Original Article

Identification of significant genes in non-small cell lung cancer by bioinformatics analyses

Xia Ye[#], Qian Gao[#], Jie Wu, Lin Zhou, Min Tao

Department of Oncology, The First Affiliated Hospital of Soochow University, Suzhou, China

Contributions: (I) Conception and design: M Tao, X Ye; (II) Administrative support: M Tao; (III) Provision of study materials or patients: L Zhou; (IV) Collection and assembly of data: J Wu; (V) Data analysis and interpretation: X Ye, Q Gao; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

[#]These authors contributed equally to this work.

Correspondence to: Min Tao. Department of Oncology, The First Affiliated Hospital of Soochow University, No. 188, Shi Zi Road, Suzhou, China. Email: taomin@suda.edu.cn.

Background: Lung cancer is the most malignant cancer featured with undesirable prognosis. It is urgent to identify novel biomarkers to improve both diagnosis and prognosis. The purpose of the study was to identify significant genes involved in lung cancer through bioinformatic methods and reveal potential underlying mechanisms.

Methods: Three datasets GSE19188, GSE27262, GSE118375, containing 122 lung cancer and 96 normal tissues, were available from GEO database. GEO2R and Venn diagram online software were applied to pick out differentially expressed genes (DEGs). Next, we used the Database for Annotation, Visualization and Integrated Discovery (DAVID) to analyze Kyoto Encyclopedia of Gene and Genome (KEGG) pathway and gene ontology (GO) enrichment, followed by protein-protein interaction (PPI) of these DEGs visualized by cytoscape. The MCODE plug-in was performed to construct a module complex of DEGs. In addition, Kaplan-Meier analysis was implemented for analysis of overall survival. To further validate the expression of these genes, Gene Expression Profiling Interactive Analysis (GEPIA) was used.

Results: A total of 149 DEGs were identified, including 127 downregulated genes and 22 upregulated genes. KEGG analysis revealed that the DEGs were mainly enriched in ECM-receptor interaction, Vascular smooth muscle contraction, and PPAR signaling pathway. GO analysis of DEGs showed that significant functional enrichment of angiogenesis, cell adhesion, and vasculogenesis. 13 genes were selected as hub genes based on MCODE, and 11 of 13 genes had a significance. The results of GEPIA were consistent with survival analysis. Furthermore, reanalysis of these genes found they were significantly enriched in ECM-receptor interaction and PI3K-Akt signaling pathway.

Conclusions: We have identified several key genes, which could be potential diagnostic and prognostic biomarker as well as therapy targets.

Keywords: Non-small cell lung cancer (NSCLC); microarray; differentially expressed genes (DEGs); bioinformatics analysis

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Introduction

Lung cancer is the most commonly diagnosed cancer and leading cause of cancer mortality worldwide, which accounts for 11.6% of total cases and 18.4% of the total deaths (1). Based on histological classification, lung cancer is categorized into non-small cell lung cancer (NSCLC, ~85%) and small-cell lung cancer (SCLC, ~15%), the former group is further classified into three common

subtypes, large-cell carcinoma, squamous cell carcinoma, and adenocarcinoma (2,3). Despite progresses achieved in therapies including surgical resection, chemotherapy, radiotherapy, and immunotherapy for NSCLC in recent years, the 5-year survival rate is still low and only 5% (4). Lack of specific molecular biomarker leads to many NSCLC patients were diagnosed at advanced stage, resulting in no long-term survival (5). Encouragingly, with the development of oncogenetics and molecular etiology of lung cancer, great progress has been made in targeted cancer therapy. For example, tyrosine kinase inhibitor (TKI), such as gefitinib and erlotinib, can block the activity of epidermal growth factor receptor (EGFR) reversibly, suppress cell proliferation and transformation, thus improve response rate and prolong survival (6). However, the clinical benefits of these targeted therapies are only restricted to a cohort of NSCLC patients with corresponding targets. Therefore, it is important to further reveal the molecular mechanisms involved in the initiation and progression of NSCLC and to identify the alternated key genes to develop more effective therapies for lung cancer.

Gene chip is a powerful and reliable technologies that can quickly yield quantitative differentially expressed genes (DEGs) and expression profiles by it (7). To date, a large number of microarray data could be explored from the Gene Expression Omnibus public database. With the rapid development of high-throughput sequencing, bioinformatics analysis has been applied in mining the pathophysiological mechanism of different cancers (8-10). In this study, we downloaded 3 NSCLC related mRNA datasets GSE19188, GSE27262, GSE118370 from GEO database to find DEGs, Subsequently, hub genes were found to be associated with survival and further validated when lung tissues compared with adjacent normal tissues. In conclusion, our study can further understand the molecular mechanism of NSCLC and provides potential useful biomarkers for diagnosis, and targeted therapy of NSCLC patients.

Methods

Microarray data

The microarray data GSE19188, GSE27262, GSE118370 used in this study were downloaded from the Gene Expression Omnibus database at NCBI (www.ncbi.nlm.nih.gov/geo/) (11), which is a openly public database. They were all based on the platform of the GPL570 [HG-U133_Plus_2] Affymetrix Human Genome U133 Plus 2.0 Array, which consisted of 91 lung cancer and 65 adjacent normal lung tissue, 25 lung cancer and 25 adjacent paired normal lung tissue, 6 lung adenocarcinoma tissues and 6 paired normal lung tissues, respectively. Data processing and identification of DEGs. The DEGs between NSCLC specimen and normal lung specimen were identified via GEO2R, which is an online tool and can be applied to screen DEGs. |logFC| >2 and adjust P<0.05 were considered as cut-off value. Venn software was applied to detect DEGs among the 3 datasets.

Gene ontology (GO) and pathway enrichment analysis of DEGs

GO and Kyoto Encyclopedia of Genes and Genomes (KEGG) (12) annotations analysis of DEGs gene were performed via the Database for Annotation, Visualization and Integrated Discovery 6.8 (DAVID6.8) (https://david.ncifcrf.gov/) (13). GO analysis is a commonly useful tool to investigate unique biological properties of DEGs that were involved, including biological processes (BP), cellular components (CC) and molecular function (MF). KEGG is an online database to integrate protein interaction network information and deal with disease, metabolism, biological pathways, and drug research. DAVID, as a comprehensive set of functional annotation tool, can integrate public bioinformatics resources and perform biological analyses of genes by clustering algorithm. P<0.05 was considered significant

Protein-protein interaction (PPI) network and module analysis

Search Tool for the Retrieval of Interacting Genes (STRING) database (https://string-db.org) (14) was applied to download the interaction information of human proteins and construct PPI network, then Cytoscape (www.cytoscape.org) (15) was used to visualize PPI network with cut-off criteria of combined score >0.4. In addition, The PPI network modules was analyzed via the Molecular Complex Detection (MCODE) app in Cytoscape based on topology (degree cutoff =2, max. Depth =100, k-core =2, and node score cutoff =0.2).

Survival analysis of crucial genes

Kaplan–Meier plotter (http://kmplot.com/) (16) is a commonly used web tool that is capable to assess the

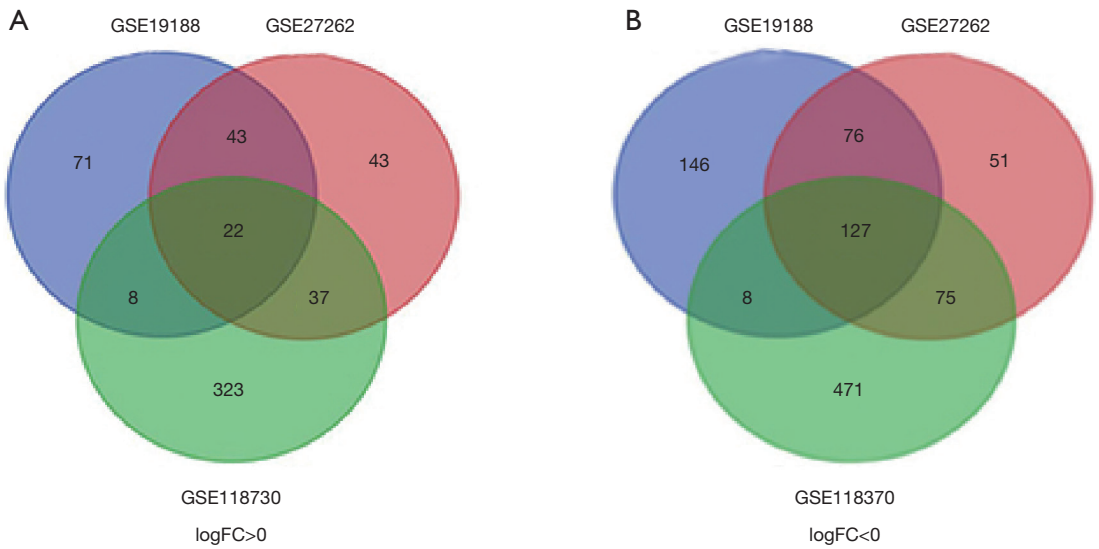

Figure 1 A total of 149 common differentially expressed genes in 3 datasets (GSE19188, GSE27260 and GSE118370) via Venn diagrams software. Different color meant different datasets. (A) Twenty-two differentially expressed genes were up-regulated in three datasets (logFC >0); (B) 127 differentially expressed genes were down-regulated in three datasets (logFC <0).

prognostic values of genes in 21 cancer patients, of which the largest dataset including breast, ovarian, lung, and gastric cancer based on GEO, EGA, and TCGA. According to the level of gene expression (high and low), the NSCLC patients were divided into two groups. The HR with 95% confidence intervals and log rank P value were computed and displayed on each plot.

RNA sequencing expression of hub gene in GEPIA

The Gene Expression Profiling Interactive Analysis (GEPIA) is online database that can analyze RNA sequencing expression. To further validate these significantly correlated genes, the GEPIA was used.

Results

Identification of DEGs in lung cancer

We used GEO2R online tool to extract 501, 474, 749 DEGs in GSE19188, GSE27262, GSE118370, of which 357, 329, 359 downregulated and 144, 145, 390 upregulated DEGs, respectively. Then, Veen diagram software was applied to identify the most reliable DEGs among 3 datasets. As shown in *Figure 1* and *Table 1*, in total, 149 DEGs that met the cut-off criteria were obtained, including 127 down-regulated and 22 were up-regulated.

GO and KEGG pathway analysis of DEGs

To further identify the potential biological functions of these 149 DEGs, DAVID online software was used to analyze GO categories. The results of GO functional enrichment analysis, as shown in *Table 2*, indicated that, as for BP, upregulated DEGs were significantly enriched in collagen catabolic process, sensory perception of sound, G2M transition of mitotic cell cycle, cell division, inner ear morphogenesis, and downregulated DEGs in angiogenesis, cell adhesion, vasculogenesis, single organismal cell-cell adhesion, response to hypoxia; for cell composition (CC) part, upregulated DEGs were particularly involved in centrosome, proteinaceous extracellular matrix, collagen trimer, spindle pole and downregulated genes in membrane raft, proteinaceous extracellular matrix, cell surface, plasma membrane, integral component of plasma membrane, external side of plasma membrane; in the MF section, the upregulated DEGs participated in extracellular matrix binding, serine-type endopeptidase activity and downregulated genes in heparin binding, receptor activity, transformation growth factor beta binding, peroxidase activity. All terms are closely associated with the tumorigenesis and development. On the other hand, KEGG pathway enrichment analysis was performed to analyze the biological functions of these genes. The most enriched KEGG pathways were as follows: ECM–receptor

Table 1 All 149 commonly differentially expressed genes were identified from three profile datasets, including 127 downregulated genes and 22 up-regulated genes in the lung tissues compared to normal tissues

DEGs	Gene names
Up-regulated	*KIF26B*, *CCNB1*, *HMGB3*, *CD24*, *CXCL13*, *GJB2*, *AURKA*, *TFAP2A*, *FERMT1*, *HMMR*, *TMPRSS4*, *HS6ST2*, *SPP1*, *SIX1*, *COL10A1*, *COL11A1*, *UGT8*, *NUF2*, *MMP1*, *NEK2*, *MMP12*, *CENPF*
Down-regulated	*HBA2///HBA1*, *RTKN2*, *EMCN*, *SOX7*, *ADARB1*, *PPP1R14A*, *WISP2*, *MFAP4*, *KCNT2*, *ERG*, *SLC6A4*, *PECAM1*, *KCNK3*, *SYNPO2*, *GIMAP8*, *OGN*, *SCARA5*, *BTNL9*, *PCAT19*, *IGSF10*, *ACVRL1*, *SCGB1A1*, *CDO1*, *CA4*, *SDPR*, *TEK*, *CLIC3*, *GRK5*, *DACH1*, *VGLL3*, *GUCY1A2*, *PALM2-AKAP2///AKAP2*, *STXBP6*, *S1PR1*, *EMP2*, *LYVE1*, *ADAMTS8*, *GDF10*, *LEPROT///LEPR*, *BCHE*, *SPOCK2*, *AKAP12*, *CD36*, *PDE5A*, *LDB2*, *ROBO4*, *SPTBN1*, *CALCRL*, *CAV1*, *PPBP*, *JAM2*, *PTPRB*, *QKI*, *FOXF1*, *ACADL*, *ANKRD29*, *AQP4*, *PIR-FIGF///FIGF*, *ITGA8*, *MT1M*, *TNNC1*, *IL1RL1*, *FAT3*, *MCEMP1*, *HBB*, *FHL1*, *RHOJ*, *THBD*, *KLF4*, *SCN7A*, *FMO2*, *ABCA8*, *MYZAP*, *AOC3*, *SFTPC*, *ADRB1*, *SEMA3G*, *TCF21*, *TGFBR3*, *HHIP*, *ADH1B*, *ARHGEF26*, *ARHGAP6*, *LINC00968*, *ASPA*, *CCL15-CCL14///CCL14*, *FABP4*, *EDNRB*, *SCN4B*, *FCN3*, *MYCT1*, *KANK3*, *STX11*, *LINC00312*, *CCDC85A*, *FAM107A*, *CCBE1*, *PGM5*, *GPX3*, *AGER*, *RGCC*, *VWF*, *MARCO*, *SEMA5A*, *ABI3BP*, *CD93*, *TIE1*, *KIAA1462*, *VIPR1*, *AGTR1*, *EPAS1*, *RAMP3*, *CLIC5*, *SLIT2*, *FHL5*, *ADAMTSL3*, *CLDN18*, *C2orf40*, *CDH5*, *PDK4*, *GPM6A*, *COL6A6*, *ANGPT1*, *SMAD6*, *TMEM100*, *DUOX1*, *AFF3*

interaction, Vascular smooth muscle contraction, PPAR signaling pathway, Adrenergic signaling in cardiomyocytes, cell adhesion molecules (CAMs) and focal adhesion (*Table 3*).

Construction PPI network and modular analysis

To further predict the interaction of the DEGs at the protein level, the PPI network was constructed, in which 102 DEGs were imported into and 47 were not contained totally. The constructed PPI network contained 204 interaction pairs. Subsequently, cytotype MCODE app was employed to identify modules. As displayed in *Figure 2*, the top 1 significant module included 13 central nodes among the 102 nodes. Among 13 central nodes, 7 including *CCNB1*, *AURKA*, *HMMR*, *SPP1*, *NUF2*, *NEK2*, *CENPF* were upregulated and 6 including *LYVE1*, *ROBO4*, *PTPRB*, *VWF*, *TIE1*, *ANGPT1* were downregulated.

Analysis of core genes by the Kaplan Meier plotter and GEPIA

In an attempt to gain insight into association between hub genes and NSCLC patients, Kaplan Meier plotter was utilized to predict the prognostic value of 13 core genes survival data. The result revealed that 11 DEGs had a significant survival while 2 had no significance (P<0.05, *Table 4* & *Figure 3*). *ROBO4* with low expression was associated with better overall survival for NSCLC patients, as well as *PTPRB*, *VWF*, *ANGPT1* (P<0.05). Additionally, high expression of *CCNB1* was associated with poorer overall survival, as well as *CCNB1*, *AURKA*, *HMMR*, *SPP1*, *NUF2*, *NEK2*, *CENPF* (P<0.05). Moreover, to further verify the expression of these DEGs, GEPIA was employed to dig up the 11 gene expression level between lung cancer and normal people. As graphed in *Table 5* & *Figure 4*, notably, the results were in line with the survival analysis above, which imply that the expression levels of the 11 hub genes are particularly associated with clinical prognosis of NSCLC patients and they may play vital roles in the progression of NSCLC.

KEGG pathway enrichment of 11 genes reanalysis

KEGG pathway was re-analyzed to investigate the possible pathway of 11 genes. Enrichment analysis showed that the module genes were mainly associated with ECM-receptor interaction and PI3K-Akt signaling pathway (*Table 6* & *Figure 5*).

Discussion

At present, the diagnosis and treatment of NSCLC is still far from satisfactory, and the number of this case is still rising year by year. It is necessary to investigate the pathogenesis and biomarker of NSCLC to provide effective treatment. Great progress has been made on the mechanism of initiation and development of NSCLC. Many experiments including vitro tumor cell lines, animal tumor models, and patients' tumor model have been done, however, NSCLC demands more comprehensive analysis because the progress of lung cancer is a multi-stage and multi-cause process. Fortunately, with the development of human genome sequencing, the high throughput and associated tumor

Table 2 Gene ontology analysis of differentially expressed genes in lung cancer, including biological processes, cellular components and molecular function

Expression	Category	Term	Count	P value	FDR
Upregulated	GOTERM_BP_DIRECT	GO:0030574~collagen catabolic process	4	6.69E-05	0.088526
	GOTERM_BP_DIRECT	GO:0007605~sensory perception of sound	4	5.82E-04	0.768065
	GOTERM_BP_DIRECT	GO:0000086~G2/M transition of mitotic cell cycle	4	6.34E-04	0.837064
	GOTERM_BP_DIRECT	GO:0051301~cell division	5	8.39E-04	1.104883
	GOTERM_BP_DIRECT	GO:0042472~inner ear morphogenesis	3	0.001902	2.490103
	GOTERM_CC_DIRECT	GO:0005813~centrosome	5	0.001285	1.321892
	GOTERM_CC_DIRECT	GO:0005578~proteinaceous extracellular matrix	4	0.003438	3.501323
	GOTERM_CC_DIRECT	GO:0005581~collagen trimer	3	0.004973	5.029117
	GOTERM_CC_DIRECT	GO:0000922~spindle pole	3	0.006912	6.926107
	GOTERM_CC_DIRECT	GO:0045120~pronucleus	2	0.00804	8.014669
	GOTERM_MF_DIRECT	GO:0050840~extracellular matrix binding	2	0.030374	27.11998
	GOTERM_MF_DIRECT	GO:0004252~serine-type endopeptidase activity	3	0.036114	31.42547
Down regulated	GOTERM_BP_DIRECT	GO:0001525~angiogenesis	13	2.78E-08	4.38E-05
	GOTERM_BP_DIRECT	GO:0007155~cell adhesion	15	1.99E-06	0.003144
	GOTERM_BP_DIRECT	GO:0001570~vasculogenesis	5	5.10E-04	0.800955
	GOTERM_BP_DIRECT	GO:0016337~single organismal cell-cell adhesion	6	5.52E-04	0.866402
	GOTERM_BP_DIRECT	GO:0001666~response to hypoxia	7	9.87E-04	1.544895
	GOTERM_CC_DIRECT	GO:0045121~membrane raft	10	6.32E-06	0.007526
	GOTERM_CC_DIRECT	GO:0005578~proteinaceous extracellular matrix	11	7.68E-06	0.009144
	GOTERM_CC_DIRECT	GO:0009986~cell surface	15	8.11E-06	0.009649
	GOTERM_CC_DIRECT	GO:0005886~plasma membrane	46	4.84E-05	0.057592
	GOTERM_CC_DIRECT	GO:0005887~integral component of plasma membrane	23	6.57E-05	0.078202
	GOTERM_CC_DIRECT	GO:0009897~external side of plasma membrane	8	4.07E-04	0.483305
	GOTERM_MF_DIRECT	GO:0008201~heparin binding	7	5.16E-04	0.682835
	GOTERM_MF_DIRECT	GO:0004872~receptor activity	7	0.002471	3.232508
	GOTERM_MF_DIRECT	GO:0050431~transforming growth factor beta binding	3	0.004425	5.719841
	GOTERM_MF_DIRECT	GO:0004601~peroxidase activity	3	0.008313	10.493

database developed and were more available to get. The integration of data by bioinformatics analyses from multiple datasets has become a vital source of data for studies of lung cancer. For example, using GSE19804 dataset, Yang *et al.* identified hub genes including *UBE2C*, *DLGAP5*, *TPX2*, *CCNB2*, *BIRC5*, *KIF20A*, *TOP2A*, *GNG11*, and *ANXA1* associated with prognosis in nonsmoking females with NSCLC patients (17). Similarly, using 4 dataset GSE21933, GSE33532, GSE44077 and GSE74706, *CCNB1*, *CCNA2*, *CEP55*, *PBK* and *HMMR* was identified and associated with poorer survival (18).

In the current study, we attempted to identify tumor related genes that contribute to NSCLC overall survival via series of database. We used bioinformatical methods based on 3 profile datasets (GSE19188, GSE27262 and GSE118370). One hundred and twenty-two lung cancer

Table 3 Kyoto Encyclopedia of Gene and Genome pathway analysis of differentially expressed genes in lung cancer

Pathway ID	Name	Count	P value	Genes	FDR
hsa04512	ECM-receptor interaction	7	1.70E-04	*VWF, CD36, COL6A6, ITGA8, COL11A1, SPP1, HMMR*	0.189451
hsa04270	Vascular smooth muscle contraction	5	0.025531	*RAMP3, AGTR1, GUCY1A2, CALCRL, PPP1R14A*	25.03342
hsa03320	PPAR signaling pathway	4	0.026133	*CD36, FABP4, ACADL, MMP1*	25.54754
hsa04261	Adrenergic signaling in cardiomyocytes	5	0.042961	*AGTR1, ADRB1, TNNC1, SCN4B, SCN7A*	38.68835
hsa04514	Cell adhesion molecules (CAMs)	5	0.046888	*CLDN18, ITGA8, PECAM1, JAM2, CDH5*	41.43357
hsa04510	Focal adhesion	6	0.047002	*VWF, CAV1, COL6A6, ITGA8, COL11A1, SPP1*	41.51176

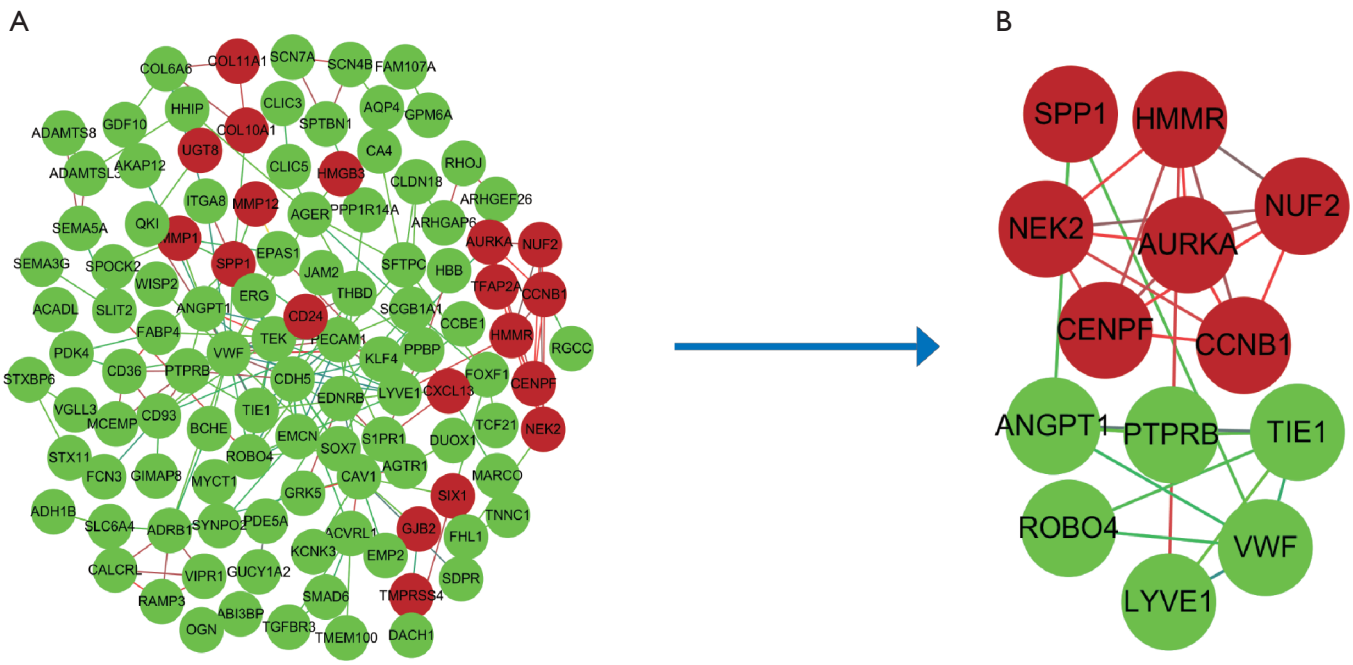

Figure 2 Protein-protein interaction network of common Differentially expressed genes constructed by Search Tool for the Retrieval of Interacting Genes online database and Module analysis. (A) Protein-protein interaction network of Differentially expressed genes. The ball represents gene; the line meant the interaction between genes. green meant down-regulated differentially expressed genes and red meant up-regulated differentially expressed genes. (B) Module analysis though cytoscape software with degree cutoff =2, node score cutoff =0.2, k-core =2, and max. Depth =100.

Table 4 The prognostic information of the 11 key differentially expressed genes

Category	Genes
Genes with significantly better survival (P<0.05)	*ROBO4*, *PTPRB*, *VWF*, *ANGPT1*
Genes with significantly worse survival (P<0.05)	*CCNB1*, *AURKA*, *HMMR*, *SPP1*, *NUF2*, *NEK2*, *CENPF*

specimens and 96 normal specimens were enrolled in this research. In particular, we were able to validate 11 genes that significantly associated with prognosis. First, we extracted 149 common DEGs yielded from 3 datasets (|logFC| >2 and adjust P value <0.05), the vast majority were down-regulated, of which including 127 downregulated and 22 upregulated genes. Next, we

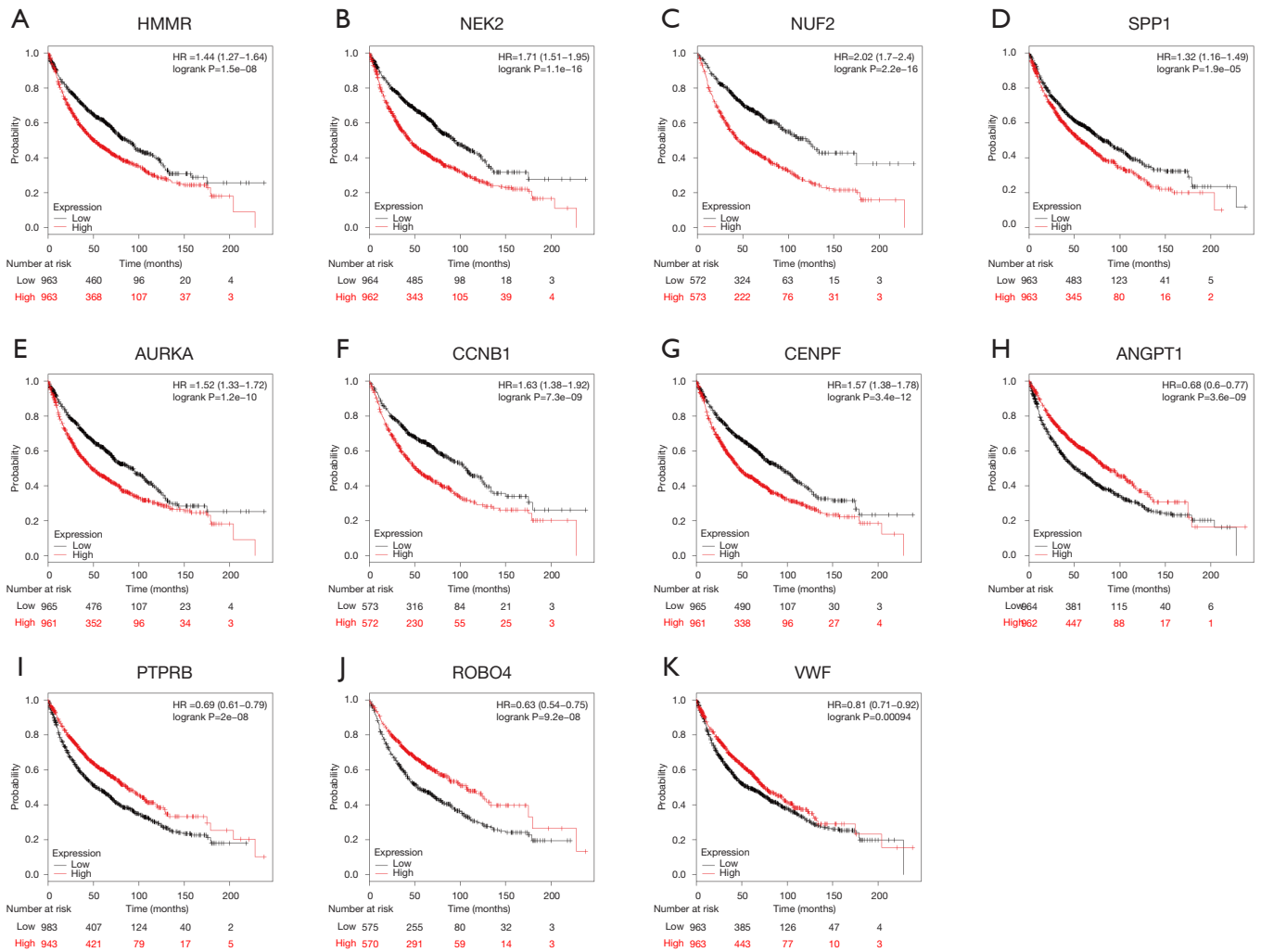

Figure 3 The prognostic information of the 13 core genes. Kaplan-Meier survival curves were generated to identify the prognostic value and 11 of 13 genes had a significantly significance (P<0.05). (A–G) High expression genes with poorer prognosis; (H–K) low expression genes with better prognosis.

Table 5 Further validation of 11 genes via Gene Expression Profiling Interactive Analysis

Category	Genes
Genes with high expressed in LC (P<0.05)	*ROBO4*, *PTPRB*, *VWF*, *ANGPT1*
Genes with low expressed in LC (P<0.05)	*CCNB1*, *AURKA*, *HMMR*, *SPP1*, *NUF2*, *NEK2*, *CENPF*

performed GO and KEGG pathway functional enrichment by DAVID online tool on these DEGs. By performing with GO enrichment analysis, the DEGs were mainly involved in angiogenesis, cell adhesion, vasculogenesis and collagen catabolic process, all these important biological progresses processes participated in the pathophysiological mechanism of NSCLC. Angiogenesis, one of hallmarks of cancer acquired during the multistep development of human tumor (19). A study showed that angiogenic switch is always activated and remains on, resulting in new vessels sprout from quiescent vasculature to help sustain neoplastic growths during tumor progression (20). As for GO cell component (CC), the DEGs were enrich in centrosome, proteinaceous extracellular matrix, plasma membrane, integral component of plasma membrane, cell surface, proteinaceous extracellular matrix and for MF, the DEGs were significantly involved in the heparin binding,

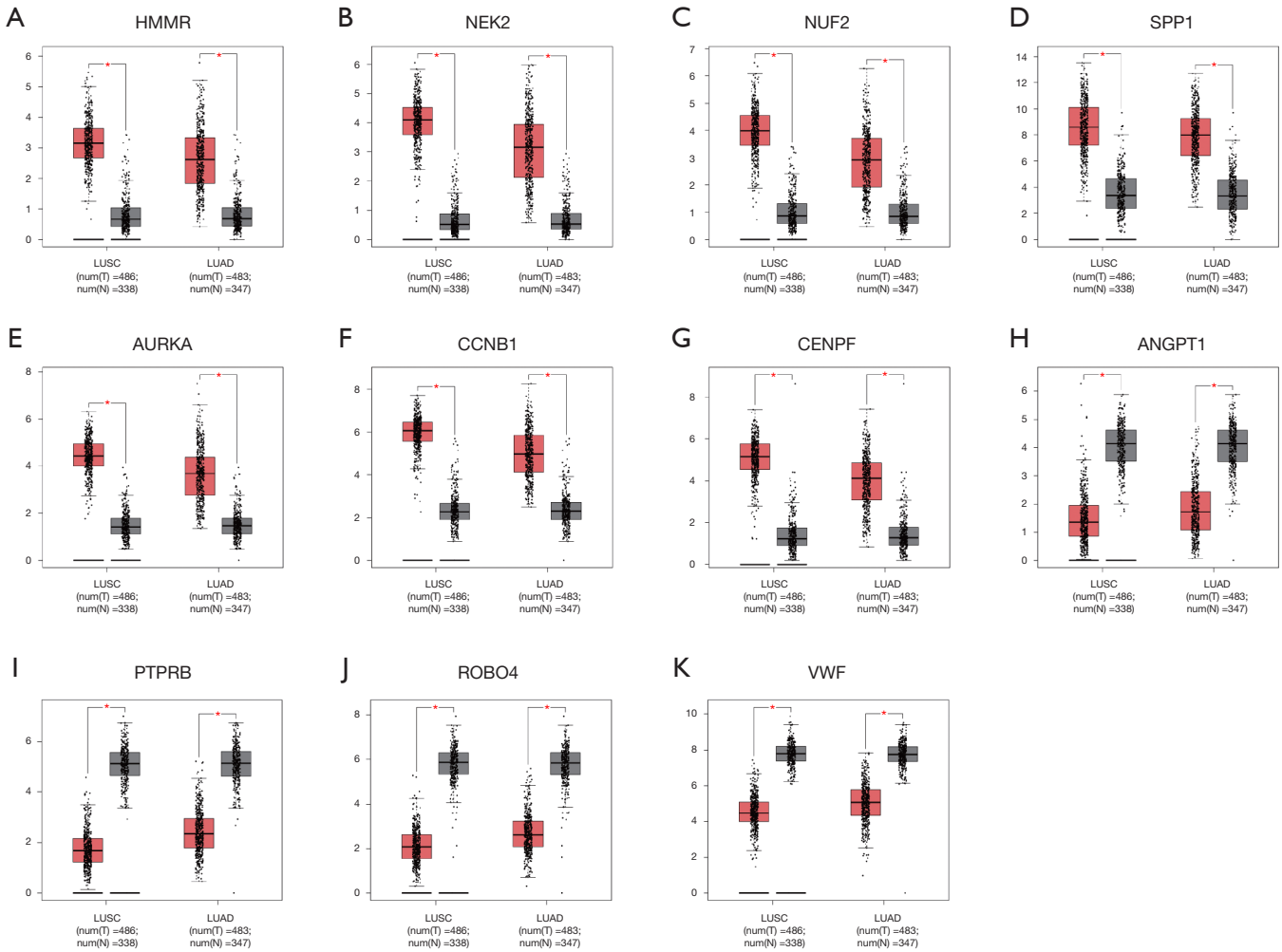

Figure 4 Significantly expressed 11 genes in lung cancer patients compared to healthy people. Eleven genes with prognostic value were analyzed by Gene Expression Profiling Interactive Analysis website. All genes had significant expression level in lung cancer specimen compared to normal specimen (*P<0.05). (A–G) High expression genes when lung squamous cell carcinoma and lung adenocarcinoma compared with normal tissues. (H–K) Low expression genes when lung squamous cell carcinoma (LUSC) and lung adenocarcinoma compared with normal tissues.

Table 6 Reanalysis of 11 candidate genes via Kyoto Encyclopedia of Gene and Genome pathway enrichment

Pathway ID	Name	Count	P value	Genes	FDR
cfa04512	ECM-receptor interaction	3	0.00238	*VWF, SPP1, HMMR*	1.67928
cfa04151	PI3K-Akt signaling pathway	3	0.03261	*VWF, ANGPT1, SPP1*	20.9804

extracellular matrix binding, serine-type endopeptidase activity. KEGG pathway enrichment analysis revealed that DEGs are mainly concentrated in the ECM-receptor interaction, Vascular smooth muscle contraction, PPAR signaling pathway. The pathways of ECM–receptor interaction is important mediators of growth, proliferation, survival, angiogenesis and migration of cancer (21), consistent with the results obtained in this study. In addition, we constructed PPI modules and identified 13 high interrelated nodes by mocode app. Subsequently, we

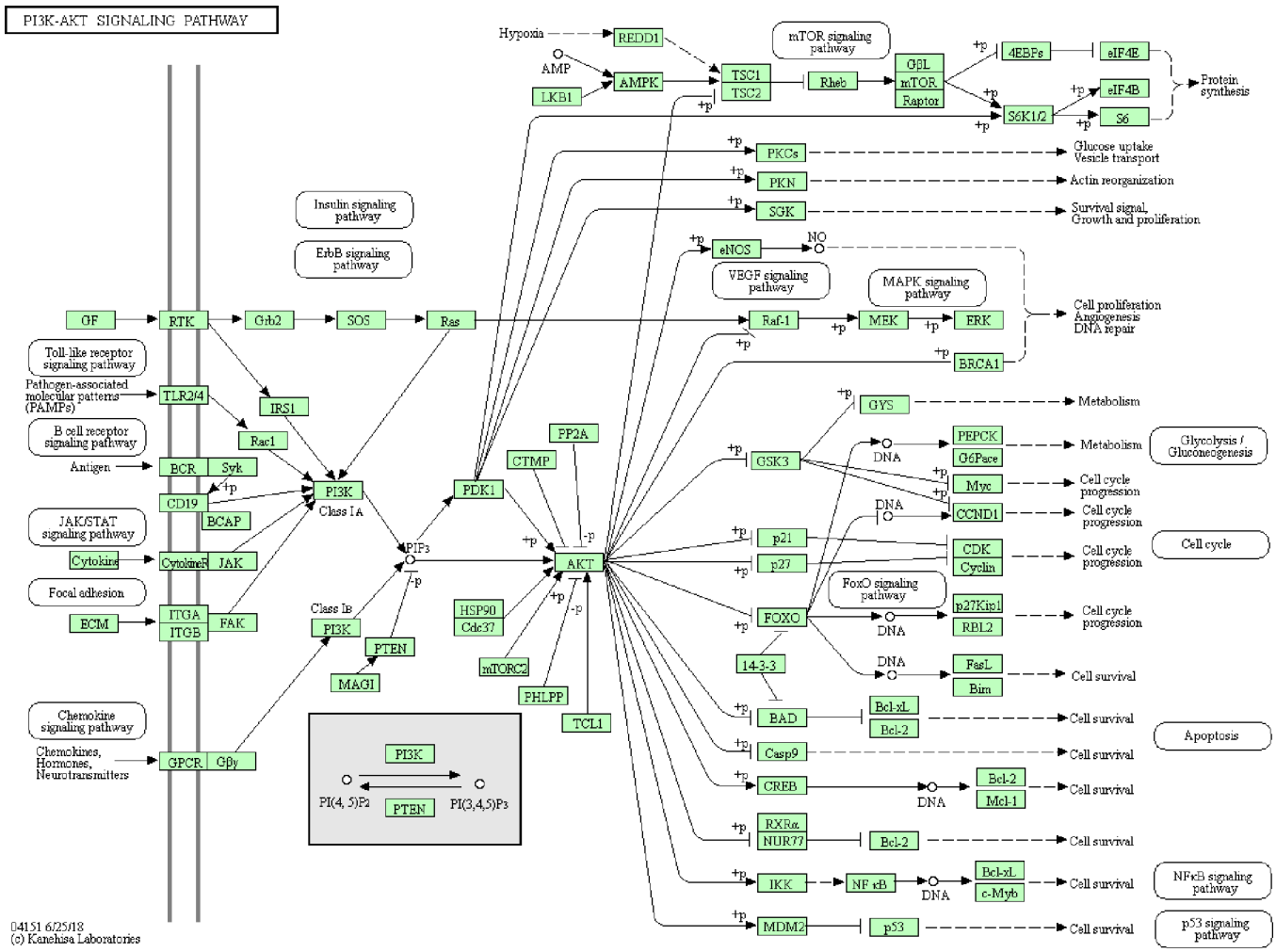

Figure 5 Re-analysis of 11 selected genes by Kyoto Encyclopedia of Gene and Genome pathway enrichment. Three genes (*VWF*, *SPP1*, and *HMMR*) were significantly enriched in the ECM-receptor interaction pathway. Three genes (*VWF*, *ANGPT1*, and *SPP1*) were significantly enriched in the PI3K-Akt signaling pathway.

performed survival of 13 genes and identified 11 related gene that significantly correlated prognosis analysis in NSCLC patients. Of the 11 genes identified, 7 genes with high expression indicated worse survival, but other 6 genes with low expression indicated better survival. In validating these 11 genes, GEPIA was applied and all genes make sense when lung cancer samples compared with normal samples. Finally, we re-analyzed 11 genes via DAVID for KEGG enrichment and found that 3 genes (*VWF*, *SPP1*, and *HMMR*) enriched in ECM-receptor interaction and 3 genes (*VWF*, *ANGPT1*, and *SPP1*) enriched in PI3K-Akt signaling pathway had a significance (P<0.05). We are particularly interested in *VWF* and *SPP1*, because they are common genes in two pathways.

VWF, Von Willebrand factor, a large multimeric plasma glycoprotein originated from endothelial cells, platelets and megakaryocytes. It has been widely known as its function in haemostasis to enables capture of platelets at sites of endothelial damage (22,23), and the function of promoting angiogenesis (24). Recent advances revealed that GATA3 can induce *VWF* upregulation in the lung adenocarcinoma vasculature by binding to the +220 GATA binding motif on the human *VWF* promoter (25) and plasma *VWF/ADAMTS*-13 ratio may act as an independent predictive factor for mortality in patients with advanced NSCLC (26). Another study indicated that *VWF* with

low expression in osteosarcoma tumors can potentially contribute to metastasis (27).

SPP1, secreted phosphoprotein 1, also called *OPN*. It is located on chromosome 4 in locus 4q13.22 and encoded by the human gene *SPP1* (28) that include seven exons and can be alternatively spliced to produce different variants (29). It can be produced by osteoclasts, endothelial cells, epithelial cells, and immune cells to play a vital role in normal and disease BP, including bone remodeling, immune regulation (30) and cell adhesion (31). It can bind to integrins and *CD44*, resulting in inflammatory disorders, autoimmune diseases, and tumorigenesis (30). In non-small cell lung cancers (NSCLC), *SPP1* induces VEGF expression and promotes tumor progression (12). Altogether, it can be a useful target and potential therapy target. Numerous studies have demonstrated that these two genes were related to distinct types of cancer, however, few papers have been studied in lung cancer. Also, *CENPF*, *PTPRB*, and *NUF2* are rarely reported after we searched these genes in PubMed online website. Taken together, our study linked to NSCLC pathogenesis could improve the understanding of underlying molecular mechanisms of NSCLC and provide useful information for future study of new anticancer in lung cancer.

Conclusions

We identified DEGs between lung cancer and normal tissues on the via bioinformatics analysis and the results revealed they may play crucial roles in the progression of lung cancer; however, Further experiments are needed to verify these predictions. Anyway, this study may provide some potential biomarkers and targets for NSCLC diagnosis and therapy.

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Availability of data and materials: The datasets generated and/or analyzed during the study are available from the corresponding author upon reasonable request.

Footnote

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at http://dx.doi.org/10.21037/tcr-19-2596). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

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