new cases were diagnosed as breast cancer in US woman in 2019 and 41,760 were predicted to die from it (1). Breast cancer has a characteristic of therapy-targeting receptors: hormone receptors (HR) that are estrogen receptor [ER: human ER α protein (NCBI protein accession NP_000116.2) encoding the *ESR1* gene (RefSeq NM_000125.4)], and progesterone receptor (PR). In addition to ER and PR is the tyrosine kinase-type receptor, erythroblastic oncogene B2 (ERBB2), also called HER2. These receptors are important for managing breast cancer in advanced stages as well as early stages. Accordingly, breast cancer is categorized into three subtypes: HR+/ERBB2-; ERBB2+ (HR+ or HR-); and triple-negative (ER-, PR-, ERBB2-). Seventy percent of breast cancer cases respond to anti-estrogen therapy through ER positivity, but one-third develop resistance within 15 years. Mutations of the *ESR1* gene have been frequently found in these cases. ERBB2+ (also called HER2+) is found in 20–25% of breast cancers and a humanized monoclonal antibody raised against ERBB2 called trastuzumab was developed. Estimated overall survival rate was 92% for trastuzumab plus doxorubicin and cyclophosphamide followed by docetaxel treatment. Triple negative (ER-, PR-, ERBB2-) accounts for 10–20% of breast cancers with the worst prognosis of the three types (2).

HMGA1 proteins: old and new proteins in breast cancer

HMGA1 (previously called HMGI/Y) is a member of the high-mobility group (HMG) proteins that were found from their high mobility characteristics during polyacrylamide electrophoresis of non-histone chromatin-associated proteins (3). The oncogenic property for these proteins was reported 35 years ago (4). Since then, proteins related to the HMG box (5), the DNA-binding motif of this protein family, have been found mostly related to cancer (6). HMGI/Y was first implicated as an oncogenic product from studies showing it expressed in a highly malignant phenotype in a ras oncogene transformed rat thyroid epithelial cell line (7) and transformed human breast epithelial cell (8). It was detected in fast proliferating, undifferentiated cells, and also in an aggressive derivative of a relatively benign prostate cell line (9). The growing number HMG proteins led to the novel nomenclature of each member (10). HMGI/Y was renamed as HMGA proteins.

The involvement of HMGA proteins in breast cancer was first documented as growth factor-induced overexpression in a highly metastatic breast cancer cell line compared to a nonmetastatic one (11). HMGA1 was shown to be related to its DNA unwinding properties as an architectural transcription factor (12), in matrix attachment of breast cancer cells (13). HMGA1 proteins have been detected as

a stem cell signature in triple negative breast cancer (14). Silencing of HMGA1 in this type of breast cancer transformed the cells from a mesenchymal-like, spindle-shape to a cuboidal, epithelial-like morphology (15).

The function of HMGA1 proteins in metastasis and stem cell signature have been studied and documented in many previous studies in many cancer types as well as breast cancer shown above. However, its contribution to estrogen-independence, tumorigenesis, and prognosis in breast cancer need to be further evaluated. Gorbounov *et al.* (16) have checked HMGA1 immunoreactivity in primary breast cancer in a large cohort of Asian women and compared it with two Western cohorts. Their staining followed current guidelines using an ER antibody raised against the C terminus (550 aa~) of human ER α protein. Immunoreactivity of HMGA1 was detected in ER-negative primary breast cancer tumors in their Korean cohort. Intriguingly, the overall survival was predicted to decrease in the high *ESR1* gene expression group by the Molecular Taxonomy of Breast Cancer International Consortium (METABRIC) cohort. It was predicted that HMGA1 proteins repress *ESR1* gene expression where HMGA1 itself is induced by estrogen (11). Thus, continuous exposure of breast cancer cells to estrogen may induce HMGA1 proteins that in turn represses its receptor. This implicates an additional role of HMGA1 proteins in ER-positive breast cancer and needs further investigation at the molecular level.

Perspectives

One clue may come from the emerging findings of HMGA1 proteins as sequence-specific RNA-binding proteins. The DNA-binding AT-hooks found in the HMG-box of HMGA1 proteins make it unique among the HMG proteins. The first evidence of its sequence-specific RNA-binding function came from its involvement in sporadic Alzheimer's disease, where it induces aberrant exon 5 skipping by preventing the release of U1 snRNP from the 5' splice site adjacent the HMGA1a RNA-binding site (17). These AT-hooks contain glycine-arginine-proline (G-R-P) tripeptide cores flanked by basic amino acids making its RNA-binding a magnitude higher than its original DNA-binding feature (18). It would be interesting if there is a specific protein modification of the AT-hook of HMGA1 proteins that differentiates their RNA- and DNA-binding, as well as the specific signal that induces each modification, since HMGA1 proteins are one of

the most highly modified protein in the human cell (19). Other examples of sequence-specific RNA-binding for HMGA1 proteins have been reported for 7SK small nuclear RNA (20), for alternative splicing of the HIV RNA genome (21), and recent reports for regulating alternative splicing of the *ESR1* gene (22,23) in ER-positive breast cancer cell lines. The protein resulting from HMGA1a-induces alternative splicing of the *ESR1* gene is an exon-skipped form of exon 1 between non-coding exon E/F and exon 2 that lacks the N terminal domain, designated as ER α 46. ER α 46 suppresses the estrogen-independent activation function 1 (AF1) activity of the full length ER α (24). ER α 46 has an intact C terminal domain that is recognized by the ER antibody that is used to detect ER positivity. Further studies similar to Gorbounov *et al.* (16) that analyze the correlation of HMGA1 proteins and ER α 46 in hormone-resistant ER-positive breast cancer may help us find the mechanism of resistance not based on *ESR1* mutations.

Taken together, the report of Gorbounov *et al.* (16) re-opens a field of research to decipher the role and therapeutic target related to ER α regulation by HMGA1 proteins in breast cancer.

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Footnote

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References

- DeSantis CE, Ma J, Gaudet MM, et al. Breast cancer statistics, 2019. *CA Cancer J Clin* 2019;69:438-51.
- Boyle P. Triple-negative breast cancer: epidemiological considerations and recommendations. *Ann Oncol* 2012;23 Suppl 6:vi7-12.
- Goodwin GH, Sanders C, Johns EW. A new group of chromatin-associated proteins with a high content of acidic and basic amino acids. *Eur J Biochem* 1973;38:14-9.
- Giancotti V, Berlingieri MT, DiFiore PP, et al. Changes in nuclear proteins on transformation of rat epithelial thyroid cells by a murine sarcoma retrovirus. *Cancer Res* 1985;45:6051-7.
- Thomas JO. HMG1 and 2: architectural DNA-binding proteins. *Biochem Soc Trans* 2001;29:395-401.
- Wunderlich V, Bottger M. High-mobility-group proteins and cancer--an emerging link. *J Cancer Res Clin Oncol* 1997;123:133-40.
- Giancotti V, Pani B, D'Andrea P, et al. Elevated levels of a specific class of nuclear phosphoproteins in cells transformed with v-ras and v-mos oncogenes and by cotransfection with c-myc and polyoma middle T genes. *EMBO J* 1987;6:1981-7.
- Reeves R, Edberg DD, Li Y. Architectural transcription factor HMGI(Y) promotes tumor progression and mesenchymal transition of human epithelial cells. *Mol Cell Biol* 2001;21:575-94.
- Bussemakers MJ, van de Ven WJ, Debruyne FM, et al. Identification of high mobility group protein I(Y) as potential progression marker for prostate cancer by differential hybridization analysis. *Cancer Res* 1991;51:606-11.
- Bustin M. Revised nomenclature for high mobility group (HMG) chromosomal proteins. *Trends Biochem Sci* 2001;26:152-3.
- Holth LT, Thorlacius AE, Reeves R. Effects of epidermal growth factor and estrogen on the regulation of the HMG-I/Y gene in human mammary epithelial cell lines. *DNA Cell Biol* 1997;16:1299-309.
- Reeves R, Nissen MS. Cell cycle regulation and functions of HMG-I(Y). *Prog Cell Cycle Res* 1995;1:339-49.
- Liu WM, Guerra-Vladusic FK, Kurakata S, et al. HMG-I(Y) recognizes base-unpairing regions of matrix attachment sequences and its increased expression is directly linked to metastatic breast cancer phenotype. *Cancer Res* 1999;59:5695-703.
- Ben-Porath I, Thomson MW, Carey VJ, et al. An embryonic stem cell-like gene expression signature in poorly differentiated aggressive human tumors. *Nat Genet* 2008;40:499-507.
- Shah SN, Cope L, Poh W, et al. HMGA1: a master regulator of tumor progression in triple-negative breast cancer cells. *PLoS One* 2013;8:e63419.
- Gorbounov M, Carleton NM, Asch-Kendrick RJ, et al. High mobility group A1 (HMGA1) protein and gene expression correlate with ER-negativity and poor outcomes in breast cancer. *Breast Cancer Res Treat* 2020;179:25-35.
- Ohe K, Mayeda A. HMGA1a trapping of U1 snRNP at an authentic 5' splice site induces aberrant exon skipping in sporadic Alzheimer's disease. *Mol Cell Biol* 2010;30:2220-8.
- Filarsky M, Zillner K, Araya I, et al. The extended AT-hook is a novel RNA binding motif. *RNA Biol* 2015;12:864-76.
- Reeves R. Molecular biology of HMGA proteins: hubs of nuclear function. *Gene* 2001;277:63-81.
- Eilebrecht S, Brysbaert G, Wegert T, et al. 7SK small nuclear RNA directly affects HMGA1 function in transcription regulation. *Nucleic Acids Res* 2011;39:2057-72.
- Tsuruno C, Ohe K, Kuramitsu M, et al. HMGA1a is involved in specific splice site regulation of human immunodeficiency virus type 1. *Biochem Biophys Res Commun* 2011;406:512-7.
- Ohe K, Miyajima S, Abe I, et al. HMGA1a induces alternative splicing of estrogen receptor alpha in MCF-7 human breast cancer cells. *J Steroid Biochem Mol Biol* 2018;182:21-6.
- Ohe K, Miyajima S, Tanaka T, et al. HMGA1a induces alternative splicing of the estrogen receptor-alpha gene by trapping U1 snRNP to an upstream pseudo-5' splice site. *Front Mol Biosci* 2018;5:52.
- Flouriort G, Brand H, Denger S, et al. Identification of a new isoform of the human estrogen receptor-alpha (hER-alpha) that is encoded by distinct transcripts and that is able to repress hER-alpha activation function 1. *EMBO J* 2000;19:4688-700.

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