



Updates in the advances of sporadic medullary thyroid carcinoma: from the molecules to the clinic

Yanhua Bai^{1#}, Dongfeng Niu^{1#}, Qian Yao¹, Dongmei Lin¹, Kennichi Kakudo²

¹Key Laboratory of Carcinogenesis and Translational Research (Ministry of Education), Department of Pathology, Peking University Cancer Hospital & Institute, Beijing, China; ²Department of Pathology and Thyroid Disease Center, Izumi City General Hospital, Izumi, Japan

Contributions: (I) Conception and design: Y Bai, D Niu, K Kakudo; (II) Administrative support: Y Bai, D Lin; (III) Provision of study materials or patients: All authors; (IV) Collection and assembly of data: All authors; (V) Data analysis and interpretation: All authors; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

[#]These authors contributed equally to this article.

Correspondence to: Yanhua Bai, MD, PhD. Key Laboratory of Carcinogenesis and Translational Research (Ministry of Education), Department of Pathology, Peking University Cancer Hospital & Institute, No. 52, Fucheng Road, Haidian District, Beijing 100142, China.

Email: byh0425@sina.com.

Abstract: Medullary thyroid carcinoma (MTC) is a rare neuroendocrine malignancy that originates in parafollicular cells. It is well-known that a quarter of MTC are involved in hereditary multiple endocrine neoplasia type 2 syndromes, whereas most MTC are sporadic. Unlike the commonly encountered gastrointestinal or pulmonary neuroendocrine tumors, most sporadic MTCs have distinct genetic alterations featured by somatic changes of either Rearranged during Transfection (*RET*) or *RAS* point mutation. The increasing application of next-generation sequencing, whole-exome sequencing, and other molecular detection techniques enables us to understand MTC comprehensively concerning its detailed molecular changes and their clinical correlations. This article reviews the advances in genetic alterations and their prognostic impact in sporadic MTC among different populations and discusses the associated tumor immune microenvironments and the potential role of immunotherapy targeting PD-L1/PD-1 in treating MTC. Furthermore, the current multikinase inhibitor targeting therapy for sporadic MTC has been summarized here and its efficacy and drug toxicity are discussed. Updates in advance of the role of calcitonin/procalcitonin/calcitonin-related polypeptide alpha (*CALCA*) gene transcripts in diagnosing and handling MTC are also mentioned. The treatment of advanced MTC is still challenging and might require a combination of several modalities.

Keywords: Sporadic medullary thyroid carcinoma (MTC); genetic alteration; Rearranged during Transfection (*RET*); *RAS*; immune microenvironment; immunotherapy; targeting therapy; multikinase inhibitor

Submitted Apr 06, 2020. Accepted for publication Aug 12, 2020.

doi: 10.21037/gs-2019-catp-21

View this article at: <http://dx.doi.org/10.21037/gs-2019-catp-21>

Introduction

Medullary thyroid carcinoma (MTC) arises from the parafollicular cells (C cells), and accounts for less than 5% of thyroid malignancies (1,2). As a neuroendocrine tumor, MTC secretes calcitonin, which leads to the elevation of serum calcitonin. Therefore, serum calcitonin becomes a vital marker to monitor the occurrence or recurrence

of MTC (3). Histologically, the frequent amyloid deposits and occasional psammoma bodies in the stroma differentiate MTC from other extrathyroid-originated neuroendocrine tumors. Unlike other neuroendocrine tumors, about a quarter of MTC appears as a predominant part of hereditary multiple endocrine neoplasia type 2 (MEN2) syndromes. With the involvement of other synchronous and metachronous lesions (including

pheochromocytoma, hyperparathyroidism, and mucosal neuroma), MEN2 is divided into three subtypes: MEN2A, MEN2B, and familial MTC (4). Most patients with MEN2A have germline mutations in the Rearranged during Transfection (*RET*) C634, whereas patients with MEN2B and FMTC have germline mutations in *RET* M918T (5,6). Somatic *RET* mutation is identified in a considerable proportion of sporadic MTCs, and *RET* M918T is the most frequent genetic change (7-19). *RAS* point mutations are usually mutually exclusive with *RET* mutations, and occur in 0-81% of *RET*-negative MTCs (12,15,17,20-25). Several independent studies have shown that *RET* and *RAS* mutations have prognostic significance (7,11,17,25-29). Hence, the prevalence of *RET* and *RAS* mutations in different populations might partly explain the difference in prognosis among different populations with sporadic MTC. Many genetic changes are found in the *RET* and *RAS*-negative cases, although their molecular mechanism and clinic significance are yet to be determined.

Immune checkpoint inhibitors targeting PD-L1/PD-1 are regarded as one of the most significant medical breakthroughs in the 21st century (30-32). Previous studies have revealed poor tumor-infiltrating lymphocytes, low expression of PD-L1 and low mutational burden in MTCs (24,33,34), which disfavor the application of immunotherapy. However, several reports from China have proved that PD-L1 expression is related to advanced disease. The positive correlation between PD-L1 expression and advanced disease may bring the dawn of immunotherapy in advanced MTC, and relevant clinical trials are expected (35,36). The American FDA has approved two multikinase inhibitor (MKI) drugs, vandetanib and cabozantinib, in the treatment of advanced MTC. A more specific *RET*-targeting tyrosine kinase inhibitor (TKI) is currently underdeveloped. The combination of TKI therapy and immunotherapy might represent a novel therapeutic option in the treatment of advanced MTC (1).

This article focuses on the advances in genetic alterations and their prognostic impact in sporadic MTC among different populations and discusses the potential role of immunotherapy targeting PD-L1/PD-1. Furthermore, the current multikinase inhibitor target therapy for sporadic MTC is summarized. Updates in advance of the role of calcitonin/procalcitonin/calcitonin-related polypeptide alpha (CALCA) gene transcripts in diagnosing and handling MTC are also mentioned.

RET mutations

The *RET* gene was firstly described in 1985, as a transforming gene encoding a receptor tyrosine kinase (37,38). *RET* is localized on 10q11.2, and consists of 21 exons (38,39). *RET* is a single-pass transmembrane protein, which contains the following functional domains: an extracellular domain with four repeated cadherin-like regions, a cysteine-rich region, a transmembrane domain, a broad intracellular tyrosine kinase domain, and a carboxy-terminal domain (allowing for three isoforms, RET 9, 43, and 51) (Figure 1) (40-42). The intracellular domain encompasses two tyrosine kinase subdomains (TKS1 and TKS2) that are involved in the activation of many intracellular signal transduction pathways (40-42).

As a receptor tyrosine kinase, *RET* plays a crucial role in cell signal transduction, and the germline mutation of *RET* leads to the destruction of cell proliferation and differentiation in tissues derived from neural crest cells, including parafollicular thyroid cells, parathyroid gland, adrenal medulla, and intestinal autonomic nerve plexus (5,40,43). The molecular mechanism of somatic *RET* mutation is supposed to be like germline mutation (44). *RET* protein performs its pivotal role in regulating cellular transformation, survival, differentiation, proliferation, and migration. The transformation is done through the binding of its ligand GFRα complex, triggering its homodimerization, phosphorylation of tyrosine residues, and initiation of several intracellular signaling cascades, including MAPK and PI3K pathways (Figure 1) (2,41,42,45,46).

Sporadic MTCs have various somatic gene point mutations and deletions. The prevalence of somatic *RET* mutations in sporadic MTCs varies from 19.4-88.9% among different populations (4,7-19,27). The lowest mutation rate of somatic *RET* is from a Chinese population, while the highest from the US population. There is a general agreement that *RET* T918M, a point mutation in the methionine residue in exon 16 corresponding to the intracellular tyrosine kinase receptor domain, is the most frequent somatic *RET* mutation, the germline mutation of which was originally known as the activating mutation in MEN 2B. Other mutations in the *RET* gene in sporadic MTC have been recognized at codons 609, 611, 618, 620, 630, 631, 632, 634, 636, 639, 641, 748, 766, 768, 876, 883, 884, 901, 908, 919, 922, and 930 at exons 10, 11, 13, 14, 15 and 16 (4,8,15,40,47-50).

There are many pieces of evidence that patients with

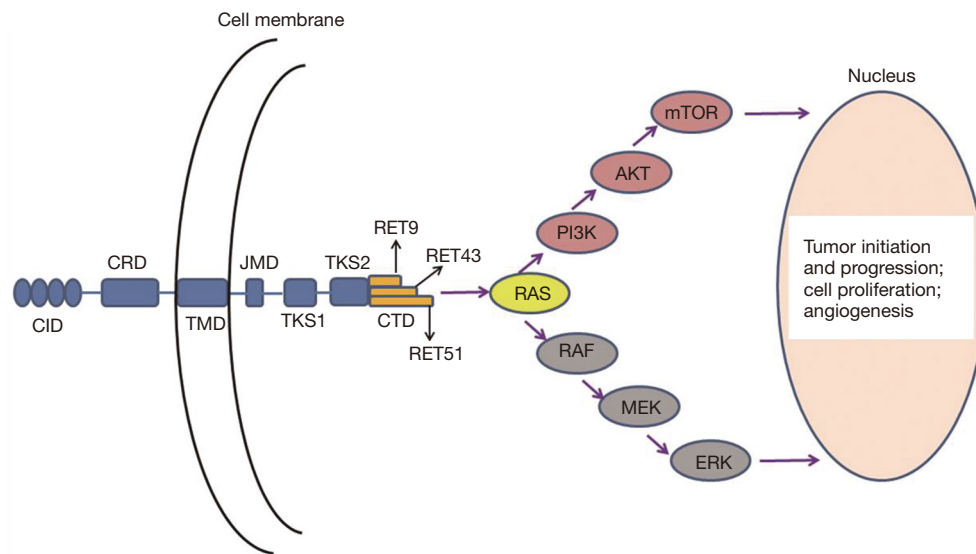


Figure 1 The structural scheme of RET protein and the principle activates downstream signal pathways. CID, cadherin-like domain; CRD, cysteine-rich domain; TMD, transmembrane domain; JMD, juxta-membrane domain; TKS, tyrosine kinase subdomain; CTD, carboxy-terminal domain.

sporadic MTC harboring *RET* mutations have a poorer prognosis than those harboring non-*RET* mutations (7,11,17,25-28,51). A statistically significant correlation was reached between *RET* mutations and the advanced tumor stage, higher T category, and lymph nodes or distant metastases and a worse patient outcome (17). In a recent meta-analysis involving 23 studies with 964 patients with MTC, *RET* mutation was determined to be associated with an elevated risk for lymph node metastasis, distant metastasis, advanced tumor stage, tumor recurrence, and patient mortality (28). Furthermore, tumors with somatic codon 918 mutations appear more aggressive than tumors with other *RET* mutations (17). Interestingly, *RET* mutations in sporadic MTCs may not necessarily lead to tumorigenesis but are essential for disease progression. This behavior is supported by the fact that the incidence of *RET* mutations in sporadic small MTC (smaller than 1 cm: microMTC) is lower than in larger tumors, and that *RET* mutations often exhibit mutational heterogeneity even in the advanced disease when present (38,52). Romei *et al.* hypothesized that either other oncogenes are responsible for most microMTCs, thereby identifying a tumor subpopulation or that *RET* mutation might or might not occur later during tumor progression (52).

Other somatic changes of *RET*, including copy number alteration and retrocopy, codon deletions,

and rearrangements, were likely to be involved in the pathogenesis and progression of MTCs (17,47,53-55). Bim *et al.* identified copy number alteration and retrocopy in the *RET* gene; determined a recurrent novel point mutation (G548V) exclusively in the somatic retrocopy of *RET* in both sporadic and hereditary MTCs and MTC cell lines, and identified retrocopies produced in somatic cells might play a role in the pathogenesis and progression of MTCs (53). A rare *RET* fusion, *MYH13* exon 35 with *RET* exon 12, was reported in a patient who died of aggressive sporadic MTC with the survival of fewer than ten months after diagnosis, which might be a novel driver genetic change, and a *RET* fusion also provides a possible target for the treatment of RET TKI (54).

RAS mutations

The RAS (P21) protein is located on the inside of the cell membrane and plays a vital role in the transduction of cell growth and differentiation signals (56). It belongs to the guanosine triphosphate (GTP) binding protein (a coupling factor of cell information transmission), which regulates information transduction through the transformation between GTP and guanosine diphosphate (GDP) (56,57). There are three members of the *RAS* gene family: *HRAS*, *KRAS*, and *NRAS* (58). The various *RAS* genes are found

on different chromosomes but have similar structures, all composed of four exons. *RAS* takes part in both MAPK and PI3K signal pathways. *RAS* point mutations in MTC occur in *HRAS* and *KRAS* within hot spots in exons 2, 3, and 4, and rarely involve *NRAS*. They are usually mutually exclusive with *RET* mutations and are present in approximately 0–81% of *RET* wild-type sporadic MTCs (15,17,20–25), suggesting that *RET* and *RAS* somatic mutations are likely mutually exclusive in MTC tumorigenesis and development. Patients harboring *RAS* mutations (*RET* negative) showed a better prognosis than those harboring *RET* mutations or presenting no mutations (17,29).

However, the outcome of patients with a somatic *RET* mutation was significantly worse than both *RAS* positive/*RET* negative and *RAS* negative/*RET* negative cases (13,17). *RAS* mutations were reported to be significantly associated with higher intensity of p-S6 expression, suggesting that the mTOR pathway was activated in such MTCs (25). These findings suggest *RAS* mutations could play a role in tumor development, which leads to a less aggressive tumor phenotype, whereas *RET* mutations could be responsible for a more aggressive phenotype associated with a worse prognosis (22,27).

Other genetic alterations

Other somatic mutations were found in a subset of patients with MTC, especially patients without *RET* and *RAS* mutations. Ciampi *et al.* identified somatic mutations in the genes of *MET*, *TP53*, *TSH* receptor, *EIF1AX*, *CHK2*, *PPM1D* (29). In a study of MTC in Taiwan's population, they identified ten novel MTC somatic mutations: *BICD2*, *DLG1*, *FSD2*, *IL17RD*, *KLHL25*, *PAPPA2*, *PRDM2*, *PSEN1*, *SCRN1*, and *TTC1*, using whole-exome sequencing (33). They further analyzed 1,152 MTCs from COSMIC data and found that most of the variants were involved in pathways of PI3k-Akt, ErbB, MAPK, mTOR, and VEGF signaling pathways, and some were included in the pathogenesis of thyroid cancer, central carbon metabolism, and microRNAs in cancer (33). *FAT1* and *FAT4*, two members of the *FAT* gene family, both located at chromosome 4q, were identified as the two most commonly mutated genes in addition to *RET* in a small cohort of 18 cases of sporadic MTCs from China by Qu *et al.* They further showed that *FAT1* and *FAT4* knockdowns could promote MTC cell proliferation, using TT and MZ-CRC-1 cell lines (15). It is a novel finding that, on the gene expression profile, MTCs could be clustered into

two molecular subtypes: the mesenchymal-like subtype, characterized by the epithelial-mesenchymal transition, and the proliferative-like subtype, associated with enrichment of cell cycle pathways. Most events of structural recurrence occur in the latter (15).

Copy-number losses in chromosome 4q and 1p cause frequent somatic changes for patients with sporadic MTC (15). A study in MD Anderson Cancer Center reported that 19% of patients with sporadic MTC had an aberrant loss of the cyclin-dependent-kinase inhibitors 2C (*CDKN2C*), found in the 1p32 chromosomal region. The aberrant *CDKN2C* loss was associated with distant metastasis at presentation and an unfavorable overall survival (OS). These findings were explained by showing that aberrant loss of *CDKNs* could lead to unrestrained phosphorylation of retinoblastoma protein and unregulated progression through the S phase of the cell cycle, thus resulting in the development of cancers. They also demonstrated the clinical impact of aberrant *CDKN2C* loss was enhanced by concomitant *RET* M918 mutation (59). Anaplastic lymphoma kinase (*ALK*) translocation, a frequent genetic alteration in anaplastic large cell lymphoma and lung adenocarcinoma, was identified in two cases by Ji *et al.* from screening 98 cases of MTC, with the partner genes of *EML4* and *GFPT1*, respectively (60). This finding may show that a rare subset of patients with MTC with *ALK* fusion might receive help from *ALK* target therapy. No *NTRK* translocation has been reported in MTC, although *NTRK* mutation has been reported in MTC (61).

Tumor microenvironment and immunotherapy

In the past decade, people have achieved gratifying results through using immunotherapy in treating many solid cancers, which was an essential milestone in the history of medical development (62). The successful findings in tumor immunotherapy are due to the discovery of immune checkpoint molecules, which are especially important in the regulation of tumor microenvironments. The tumor microenvironment is the internal environment in which tumor cells are produced and live. It consists of not only tumor cells but also fibroblasts, immune and inflammatory cells, and other cells in close contact with tumor cells, as well as adjacent cell stroma, microvessels, and infiltrated biological molecules (42,63–65). Malignant cells actively take part in the reconstruction of a pre-existing matrix, creating a new microenvironment that can be characterized

by inflammation or desmoplastic characteristics. Tumor microenvironment and cancer cells interact and influence each other. They are thereby promoting tumor progression and metastasis (42,63-65). It has been proposed to classify tumor microenvironments into four groups on their status of PD-L1 and tumor-infiltrating lymphocytes to streamline immunotherapy (31).

MTCs often manifest as few tumor-infiltrating immune cells with low expression of PD-L1 both in the tumor and in the stroma cells (34,35), and a low mutational rate (24). However, there is also evidence showing a positive immune reactivity of MTC. In a large cohort of 201 consecutive Chinese patients with MTC, Shi *et al.* demonstrated PD-L1 positivity was associated with aggressive clinicopathologic features (a larger tumor size, lymph node metastases, and advanced TNM staging) and is independently predictive of structural recurrence, and biochemical recurrence/persistent disease (36). Furthermore, they revealed a higher rate of PD-L1 expression in patients with incurable recurrence (36). Bi *et al.* have reported similar findings (35). They detected the expression of PD-L1 and PD-1 in 87 cases of MTC and found PD-L1 positivity was significantly correlated with distant metastases at the surgery. That co-expression of PD-1 and PD-L1 in MTC was correlated with advanced pathologic TNM stage III/IV and distant metastases at the surgery. The positive correlation between PD-1/PD-L1 and prognosis suggests that immunotherapy targeting PD-L1/PD-1 might be effective in treating advanced MTC. Several ongoing trials of immunotherapy in treating advanced MTC are under study, but no definite result has been reported. Although the FDA has approved several anti-angiogenic multikinase inhibitors (aaMKIs) in treating advanced MTC (discussed below), novel methods are still required to treat patients who fail to respond to aaMKI. Immunotherapy might be a choice in the future (66).

Targeting therapy in MTC

By far, surgery is still to be the only therapeutic method in treating most MTCs. Although radioactive iodine and TSH-suppressive therapies can treat differentiated thyroid carcinoma, they have no role in the management of MTC since the neuroendocrine-derived tumor cells do not concentrate iodine or respond to TSH in the same manner that follicular cells do. With the development of targeted therapy in treating advanced or unresectable MTC two aaMKIs, vandetanib, and the American FDA approved cabozantinib for the treatment of advanced MTC in 2011

and 2012, respectively. In the phase III trials of both drugs, patients with metastatic MTC had a significantly longer median progression-free survival (PFS) in the treatment group than in the placebo group (67-69). Both of the drugs did not show their efficacy in improving the OS of patients (67-70). Nevertheless, both studies met their pre-specified primary endpoints for improving PFS and were thus approved by the FDA as the first effective drugs for the treatment of advanced MTC.

Further, evidence showed that cabozantinib provided the most significant clinical benefit to patients with *RET* M918T or *RAS* mutation (71). The most common adverse drug reactions for vandetanib and cabozantinib were diarrhea, rashes, nausea, hypertension, and headaches (67,69,72,73). It is worth noting QT prolongation represented a rare but critical adverse event in the therapy of vandetanib, which potentially evolved into torsade de pointes and sudden death (67,73). Hence, the drug can only be prescribed by qualified physicians and should be used under close surveillance. Despite the advances in the management of metastatic MTC, aaMKIs show significant off-target toxicity and limited efficacy (67,69). LOXO-292 and BLU-667, a new generation of small-molecule TKIs, are highly selective RET inhibitors, and both of them showed better therapeutic efficacy than approved MKIs in the preliminary studies (74-77). At present, clinical trials of both are currently under evaluation (76).

The role of calcitonin/procalcitonin/CALCA gene transcripts in diagnosing and handling MTC

Calcitonin is a hormone that regulates the level of serum calcium, secreted by the C cells of the thyroid and metabolized by the kidneys (78). Calcitonin is regarded as a sensitive marker for diagnosing MTC, and is used for postoperative monitoring of MTC patients (3,79-81). However, the role of routine calcitonin screening in patients with thyroid nodules is still questionable, since the incidence of MTC is low and other non-neoplastic and neoplastic conditions may cause the increase of serum calcitonin (79,82,83). Calcitonin measurement in aspiration needle washout of thyroid nodules and neck lesions is reported to have a higher sensitivity than traditional cytology in diagnosing MTC. It may represent an ancillary tool utilized in patients with high serum calcitonin to avoid false-negative MTC by cytology (83,84). A large-scale retrospective analysis shows that postoperative biochemical remission of serum calcitonin is significantly correlated with

a decrease of the 5-year recurrence rate, but not with the improved 5-year survival rate (81).

Procalcitonin is the precursor of calcitonin. It is widely used as an indicator of severe infection, sepsis, and multiple organ failure (85-87). In recent years, a growing body of research has shown that procalcitonin presents an equivalent or even superior alternative of calcitonin in the management of MTC (88-92). Camacho *et al.* show that the expression of CALCA mRNA from the peripheral blood presents higher sensitivity, specificity, together with higher positive predictive value, and negative predictive value than did serum calcitonin (93). Thus, the detection of CALCA mRNA expression could serve as an alternative to the calcitonin-stimulation test since the former is more convenient to perform (93).

Conclusions

Advanced MTC is still one of the most challenging malignancies for clinicians. With the increasing development of molecular detection techniques, many genetic alterations have been discovered, while *RET* and *RAS* point mutations are still the most frequent and most significant changes. There is growing evidence that genetic changes might be used to predict the clinical outcome of patients with sporadic MTC, on different *RET* point mutations and *RAS* mutations. The in-depth understanding of molecular mechanisms and immune microenvironments of sporadic MTC has prompted the development of targeted therapy and immunotherapy. It is promising that the combination of TKI-targeted therapy and immune checkpoint inhibitors might be a novel therapeutic approach for treating patients with advanced MTC according to the individual tumor mutation profile and tumor microenvironment.

Acknowledgments

Funding: This article was supported by the Beijing Municipal Administration of Hospitals Incubating Program, Code: PX2018042.

Footnote

Provenance and Peer Review: This article was commissioned by the Guest Editor (Kennichi Kakudo) for the series “Asian and Western Practice in Thyroid Pathology: Similarities and Differences” published in *Gland Surgery*. The article

was sent for external peer review organized by the Guest Editor and the editorial office.

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <http://dx.doi.org/10.21037/gs-2019-catp-21>). The series “Asian and Western Practice in Thyroid Pathology: Similarities and Differences” was commissioned by the editorial office without any funding or sponsorship. KK served as the unpaid Guest Editor of the series and serves as an unpaid editorial board member of *Gland Surgery* from Sep 2018 to Aug 2020. YB reports grants from Beijing Municipal Administration of Hospitals Incubating Program, during the conduct of the study. The authors have no other conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work, including ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Open Access Statement: This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

References

1. Cakir M, Grossman AB. Medullary thyroid cancer: molecular biology and novel molecular therapies. *Neuroendocrinology* 2009;90:323-48.
2. Giunti S, Antonelli A, Amorosi A, et al. Cellular signaling pathway alterations and potential targeted therapies for medullary thyroid carcinoma. *Int J Endocrinol* 2013;2013:803171.
3. Grozinsky-Glasberg S, Benbassat CA, Tsvetov G, et al. Medullary thyroid cancer: a retrospective analysis of a cohort treated at a single tertiary care center between 1970 and 2005. *Thyroid* 2007;17:549-56.
4. Hofstra RM, Landsvater RM, Ceccherini I, et al. A mutation in the RET proto-oncogene associated with multiple endocrine neoplasia type 2B and sporadic medullary thyroid carcinoma. *Nature* 1994;367:375-6.

5. Eng C. RET proto-oncogene in the development of human cancer. *J Clin Oncol* 1999;17:380-93.
6. Zhou Y, Zhao Y, Cui B, et al. RET proto-oncogene mutations are restricted to codons 634 and 918 in mainland Chinese families with MEN2A and MEN2B. *Clin Endocrinol (Oxf)* 2007;67:570-6.
7. Dvorakova S, Vaclavikova E, Sykorova V, et al. Somatic mutations in the RET proto-oncogene in sporadic medullary thyroid carcinomas. *Mol Cell Endocrinol* 2008;284:21-7.
8. Marsh DJ, Learoyd DL, Andrew SD, et al. Somatic mutations in the RET proto-oncogene in sporadic medullary thyroid carcinoma. *Clin Endocrinol (Oxf)* 1996;44:249-57.
9. Kebebew E, Ituarte PH, Siperstein AE, et al. Medullary thyroid carcinoma: clinical characteristics, treatment, prognostic factors, and a comparison of staging systems. *Cancer* 2000;88:1139-48.
10. Romei C, Cosci B, Renzini G, et al. RET genetic screening of sporadic medullary thyroid cancer (MTC) allows the preclinical diagnosis of unsuspected gene carriers and the identification of a relevant percentage of hidden familial MTC (FMTC). *Clin Endocrinol (Oxf)* 2011;74:241-7.
11. Mian C, Pennelli G, Barollo S, et al. Combined RET and Ki-67 assessment in sporadic medullary thyroid carcinoma: a useful tool for patient risk stratification. *Eur J Endocrinol* 2011;164:971-6.
12. Nikiforova MN, Wald AI, Roy S, et al. Targeted next-generation sequencing panel (ThyroSeq) for detection of mutations in thyroid cancer. *J Clin Endocrinol Metab* 2013;98:E1852-60.
13. Simbolo M, Mian C, Barollo S, et al. High-throughput mutation profiling improves diagnostic stratification of sporadic medullary thyroid carcinomas. *Virchows Arch* 2014;465:73-8.
14. Sharma BP, Saranath D. RET gene mutations and polymorphisms in medullary thyroid carcinomas in Indian patients. *J Biosci* 2011;36:603-11.
15. Qu N, Shi X, Zhao JJ, et al. Genomic and Transcriptomic Characterization of Sporadic Medullary Thyroid Carcinoma. *Thyroid* 2020;30:1025-36.
16. Wei S, LiVolsi VA, Montone KT, et al. Detection of Molecular Alterations in Medullary Thyroid Carcinoma Using Next-Generation Sequencing: an Institutional Experience. *Endocr Pathol* 2016;27:359-62.
17. Ciampi R, Mian C, Fugazzola L, et al. Evidence of a low prevalence of RAS mutations in a large medullary thyroid cancer series. *Thyroid* 2013;23:50-7.
18. Romei C, Casella F, Tacito A, et al. New insights in the molecular signature of advanced medullary thyroid cancer: evidence of a bad outcome of cases with double RET mutations. *J Med Genet* 2016;53:729-34.
19. Hedayati M, Zarif Yeganeh M, Sheikholeslami S, et al. Diversity of mutations in the RET proto-oncogene and its oncogenic mechanism in medullary thyroid cancer. *Crit Rev Clin Lab Sci* 2016;53:217-27.
20. Oczko-Wojciechowska M, Pfeifer A, Rusinek D, et al. The prevalence of somatic RAS mutations in medullary thyroid cancer - a Polish population study. *Endokrynol Pol* 2015;66:121-5.
21. Boichard A, Croux L, Al Ghuzlan A, et al. Somatic RAS mutations occur in a large proportion of sporadic RET-negative medullary thyroid carcinomas and extend to a previously unidentified exon. *J Clin Endocrinol Metab* 2012;97:E2031-5.
22. Moura MM, Cavaco BM, Pinto AE, et al. High prevalence of RAS mutations in RET-negative sporadic medullary thyroid carcinomas. *J Clin Endocrinol Metab* 2011;96:E863-8.
23. Accardo G, Conzo G, Esposito D, et al. Genetics of medullary thyroid cancer: An overview. *Int J Surg* 2017;41:S2-6.
24. Agrawal N, Jiao Y, Sausen M, et al. Exomic sequencing of medullary thyroid cancer reveals dominant and mutually exclusive oncogenic mutations in RET and RAS. *J Clin Endocrinol Metab* 2013;98:E364-9.
25. Lyra J, Vinagre J, Batista R, et al. mTOR activation in medullary thyroid carcinoma with RAS mutation. *Eur J Endocrinol* 2014;171:633-40.
26. Fussey JM, Vaidya B, Kim D, et al. The role of molecular genetics in the clinical management of sporadic medullary thyroid carcinoma: A systematic review. *Clin Endocrinol (Oxf)* 2019;91:697-707.
27. Elisei R, Cosci B, Romei C, et al. Prognostic significance of somatic RET oncogene mutations in sporadic medullary thyroid cancer: a 10-year follow-up study. *J Clin Endocrinol Metab* 2008;93:682-7.
28. Vuong HG, Odate T, Ngo HTT, et al. Clinical significance of RET and RAS mutations in sporadic medullary thyroid carcinoma: a meta-analysis. *Endocr Relat Cancer* 2018;25:633-41.
29. Ciampi R, Romei C, Ramone T, et al. Genetic Landscape of Somatic Mutations in a Large Cohort of Sporadic Medullary Thyroid Carcinomas Studied by Next-Generation Targeted Sequencing. *iScience* 2019;20:324-36.

30. Sznol M, Chen L. Antagonist antibodies to PD-1 and B7-H1 (PD-L1) in the treatment of advanced human cancer. *Clin Cancer Res* 2013;19:1021-34.
31. Teng MW, Ngiow SF, Ribas A, et al. Classifying Cancers Based on T-cell Infiltration and PD-L1. *Cancer Res* 2015;75:2139-45.
32. Tang H, Qiao J, Fu YX. Immunotherapy and tumor microenvironment. *Cancer Lett* 2016;370:85-90.
33. Chang YS, Chang CC, Huang HY, et al. Detection of Molecular Alterations in Taiwanese Patients with Medullary Thyroid Cancer Using Whole-Exome Sequencing. *Endocr Pathol* 2018;29:324-31.
34. Bongiovanni M, Rebecchini C, Saglietti C, et al. Very low expression of PD-L1 in medullary thyroid carcinoma. *Endocr Relat Cancer* 2017;24:L35-8.
35. Bi Y, Ren X, Bai X, et al. PD-1/PD-L1 expressions in medullary thyroid carcinoma: Clinicopathologic and prognostic analysis of Chinese population. *Eur J Surg Oncol* 2019;45:353-8.
36. Shi X, Yu PC, Lei BW, et al. Association Between Programmed Death-Ligand 1 Expression and Clinicopathological Characteristics, Structural Recurrence, and Biochemical Recurrence/Persistent Disease in Medullary Thyroid Carcinoma. *Thyroid* 2019;29:1269-78.
37. Takahashi M, Ritz J, Cooper GM. Activation of a novel human transforming gene, *ret*, by DNA rearrangement. *Cell* 1985;42:581-8.
38. Eng C, Mulligan LM, Healey CS, et al. Heterogeneous mutation of the RET proto-oncogene in subpopulations of medullary thyroid carcinoma. *Cancer Res* 1996;56:2167-70.
39. Pasini B, Hofstra RM, Yin L, et al. The physical map of the human RET proto-oncogene. *Oncogene* 1995;11:1737-43.
40. de Groot JW, Links TP, Plukker JT, et al. RET as a diagnostic and therapeutic target in sporadic and hereditary endocrine tumors. *Endocr Rev* 2006;27:535-60.
41. Airaksinen MS, Saarma M. The GDNF family: signalling, biological functions and therapeutic value. *Nat Rev Neurosci* 2002;3:383-94.
42. Castellone MD, Melillo RM. RET-mediated modulation of tumor microenvironment and immune response in multiple endocrine neoplasia type 2 (MEN2). *Endocr Relat Cancer* 2018;25: T105-19.
43. Ponder BA. The phenotypes associated with *ret* mutations in the multiple endocrine neoplasia type 2 syndrome. *Cancer Res* 1999;59:1736s-41s; discussion 1742s.
44. Ameer N, Lacroix L, Roucan S, Roux V, et al. Aggressive inherited and sporadic medullary thyroid carcinomas display similar oncogenic pathways. *Endocr Relat Cancer* 2009;16:1261-72.
45. Lai AZ, Gujral TS, Mulligan LM. RET signaling in endocrine tumors: delving deeper into molecular mechanisms. *Endocr Pathol* 2007;18:57-67.
46. Boikos SA, Stratakis CA. Molecular mechanisms of medullary thyroid carcinoma: current approaches in diagnosis and treatment. *Histol Histopathol* 2008;23:109-16.
47. Alemi M, Lucas SD, Sällström JF, et al. A complex nine base pair deletion in RET exon 11 common in sporadic medullary thyroid carcinoma. *Oncogene* 1997;14:2041-5.
48. Shirahama S, Ogura K, Takami H, et al. Mutational analysis of the RET proto-oncogene in 71 Japanese patients with medullary thyroid carcinoma. *J Hum Genet* 1998;43:101-6.
49. Kalinin VN, Amosenko FA, Shabanov MA, et al. Three novel mutations in the RET proto-oncogene. *J Mol Med* 2001;79:609-12.
50. Jindrichova S, Kodet R, Krskova L, et al. The newly detected mutations in the RET proto-oncogene in exon 16 as a cause of sporadic medullary thyroid carcinoma. *J Mol Med* 2003;81:819-23.
51. Romei C, Elisei R, Pinchera A, et al. Somatic mutations of the *ret* protooncogene in sporadic medullary thyroid carcinoma are not restricted to exon 16 and are associated with tumor recurrence. *J Clin Endocrinol Metab* 1996;81:1619-22.
52. Romei C, Ugolini C, Cosci B, et al. Low prevalence of the somatic M918T RET mutation in micro-medullary thyroid cancer. *Thyroid* 2012;22:476-81.
53. Bim LV, Navarro FCP, Valente FOF, et al. Retroposed copies of RET gene: a somatically acquired event in medullary thyroid carcinoma. *BMC Med Genomics* 2019;12:104.
54. Grubbs EG, Ng PK, Bui J, et al. RET fusion as a novel driver of medullary thyroid carcinoma. *J Clin Endocrinol Metab* 2015;100:788-93.
55. Ciampi R, Romei C, Cosci B, et al. Chromosome 10 and RET gene copy number alterations in hereditary and sporadic Medullary Thyroid Carcinoma. *Mol Cell Endocrinol* 2012;348:176-82.
56. Adjei AA. Blocking oncogenic Ras signaling for cancer therapy. *J Natl Cancer Inst* 2001;93:1062-74.
57. Boguski MS, McCormick F. Proteins regulating Ras and its relatives. *Nature* 1993;366:643-54.
58. Barbacid M. *ras* genes. *Annu Rev Biochem* 1987;56:779-827.

59. Grubbs EG, Williams MD, Scheet P, et al. Role of CDKN2C Copy Number in Sporadic Medullary Thyroid Carcinoma. *Thyroid* 2016;26:1553-62.
60. Ji JH, Oh YL, Hong M, et al. Identification of Driving ALK Fusion Genes and Genomic Landscape of Medullary Thyroid Cancer. *PLoS Genet* 2015;11:e1005467.
61. Gimm O, Greco A, Hoang-Vu C, et al. Mutation analysis reveals novel sequence variants in NTRK1 in sporadic human medullary thyroid carcinoma. *J Clin Endocrinol Metab* 1999;84:2784-7.
62. Boutros C, Tarhini A, Routier E, et al. Safety profiles of anti-CTLA-4 and anti-PD-1 antibodies alone and in combination. *Nat Rev Clin Oncol* 2016;13:473-86.
63. Egeblad M, Nakasone ES, Werb Z. Tumors as organs: complex tissues that interface with the entire organism. *Dev Cell* 2010;18:884-901.
64. Hanahan D, Coussens LM. Accessories to the crime: functions of cells recruited to the tumor microenvironment. *Cancer cell* 2012;21:309-22.
65. McAllister SS, Weinberg RA. The tumour-induced systemic environment as a critical regulator of cancer progression and metastasis. *Nat Cell Biol* 2014;16:717-27.
66. Hadoux J, Pacini F, Tuttle RM, et al. Management of advanced medullary thyroid cancer. *Lancet Diabetes Endocrinol* 2016;4:64-71.
67. Wells SA, Robinson BG, Gagel RF, et al. Vandetanib in patients with locally advanced or metastatic medullary thyroid cancer: a randomized, double-blind phase III trial. *J Clin Oncol* 2012;30:134-41.
68. Elisei R, Schlumberger MJ, Müller SP, et al. Cabozantinib in progressive medullary thyroid cancer. *J Clin Oncol* 2013;31:3639-46.
69. Krajewska J, Olczyk T, Jarzab B. Cabozantinib for the treatment of progressive metastatic medullary thyroid cancer. *Expert Rev Clin Pharmacol* 2016;9:69-79.
70. Tappenden P, Carroll C, Hamilton J, et al. Cabozantinib and vandetanib for unresectable locally advanced or metastatic medullary thyroid cancer: a systematic review and economic model. *Health Technol Assess* 2019;23:1-144.
71. Sherman SI, Clary DO, Elisei R, et al. Correlative analyses of RET and RAS mutations in a phase 3 trial of cabozantinib in patients with progressive, metastatic medullary thyroid cancer. *Cancer* 2016;122:3856-64.
72. Milling RV, Grimm D, Krüger M, et al. Pazopanib, Cabozantinib, and Vandetanib in the Treatment of Progressive Medullary Thyroid Cancer with a Special Focus on the Adverse Effects on Hypertension. *Int J Mol Sci* 2018;19:3258.
73. Hu MI, Elisei R, Dedecjus M, et al. Safety and efficacy of two starting doses of vandetanib in advanced medullary thyroid cancer. *Endocr Relat Cancer* 2019;26:241-50.
74. Subbiah V, Gainor JF, Rahal R, et al. RETPrecision Targeted Therapy with BLU-667 for -Driven Cancers. *Cancer Discov* 2018;8:836-49.
75. Subbiah V, Velcheti V, Tuch BB, et al. Selective RET kinase inhibition for patients with RET-altered cancers. *Ann Oncol* 2018;29:1869-76.
76. Ceolin L, Duval MADS, Benini AF, et al. Medullary thyroid carcinoma beyond surgery: advances, challenges, and perspectives. *Endocr Relat Cancer* 2019;26:R499-518.
77. Zhao Z, Fu T, Gao J, et al. Identifying novel oncogenic RET mutations and characterising their sensitivity to RET-specific inhibitors. *J Med Genet* 2020. [Epub ahead of print].
78. Felsenfeld AJ, Levine BS. Calcitonin, the forgotten hormone: does it deserve to be forgotten? *Clin Kidney J* 2015;8:180-7.
79. Verbeek HH, de Groot JWB, Sluiter WJ, et al. Calcitonin testing for detection of medullary thyroid cancer in people with thyroid nodules. *Cochrane Database Syst Rev* 2020;3:CD010159.
80. Danila R, Livadariu R, Branisteanu D. Calcitonin revisited in 2020. *Acta Endocrinol (Buchar)* 2019;15:544-8.
81. Jung KY, Kim SM, Yoo WS, et al. Postoperative biochemical remission of serum calcitonin is the best predictive factor for recurrence-free survival of medullary thyroid cancer: a large-scale retrospective analysis over 30 years. *Clin Endocrinol (Oxf)* 2016;84:587-97.
82. Silvestre C, Sampaio Matias J, Proença H, et al. Calcitonin Screening in Nodular Thyroid Disease: Is There a Definitive Answer? *Eur Thyroid J* 2019;8:79-82.
83. Costante G, Meringolo D. Calcitonin as a biomarker of C cell disease: recent achievements and current challenges. *Endocrine* 2020;67:273-80.
84. Trimboli P, Cremonini N, Ceriani L, et al. Calcitonin measurement in aspiration needle washout fluids has higher sensitivity than cytology in detecting medullary thyroid cancer: a retrospective multicentre study. *Clin Endocrinol (Oxf)* 2014;80:135-40.
85. Iankova I, Thompson-Leduc P, Kirson NY, et al. Efficacy and Safety of Procalcitonin Guidance in Patients With Suspected or Confirmed Sepsis: A Systematic Review and Meta-Analysis. *Crit Care Med* 2018;46:691-8.
86. Dou YH, Du JK, Liu HL, et al. The role of procalcitonin in the identification of invasive fungal infection-a systemic

- review and meta-analysis. *Diagn Microbiol Infect Dis* 2013;76:464-9.
87. Honore PM, David C, Attou R, et al. Procalcitonin to allow early detection of sepsis and multiple organ failure in severe multiple trauma: beware of some confounders. *Crit Care* 2020;24:9.
 88. Algeciras-Schimmich A, Preissner CM, et al. Procalcitonin: a marker for the diagnosis and follow-up of patients with medullary thyroid carcinoma. *J Clin Endocrinol Metab* 2009;94:861-8.
 89. Woliński K, Kaznowski J, Klimowicz A, et al. Diagnostic value of selected biochemical markers in the detection of recurrence of medullary thyroid cancer - comparison of calcitonin, procalcitonin, chromogranin A, and carcinoembryonic antigen. *Endokrynol Pol* 2017;68:434-7.
 90. Giovanella L, Imperiali M, Piccardo A, et al. Procalcitonin measurement to screen medullary thyroid carcinoma: A prospective evaluation in a series of 2705 patients with thyroid nodules. *Eur J Clin Invest* 2018;48:e12934.
 91. Walter MA, Meier C, Radimerski T, et al. Procalcitonin levels predict clinical course and progression-free survival in patients with medullary thyroid cancer. *Cancer* 2010;116:31-40.
 92. Machens A, Lorenz K, Dralle H. Utility of serum procalcitonin for screening and risk stratification of medullary thyroid cancer. *J Clin Endocrinol Metab* 2014;99:2986-94.
 93. Camacho CP, Lindsey SC, Melo MC, et al. Measurement of calcitonin and calcitonin gene-related peptide mRNA refines the management of patients with medullary thyroid cancer and may replace calcitonin-stimulation tests. *Thyroid* 2013;23:308-16.

Cite this article as: Bai Y, Niu D, Yao Q, Lin D, Kakudo K. Updates in the advances of sporadic medullary thyroid carcinoma: from the molecules to the clinic. *Gland Surg* 2020;9(5):1847-1856. doi: 10.21037/gs-2019-catp-21