



A narrative review on the main chemical constituents and bioactivity of *Camellia nitidissima* Chi

Hanyu Zheng^{1,2}, Qizhen Du², Junfeng Yin¹, Ying Gao¹

¹Tea Research Institute Chinese Academy of Agricultural Sciences, Hangzhou, China; ²College of Food and Health, Zhejiang A&F University, Linan, China

Contributions: (I) Conception and design: H Zheng, Y Gao; (II) Administrative support: Q Du, J Yin; (III) Provision of study materials or patients: H Zheng; (IV) Collection and assembly of data: H Zheng, Y Gao; (V) Data analysis and interpretation: H Zheng, Y Gao; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

Correspondence to: Ying Gao. Tea Research Institute Chinese Academy of Agricultural Sciences, Hangzhou, China. Email: yinggao@tricaas.com.

Background and Objective: *Camellia nitidissima* Chi (*C. nitidissima*) is an herb used in traditional Chinese medicine. In recent years, researches of *Camellia nitidissima* Chi have boomed. Various components in *Camellia nitidissima* Chi have been identified and some of its health-beneficial functions have been demonstrated. The review aims to supply an overview of the main chemical components and health-beneficial functions of *Camellia nitidissima* Chi.

Methods: The references in this review are mainly collected from published books and the scientific literature databases including Web of science, PubMed, and China National Knowledge Infrastructure, with a timeframe from January 1986 to March 2022, containing English and Chinese references.

Key Content and Findings: Chemical analysis reveals that *Camellia nitidissima* Chi contains a variety of chemical constituents, including phenolic compounds, saponins, polysaccharides, and other substances. Phenolic compounds, particularly flavonoids, are the most well-studied components in *Camellia nitidissima* Chi. Many of them are bioactive and contribute to the health-beneficial functions of *Camellia nitidissima* Chi. Plenty of studies confirm that *Camellia nitidissima* Chi have effects on antioxidant, anti-cancer, anti-hyperglycemia, anti-hyperlipidemia, anti-allergy, and anti-depression. Part of the underlying mechanisms are unveiled.

Conclusions: Based on current research progress, *Camellia nitidissima* Chi has the potential to be applied in dietary supplements and even medications. More studies are needed to further figure out its working mechanisms and assess its effectiveness in humans.

Keywords: *Camellia nitidissima* Chi (*C. nitidissima*); chemical profile; health-beneficial functions; herb; traditional Chinese medicine

Received: 09 March 2022; Accepted: 04 July 2022; Published: 30 September 2022.

doi: 10.21037/lcm-22-9

View this article at: <https://dx.doi.org/10.21037/lcm-22-9>

Introduction

Camellia nitidissima Chi (*C. nitidissima*) is a precious plant species with high ornamental value because of its canary yellow flowers (1,2). Wild *C. nitidissima* is predominately distributed in the mountains of Southwest China (particularly in Guangxi Zhuang autonomous region) and North Vietnam (3,4). *C. nitidissima* has been introduced

and cultivated in Japan, Australia, North America, and several provinces in China (5). Guangxi is the main planting area of *C. nitidissima* in China and the acreage reaches over 2,000 hectares. The annual productions of *C. nitidissima* fresh leaves and flowers in Guangxi are ~15,000 and ~225 tons, respectively. The economic value of the *C. nitidissima* industry exceeds 2 billion RMB per year, making it one of the pillar industries in Guangxi (source from the forestry

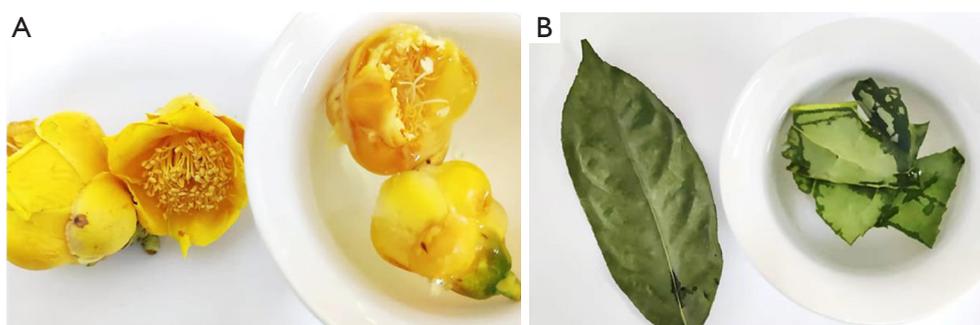


Figure 1 Freeze-dried *Camellia nitidissima* Chi flowers (A) and leaves (B).

bureau of Fangchenggang City, Guangxi, China).

C. nitidissima was first discovered by the Chinese botanist Jinglie Zuo in Fangchenggang City (Guangxi, China) in 1933, initially named as *Theopsis chrysantha* Hu by the Chinese botanist Jingwen Qi in 1948 (5), and later revised as *Camellia nitidissima* Chi in Flora Reipublicae Popularis Sinicae (6). The plant belongs to Theaceae *Camellia* section *Chrysantha* Chang, genetically close to *Camellia sinensis*, *Camellia semiserrata*, and *Camellia oleifera*.

Although botanically classified in the last century, *C. nitidissima* has a long history of use in traditional medicine. It is recorded in *Ben Cao Gang Mu*, a sixteenth-century Chinese encyclopedia of medical matter and natural history. It exhibits activities in detoxifying, promoting diuresis, and reducing puffiness. It also helps in the treatment of dysentery and pharyngitis. Besides its medical use, *C. nitidissima* is utilized in daily life for beverages. Freeze-dried *C. nitidissima* flowers are the most common product of *C. nitidissima* in the commercial market (Figure 1A). *C. nitidissima* leaves (Figure 1B) are less popular, which are mainly consumed by ethnic minorities in Guangxi to prepare decoctions for the nourishment. In 2010, *C. nitidissima* was approved as a new resource food by the Ministry of Health of China, providing a bright future for its applications in medicinal food and dietary supplements.

Phytochemical studies have shown that *C. nitidissima* contains a variety of active ingredients, such as phenolic compounds, saponins, polysaccharides, volatiles, mineral elements, and amino acids (7). Biological studies have demonstrated that *C. nitidissima* exhibits antioxidant and anticancer activities *in vitro* and *in vivo* (7). In addition, *C. nitidissima* exhibits lipid-lowering and immunomodulatory activities in animal models (8). In the past decade, several novel compounds in *C. nitidissima* have been identified and proved to be bioactive (7). In the review, research progresses

on the constituents and health-beneficial properties of *C. nitidissima* in recent years are summarized. It is hoped that the review will cause more readers' interest in *C. nitidissima* and inspire them to think about the future prospects of *C. nitidissima*. It is also hoped that the review will help scientists find out the promising directions for further researches and applications of *C. nitidissima*.

We present the following article in accordance with the Narrative Review reporting checklist (available at <https://lcm.amegroups.com/article/view/10.21037/lcm-22-9/rc>).

Methods

The references in this review are mainly collected from published books and the scientific literature databases including Web of science, PubMed, and China National Knowledge Infrastructure (CNKI), with a timeframe from January 1986 to March 2022, containing English and Chinese references. The search was conducted between January 3, 2022 to March 8, 2022. Search terms included "*Camellia nitidissima*" and its Chinese characters "金花茶". The detailed search strategy is listed in Table 1.

Main chemical constituents of *C. nitidissima*

Nowadays, it is generally accepted that herbs play an important role in the prevention and treatment of diseases by virtue of its functional ingredients. There are many studies describing the chemical constituents in *C. nitidissima* flowers and leaves (1,2,8). Comparatively, less is known about the chemical constituents in other parts of *C. nitidissima*. Though different in contents, the classes of compounds found in flowers and leaves are similar, including phenolic compounds, saponins, polysaccharides, and other substances.

Table 1 The search strategy summary

Items	Specification
Date of search	January 3, 2022–March 8, 2022
Databases and other sources searched	Databases: Web of science, PubMed, and China National Knowledge Infrastructure (CNKI) Published books
Search terms used	Search terms: <i>Camellia nitidissima</i> (for English databases, including Web of science and PubMed), 金花茶 (for the Chinese database CNKI)
Timeframe	From January 1986 to March 2022
Inclusion and exclusion criteria	Inclusion and exclusion criteria: (I) articles in English and Chinese languages; (II) article types were research articles and reviews
Selection process	Hanyu Zheng and Ying Gao conducted the selection together and consensus was obtained after a discussion among all authors

Phenolic compounds

Phenolic compounds are a diverse group of bioactive secondary metabolites characterized by their structures having at least one phenol unit. Flavonoids, phenolic acids, and lignans are important phenolic compounds in *C. nitidissima*.

Flavonoids

Flavonoids are low-molecular-weight polyphenolic substances characterized by the flavan nucleus. Many flavonoids are with physiological functions such as antioxidant, antiviral, anti-inflammatory, hypotensive, and lipid-lowering activity (9). Flavonoids are classified into 12 major subclasses, six of which, namely flavonols, flavones, flavan-3-ols, anthocyanidins, flavanones, and isoflavones, are widely distributed in the diet.

The flavonoid content of *C. nitidissima* is not the highest among members in Theaceae *Camellia* section *Cbrysantha* Chang (10,11). In *C. nitidissima*, more flavonoids are accumulated in flowers rather than leaves. A research showed that the flavonoid content of *C. nitidissima* flowers reached 8.5%, which was 37 times to that of leaves. The flavonoid content of *C. nitidissima* flowers varies in different stages, decreased in the order of semi-open stage > fish-mouth stage ≈ blooming stage > withering stage ≈ budding stage (12). The flavonoid content of *C. nitidissima* leaves decreases as the leaves grow. Compared with young leaves, about 69% and 77% of flavonoids were lost in one-year-old leaves and two-year-old leaves, respectively (13). *C.*

nitidissima contains more water-soluble flavonoids than alcohol-soluble flavonoids. Tang (11) found that the flavonoid content of *C. nitidissima* water extract was much higher than that of *C. nitidissima* alcohol extract. At present, quite a number of flavonoids in *C. nitidissima* have been identified.

Flavones are a class of flavonoids based on the backbone of 2-phenylchromen-4-one and flavonols are a class of flavonoids that have the 3-hydroxyflavone backbone. Kaempferol, quercetin, apigenin, and their glycosides are main flavones and flavonols in *C. nitidissima*. Aromadendrin, dihydroquercetin, dihydrokaempferol, and isorhamnetin glucosides are also detected (2,14,15). Glucose and rhamnose are the two most common sugars to form glycosides in *C. nitidissima*. Peng *et al.* (16) used repeated silica gel column chromatography, Sephadex LH-20 column chromatography, ODS column chromatography, repeated recrystallization and other strategies to separate and purify the chemical components of *C. nitidissima*. Seven of the thirteen obtained compounds belonged to flavones and flavonols, including quercetin, quercetin-7-O-β-D-glucopyranoside, quercetin-3-O-β-D-glucopyranoside, rutin, vitexin, kaempferol, and kaempferol-3-O-β-D-glucopyranoside. In addition to usual monoglycosides, several diglycosides and triglycosides are identified. Some glycosides are modified with the acetyl moiety and/or coumaroyl moiety. For example, Yang *et al.* (17,18) isolated two acetyl flavonol glycosides from *C. nitidissima* flowers, namely kaempferol 3-O-[2,3,4-tri-

O-acetyl- α -L-rhamnopyranosyl-(1/3)-2,4-di-O-acetyl- α -L-rhamnopyranosyl-(1/6)]- β -D-glucopyranoside and kaempferol 3-O-[2,3,4-tri-O-acetyl- α -L-rhamnopyranosyl-(1/3)-4-O-acetyl- α -L-rhamnopyranosyl-(1/6)]- β -glucopyranoside, which showed remarkable inhibitory effects on the advanced glycation end-products (AGEs) formation. To be mentioned, a glycoside dimer called kaempferol-3-O-glycosyl-4'-kaempferol-3-O-glycoside was identified in *C. nitidissima* flowers, which was rare in other plants.

Flavan-3-ols are derivatives of flavans that possess a 2-phenyl-3,4-dihydro-2H-chromen-3-ol skeleton. Epicatechin and catechin, two flavan-3-ols distributed in many plants, are found in *C. nitidissima*. Gallocatechin gallate, epigallocatechin (EGC), epicatechin gallate (ECG), catechin gallate, epigallocatechin gallate, and gallocatechin, which are unusual in plants out of the Theaceae family, are detected in *C. nitidissima* flowers (19). Among these catechins, the abundance of epicatechin is the highest in flowers, followed by EGC and ECG. The contents of catechins in different parts of flowers are not the same. Stamens and petals contain less catechins than sepals. In leaves, catechin and epicatechin are also detected (20). However, the contents of catechins and epicatechins in leaves are much lower than that in flowers. Old leaves accumulate more catechins than young leaves (21). Besides catechin monomers, procyanidins, which refer to the polymers of catechins, exist in *C. nitidissima* as well. So far, procyanidin dimers, trimers, tetramers, and pentamers have been identified in *C. nitidissima* flowers (22). Yang (23) identified a unique procyanidin tetramer in *C. nitidissima* flowers, which was catechin-4 \rightarrow 8-catechin-4 \rightarrow 8-catechin-3 \rightarrow 7-catechin, and named it nitidissimol A.

Anthocyanins are water-soluble vacuolar pigments which are responsible for the vivid colors in plant tissues. Compared with *C. nitidissima* flowers, more anthocyanins are in leaves. Young leaves have more anthocyanins than old leaves. Li *et al.* (21) identified two anthocyanins in *C. nitidissima*, which were pelargonium-3-O-glucoside and cyanidin-3-O-glucoside. The former one existed in both flowers and leaves of *C. nitidissima*. The latter one was only detected in leaves and was considered to contribute to the purple color of new leaves.

Phenolic acids

Phenolic acids are phenols that contain a carboxylic acid. Gallic acid, chlorogenic acid, salicylic acid, and protocatechuic acid, which are common phenolic acids in

plants, are found in the flowers of *C. nitidissima* (23). Ellagic acid is a phenolic acid with outstanding antioxidant and anti-proliferative properties. Multiple ellagic acid derivatives are identified in the leaves of *C. nitidissima*. Yu (24) demonstrated the presence of ellagic acid and four ellagic acid derivatives, including 3'-methy-4'-glucoside-ellagic acid, okicamelliaside, 3'-methyellagic acid, and 3,4-O,O-methylidyne-ellagic acid, in the leaves of *C. nitidissima*. Mo *et al.* (25) identified five ellagic acid derivatives in the leaves of *C. nitidissima*, which were 3,4-methylenedioxy-3'-O-methyl-4'-O-(6'-O-acetyl-glucoside) ellagic acid, okicamelliaside, 3,4-O,O-methylidyne-ellagic acid, ellagic acid-4-O- β -D-glucopyranoside, and 3,4-methylenedioxy-3'-O-methyl-4'-O-glucoside ellagic acid. Among these compounds, okicamelliaside is the relatively abundant one, whose content ranges from 0.51% to 1.33% (26). Notably, little study on the ellagic acid derivatives in *C. nitidissima* flowers was found.

Lignans

Lignans are known to be minor constituents of many plants, often recognized as phytoestrogens. Lignans are derived from phenylalanine, consisting of two phenol units linked by four carbons. Lignans can polymerize to lignin to building the plant cell wall. Lignans often occur in the glycosidic form. They can be metabolized by intestinal bacteria to form mammalian lignans, which have cytostatic activity (27). Zhang *et al.* (28) isolated and purified lignans from *C. nitidissima* flowers using silica gel, Sephadex LH-20 gel, C18 reversed silica gel, and semi-preparative high performance liquid chromatography (HPLC). Eight lignans were identified, which were eudesmin, (+)-diasyringaresinol, (+)-isoeucommin A, pinoresinol 4-O-glucoside, 7S, 8R, 8'R-(-)-lariciresinol-4'-O-D-glucopyranoside, (+)-isolariciresinol 9-O- β -D-glucopyranoside, (+)-isolariciresinol 9'-O- β -D-glucopyranoside, and 3',4-O-dimethylcedrusin. All of them have been reported to possess bioactivity.

Based on current studies, a table (Table 2) concludes phenolic compounds observed in *C. nitidissima* are prepared. Their molecular structures are presented in Figure 2.

Saponins

Saponins are plant-derived organic chemicals that have a foamy quality when agitated in water. Booming evidences indicate that various saponins have biological activity, such as anti-cancer, lipid-lowering, and anti-bacteria (29).

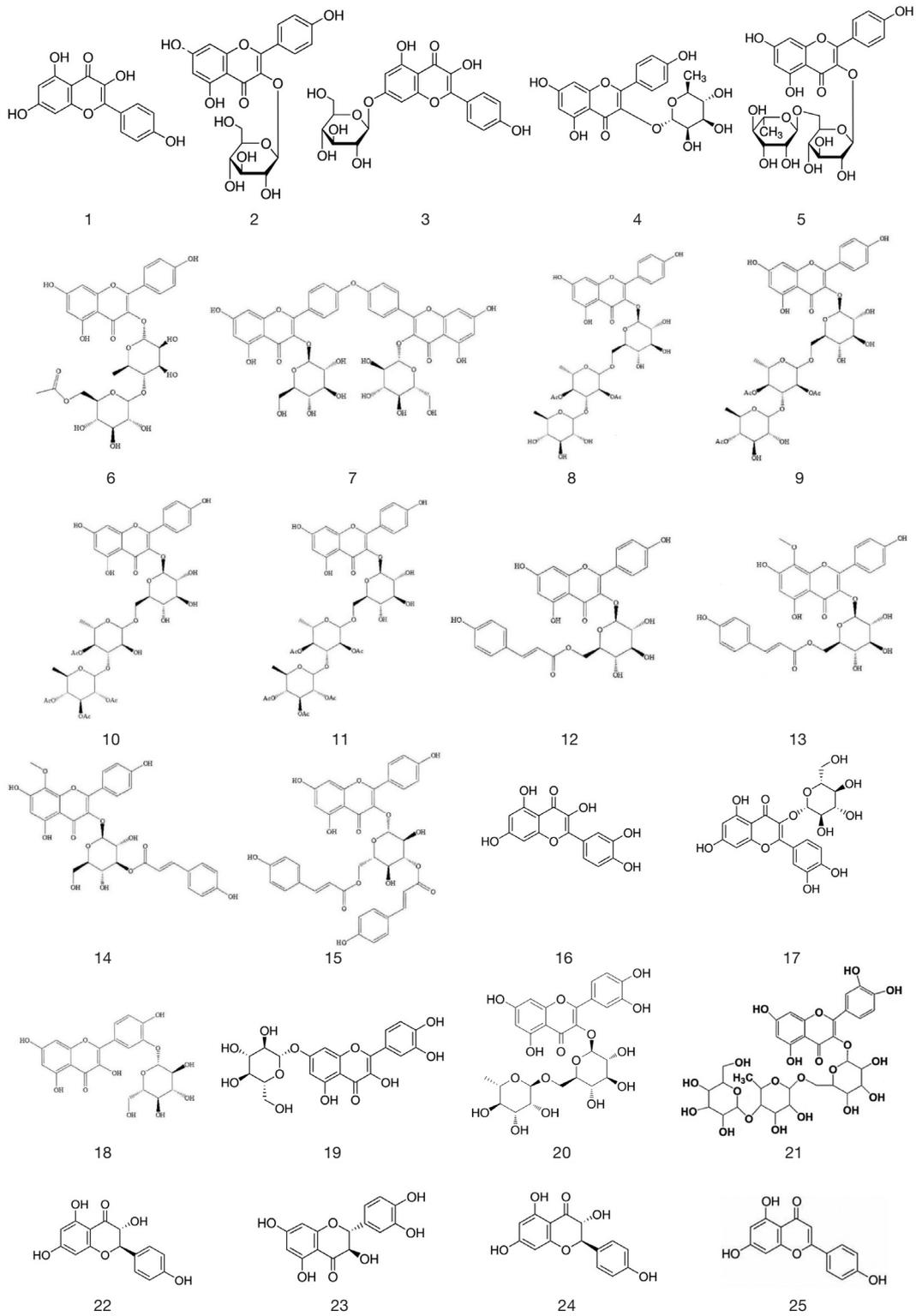
Table 2 Phenolic compounds in *Camellia nitidissima* Chi

Classes/compound No.	Compound names	References
Flavonoids		
Flavonols		
1	Kaempferol	(16)
2	Kaempferol-3-O- β -D-glucoside	(23)
3	Kaempferol-7-O- β -D-glucoside	(23)
4	Kaempferol-3-O-rhamnoside	(23)
5	Kaempferol-3-O- β -D-rutinoside	(23)
6	Multiflorin C	(23)
7	Kaempferol 3-O-glucosyl-4'-kaempferol 3-O-glycoside	(23)
8	Kaempferol 3-O-[α -L-rhamnopyranosyl-(1 \rightarrow 3)-2,4-di-O-acetyl- α -L-rhamnopyranosyl-(1 \rightarrow 6)]- β -D-glucopyranoside	(23)
9	Kaempferol 3-O-[4-O-acetyl- α -L-rhamnopyranosyl-(1 \rightarrow 3)-2,4-di-O-acetyl- α -L-rhamnopyranosyl-(1 \rightarrow 6)]- β -D-glucopyranoside	(23)
10	Kaempferol 3-O-[2,3,4-tri-O-acetyl- α -L-rhamnopyranosyl-(1 \rightarrow 3)-4-O-acetyl- α -L-rhamnopyranosyl-(1 \rightarrow 6)]- β -D-glucopyranoside	(23)
11	Kaempferol 3-O-[2,3,4-tri-O-acetyl- α -L-rhamnopyranosyl-(1 \rightarrow 3)-2,4-di-O-acetyl- α -L-rhamnopyranosyl-(1 \rightarrow 6)]- β -D-glucopyranoside	(23)
12	Kaempferol 3-O-(6"-O-trans-p-coumaroyl)- β -D-glucopyranoside	(23)
13	Kaempferol 8-methoxy-3-O-(6"-O-trans-p-coumaroyl)- β -D-glucopyranoside	(23)
14	Kaempferol 8-methoxy-3-O-(3"-O-trans-p-coumaroyl)- β -D-glucopyranoside	(23)
15	Kaempferol 3-O-(3",6"-O-trans-p-coumaroyl)- β -D-glucopyranoside	(23)
16	Quercetin	(23)
17	Quercetin-3-O- β -D-glucoside	(23)
18	Quercetin-3'-O- β -D-glucoside	(23)
19	Quercetin-7-O- β -D-glucoside	(23)
20	Rutin	(23)
21	Quercetin-3-O-[α -L-rhamnosyl-(1 \rightarrow 2)- β -D-glucosyl]-5-O- β -D-glucoside	(9)
22	Dihydrokaempferol	(2)
23	Dihydroquercetin	(2)
24	Aromadendrin	(14)
Flavones		
25	Apigenin	(24)
26	Vitexin	(7)
27	Apigenin 6,8-di-C- β -glucopyranoside	(24)
28	Apigenin 6-C-pentoside-8-C-hexoside	(9)
29	Luteolin-7-O-rutinoside	(24)

Table 2 (continued)

Table 2 (continued)

Classes/compound No.	Compound names	References
Flavan-3-ols		
30	Epicatechin	(19)
31	Catechin	(19)
32	Gallocatechin gallate	(19)
33	Epigallocatechin	(19)
34	Epicatechin gallate	(19)
35	Catechin gallate	(19)
36	Epigallocatechin gallate	(19)
37	Gallocatechin	(19)
38	Procyanidin dimer	(22)
39	Procyanidin trimer	(22)
40	Procyanidin tetramer	(22)
41	Procyanidin pentamer	(22)
Anthocyanins		
42	Pelargonium-3-O-glucoside	(21)
43	Cyanidin-3-O-glucoside	(21)
Phenolic acids		
44	Gallic acid	(20)
45	Salicylic acid	(23)
46	Chlorogenic acid	(23)
47	Protocatechuic acid	(23)
48	Ellagic acid	(24)
49	3'-Methy-4'-glucoside-ellagic acid	(24)
50	Okicamelliaside	(24)
51	3'-Methyellagic acid	(24)
52	3,4-O,O-Methylidyne-ellagic acid	(24)
53	3,4-Methylenedioxy-3'-O-methyl-4'-O-(6'-O-acetyl-glucoside) ellagic acid	(25)
54	Ellagic acid-4-O-β-D-glucopyranoside	(25)
55	3,4-Methylenedioxy-3'-O-methyl-4'-O-glucoside ellagic acid	(25)
Lignans		
56	Eudesmin	(28)
57	(+)-Diasyringaresinol	(28)
58	(+)-Isoeucommin A	(28)
59	Pinoresinol 4-O-glucoside	(28)
60	7S, 8R, 8'R-(-)-lariciresinol-4'-O-D-glucopyranoside	(28)
61	(+)-Isolariciresinol 9-O-β-D-glucopyranoside	(28)
62	(+)-Isolariciresinol 9'-O-β-D-glucopyranoside	(28)
63	3', 4-O-Dimethylcedrusin	(28)



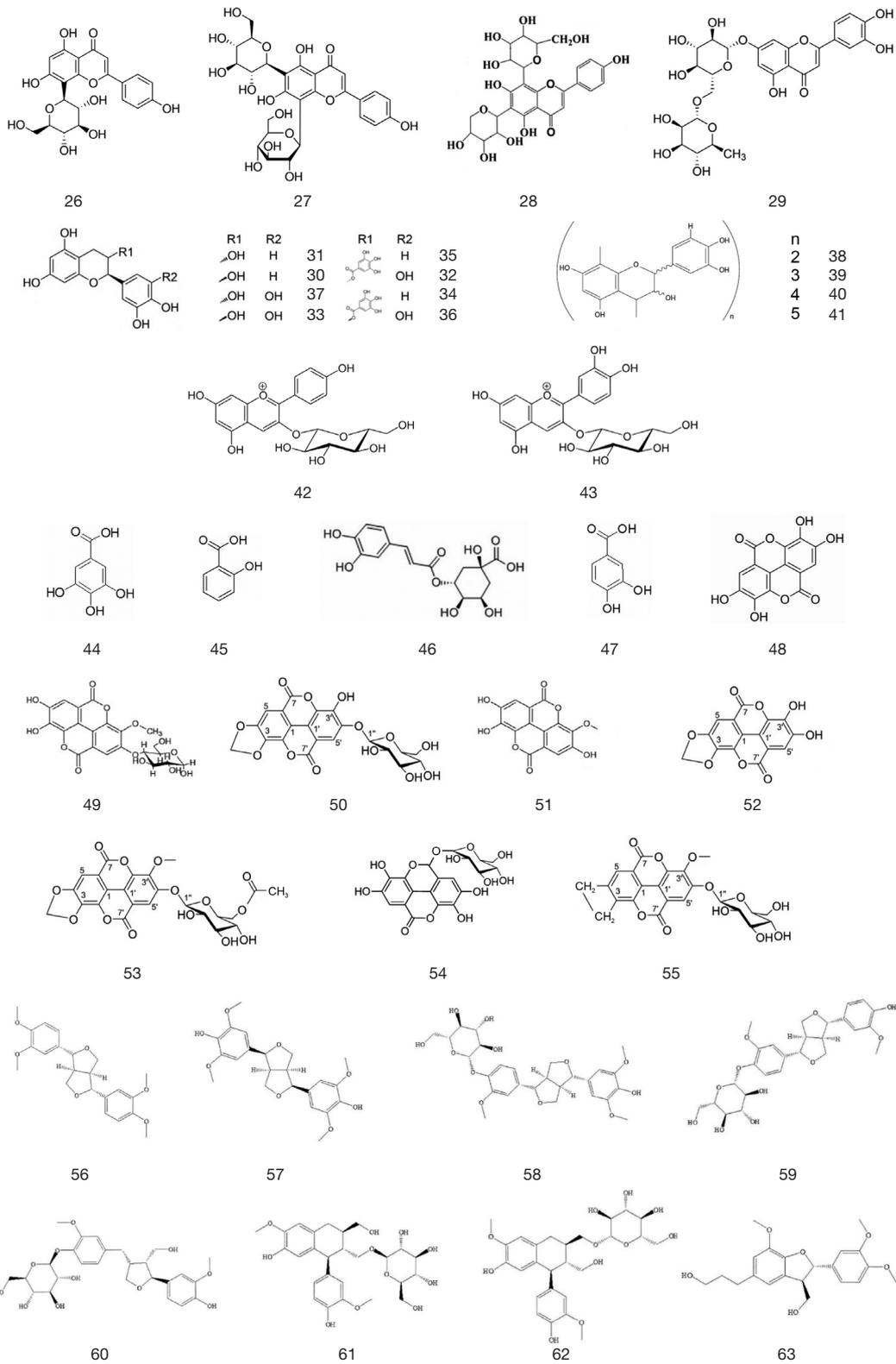


Figure 2 Chemical structures of phenolic compounds in *Camellia nitidissima* Chi. A code is marked under each molecular structure. The name of the chemical can be found via checking the code in Table 2.

Saponins are characterized by their structure containing a steroid or triterpene aglycone and one or more sugar moieties. Steroidal saponins almost exclusively occur in monocotyledonous angiosperms, while triterpenoid saponins more frequently occur in dicotyledonous angiosperms (30).

Saponins are observed in different organs of *C. nitidissima* and the saponin contents were flowers > fruit shells > leaves > buds (31). Saponins in *C. nitidissima* mainly belong to ursane-type tetracyclic triterpenoids, lupane-type pentacyclic triterpenes, and oleanolane-type pentacyclic triterpenes. Su *et al.* (32) isolated three ginsenosides from *C. nitidissima* leaves, which were ginsenoside Rg1, ginsenoside F1, and ginsenoside F5. Wei *et al.* (33) demonstrated the presence of ilexside II in the water extract of *C. nitidissima* leaves. Mo (34) identified a new dammarane-type saponin from *C. nitidissima*, i.e. (3 β ,6 α ,12 β)-3,6,12-trihydroxydammar-24-en-20-yl-2-O- β -D-glucopyranosyl-(2 \rightarrow 1)-O- β -D-glucopyranosyl-(2 \rightarrow 1)-O- α -L-rhamnopyranoside, and proved its anti-tumor activity. Yang (23) identified an oleanolane-type triterpene from *C. nitidissima* flowers, i.e., 3-O- β -D-galactopyranosyl-(1 \rightarrow 2)- β -D-glucuronopyranosyl-21 β ,22 α -di-O-angeloyl barringtonol C. Qi (14) identified four saponins from *C. nitidissima* leaves, i.e., 3 β -acetoxy-20-lupanol, 3 β ,6 α ,13 β -trihydroxyolean-7-one, 22 α -angeloyl-A1-barrigenol, and rubiprasin.

Polysaccharides

Polysaccharides are a group of biomolecules that are essential to all living organisms and are structurally composed of aldoses or ketoses linked by glycosidic bonds (35). They are widely distributed in plants, animals, algae, and microorganisms. Polysaccharides have a variety of biological activities, such as antioxidant, anti-hyperglycemia, anti-hyperlipidemia, anti-inflammation, anti-cancer, and immune enhancement (36).

Niu *et al.* (37) measured the polysaccharide content in the flowers, leaves, buds, and fruit shells of *C. nitidissima*, which were 32.88, 29.48, 35.89, and 30.02 g/kg, respectively. Tian (38) analyzed the sugar compositions of *C. nitidissima* polysaccharides, indicating that the *C. nitidissima* polysaccharides were composed of glucose, galactose, arabinose, mannose, rhamnose, and xylose. The former four monosaccharides were main components, accounting for 31%, 27%, 21%, and 13%, respectively. Some *C. nitidissima* polysaccharides not only contain monosaccharides, but

also combine with galacturonic acid (39). Gong *et al.* (40) obtained three *C. nitidissima* polysaccharides, i.e., TPS1, TPS2, and TPS3, using water extraction, alcohol precipitation, and DEAE cellulose anion exchange chromatography. TPS1 is composed of glucose, galactose, and arabinose. TPS2 and TPS3 are composed of rhamnose, galacturonic acid, galactose, and arabinose. TPS3 contained more galacturonic acid than TPS2. Among them, the antioxidant activity of TPS3 was the best. It implies that polysaccharides with higher content of galacturonic acid tend to possess higher antioxidant capacity. One of the mechanisms is that galacturonic acid has electron-withdrawing groups, such as carboxyl and hydroxyl groups, which provide more hydrogen ions to neutralize free radicals (41).

Tian *et al.* (42) isolated six polysaccharides from *C. nitidissima*, three of which belonged to neutral polysaccharides and three of which belonged to pectins. Structural analysis suggested that the three pectins were probably composed of a hairy region which had a backbone of alternating galacturonic acid and α -L-rhamnosyl residues and a smooth region which had a backbone of galacturonic acid residues (42). Lin *et al.* (43) analyzed the structure of a *C. nitidissima* polysaccharide, and the results suggested that the polysaccharide was composed of a smooth region with highly methyl esterified galacturonic acid residues and three hairy regions with different chemical structures. Due to these structural characteristics, it is no wonder that several *C. nitidissima* polysaccharides are digestion-resistant. Gong *et al.* (44) investigated the digestibility of three polysaccharides and found none of them were digestible. However, all of them showed prebiotic activity. They promoted the proliferations of *Lactobacillus* and *Bifidobacterium*, and increased the production of short-chain fatty acids.

Others

C. nitidissima contains multiple mineral elements, such as Ca, Mg, Na, K, P, Cu, Fe, Zn, B, Mn, Ni, and Mo. Some trace elements, such as Se, Sr, Cr, Ge, Co, Ga, and V, are also detected.

C. nitidissima are abundant in free amino acids. Zhao *et al.* (45) found that there were 16 types of amino acids in *C. nitidissima* leaves, including 7 essential amino acids. The content of free amino acids was 6% in old leaves and 5.37% in young leaves. Essential amino acids accounted for about 42% of total free amino acids in *C. nitidissima*

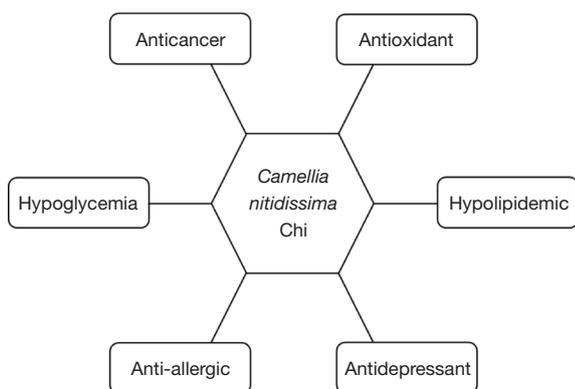


Figure 3 Main bioactivity of *Camellia nitidissima* Chi.

leaves. The content and composition of free amino acids in *C. nitidissima* flowers are not quite the same as that in *C. nitidissima* leaves. Huang *et al.* (46) revealed that 17 types of amino acids were observed in *C. nitidissima* flowers. The content of free amino acids in *C. nitidissima* flowers ranged from was 4.32% to 5.46%, reaching the top at the fish-mouth stage. Essential amino acids accounted for 38% of total free amino acids in *C. nitidissima* flowers.

Volatiles are components which contribute to the aroma. Some volatiles also act as bioactive compounds, playing roles in anti-bacteria, anti-virus, anti-depression, and so on. Though almost odorless, 45 volatiles were identified in *C. nitidissima* flowers (47). Elaidic acid, palmitic acid, and stearic acid accounted for over 30% of total volatiles. (E,E)-2,4-heptadienal, (E,E)-2,4-decadienal, and geranyl acetone, each possessed over 1.7% of total volatiles. Huang *et al.* (48) identified 37 volatiles in *C. nitidissima* leaves. Benzoic acid-2-hydroxy-methyl ester was the major volatile, accounting for 26.91% of total volatiles. Benzyl alcohol, cis-octahydropentalene, cis-linaloloxide, phenylethyl alcohol, and 2,6-dimethyl-3,7-octadiene-2,6-diol were relatively abundant in *C. nitidissima* leaves.

Phytosterols are a family of molecules related to cholesterol and serve as structural components of biological membranes of plants. α -spinasterol, α -spinasteryl- β -D-glucopyranoside, β -sitosterol, and stigmasta-7,22-diene-3-O-[α -L-arabinopyranosyl(1 \rightarrow 2)]- β -D-galactopyranoside are observed in *C. nitidissima* (14,23).

Main bioactivity of *C. nitidissima*

Although *C. nitidissima* has been traditionally used as an herbal medicine and regarded as health-beneficial, scientific

researches on the bioactivity of *C. nitidissima* have been merely conducted in recent two decades. Evidences indicate that *C. nitidissima* is potent in antioxidant, anti-cancer, anti-hyperglycemia, and anti-hyperlipidemia (7). Particularly, it works excellent in anti-allergy and anti-depression (7). In next subsections, the main bioactivity (Figure 3) and possible underlying mechanisms are introduced.

Antioxidant activity

At present, there are many studies on the antioxidant activity of *C. nitidissima*. Wei *et al.* (49) proved that the ethanol extract of *C. nitidissima* leaves had hydroxyl radical (\cdot OH) scavenging, superoxide anion radical ($O_2^{\cdot-}$) scavenging, 2,2-diphenyl-1-picrylhydrazyl radical (DPPH \cdot) scavenging activity and reducing power. Qin *et al.* (50) found that the water extract of *C. nitidissima* leaves dose-dependently scavenged \cdot OH and $O_2^{\cdot-}$. At the concentration of 1.25 mg/mL, $O_2^{\cdot-}$ was completely scavenged. Yang *et al.* (18) found that the n-butanol extract of *C. nitidissima* flowers had a strong inhibitory effect on AGEs. Wen *et al.* (51) used 95% ethanol to extract the leaves, stamens, buds and petals of *C. nitidissima*. All the above parts had antioxidant activity, and the buds had the strongest antioxidant activity while the leaves had the weakest.

Phenolic compounds are vital for the antioxidant activity of *C. nitidissima*. The *C. nitidissima* flavonoids showed good antioxidant activity with an IC_{50} of 0.070 mg/mL for the scavenging of DPPH \cdot and an IC_{50} of 0.679 mg/mL for the scavenging of \cdot OH (52). Song *et al.* (15) analyzed the total phenolic content and antioxidant capacity of six types of *C. nitidissima* leaves by HPLC and liquid chromatography-electrospray ionization-tandem mass spectrometry (LC-ESI-MS), and concluded that the antioxidant capacity was correlated with the total phenolic content. Kaempferol derivatives and quercetin derivatives, well-known for their antioxidant activity, were abundant in *C. nitidissima* and possibly contributed to the superior antioxidant activity of *C. nitidissima*. Song *et al.* (15) found that the dichloromethane and ethyl acetate fractions of *C. nitidissima* were similarly effective in inhibiting the formation of AGEs in the bovine serum albumin (BSA)-glucose reaction system, while the ethyl acetate fraction was more effective in inhibiting the formation of AGEs in the BSA-methylglyoxal reaction system. Mono- and di-methylglyoxal quercetin adducts were detected in the reaction systems, suggesting that quercetin derivatives inhibited the formation of AGEs by scavenging

methylglyoxal. Catechins inhibited the formation of AGEs using the same strategy, i.e., by reacting with methylglyoxal to form adducts (23).

Saponins also play a role in the antioxidant activity of *C. nitidissima*. Ning *et al.* (53) used XAD16 macroporous adsorbent resin to isolate and purify *C. nitidissima* saponins. The crude saponin extract effectively scavenged a variety of free radicals and even worked better than Vitamin C in scavenging $\cdot\text{OH}$ and H_2O_2 . Su *et al.* proved that ginsenoside F1, which was extracted from *C. nitidissima* leaves, protected HepG2 cells from H_2O_2 -induced oxidative damage by increasing the superoxide dismutase (SOD) activity (54,55). The results provided a scientific basis for the effectiveness of *C. nitidissima* on antioxidation in cells.

Polysaccharides from *C. nitidissima* show antioxidant activity as well. Song *et al.* (56) isolated three polysaccharides with the β -pyranose configuration from *C. nitidissima* leaves and demonstrated that the antioxidant activity of neutral polysaccharide was weaker than that of the two acidic polysaccharides, implying the polarity affected the antioxidant activity of polysaccharides. He (41) proved that *C. nitidissima* polysaccharides with higher content of glucuronic acid exhibited stronger antioxidant activity, which fitted the theory.

Anticancer activity

Multiple researches have proved that *C. nitidissima* exhibits its anticancer activity not only by preventing the initiation and promotion of cancer, but also the progression of cancer.

C. nitidissima has the potential to be a chemoprevention agent. Daily consumption of diet containing 5% *C. nitidissima* leaves or 5% *C. nitidissima* leave extract significantly decreased diethylnitrosamine-induced precancerous lesion of liver in rats (57). Daily intra-gastric administration of *C. nitidissima* leave extract for 73 weeks effectively reduced the incidence of aflatoxin B1-induced hepatocellular carcinoma and delayed aflatoxin B1-induced hyperplasia in rats (58). The underlying mechanisms included the inhibition of *C. nitidissima* leave extract on cytochrome P450 enzyme 3A4 (CYP3A4) and glutathione S-transferase π (GST- π), two enzymes mediating the metabolism of aflatoxin B1, as well as the down-regulation of the aflatoxin B1-induced expression of signal transducer and activator of transcription 3 (STAT3), a transcriptional factor well-validated to promote tumorigenesis. Moreover, Sai (59) found that the intra-gastric administration of *C. nitidissima* flower extract for 16 weeks reduced the

incidence of lung tumors by about 13% in an uratan-treated mouse model. The catalase and SOD activity were increased while the malondialdehyde level was decreased in the *C. nitidissima* flower extract group. The interleukin-2 and tumor necrosis factor α levels were also increased. It suggested that *C. nitidissima* flower extract exerted the chemopreventive activity via enhancing the antioxidant and immunomodulatory activity of mice.

In addition to preventing cancer, *C. nitidissima* directly inhibits cancer via affecting the proliferation, apoptosis, cell cycle, and migration of cancer cells.

C. nitidissima reduces the proliferation of various cancer cells *in vitro*, for example, gastric carcinoma MGC-803 cells (60), esophageal squamous carcinoma Eca-109 cells (61), leukemia U937 cells (62), cervical carcinoma Hela cells (63) and prostate cancer PC-3 cells (64). The effective parts of *C. nitidissima* include flowers, leaves, and seeds. Yu *et al.* (65) showed that the alcoholic extracts of *C. nitidissima* flowers, seeds, and leaves inhibited the proliferation of U937 cells, and the first two extracts also dose-dependently inhibited human colon cancer HCT116 cells.

C. nitidissima extracts promote the apoptosis of cancer cells. Zhao (63) found that *C. nitidissima* flower extract time-dependently and dose-dependently triggered the apoptosis of Hela cells. Sai (59) demonstrated that the *C. nitidissima* flower extract upregulated the expression of Bax, a pro-apoptotic protein in the Bcl-2 family, led to the depolarization of the mitochondrial membrane potential, initiated the mitochondrial apoptotic pathway, and eventually caused the intrinsic apoptosis of lung carcinoma A549 cells.

In some cases, *C. nitidissima* extracts induce cell cycle arrest of cancer cells. Li (60) proved that *C. nitidissima* flower extract dose-dependently halted MGC-803 cells at the S and G2 cell cycle phases. Shen (66) demonstrated that *C. nitidissima* ethanol extract blocked the cell cycle of nasopharyngeal carcinoma CNE-2 cells at the G1 phase, induced apoptosis by activating Caspase-3, and down-regulated the expression of vascular endothelial growth factor C and vascular endothelial growth factor receptor 3, two molecules which were associated with the migration of CNE-2 cells.

At present, most researches on the anticancer property of *C. nitidissima* are carried out using crude extracts. Only a few studies pinpoint the anticancer components of *C. nitidissima*. Learning from current studies, saponins may be key anticancer components in *C. nitidissima*. Mo (34)

proved that a dammarane-type saponin from *C. nitidissima* effectively inhibited the growth of Bel-7402 and SMMC-7721 cells *in vitro*. Jing *et al.* (8) verified that $3\beta,6\alpha,13\beta$ -trihydroxyolean-7-one, which was a saponin extracted from *C. nitidissima*, showed potential cytotoxic activity against SGC7901 cells *in vitro*. 22α -Angeloyl-A1-barrigenol, another saponin from *C. nitidissima*, significantly inhibited A549 cells, human gastric carcinoma HGC-27 cells, human breast cancer MDA-MB-435 cells, and human colorectal cancer SW620 cells (8).

Although *C. nitidissima* has toxicity to cancer cells, it does not hurt normal cells. Li (67) found that *C. nitidissima* flowers extract had no toxic side effects on human normal liver HL-7702 cells. Sai (59) demonstrated that *C. nitidissima* extract had no subchronic toxicity in mice. The above results reveal that *C. nitidissima* is highly selective and effective in inhibiting cancer cells, meanwhile it has low toxicity to normal cells and causes little side effects. It suggests that this plant has a great potential as an anticancer drug candidate.

Hypoglycemic and hypolipidemic activity

Overnutrition is a form of malnutrition in which the intake of nutrients is oversupplied (68). It adversely affects health, causing symptoms like hypoglycemia and hypolipidemia. It may further increase the risks of chronic metabolic diseases, such as diabetes and atherosclerosis.

C. nitidissima leave extracts have excellent hypoglycemic activity. *C. nitidissima* leave n-butanol and ethyl acetate extracts increased the glucose consumption of insulin-resistant HepG2 cells and decreased the fasting blood glucose and postprandial blood glucose levels in type 2 diabetic mice (69). In another type 2 diabetic mouse model, intra-gastric administration of *C. nitidissima* leave extract for 28 days increased the insulin level, attenuated pancreatic injury, and promoted the accumulation of hepatic glycogen (70). Feng *et al.* investigated the effect of *C. nitidissima* leave extract capsules on lowering blood glucose in diabetic patients. The results supported that *C. nitidissima* leave extract capsules were effective for the adjuvant therapy of diabetes (71).

C. nitidissima flower extracts display hypolipidemic activity. *C. nitidissima* flower extract significantly decreased oleic acid-induced lipid accumulation in HepG2 cells by inhibiting the mRNA expression of lipogenesis-related fatty acid synthase, 3-hydroxy-3-methyl glutaryl coenzyme A reductase, and glycerol-3-phosphate acyltransferase

genes. It significantly reduced the total triglycerides, total cholesterol, and low-density lipoprotein cholesterol, while increased the high-density lipoprotein cholesterol in serum of hyperlipidemic mice (72). Phenolic compounds may play an important role in it. Zhang (9) observed that *C. nitidissima* flower flavonoid extract decreased food intake by upregulating the secretion of glucagon-like peptide-1, a hormone negatively regulating the appetite. It inhibited the activity of α -amylase, α -glucosidase, pancreatic lipase, and cholesterol esterase, and decreased the solubility of cholesterol micelles, thus interfering the digestion and absorption of carbohydrates and lipids. In high-fat-diet-induced rats, it reduced lipogenesis, promoted lipolysis and lipid oxidation, attenuated triglycerides and cholesterol accumulation in serum and liver, and alleviated hepatic lipotoxicity. It improved impaired glucose tolerance and restored insulin sensitivity. Additionally, it alleviated high-fat diet-induced dysbiosis.

Anti-allergic activity

Allergy is a number of conditions caused by hypersensitivity of the immune system. Type I hypersensitivity, known as the immediate-type reaction, can be triggered by pollen, foods, drugs, and insect stings. It involves immunoglobulin E (IgE)-mediated release of antibodies against the antigen, degranulation of mast cell, and release of inflammatory factors (e.g., histamine), resulting in symptoms like itch, edema, and pain (73).

C. nitidissima leave water extract and *C. nitidissima* fruit peel ethyl acetate extract effectively alleviated ovalbumin and Al(OH)₃ mixture-induced type I allergy in mice (74). The serum IgE and leukotriene levels were reduced, the number of eosinophils in blood and bronchoalveolar lavage fluid were decreased, and the inflammation in lung was attenuated.

Okicamelliaside, an ellagic acid derivative which exists in *Camellia japonica* and *C. nitidissima* leaves, is considered to be the major anti-allergic agent in *C. nitidissima*. It was 12,000 times more potent than ketotifen fumarate, an antihistamine drug, in inhibiting the degranulation of RBL-2H3 cells (75). It significantly inhibited the vascular hyperpermeability in a passive cutaneous anaphylaxis mouse model. Further study indicated that okicamelliaside inhibited antigen-IgE-Fc ϵ RI-induced activation of the Lyn-Syk-LAT-PLC γ -1 pathway, blocked the release of Ca²⁺, decreased the expression of proinflammatory cytokines (e.g., interleukin-4 and interleukin-13), cytokine-producing

signaling factors, and prostaglandin-endoperoxidase 2, resulting in the suppression of allergic inflammation.

Kaempferol 3-O- β -D-glucosyl(1 \rightarrow 3) [α -L-rhamnosyl(1 \rightarrow 6)]-(2-O-E-*p*-coumaroyl- β -D-glucoside), a flavonol glucoside obtained from *C. nitidissima* water extract, is another promising anti-allergic agent in *C. nitidissima*. It significantly inhibited lipoxygenase activity and leukotriene production *in vitro* (76).

Antidepressant activity

Depression is a common but serious mood disorder. It causes a persistent feeling of sadness and loss of interest. Nowadays, the stress of life increases. Along with it, is the increasing incidence of depression. Current clinical antidepressant drugs are chemical synthetic drugs, which shows several side effects. Therefore, novel antidepressant drugs with less toxic side effects have come into the limelight.

C. nitidissima contains a variety of natural active ingredients with antidepressant effects, such as quercetin (77), kaempferol (78) and ginsenoside Rg1 (79). *C. nitidissima* extract significantly decreased corticosterone-induced apoptosis of differentiated PC12 neuronal cells by increasing the expression of brain-derived neurotrophic factor (BDNF) via the protein kinase A-cAMP-response element binding protein signaling pathway (80). It indicated that *C. nitidissima* extract was capable of protecting neurons. In a chronic unpredictable mild stress rat model, *C. nitidissima* extract alleviated the decrease of body weight and loss of interest in sucrose. Immunohistochemistry staining and Hematoxylin and Eosin staining confirmed that *C. nitidissima* extract attenuated the hippocampus injury by increasing the expression of BDNF. Serum corticosterone and adrenocorticotrophic hormone, which were increased under depression, was decreased. At the same time, serum SOD and glutathione peroxidase activity were increased while serum malondialdehyde levels were decreased, implying *C. nitidissima* extract attenuated depression-induced oxidative stress in the body. In mice, *C. nitidissima* extract also effectively alleviated the depression symptoms. Compared with mice in the model group, the brain and serum serotonin, dopamine, and norepinephrine levels of mice administering *C. nitidissima* extract were increased. These results suggest that *C. nitidissima* extract displays its antidepressant activity via multiple targets and it has the promise to be applied in the treatment of depression.

Conclusions

C. nitidissima, though merely being taxonomically classified within a century, has a long history being used as an herb. Chemical analysis reveals that phenolic compounds, saponins, and polysaccharides are important components in *C. nitidissima*. Some of them are unique in *C. nitidissima* (e.g., nitidissimol A and some complex flavonol glucosides), some are featuring components in the *Camellia* genus (e.g., okicamelliaside), and some are commonly distributed in plants (e.g., kaempferol, quercetin, and epicatechin). Biological experiments prove that *C. nitidissima* exhibits multiple physiological functions, particularly in antioxidant, anti-cancer, anti-hyperglycemia, anti-hyperlipidemia, anti-allergy, and anti-depression. Scientific evidences of the pharmacological value of *C. nitidissima* and corresponding chemical basis are partially established.

It is noteworthy that most of the current biological studies of *C. nitidissima* are based on crude extract without a clear description of the chemical profile, which makes it difficult to figure out the predominant bioactive component. Some bioactivity of *C. nitidissima* is verified *in vitro*, whether it works *in vivo* or not still remains unknown. In addition, current understanding of the molecular mechanisms of *C. nitidissima* is relatively preliminary. In the future, more attentions should be drawn on the bioactivity of individual component in *C. nitidissima*. The assessments are recommended to be conducted both *in vitro* and *in vivo*. Detailed working mechanisms are encouraged to be explored. Researches of pharmacokinetics and pharmacodynamics are also necessary. By ascertaining these properties of individual component, investigations on interactions between bioactive components can be carried out more easily. The above information will help us better understand why *C. nitidissima* is capable of a specific physiological function and how it exhibits the activity.

It is also aware that some empirical therapeutic activity of *C. nitidissima* still lacks scientific proof. Little is known about the activity of some special compounds in *C. nitidissima*. Future researches on these aspects are needed. The results will certainly enhance the knowledges of *C. nitidissima* and benefit the applications of *C. nitidissima* in health industry.

Acknowledgments

Funding: This research was supported by the China Agriculture Research System of MOF and MARA (CARS-

19) and the Innovation Project for the Chinese Academy of Agricultural Sciences.

Footnote

Reporting Checklist: The authors have completed the Narrative Review reporting checklist. Available at <https://lcm.amegroups.com/article/view/10.21037/lcm-22-9/rc>

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <https://lcm.amegroups.com/article/view/10.21037/lcm-22-9/coif>). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Open Access Statement: This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

References

- Zhou XW, Fan ZQ, Chen Y, et al. Functional analyses of a flavonol synthase-like gene from *Camellia nitidissima* reveal its roles in flavonoid metabolism during floral pigmentation. *J Biosci* 2013;38:593-604.
- Wang W, Liu H, Wang Z, et al. Phytochemicals from *Camellia nitidissima* Chi inhibited the formation of advanced glycation end-products by scavenging methylglyoxal. *Food Chem* 2016;205:204-11.
- Chang B, Huang G. The classification and geographic distribution of *Camellia nitidissima*. *Journal of Wuhan Botanical Research* 1986;4:31-42.
- Zhang X, Feng J, Su S, et al. Hepatoprotective effects of *Camellia nitidissima* aqueous ethanol extract against CCl₄-induced acute liver injury in SD rats related to Nrf2 and NF- κ B signalling. *Pharm Biol* 2020;58:239-46.
- Liang SY, Lu MZ, Huang LD. The cultivation and utilization of *Camellia nitidissima*. Beijing: China Forestry Press, 2005.
- Ye CX. Notes on the change of the scientific name of *Camellia nitidissima*. *Guangxi Plants* 1997;4:22-6.
- Dong Y, He, Xiao Y, et al. *Camellia nitidissima* C.W. Chi: a review of botany, chemistry, and pharmacology. *Phytochemistry Reviews Proceedings of the Phytochemical Society of Europe* 2018;17:327-49.
- Jing Q, Ruo FS, Jian MY, et al. Chemical constituents from leaves of *Camellia nitidissima* and their potential cytotoxicity on SGC7901 cells. *Chinese Herbal Medicines* 2016;8:80-4.
- Zhang HL. Anti-food-induced obesity effect and mechanism of *Camellia nitidissima* flavonoids. Guangdong Ocean University, 2020.
- Huang YL, Wen YX, Liu JL, et al. Determination of total flavonoids in five kinds of *Camellia nitidissima*. *China Science and Technology of Traditional Chinese Medicine* 2009;16:38-9.
- Tang Q, Luo YY, Huang LD, et al. Determination of Chemical Constituents in Section *Chrysanthemum*. *Lishizhen Medicine and Materia Medica Research* 2009;20:769-71.
- Huang YZ, Chen JY, Zhang WJ, et al. Comparison of quality and yield of *Camellia nitidissima* in different flowering stages. *Fujian Agricultural Journal* 2021;36:899-908.
- Huang XX, Zou R, Hu XH, et al. Comparison of total flavonoids content in 14 species of *Camellia nitidissima* sect. *Chrysanthemum*. *Guihaia* 2011;31:281-4.
- Qi J. Isolation, identification and activity evaluation of chemical constituents of *Camellia nitidissima*. Nanjing University of Science and Technology, 2016.
- Song L, Wang X, Zheng X, et al. Polyphenolic antioxidant profiles of *Camellia nitidissima*. *Food Chem* 2011;129:351-7.
- Peng X, Yu DY, Feng BM, et al. Study on the chemical constituents of *Camellia nitidissima*. *Guihaia* 2011;31:550-3.
- Yang R, Guan Y, Wang W, et al. Antioxidant capacity of phenolics in *Camellia nitidissima* Chi flowers and their identification by HPLC Triple TOF MS/MS. *PLoS One* 2018;13:e0195508.
- Yang R, Wang WX, Chen HJ, et al. The inhibition of advanced glycation end-products by five fractions and three main flavonoids from *Camellia nitidissima* Chi flowers. *J Food Drug Anal* 2018;26:252-9.
- Jiang LN, Li JY, Fan ZQ, et al. Analysis of polyphenol components in flowers of *Camellia nitidissima* plants.

- Forestry Science Research 2020;33:117-26.
20. Yan DM, Li RJ. Determination of 5 kinds of phenolic substances in *Camellia nitidissima* by high performance liquid chromatography. *Journal of Henan University of Technology (Natural Science Edition)* 2010;31:59-62.
 21. Li XL, Wang JT, Sun ZY, et al. UPLC-QTOF-MS analysis of *Camellia nitidissima* flowers and leaves. *Scientific Research in Forestry* 2018;31:83-8.
 22. Zhang HL, Yu QT, Wu QX, et al. On-line screening of flavonoids from *Camellia nitidissima* and in vivo antioxidant activity based on iron ion interaction combined with high performance liquid chromatography. *Research and Development of Natural Products* 2020;32:719-26.
 23. Yang R. The chemical constituents of *Camellia nitidissima* and its biological activities based on quorum sensing inhibition. *Nanjing University of Science and Technology*, 2019.
 24. Yu J. Analysis of polyphenolic compounds in *Camellia nitidissima* leaves. *Guangxi University of Traditional Chinese Medicine*, 2017.
 25. Mo JG, Chen QH, Huang Y, et al. A new ellagic acid compound in *Camellia nitidissima*. *Chinese Herbal Medicine* 2018;49:75-9.
 26. Cheng CJ, Cong LF, Li ZQ, et al. Screening, preparation and investigation of antitumor activity of Okicamelliaside in *Camellia nitidissima*. *Traditional Chinese Medicine in Tianjin* 2020;37:1425-30.
 27. Wcislo G, Szarlej-Wcislo K. *Colorectal Cancer Prevention by Wheat Consumption: A Three-Valued Logic-True, False, or Otherwise?* Elsevier Inc., 2014.
 28. Zhang PP, Wang ZN, Yang R, et al. Analysis of chemical constituents of lignans in *Camellia nitidissima*. *Acta Tropical Biology* 2020;11:296-300, 323.
 29. AAT Intégral. *Saponins: properties, applications and processing*; 2010.
 30. Sparg SG, Light ME, van Staden J. Biological activities and distribution of plant saponins. *J Ethnopharmacol* 2004;94:219-43.
 31. Niu GJ, Xing JH, Zhu S, et al. Determination of active components and antioxidant activity of *Camellia nitidissima*. *Journal of Forestry and Environment* 2015;35:165-8.
 32. Su L, Mo JG, Wei YL, et al. Study on saponins components of *Camellia nitidissima*. *Chinese Herbal Medicine* 2012;43:3.
 33. Wei JB, Nong CL, Su ZH, et al. A preliminary study on in vitro antitumor activity and material basis of *Camellia nitidissima*. *Chinese Journal of Experimental Formulas* 2014;20:169-74.
 34. Mo JG. *Camellia saponin A and its preparation method and antitumor use*. Guangxi Zhuang Autonomous Region, Guangxi Zhuang Autonomous Region Analysis and Testing Research Center, 2017.
 35. Ullah S, Khalil AA, Shaukat F, et al. Sources, Extraction and Biomedical Properties of Polysaccharides. *Foods* 2019;8:304.
 36. Yu Y, Shen M, Song Q, et al. Biological activities and pharmaceutical applications of polysaccharide from natural resources: A review. *Carbohydr Polym* 2018;183:91-101.
 37. Niu GJ, Zhu S, Chen QY, et al. Determination and in vitro antioxidant activity of polysaccharides from different parts of *Camellia nitidissima*. *Chinese Journal of Experimental Formulas* 2014;20:168-72.
 38. Tian XC. *Isolation, purification and chemical structure study of polysaccharides from Camellia nitidissima*. Guangdong Ocean University; 2011.
 39. Wang DF, Li J, Wang CH. Study on the component and immune activity of polysaccharides from tea. *J Tea Sci* 2000;20:45-50.
 40. Gong W, Tang J, Wei YY, et al. Isolation, purification, structural characterization and in vitro antioxidant activity of polysaccharides from *Camellia nitidissima*. *Food and Machinery* 2021;37:184-90.
 41. He Z. *Isolation, Purification, Antioxidant and Immune Activity of Ashwagandha Polysaccharide*. Beijing: Chinese Academy of Forestry Sciences, 2014.
 42. Tian XC, Qin XM, Lin HJ, et al. Study on the physicochemical properties of polysaccharides from *Camellia nitidissima*. *Chinese Journal of Foodstuffs* 2011;11:47-52.
 43. Lin HJ, Tian XC, Qin XM, et al. Analysis of chemical structure characteristics of single polysaccharide components in *Camellia nitidissima*. *Food Science* 2013;34:6.
 44. Gong W, Tang J, Wei YY, et al. Study on in vitro digestion and fermentation characteristics of polysaccharides from *Camellia nitidissima*. *Food Industry Science and Technology* 2021;42:9.
 45. Zhao HJ, Luo ZR, Ding YL, et al. Analysis of nutrients in old and young leaves of *Camellia nitidissima*. *Journal of Inner Mongolia Agricultural University (Natural Science Edition)* 2016;37:52-6.
 46. Huang YZ, Chen JY, Zhang WJ, et al. Comparison of quality and yield of *Camellia nitidissima* in different flowering stages. *Fujian Agricultural Journal* 2021;36:10.
 47. Wei Q, Zhang LY. Comparative analysis of two kinds of

- Camellia nitidissima aroma components. *Modern Food Science and Technology* 2013;29:668-72.
48. Huang YL, Chen YY, Wen YX, et al. GC-MS analysis of volatile components in *Camellia nitidissima*. *Food Science and Technology* 2009;34:257-60.
 49. Wei X, Huang XX, Jiang YS, et al. Comparison of antioxidant activities of plant extracts from three *Camellia nitidissima* groups. *China Journal of Traditional Chinese Medicine* 2011;36:639-41.
 50. Qin XM, Lin HJ, Ning EC, et al. Antioxidant activity of water extracts from *Camellia nitidissima*. *Food Science and Technology* 2008;(02):189-91.
 51. Wen J, Liang W, Wang XC, et al. Study on the chemical constituents and anti-inflammatory and antioxidant activities of *Camellia nitidissima*. *Chinese Journal of Medicinal Chemistry* 2020;30:487-92.
 52. Xu JH, Liu MM, Qi DJ, et al. Optimization of aqueous two-phase extraction of total flavonoids from *Camellia nitidissima* and analysis of antioxidant activity. *Food Industry Science and Technology* 2022;43:155-61.
 53. Ning EC, Xin M, Wei L, et al. Antioxidative activity of saponins from *Camellia nitidissima*. *Food Science and Technology* 2009;34:197-9.
 54. Su L, Mo JG, Wei YL, et al. Chemical constituents of saponins from leaves of *Camellia nitidissima*. *Chinese Traditional and Herbal Drugs* 2012;43:877-9.
 55. Wen L, Guo XB, Liu RH, et al. Phenolic contents and cellular antioxidant activity of Chinese hawthorn "*Crataegus pinnatifida*". *Food Chem* 2015;186:54-62.
 56. Song LL, Wen G, Huo SH, et al. Isolation, purification, structural characteristics and antioxidant activity of polysaccharides from turmeric. *Food and Fermentation Industry* 2020;46:73-9.
 57. Tang XL, Fu JY, Duan XX, et al. Preliminary study on the inhibitory effect of *Camellia nitidissima* and *Ginkgo biloba* on 2-ethylnitrosamine-induced precancerous lesions in rats. *Journal of Practical Cancer* 2007;22:4.
 58. Ou C, Cao J, Yang C, et al. The chemopreventive effect and mechanism of *Camellia nitidissima* in inhibiting aflatoxin B1-induced liver cancer in rats. *Proceedings of the National Academic Conference on Tumor Epidemiology and Tumor Etiology Collection* 2011:131-3.
 59. Sai X. Preliminary study on the preventive effect of *Camellia nitidissima* extract against lung cancer and its mechanism. *Dalian University of Technology*, 2018.
 60. Li L. Effects of *Camellia nitidissima* water extract on the proliferation and cycle of human gastric cancer MGC-803 cells. *Nanning: Guangxi Medical University*, 2013.
 61. Dai L, Li JL, Liang XQ, et al. Flowers of *Camellia nitidissima* cause growth inhibition, cell-cycle dysregulation and apoptosis in a human esophageal squamous cell carcinoma cell line. *Mol Med Rep* 2016;14:1117-22.
 62. He GL, Wang CY, Tan H, et al. Study on the inhibitory effect of the extract of *Camellia nitidissima* on human monocytic leukemia cell line U937 cells. *Bright Traditional Chinese Medicine* 2014;29:1382-4.
 63. Zhao YH. Effects of *Camellia nitidissima* water extract on the proliferation and apoptosis of human cervical cancer Hela cells. *Lanzhou: Lanzhou University*, 2015.
 64. Han LC, Shi LY, Yu DY, et al. Experimental study on the inhibitory effect of *Camellia nitidissima* seeds on hormone-related tumors in vitro. *Shi Zhen Guo Yi Guo Yao* 2009;20:3146-8.
 65. Yu DY, Shi ZS, Shi LY, et al. Experimental observation on the proliferation inhibition of U937 and HCT116 cells by extracts of *Camellia nitidissima*, seeds and leaves. *Chinese Patent Medicine* 2013;35:2005-7.
 66. Shen J. *Camellia nitidissima* inhibits the growth of human poorly differentiated nasopharyngeal carcinoma cells and the expression of VEGF-C\VEGFR-3 in vitro. *Guilin Medical College*, 2011.
 67. Li CY. Effects of different concentrations of *Camellia nitidissima* and *camellia* on precancerous lesions of rat liver induced by diethylnitrosamine. *Guangxi Medical University*, 2007.
 68. Mathur P, Pillai R. Overnutrition: Current scenario & combat strategies. *Indian J Med Res* 2019;149:695-705.
 69. Chen LM, Wang YJ, Xiao YM, et al. Screening of the hypoglycemic activity of Zhuang medicine *Camellia nitidissima*. *Modern Chinese Medicine Research and Practice* 2017;31:5.
 70. Xia X, Pan CS, Huang L, et al. Effects of *Camellia nitidissima* on pancreatic function in diabetic mice. *Shi Zhen Chinese Medicine and Chinese Medicine* 2013;24:3.
 71. Feng Q, Wang ZP, Xu K, et al. Clinical observation on adjuvant treatment of type 2 diabetes mellitus with *Camellia nitidissima* Capsule. *Guangxi Traditional Chinese Medicine* 2015;38:32-3.
 72. Zhang Ping. Study on the hypolipidemic effect of the flower extract of *Camellia nitidissima*. *Dalian University of Technology*, 2015.
 73. Abbas M, Moussa M, Akel H. Type I Hypersensitivity Reaction 2022.
 74. Wang YQ, Peng X, Tang Q, et al. Screening of plant anti-IgE-mediated type I allergic reaction in *Camellia*

- nitidissima. Zhongnan Pharmacy 2009;7:4.
75. Onodera K, Tsuha K, Yasumoto-Hirose M, et al. Okicamelliaside, an extraordinarily potent anti-degranulation glucoside. Biosci Biotech Bioch 2010;2532-4.
76. Shi LY, Yu DY. A kind of *Camellia nitidissima* flavonoid glycoside and its preparation method and use [patent]. Liaoning Province: CN103951723B, 2015.
77. Wang YQ, Wang YH, Zou MS, et al. Research progress on the antidepressant effect and mechanism of quercetin and its glycoside derivatives. Chinese Herbal Medicine 2022;53:1548-57.
78. Liang YD, Tan YG, Zhang S, et al. Effect and mechanism of kaempferol on depression-like behavior in aged rats with chronic stress and depression. Chinese Journal of Clinical Pharmacology 2020;36:4028-30 .
79. Huang Q, Chu SF, Lian XY, et al. Antidepressant effect of ginsenoside Rg1 and its mechanism of action. Acta Neuropharmacologica Sinica 2013:1-11.
80. He DY. Study on the antidepressant effect of the medicinal leaf extract of *Camellia nitidissima*. Dalian University of Technology, 2018.

doi: 10.21037/lcm-22-9

Cite this article as: Zheng H, Du Q, Yin J, Gao Y. A narrative review on the main chemical constituents and bioactivity of *Camellia nitidissima* Chi. Longhua Chin Med 2022;5:29.