

Epidermal growth factor-like repeats and discoidin I-like domains 3: a multifaceted oncoprotein at the crossroad of MAPK and TGFbeta pathways in human hepatocellular carcinoma

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Introduction

Hepatocellular carcinoma (HCC) is a frequent human cancer with 0.25-1 million of newly diagnosed cases each year (1-3). Major risk factors associated with the development of HCC are chronic hepatitis B virus (HBV) and hepatitis C virus (HCV) infections, alcoholic hepatitis, aflatoxin B1 (3-5), and some inherited diseases (6). HCC is a fatal disease, with a life expectancy of about 6 months from the time of diagnosis (6). Early liver lesions could be detected by ultrasonography and efficiently treated by resection or radiofrequency ablation (7). However, only a minority of cases is eligible to these treatment modalities due to the late diagnosis of the disease (2,7,8). In addition, therapies with pharmacological agents or alternative approaches, including percutaneous ethanol injection, transarterial chemo-embolization or yttrium-90 microspheres, do not improve significantly the prognosis of patients with advanced disease (2,7,8).

The evaluation of the molecular mechanisms and the identification of prognostic categories of HCC are difficult due to HCC heterogeneity, which results from complex relationships between genetic, etiologic, and environmental risk factors (6). A better understanding of HCC molecular pathogenesis may hasten the identification of new prognostic markers and the development of novel diagnostic and therapeutic strategies against this disease (6,9).

Biological ad clinical behavior of HCC may be largely influenced by both genetic and epigenetic alterations of a number of genes and signaling pathways (6,10). The remodeling of microenvironment (11,12) surrounding HCC may also affect HCC biological behavior, thus influencing patients' outcome (13). This is an important facet of the complex mechanisms involved in tumor progression. Different proteins of the extracellular matrix (ECM) may affect cell growth, migration, invasion, anoikis and metastasis (13-17) by binding to specific receptors of cancer cells or interfering with the binding of specific cytokines (18).

The epidermal growth factor-like repeats and discoidin I-like domains 3 (EDIL3) protein

EDIL3, also known as endothelial cell locus (DEL-1), is a secreted ECM protein isolated and identified from embryonic mouse lung in 1998 (19). EDIL3, secreted by embryonic endothelial cells and hypertrophic chondrocytes (20), was firstly characterized in vascular morphogenesis (21).

EDIL3 is a glycoprotein composed of five domains: three epidermal growth factor (EGF)-like repeats (E1, E2, E3), and two discoidin I-like domains (C1, C2). In particular, the second EGF repeat contains an Arg-Gly-Asp (RGD) motif (*Figure 1*) (19,20). It has been shown that the C-terminus of the C1 domain is essential for the organization of EDIL3 into the ECM and that all the E repeat domains and the N-terminus of the C1 domain play supportive roles for this organization (20).

At the cellular level, EDIL3 exerts numerous, important roles. Through the interaction of the Arg-Gly-sp tripeptide

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Figure 1 Schematic representation of the EDIL3 glycoprotein showing three epidermal growth factor (EGF)-like repeats (E1, E2, E3) and two discoidin I-like domains (C1, C2). The second EGF repeat contains an Arg-Gly-Asp (RGD) motif.

with the $\alpha\nu\beta3$ integrin, EDIL3 induces clustering of integrin receptors, endothelial attachment, and migration as well as focal contact and phosphorylation of different molecules involved in cell signaling, including p125FAK and MAP kinase (22). Moreover, EDIL3 plays a pivotal role in inflammatory and immune responses, where leukocyte adhesion to endothelium, crucial for leukocyte recruitment, requires numerous adhesion molecules expressed on leukocytes and endothelial cells. Indeed, EDIL3 acts an anti-adhesive factor that interferes with the integrin LFA-1dependent leukocyte-endothelial adhesion, thus preventing leukocyte adhesion to the endothelium (23,24).

EDIL3 is also a potent pro-angiogenic factor, as it significantly contributes to vessel wall remodeling and development during angiogenesis (25), mediates endothelial cell attachment and migration (26), and induces mesentery and cerebral angiogenesis in mice (27,28). Both animal experiments and clinical studies have demonstrated that EDIL3 gene therapy is effective in the presence of an ischemic disease (29-31).

EDIL3 is expressed in brain, heart, small intestine and kidney tissues, but not in colon, liver, or lung of human adults (26). In addition, EDIL3 is expressed in primary human tumors, such as lung (32), bladder (33), pancreas (34), liver (35), breast, and colon cancer, and melanomas (26), and in many tumor cell lines (28). Furthermore, EDIL3 levels have been associated with the progression and prognosis of lung cancer (32), bladder cancer (33), and pancreatic ductal adenocarcinoma (35). Interestingly, EDIL3 was recently shown to be a novel biomarker for early breast cancer detection (36).

EDIL3, epithelial-to-mesenchymal transition (EMT), and integrin signaling

EMT is involved in different physiological events, including blastocyst implantation, generation of the neural crest, normal wound healing (37,38) as well as in pathological events such as pathological wound healing, tissue fibrosis

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and carcinogenesis (39,40).

EMT consists of the loss of typical epithelial features, such as cell polarity, intercellular junctions, and ability to synthesize basement membranes, associated with the development of a fibroblastic morphology with rearrangement of the actin cytoskeleton and changes in cell surface matrix receptors, such as integrins. As a consequence, cells form filopodia, migrate, and synthesize ECM (39,41). Three types of EMT have been described: (I) type 1, which occurs during earliest stages of development; (II) type 2, occurring in mature epithelial tissues, generally triggered by inflammation or wound-healing responses, which may induce fibrosis; (III) type 3, which is associated with cancer progression (38).

Tumors contain a subpopulation of cells characterized by the loss of epithelial features and the acquisition of the mesenchymal-like migratory phenotype. These cells, known as cancer stem cells (CSCs), are able to self-renew and regenerate the tumor mass. CSCs are crucial to the development of invasive carcinomas and metastasis (38-41). Nonetheless, tumor cells disseminated into target organs may undergo mesenchymal-epithelial transition (MET), which would also favor metastasis formation (42,43).

EMT is regulated by TGF- β through different mechanisms. Nuclear translocation of SMAD complexes, formed in the canonical TGF- β cascade (6), stimulates the expression of different mesenchymal genes, while repressing epithelial gene transcription (*Figure 2*). Furthermore, TGF- β signaling activates integrin-linked kinase (ILK), which phosphorylates GSK-3 β and AKT (serine/threonine protein kinase), with consequent nuclear translocation of β -Catenin and activation of different transcription factors involved in EMT (44). EMT is also induced trough the ERK/MAP kinase, Rho GTPase and the PI3 kinase/AKT pathways following TGF- β receptor activation (*Figure 2*) (45,46).

TGF- β is synthesized in a complex pathway: precursor forms of TGF- β 1 and TGF- β 3 are linked to a latencyassociated peptide (LAP), containing an RGD motif that may be activated by $\alpha\nu\beta$ 1, 3, 5, 6, and 8 integrins and interacts with RGD (*Figure 2*) (47-50). This is followed by the interaction of mature TGF- β with its receptor and the activation of different signals that, at DNA level, induce the activation of mesenchymal markers (i.e., integrins, N-Cadherin, fibronectin, collagen) and inhibition of epithelial markers (i.e., CDH1, claudins, occludins, desmoplakin), and integrin activation (38).

EDIL3 binding to $\alpha\nu\beta3$ integrin by the RGD motif (*Figure 2*) prevents apoptosis of endothelial cells, thus

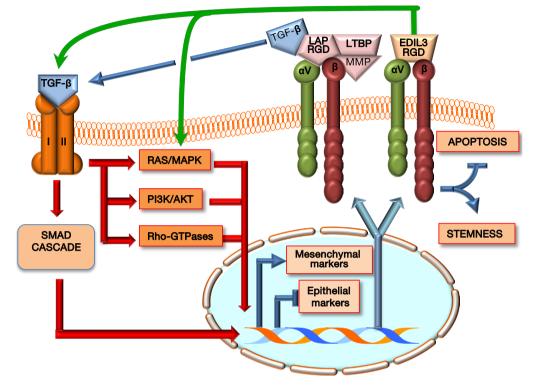


Figure 2 The regulatory circuitry of $\alpha\nu\beta$ integrins. The interaction of ECM protein, EDIL3, and the LAP protein of the TGF- β -immature complex with $\alpha\nu\beta$ integrin is followed by activation of TGF- β , RAS/ERK, PI3K/AKT and Rho/GTPases signaling pathways. This leads to the up-regulation of the mesenchymal markers, the down-regulation of the epithelial markers, and the up-regulation of integrins, with consequent decrease in cell death and acquisition of the molecular and morphologic changes of stemness and EMT. Arrows indicate activation; blunt arrows indicate inhibition. LAP, latency activated peptide; LTBP, latent transforming growth factor β binding protein; MMP, metalloproteinase; EMT, epithelial-to-mesenchymal transition.

favoring cancer vascularization and potentiating cancer cell proliferation and invasion (51). This effect is mediated specifically through the crosstalk with FAK/ERK and AKT signaling (51). Integrins, as primary receptors involved in cell-matrix adhesion, may strongly influence the ability of cancer cells to survive in specific sites. Interestingly, it has been observed that in some cases integrin receptors can also function in the absence of ligand binding to promote stemness and survival (52). Thus, the interplay between TGF- β and integrin signaling, occurring downstream of initial TGF- β receptor activation, regulates various cellular processes (53), including different signaling pathways that are able to override the tumor suppressing functions of TGF- β (54-56).

EDIL3 and HCC

Mounting evidence supports an important role of EDIL3

in HCC. According to recent data, indeed, EDIL3 activity is crucial for the interaction between HCC cells and endothelial cells (28), and may accelerate tumor growth by stimulating angiogenesis (57). *EDIL3* gene is overexpressed in HCC (35) and predicts poor prognosis of HCC patients (13,35,58). Interestingly, recent studies suggest that autocrine EDIL3 may contribute to a receptive microenvironment for the survival of detached HCC cells by promoting anoikis resistance (13). This intriguing finding suggests that activation of integrin signaling pathways by EDIL3 may contribute to HCC cell spreading. Furthermore, the accumulation of tumor-produced EDIL3 in the microenvironment represents an advantage for anchorage-independent growth of tumor cells.

These observations have been confirmed and extended in an interesting publication by Xia and coworkers (59). As a first approach to establish the role of EDIL3 as regulator of EMT in HCC, the authors evaluated the correlation of EDIL3 expression with that of mesenchymal and epithelial markers, using independent published microarray data for liver cancer cell lines. Noticeably, the authors found a positive correlation between EDIL3 levels and the expression of the mesenchymal marker vimentin (VIM), and a negative correlation with the epithelial marker E-cadherin (CDH1). Accordingly, forced EDIL3 expression in Huh7 cells led to the acquisition of a fibroblastic elongated phenotype associated with a fall in the expression of the epithelial marker CDH1 and up-regulation of the mesenchymal marker VIM. The opposite occurred when EDIL3 expression was inhibited by specific siRNA in HLE cells. In the latter case, morphologic changes indicative of MET were found. Further support to the role of EDIL3 as regulator of EMT in HCC was obtained by the evaluation of different phenotypic properties linked to EMT. Indeed, the migration and invasion properties of Huh7 cells, characterized by lower EDIL3 expression and an epithelial phenotype, were significantly lower than that of HLE cells, which exhibit high EDIL3 expression and a mesenchymal phenotype. The modulation of EDIL3 expression strongly influenced HCC cell migration, invasion, and HCC angiogenesis in the same cells, as evaluated by in vitro endothelial recruitment and capillary tube formation assays.

Interestingly, an epigenetic mechanism was found to be responsible for EDIL3 deregulation in HCC. Specifically, the authors identified microRNA (miR)-137 as a critical, negative regulator of EDIL3. In particular, Xia *et al.* observed the downregulation of miR-137 in HCC samples from patients exhibiting early recurrent disease, when compared to samples from patients with non-recurrent HCC. The decrease in miR-137 expression was correlated with the up-regulation of EDIL3 expression. Subsequent *in vitro* experiments showed that miR-137 triggers EDIL3 downregulation, inhibits HCC cell invasion, and induces endothelial cell capillary tube formation.

In accordance with previous studies on the relationships between TGF- β an integrin expression (60), TGF- β 1 levels were found to be significantly increased in HuH7 and PLC/PRF/5 HCC cells stably transfected with EDIL3. Using the data reported in the Cancer Cell Line Encyclopedia dataset (http://www.broadinstitute.org/ ccle), the authors compared two groups of liver cancer cells displaying high and low EDIL3 expression, respectively. This allowed the study of the correlation of differentially expressed genes with EDIL3 expression levels. Significant correlation was observed for the expression of TGF β 111 and TGF β 2, suggesting a regulation of TGF- β signaling through binding to $\alpha v \beta$ 3 integrin in liver cancer cells. Further analysis showed that pseudopodium-enriched atypical kinase 1 (PEAK1)-associated regulatory signaling interacts with EDIL3 through the SRC family kinases. Importantly, overexpression of EDIL3 not only significantly enhanced the expression of PEAK1, but also induced the phosphorylation of SRC, ERK and SMAD2, suggesting the activation of ERK and TGF- β signaling.

These important observations by Xia *et al.* confirm and extend to the HCC field previous observations (38) indicating the existence of a regulatory circuitry for EMT (*Figure 2*). In this circuitry, the ECM protein EDIL3 interacts with $\alpha\nu\beta$ integrin, thus inducing the activation of TGF- β and RAS/ERK cascades. Once activated, the TGF- β and RAS/ERK pathways trigger the up-regulation of mesenchymal marker and integrins, while promoting the down-regulation of epithelial markers. These molecular events are associated with cell death decrease and acquisition of the molecular and morphologic changes of stemness and EMT by cancer cells.

Concluding remarks

A growing body of experimental and clinical observations points to a pivotal role of EDIL3 protein in HCC progression and patient's prognosis. The study by Xia et al., in particular, indicates that EDIL3 significantly contributes to many traits of HCC cells, namely uncontrolled growth, resistance to apoptosis, migration, invasion, and angiogenesis. At the clinical level, EDIL3 up-regulation results in early tumor recurrence and poor outcome. Intriguingly, it has been demonstrated that EDIL3 lies at the crossroad of numerous oncogenic pathways, including the ERK/MAPK, TGF-β, and integrin signaling cascades. Consequently, EDIL3 suppression might result in the concomitant inhibition of multiple oncogenic stimuli, whose inactivation could be highly deleterious for the survival of HCC cells. Based on these important findings, additional efforts should be devoted to elucidate the function of EDIL3 in liver cancer as well as to develop novel therapeutic approaches aimed at suppressing EDIL3 activity for the treatment of this pernicious disease.

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Footnote

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