



Development and validation of the diagnostic accuracy of artificial intelligence-assisted ultrasound in the classification of splenic trauma

Xue Jiang¹, Yukun Luo², Xuelei He³, Kun Wang⁴, Wenjing Song¹, Qinggui Ye¹, Lei Feng¹, Wei Wang¹, Xiaojuan Hu¹, Hua Li⁵

¹Department of Ultrasound Diagnosis, The Fourth Medical Center of PLA General Hospital, Beijing, China; ²Department of Ultrasound Diagnosis, The First Medical Center of PLA General Hospital, Beijing, China; ³School of Information Sciences and Technology, Northwest University, Xi'an, China; ⁴CAS Key Laboratory of Molecular Imaging, the State Key Laboratory of Management and Control for Complex Systems, Institute of Automation, Chinese Academy of Sciences, Beijing, China; ⁵Beijing Mindray Medical Instrument Co., Ltd., Beijing, China

Contributions: (I) Conception and design: X Jiang, Y Luo, K Wang; (II) Administrative support: Y Luo; (III) Provision of study materials or patients: L Feng, W Wang, H Li; (IV) Collection and assembly of data: W Song, Q Ye, X Hu; (V) Data analysis and interpretation: X Jiang, X He; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

Correspondence to: Yukun Luo, MD. Department of Ultrasound Diagnosis, The First Medical Center of PLA General Hospital, Beijing, China. Email: lyk301@163.com.

Background: Spleen is the most vulnerable organ in abdominal trauma. Ultrasound (US) has become an important examination method for splenic trauma. However, the sensitivity of routine US in the diagnosis of splenic trauma is low. Contrast-enhanced ultrasound (CEUS) can improve the sensitivity, but it also has some limitations. This study sought to explore the application value of artificial intelligence (AI)-assisted US in the classification of splenic trauma.

Methods: The splenic injuries of Bama miniature pigs were established. A large number of ultrasonic images were collected. Then, 3-fold cross validation (CV) was used to establish the animal models. Next, clinical ultrasonic images were collected at multiple centers. All injuries were diagnosed by CEUS, enhanced CT or surgery. We used animal models to fine tune a small amount of human data, and then established the final AI splenic trauma recognition model. The whole model was constructed by averaging the prediction ability of the 3 fine-tuned models. Finally, 2 doctors' recognition US results of splenic trauma were compared to the AI recognition results. The area under the curve (AUC), sensitivity, specificity, negative predictive value, and positive predictive value were used to evaluate the diagnostic performance in diagnosis of spleen trauma.

Results: (I) Based on the receiver operating characteristic (ROC) curves, the test cohort 1 (AUC =0.90) and 2 (AUC =0.84) had a similar performance. Based on the decision curve analysis (DCA) curves, while threshold smaller than 0.8, the proposed model had better performance on test cohort 1 than test cohort 2. Test cohort 1 had higher sensitivity (0.82 vs. 0.71, $P<0.01$) and higher specificity (0.88 vs. 0.81, $P<0.01$) than test cohort 2. (II) The diagnostic accuracy of the AI model was higher than that of doctor 1 (0.82 vs. 0.62, $P<0.001$) and doctor 2 (0.82 vs. 0.66, $P<0.001$), and its specificity was higher than that of doctor (0.88 vs. 0.78, $P=0.001$).

Conclusions: AI-assisted US diagnosis of splenic trauma can significantly improve the ultrasonic diagnosis rate. We still need to increase the number of samples to further improve the diagnostic efficiency of the model.

Keywords: Trauma; splenic injury; deep learning; ultrasound

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Introduction

Due to its fragile texture and abundant blood supply, the spleen is the most vulnerable organ in abdominal trauma. Splenic injury accounts for about 13–25% of abdominal trauma (1). The early symptoms of splenic trauma are not obvious in some patients, but splenic trauma is often accompanied by serious organ problems. If patients are not diagnosed and treated in a timely manner, their lives may be endangered (2-4). Presently, preliminary screening by imaging is the main way to quickly evaluate splenic injury. Imaging examination methods mainly include computed tomography (CT) and ultrasound (US) examinations. CT has a number of advantages, including that it has a high coincidence rate and is not affected by gas interference or patient breathing during the examination. However, CT examinations also have certain limitations. As CT is radioactive, children and pregnant women have an increased risk of exposure to radiation. Additionally, a certain rate of adverse reactions occurs in enhanced CT.

US has become an important examination method for splenic trauma. It can be applied to rapid injury assessments in pre-hospital or emergency rooms and can be used as an evaluation method for post-traumatic treatment. It can also be repeatedly performed to observe the recovery of organs (5,6). US can reveal whether there is organ damage by displaying the continuity of the organ capsule, the presence of subcapsular hematoma, and changes to the echo of the parenchyma. Additionally, US can also indicate whether there is organ damage through some indirect signs, such as retroperitoneal hematoma and peritoneal effusion. Color doppler US can detect abnormal blood supply in the lesions, which improves the detection rate of parenchymal organ injury. However, US examinations are operator dependent and cannot detect some mild traumas. It has been reported in the literature that the sensitivity of conventional US in the diagnosis of parenchymal organ injury is only about 41% (7,8).

Contrast-enhanced ultrasound (CEUS) can detect parenchymal organ injury and active bleeding from multiple abdominal injuries, improve the accuracy of ultrasonic examinations of splenic trauma, provides a more reliable evaluation method for parenchymal organ injury, and makes up for the shortcomings of conventional US (9). Studies have shown that CEUS is as accurate as CT in the detection and staging of traumatic splenic injury (10-12). However, CEUS is an invasive examination, and the contrast agent is expensive; thus, it has some limitations.

As a leading technology for the future, artificial intelligence (AI) is increasingly being applied to all fields.

As a popular analysis method of machine learning, deep learning can represent massive data by constructing multi-layer artificial neural networks. Rapid improvement of graphics processing capacity has made the development of more advanced algorithm possible. Such algorithms have a stable and powerful image analysis ability (13,14). Thus, they have been widely used in the diagnosis and recognition of ultrasonic images of various organs, and have been proven to have high accuracy (15-17).

However, deep learning requires a large amount of image data. To solve this problem, we applied the method of transfer learning. The performance of pig spleen in ultrasound is similar to that of human, and the performance of spleen trauma in conventional ultrasound and contrast-enhanced ultrasound is also similar to that of human spleen trauma (18). Therefore, we learn the low-level ultrasonic damage characteristics by building animal models. On the premise that the low-level ultrasonic damage characteristics are unchanged, we migrate the animal models, retrain with a small number of human spleen ultrasonic images, and learn the high-level human spleen damage characteristics, so as to achieve the effect of building a human spleen damage identification model. Although there was no migration from animal to human model in the past studies, However, there are examples of using natural image modeling to migrate medical images (19), using data modeling of other organs or modalities to migrate to lung segmentation models (20), and using 2D image modeling to migrate to 3D images (21). Therefore, we assume that animal models can be established first by collecting a large number of animal spleen trauma images and fine-tuning through human data, so as to make up for the lack of human data. At present, there is no report on the AI-assisted US diagnosis of splenic trauma. This study combined animal experiments and clinical US image data, and applied a deep-learning method to establish a CEUS splenic trauma classification model. We present the following article in accordance with the TRIPOD reporting checklist (available at <https://atm.amegroups.com/article/view/10.21037/atm-22-3767/rc>).

Methods

Experimental animals

The animal experiments were performed under a project license (No. 2021KY033-KS001) granted by the Medical Ethics Committee of PLA General Hospital, in compliance with National Laboratory Animal Management Regulations and guidelines for the care and use of animals.

In total, 20 Bama miniature pigs, male and female, weighing 15–25 kg, were anesthetized by intramuscular injection with 50 mg/mL of Shutai 50 (tiletamine hydrochloride 125 mg and zolazepam hydrochloride 125 mg) (0.1 mL/kg). After being anesthetized, each animal was fixed on the operating table, the vein access was established in the ear marginal vein, and the abdominal skin was prepared. Heart rate, blood oxygen, and other indicators were monitored. We examined the injuries in the condition of gray scale. Then we collected normal-spleen images in different sections before establishing the trauma model. About 100 images were collected for each organ, and a total of 2,225 spleen images were collected. After the image acquisition, the spleen was impacted by external force to simulate blunt organ injury and establish a splenic trauma model. A total of 49 splenic trauma foci were established.

We used the Mindray M9 portable ultrasonic diagnostic instrument (Mindray Company, China), which has a convex array probe C5-1s, and a probe frequency of 1–5 MHz, and the large ultrasonic diagnostic instrument, Philip EPIQ 7 (Philips, Netherlands), which has a convex array probe C5-1, and a probe frequency of 1–5 MHz. After the establishment of the wound lesions, US contrast agent was administered. The contrast agent used was the SonoVue contrast agent (Bracco, Italy). Normal saline (5 mL) was dissolved in each contrast agent and shaken. The probe was placed in the abdomen, and at the trauma site, the parameters of the ultrasonic instrument were adjusted, and the same parameters were applied to the same trauma focus. The contrast medium (1 mL) was injected into the spleen through the ear vein, followed by normal saline (5 mL). In the arterial phase, the damaged part was observed, the size of the wound was measured, and the dynamic angiography images were collected. The ultrasonic instrument was adjusted to the gray-scale condition, the wound location shown by the contrast agent was explored, its location, boundary, echo, and other ultrasonic manifestations were observed, the size of the wound was measured, and the scope of the wound was determined according to the results of the CEUS. About 100 gray-scale images were collected from different angles and sections of each trauma focus, and 4,280 trauma images were collected from the spleens.

Clinical image collection

The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). The study was approved by ethics committee of PLA General Hospital

(No. S2020-323-01) and informed consent was taken from all the patients. Ultrasonic images of splenic trauma from PLA General Hospital and Beijing Chaoyang Emergency Rescue Center were collected retrospectively (2015.4.10–2021.8.18). All injuries were diagnosed by CEUS, enhanced CT or surgery. A total of 548 images of 149 patients with splenic trauma were collected. Additionally, 652 images of 652 normal spleen patients were collected. After excluding the poor-quality images, 505 spleen-trauma images and 559 normal-spleen images remained. A small number of images (125 images, including 58 spleen-trauma images and 67 normal-spleen images) were randomly selected as the training set. The remaining images were divided into test set 1 and test set 2. The 274 images (133 spleen-trauma images and 141 normal-spleen images) collected at the 1st and 4th Medical Centers of PLA General Hospital were used as test set 1, and the 665 images (314 spleen-trauma images and 351 normal-spleen images) collected by the Beijing Chaoyang Emergency Rescue Center were used as test set 2.

Image annotation

Image annotation was undertaken to establish the classification model, and the normal spleen and spleen-trauma images were sketched with sketching software. For the normal-spleen images, we only needed to draw the outline of the spleen and the edge of the spleen as accurately as possible along the spleen envelope. For the spleen-trauma images, we needed to draw the outline of the spleen and trauma at the same time. The outline of the wound was based on the location and scope of the wound as determined by the results of the CEUS or enhanced CT.

US model building

US static images were collected in JPEG format. Each image was resized to 224×224 pixels. The MobileNet V2 structure was used to construct the US model. The animal CV models were trained by 3-fold cross validation (CV) in the animal training cohorts and animal validation cohorts. Next, some human data were used to fine tune the animal CV models. The whole model was constructed by averaging the prediction ability of the 3 fine-tuned models.

Model construction

The static model was a structure of MobileNet V2

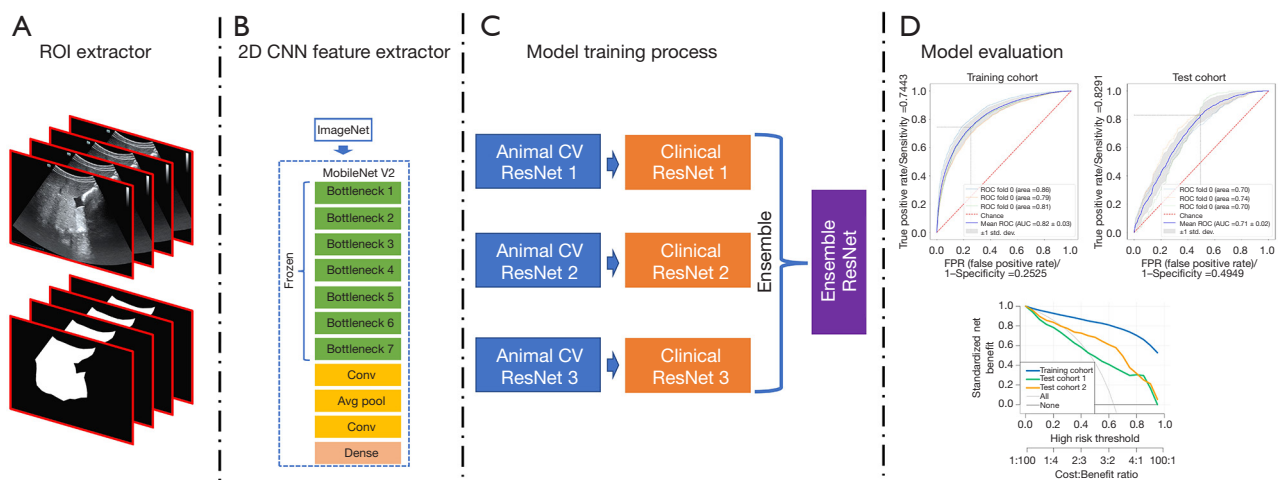


Figure 1 The model construction process employed in this study. 3-fold CV was used to construct the animal CV models for the animal training cohort. A few human images were used to fine tune the animal model and establish the human model. An inter-test cohort and an independent test cohort were used to evaluate the human model. ROI, region of interest; CNN, convolutional neural network; CV, cross validation; ROC, receiver operating characteristic; AUC, area under the curve.

(Appendix 1). MobileNet V2 was used to extract the spatial features (see Figure 1).

MobileNet V2

The structure of MobileNet V2 (22) is listed in Table S1, and had 7 bottleneck stages. All of the bottleneck stages were constructed by a 2-dimensional convolution layer, batch normalization layer, ReLU activation layer, and depth-wise convolution layer (see Figure 1).

Image evaluation

In this study, 2 experienced doctors (with 10 and 12 years of US examination experience respectively) performed blind evaluations of 382 splenic injury images from clinical test set 2 and classified each image as traumatic or non-invasive. We then compared the US doctors' image recognition results to those of the CEUS model to evaluate the performance of the model.

Statistical analysis

We used *t*-tests for the continuous data and χ^2 -tests for the categorical data to compare the baseline characteristics between test cohorts 1 and 2. A receiver operating characteristic (ROC) curve analysis was conducted to examine the prediction performance of the model. In the

CV process, the validation cohorts were used to evaluate the performance of the animal model. Test cohorts 1 and 2 were used to verify the predictive efficacy of the model. Statistical analyses were undertaken using R software (version 3.3.1; R 21 Foundation for Statistical Computing, Vienna, Austria). All the statistical tests were 2-sided, and the statistical significance level was set at $P < 0.05$. The AUC, sensitivity, specificity, negative predictive value, and positive predictive value were used to evaluate the diagnostic performance in diagnosis of splenic trauma.

Results

Results of the animal model

We used 3-fold CVs to establish the animal models. A total of 5,087 images (3,275 spleen-trauma images and 1,812 normal-spleen images) were selected for the 1st training set, and a total of 1,079 images (666 spleen-trauma images and 413 normal-spleen images) were selected for the validation set). Next, 5,549 images (3,532 spleen-trauma images and 2,017 normal-spleen images) were selected for the 2nd training set, and 617 images (409 spleen-trauma images and 208 normal-spleen images) were selected for the validation set. Finally, 5,541 images (3,515 spleen-trauma images and 2,026 normal-spleen images) were selected for the 3rd-fold CV training set, and 625 images (426 spleen-trauma images and 199 normal-spleen images) were selected for

Table 1 Classification performance of animal model

Data	Cohort	AUC	Accuracy	Sensitivity	Specificity	PPV	NPV
Fold 1	Training cohort	0.86 (0.86–0.87)	0.77 (0.77–0.78)	0.76 (0.74–0.76)	0.81 (0.80–0.82)	0.88 (0.87–0.88)	0.65 (0.63–0.66)
	Test cohort	0.70 (0.68–0.71)	0.65 (0.64–0.67)	0.66 (0.64–0.73)	0.63 (0.55–0.66)	0.74 (0.72–0.76)	0.54 (0.52–0.57)
Fold 2	Training cohort	0.79 (0.79–0.80)	0.72 (0.71–0.73)	0.70 (0.69–0.75)	0.75 (0.70–0.76)	0.83 (0.81–0.84)	0.58 (0.58–0.62)
	Test cohort	0.74 (0.72–0.76)	0.72 (0.70–0.74)	0.74 (0.71–0.81)	0.67 (0.59–0.71)	0.81 (0.79–0.84)	0.57 (0.54–0.63)
Fold 3	Training cohort	0.81 (0.81–0.82)	0.73 (0.73–0.76)	0.71 (0.69–0.78)	0.77 (0.71–0.80)	0.84 (0.82–0.86)	0.61 (0.59–0.65)
	Test cohort	0.70 (0.67–0.72)	0.79 (0.78–0.81)	0.93 (0.92–0.97)	0.48 (0.43–0.51)	0.79 (0.77–0.81)	0.77 (0.74–0.87)

Data in parentheses are 95% CIs. AUC, area under the curve; PPV, positive predictive value; NPV, negative predictive value.

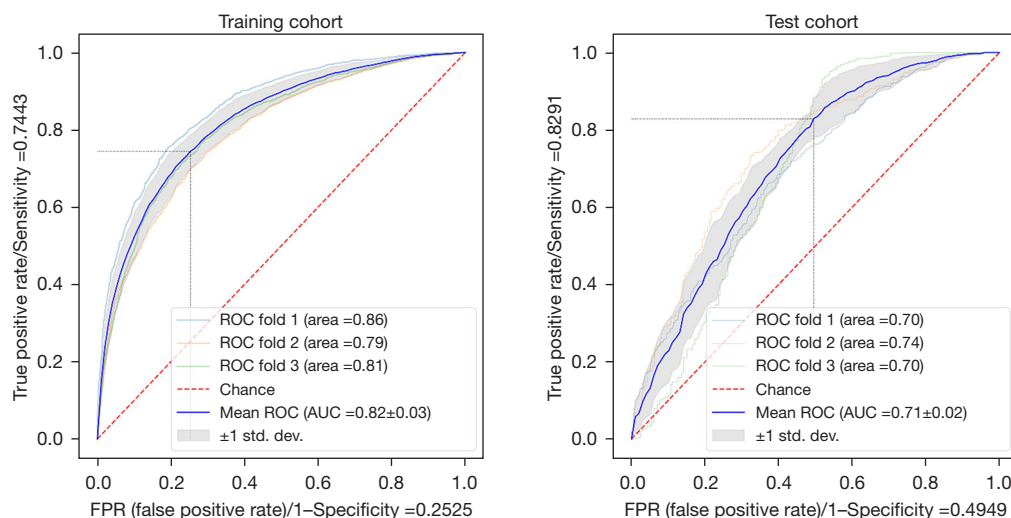


Figure 2 The ROC curves of the animal model for the training and test cohorts. ROC, receiver operating characteristic; AUC, area under the curve.

the validation set. The areas under the curve (AUCs) of the 3rd-fold model were 0.81 for the training set and 0.70 for the test set (see *Table 1*, *Figure 2*).

Clinical model establishment

To evaluate the performance of the proposed model, the ROC and DCA curves were plotted (see *Figure 3*). The ROC curve results showed that the performance of test cohorts 1 (AUC =0.84) and 2 (AUC =0.90) was similar. Based on the DCA curves, while the threshold was <0.8, the proposed model performed better on test cohort 2 than test cohort 1. As *Table 2* shows, the test cohort 2 had higher sensitivity (0.82 *vs.* 0.71, $P<0.01$) and higher specificity (0.88 *vs.* 0.81, $P<0.01$) than test cohort 1.

Doctor's US recognition results

Next, 2 doctors examined all the normal and abnormal images and divided the recognition results into “traumatic” and “non-invasive”. Taking the final modeling results from test cohort 2 as the standard for convolutional neural network (CNN) recognition, *Figure 3* shows the ROC curves of the images recognized by the CNN model and doctors. The accuracy of image recognition by doctor 1 was 0.70 (95% CI: 0.66–0.73), the sensitivity was 0.62 (95% CI: 0.56–0.67), the specificity was 0.78 (95% CI: 0.73–0.83), the positive predictive value was 0.76 (95% CI: 0.71–0.81), and the negative predictive value was 0.64 (95% CI: 0.60–0.69). While the accuracy of image recognition by doctor 2 was 0.74 (95% CI: 0.70–0.77), the sensitivity was 0.66 (95% CI:

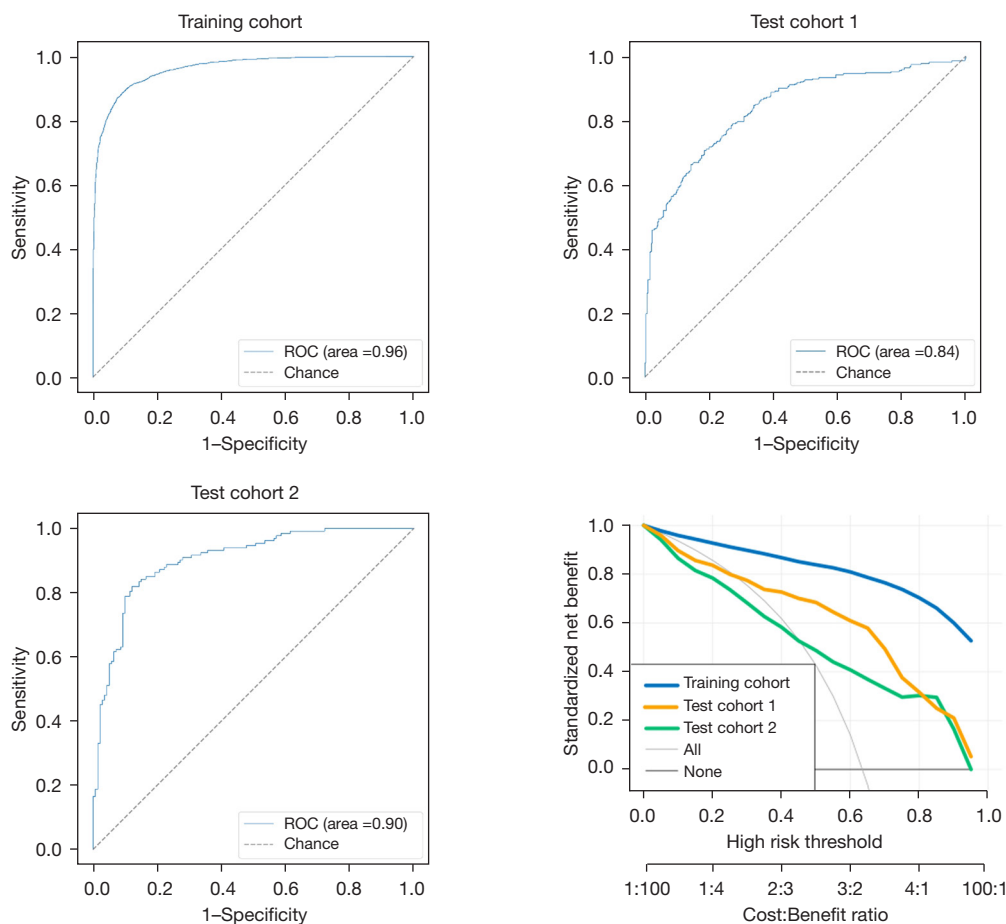


Figure 3 The ROC and DCA curves of the human model for the training cohort, test cohort 1, and test cohort 2. ROC, receiver operating characteristic; DCA, decision curve analysis.

Table 2 Classification performance of human model

Data	AUC	Accuracy	Sensitivity	Specificity	PPV	NPV
Animal cohort	0.96 (0.96–0.96)	0.89 (0.89–0.90)	0.89 (0.87–0.90)	0.91 (0.89–0.93)	0.94 (0.94–0.96)	0.82 (0.80–0.84)
Test cohort 1	0.84 (0.83–0.86)	0.76 (0.75–0.78)	0.71 (0.65–0.87)	0.81 (0.65–0.87)	0.77 (0.69–0.83)	0.76 (0.73–0.85)
Test cohort 2	0.90 (0.89–0.92)	0.85 (0.83–0.87)	0.82 (0.78–0.87)	0.88 (0.83–0.92)	0.87 (0.82–0.90)	0.84 (0.81–0.87)

Data in parentheses are 95% CIs. AUC, area under the curve; PPV, positive predictive value; NPV, negative predictive value.

0.61–0.71), the specificity was 0.82 (95% CI: 0.78–0.86), the positive predictive value is 0.80 (95% CI: 0.75–0.85), and the negative predictive value was 0.68 (95% CI: 0.63–0.73). The diagnostic sensitivity of the deep-learning model was higher than that of doctor 1 (0.82 *vs.* 0.62, $P < 0.001$) and doctor 2 (0.82 *vs.* 0.66, $P < 0.001$), and the specificity was higher than that of doctor 1 (0.88 *vs.* 0.78, $P = 0.001$) and doctor 2 (0.88 *vs.* 0.82, $P = 0.03$). The final modeling results

for the deep-learning test set 2 are shown in *Figure 4* and *Table 3*.

Discussion

As an important imaging method for the injury evaluation of patients with splenic trauma, US plays a very important role in the diagnosis of trauma. In pre-hospital treatment,

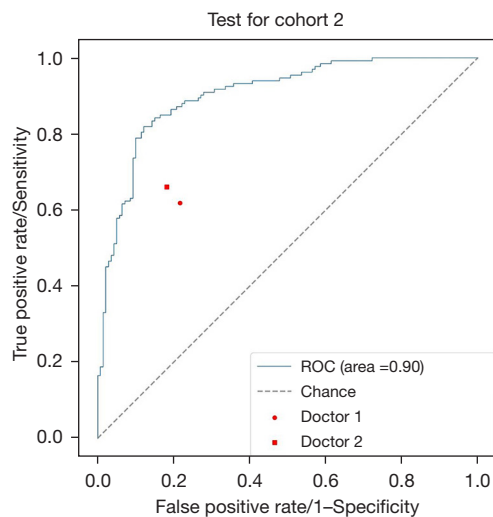


Figure 4 The ROC curves of the human model for test cohort 2, doctor 1, and doctor 2. ROC, receiver operating characteristic.

it can help save lives, as it enables the injuries of patients to be evaluated and medical interventions to be conducted quickly and accurately.

Focused assessment with sonography for trauma (FAST), which is a bedside assessment tool for patients with abdominal trauma, is widely used in clinical practice. This method is mainly used for the assessment of intraperitoneal hemorrhages caused by injuries to the liver, spleen or other organs in patients with trauma. It has high sensitivity to intraperitoneal hemorrhage and can indirectly assess abdominal organ injuries (23). In the diagnosis of abdominal trauma, AI can be used to diagnose organ trauma by FAST (24,25). However, in FAST, it is often difficult to distinguish between peritoneal effusion caused by abdominal organ injury and peritoneal effusion caused by other factors (e.g., tumor or liver cirrhosis), and it is impossible to determine which organ has trauma.

Conventional US diagnosis of organ trauma is not only operator dependent, but also often miss minor traumas. The sensitivity of US diagnosis is relatively low. Thus, as an important examination method of abdominal trauma, it is urgent to improve the sensitivity of ultrasonic diagnosis of visceral trauma. However, there is no relevant report on evaluations of the presence or absence of trauma foci in organs. In this study, AI was used to identify traumatic lesions that are difficult to identify by conventional gray-scale US through feature extraction, internal echo changes of parenchymal organs, and subtle texture changes of images. Its diagnosis accuracy was significantly higher than

that of the US doctors.

The establishment of the AI model required a large number of ultrasonic images, but ultrasonic images of splenic trauma are difficult to collect. Thus, we first established an animal model of splenic trauma, and found that the performance of image features of the pig spleen under US was similar to that of human. In the process of establishing the model, to establish different degrees of trauma, we used different forces to impact the spleen area. Thus, some trauma foci were difficult to identify in gray-scale US images, and the trauma lesions needed to be identified by CEUS. Part of the trauma included splenic rupture. In US images, splenic trauma can be classified as hypoechoic, isoechoic, or hyperechoic echogenicity, heterogeneous echogenicity, or other. To simulate different levels of splenic trauma to the greatest extent possible, we obtained a large number of animal spleen-trauma images and established an AI model of splenic trauma through deep learning. For the human spleen-trauma images, we used a multi-center method to gather images collected by different machines and doctors to increase the number of images. Next, we fine-tuned the animal model. To evaluate the performance of this model, we established a test cohort 1 and an independent test cohort 2. We proposed a transfer-learning model with a MobileNet V2 structure and a pre-trained ImageNet weight to predict the splenic trauma from the US images. Transfer learning can not only transfer the information in the original model established by a large amount of data to less data, but can also shorten the training time and achieve higher performance. The transfer-learning time is shorter than the original learning process time required to build the source model (26,27). The model can learn higher features by pre-trained weight. Our results showed that the model was able to predict splenic trauma from US images even in different center cohorts.

This study had some limitations. First, due to the morphological difference in the spleens of different patients, it was necessary to crop the spleen region for further analysis. This will become more automatized in further research. Second, the model was constructed using animal US images, and more clinical images are needed for further correction. Finally, more data from different center cohorts are needed for further study.

In summary, establishing an AI splenic trauma model through transfer learning could significantly improve the ultrasonic diagnostic sensitivity of splenic trauma. Such a model could help clinicians make clear judgments for patients with splenic trauma, especially pre-hospital

Table 3 Diagnostic performance of the CNN model and radiologists.

Data	Accuracy	Sensitivity	Specificity	PPV	NPV
Test cohort 2	0.85 (0.83–0.87)	0.82 (0.78–0.87)	0.88 (0.83–0.92)	0.87 (0.82–0.90)	0.84 (0.81–0.87)
Doctor 1	0.70 (0.66–0.73)	0.62 (0.56–0.67)	0.78 (0.73–0.83)	0.76 (0.71–0.81)	0.64 (0.60–0.69)
Doctor 2	0.74 (0.70–0.77)	0.66 (0.61–0.71)	0.82 (0.78–0.86)	0.80 (0.75–0.85)	0.68 (0.63–0.73)

Data in parentheses are 95% CIs. CNN, convolutional neural network; PPV, positive predictive value; NPV, negative predictive value.

patients. When enhanced CT and other examinations cannot be carried out, routine US examination and AI-assisted diagnosis can be used to quickly evaluate injuries, which is conducive to the clinical adoption of a reasonable treatment plan and is of great significance to improve the survival rate of patients. This method is convenient and non-invasive and can be applied to pre-hospital diagnoses, post-treatment injury evaluations, and regular follow-ups. We still need to collect additional clinical data from more centers and adjust the original model to increase the accuracy of the diagnoses.

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Footnote

Reporting Checklist: The authors have completed the TRIPOD reporting checklist. Available at <https://atm.amegroups.com/article/view/10.21037/atm-22-3767/rc>

Data Sharing Statement: Available at <https://atm.amegroups.com/article/view/10.21037/atm-22-3767/dss>

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <https://atm.amegroups.com/article/view/10.21037/atm-22-3767/coif>). HL is from Beijing Mindray Medical Instrument Co., Ltd. The other authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The animal experiments were performed under a project license

(No. 2021KY033-KS001) granted by the Medical Ethics Committee of PLA General Hospital, in compliance with National Laboratory Animal Management Regulations and guidelines for the care and use of animals. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). The study was approved by ethics committee of PLA General Hospital (No. S2020-323-01) and informed consent was taken from all the patients.

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Appendix 1

Model Construction

The Static Model is a combination of MobileNet V2. The MobileNet V2 is used to extract the spatial features, which is shown in *Figure S1*.

MobileNet V2

The structure of MobileNet V2 (22) is detailed in *Table S1*, which had 7 bottleneck stages. All of the bottleneck stages were constructed by a 2-dimensional convolution layer, batch normalization layer, ReLU activation layer, and depth-wise convolution layer (see *Figure S1*).

Mathematics for layers

Convolution

Convolution is a basic operation used to extract the local features in an image (28), which can be expressed as:

$$g(i, j) = \sum_{k,l} f(i+k, j+l)h(k, l)$$

where $f(i, j)$ is the original image, $h(k, l)$ is the convolution kernel, and $g(i, j)$ is the feature image.

Batch normalization

Batch normalization (BN) is a non-parametric operation, which is used to accelerate the training process and address the overfit problem (29). The BN is a practical method to regularize a model and enables higher learning rates. The formulation can be expressed as:

$$\hat{x} = \frac{x - E[x]}{\sqrt{\text{Var}[x] + \epsilon}}$$

Where $E[x]$ is the expectation over training mini-batches, and $\text{Var}[x] = \frac{m}{m-1} \dot{E}_B[\sigma_b^2]$ is the unbiased variance estimate over training mini-batches of size m and sample variances σ_b^2 . This operation is used to follow the convolution operation.

Non-linear activation

Non-linear activation is a non-linear function that can transform a linear system to a non-linear system. The operation usually uses a map function to enhance the fitting ability of the model. Sigmoid function and ReLU function (30) were used in our models, and both of these operations were element-wise. Sigmoid function is formulated as:

$$f(x) = \frac{1}{1 + e^{-x}}$$

and this activation is used in the last dense layer to match the binary cross entropy loss. The ReLU function is expressed as:

$$f(x) = \max(x, 0)$$

and this operation is used in each BN operation.

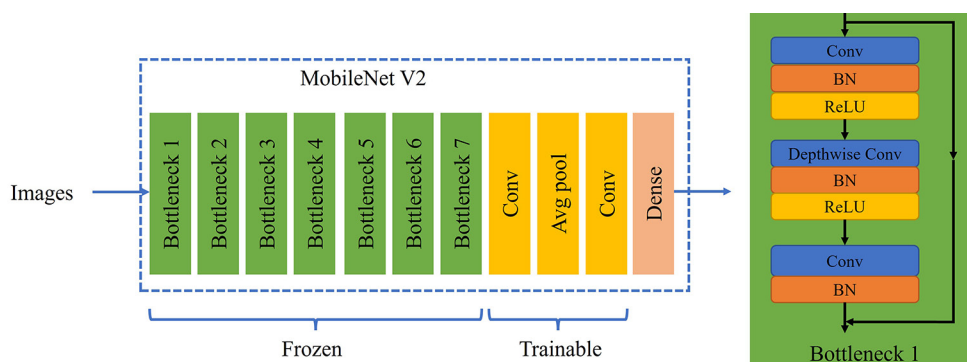


Figure S1 The structure of the static model.

Table S1 Structure of ResNet50

Layers	Output Size	Channel Size	Stride	Expansion
Convolution	112*112	32	2	–
Bottleneck 1	112*112	16	1	1
Bottleneck 2	56*56	24	2	6
Bottleneck 3	28*28	32	2	6
Bottleneck 4	14*14	64	2	6
Bottleneck 5	14*14	96	1	6
Bottleneck 6	7*7	160	2	6
Bottleneck 7	7*7	320	1	6
Conv	7*7	1280	1	–
Average Pool	1*1	–	–	–
Conv	1*1	1	–	–
Dense	1	–	–	–

Pooling

Pooling is a method to reduce the parameters and redundant information. Due to the static property of images, local features can be described by the statistics of local regions. Thus, the operation paid attention to the whole image domain and ignored the local information, which led to the invariance in translation, rotation, and scale. In this study, we use 2-dimensional maximum pooling with strides 2×2 and 2-dimensional global average pooling with strides 2×2 in the first convolution stage and final part.

Constraint

Weight constraint is a way to control the weight that needs to be trained. A weight constraint can keep the updated weight in a small range, which is a strategy for reducing overfit. The ResNet50 is based on ImageNet weights used in our study. Thus, only the last dense layer needed to be set. In this study, the weight constraint was used as the UnitNorm, which can be expressed as:

$$\text{norm}(w) = 1$$

Table S2 The hyper parameters for training process.

Model	LR	BS	Eq	Dropout
Animal Model	1e-07	64	60	0.5
Human Model	1e-07	64	60	0.5

LR, learning rate; BS, batch size; Ep, epoch.

Depth-wise separable convolution

Depth-wise separable convolution is a two-step convolution operation to reduce the parameters of convolution (28), which can be expressed as depth-wise convolution and pointwise convolution.

Depth-wise convolution can be expressed as:

$$g(i,j,c) = \sum_{k,l} f(i+k, j+l, c)h(k,l,c)$$

Where $f(i,j,c)$ is the original image for channel c , $h(k,l,c)$ is the convolution kernel with c filters, and $g(i,j,c)$ is the feature image. Pointwise convolution can be expressed as:

$$g(i,j,M) = \sum_{k,l,c} f(i+k, j+l, c)h(k,l,M)$$

Where $f(i,j,c)$ is the original image for channel c , $h(k,l,c)$ is the $1*1$ convolution kernel with M filters, and $g(i,j,M)$ is the feature image.

Training details

In this study, a clinical model was trained. The model was trained in 2 steps. First, it was trained using animal US images. Second, the model was fine-tuned using a few human US images. For the 1st step, 3-fold CV was used for the animal US images. For the 2nd step, each fold CV model was fine-tuned with the last 3 layers from the same human US images and averaged to obtain the final clinical model. The training process employed an optimization process to obtain the parameters in the deep-learning model. In our study, 4 deep-learning models were trained by gray scale (GS) images. The data were augmented to overcome the overfit in the training process. The augmentation included vertical and horizontal random flipping, random brightness, and random contrast. The classification model sought to predict 2 classes of images. In the training set, binary cross entropy loss was used for the classification, which can be expressed as:

$$L = -\frac{1}{n} \sum_n [y \ln x + (1-y) \ln(1-x)]$$

where the sample number is n , x is the prediction, and y is the ground truth. An algorithm for the 1st-order gradient-based optimization of stochastic objective functions, based on adaptive estimates of lower-order moments (Adam) (31), served as the optimizer. The hyper parameter learning rate (LR), batch size (BS) and epoch (Ep) are listed in *Table S2*.

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