



Bioinformatics based exploration of hsa-miR-194-5p regulation of CHD4 through PI3K/AKT signal pathway to enhance tumor resistance to apoptosis due to loss of nests and participate in poor prognosis of oral squamous cell carcinoma

Qin Li^{1#}, Kai Wang^{2#}, Yi Shen³, Chaosheng Lin⁴, Jie Miao⁵, Xun Hu⁵

¹Department of Stomatology, Eye & ENT Hospital, Fudan University, Shanghai, China; ²Department of Stomatology, Gongli Hospital, The Second Military University, Shanghai, China; ³Department of General Surgery, Ruijin Hospital Luwan Branch, Shanghai Jiaotong University School of Medicine, Shanghai, China; ⁴Department of Oral and Maxillofacial Surgery, Ruijin Hospital, Shanghai Jiaotong University School of Medicine, Shanghai, China; ⁵Department of Stomatology, Ruijin Hospital Luwan Branch, Shanghai Jiaotong University School of Medicine, Shanghai, China

Contributions: (I) Conception and design: Q Li, J Miao, X Hu; (II) Administrative support: X Hu; (III) Provision of study materials or patients: K Wang; (IV) Collection and assembly of data: Y Shen; (V) Data analysis and interpretation: C Lin; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

[#]These authors contributed equally to this work.

Correspondence to: Jie Miao; Xun Hu. Department of Stomatology, Ruijin Hospital Luwan Branch, Shanghai Jiaotong University School of Medicine, Shanghai, China. Email: miaojie0504@163.com; Doctorhu1978@163.com.

Background: Recent evidence shows that CHD4 is involved in a variety of biological events of tumors. Our aim was to investigate the correlation between CHD4 and oral squamous cell carcinoma (OSCC).

Methods: After CHD4 was screened as a differentially expressed gene in The Cancer Genome Atlas (TCGA) database, the correlations of its expression level with the clinical parameters and prognosis of patients with OSCC were analyzed. The outcomes of the multivariate analysis were used to construct a nomogram, and the accuracy of the model was evaluated with the calibration curve. The GeneMANIA and STRING databases were used to generate network diagrams depicting interactions of genes with CHD4, and heat maps of genes co-expressed with CHD4 were generated using the TCGA database. TargetScan was then used to look into the miRNAs that interact with the 3' untranslated region of CHD4 mRNA. Finally, GSEA enrichment analysis was used to explore the possible mechanism.

Results: The differentially expressed molecule CHD4 was screened by TCGA database for OSCC. CHD4 was overexpressed in OSCC tumor tissues, and OSCC patients with low expression of CHD4 have better OS and DSS. The nomogram had a C-index of 0.575 (0.548–0.602), which indicated some degree of predictive reliability. CHD4 has certain correlation with exons of OSCC related genes, including *TP53*, *NOTCH1*, *CASP8*, *PTEN*, *TP63*, *ANXA1*, *CDH1*, *CTNNB1*, *GDF15* and *EGFR*. According to the TargetScan database, hsa-miR-194-5p is the miRNA that regulates CHD4 upstream the most. GSEA analysis showed that CHD4 may participate in the poor prognosis of OSCC by participating in PI3K/AKT pathway, protein adhesion regulation, MAPK pathway, cytokine and inflammatory response regulation, angiogenesis and platelet regulation.

Conclusions: The decreased expression of CHD4 may indicate a better prognosis in OSCC patients and could serve as a novel predictive biomarker for OSCC. Also, hsa-miR-194-5p was found to contribute to the poor prognosis of OSCC by regulating CHD4 and enhancing tumor anoikis resistance via the PI3K/AKT signaling pathway. These findings suggest that CHD4 might be a therapeutic target for the effective treatment of OSCC.

Keywords: CHD4; oral squamous cell carcinoma (OSCC); PI3K/AKT pathway; anoikis; bioinformatics analysis

Submitted Dec 02, 2022. Accepted for publication Jan 11, 2023. Published online Jan 31, 2023.

doi: 10.21037/atm-22-6332

View this article at: <https://dx.doi.org/10.21037/atm-22-6332>

Introduction

Oral squamous cell carcinoma (OSCC) refers to squamous cell carcinoma originating in the oral cavity, including the cheeks, lips, mouth floor, tongue, and palate. It is one of the common head and neck malignant tumors worldwide (1,2). There are almost 300,000 new cases of OSCC each year, and about 170,000 related deaths (3). The delayed diagnosis of oral cancer is strongly linked to its high fatality rate (4).

Despite advancements in treatment, the 5-year survival rate of oral cancer is still less than 50%, making the disease a severe health concern. At present, the treatment of patients with advanced or lymph node metastasis mainly depends on surgery combined with radiotherapy and chemotherapy, but these treatments often lead to serious side effects. At present, it has been found that a variety of “oncogenes”, “tumor suppressor genes” and multiple signal pathways are related to the occurrence and development of OSCC. For example, Sun *et al.* found that procyanidin B2 could inhibit cell growth and angiogenesis in OSCC via the vascular endothelial growth factor (VEGF)/VEGF receptor 2 (VEGFR2) pathway (5). Snail and Slug cooperate in EMT and tumor metastasis in oral tongue squamous cell

carcinoma through MiR-101 mediated EZH2 axis (6). Carnosic acid inhibits the development of OSCC through mitochondrial mediated apoptosis (7). Commonly used diagnostic methods, such as serum tumor markers and imaging, are not ideal for OSCC and do not meet all clinical needs. For example, CYFRA21-1 as a serum tumor marker for follow-up patients with squamous cell lung cancer and oropharyngeal squamous cell carcinoma (8). GDF 15 as an anti apoptosis, diagnostic and prognostic marker of OSCC (9). Hyperion imaging system can reveal heterogeneous tumor microenvironment of T1N0M0 OSCC patients (10). Indocyanine green fluorescent navigation technology can determine the safe boundary of advanced OSCC (11). But these technologies have certain limitations. Therefore, screening of markers associated with the early diagnosis and poor prognosis of OSCC is crucial for improving the therapeutic effect and prognosis of patients with this disease.

The human chromodomain helicase DNA-binding (CHD) protein family is a class of chromatin remodeling complexes. Six members of the family have been discovered to date: CHD1, CHD2, CHD3, CHD4, CHD5, and CHD6. All CHD proteins are members of the SWI2/SNF2-related ATPase superfamily. Among them, CHD4 and CHD5 together form the second subfamily of this family according to the conservation of the coding protein sequence (12,13). CHD proteins have a DNA-binding domain at their C termini, opposite a chromatin regulatory domain at their N termini. Different components of the SWI/SNF complex (brahma-related gene 1) and RET finger protein are bound at the N- and C-termini of CHD4, respectively, and each has its own transcriptional activity. Sequence analysis of the fluoroenzyme reporter gene showed that the N-terminal of CHD4 had strong trans activation ability, while its C-terminal had transcriptional inhibition activity. On the one hand, CHD4 and BRG1 are interrelated, and the bromo domain of BRG1 strongly inhibits the trans activation of the N-terminal of CHD4. On the other hand, CHD4 and RET finger proteins play a common role in transcriptional inhibition. CHD4 interacts with both trans-activators and repressors, and is directly

Highlight box

Key findings

- CHD4 plays an important role in the occurrence and development of various tumors. Our study found that hsa-miR-194-5p can regulate CHD4 and enhance the tumor resistance to apoptosis through PI3K/AKT signaling pathway, thus participating in the poor prognosis of oral squamous cell carcinoma.

What is known and what is new?

- The mortality of oral cancer is high. It is important to screen the markers related to OSCC diagnosis and treatment.
- The decreased expression of CHD4 can improve the prognosis of OSCC patients and provide a therapeutic target for the treatment of OSCC.

What is the implication, and what should change now?

- In the future, more *in vitro* and *in vivo* experiments are needed to further determine the relevant mechanism of action.

related to chromatin remodeling proteins, thus providing a new perspective on the form of multiprotein super complexes involved in transcriptional regulation (14).

Our study mainly aimed to explore the expression of CHD4 in The Cancer Genome Atlas (TCGA) database and its role in predicting survival in patients with OSCC. A bioinformatics analysis was also performed to predict upstream regulatory genes of CHD4 in OSCC to investigate its effect on the behavior of tumor cells and the underlying mechanisms. We present the following article in accordance with the TRIPOD reporting checklist (available at <https://atm.amegroups.com/article/view/10.21037/atm-22-6332/rc>).

Methods

Analysis of CHD4 expression in the TCGA database

The expression of CHD4 was analyzed in pan-cancer and in OSCC tumor and adjacent tissues in the TCGA database. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013).

Analysis of the relationship between CHD4 and clinical parameters of patients with OSCC in the TCGA database

Data of clinical parameters of patients with OSCC were retrieved from the TCGA database, and then the correlations between CHD4 expression and these clinical parameters and patient prognosis were analyzed.

Construction and evaluation of a nomogram

A nomogram was constructed using the data from the multivariate analysis. A calibration curve was generated to determine the predictive value of the nomogram.

Construction of a protein-gene interaction network map

GeneMANIA was used to construct a gene-gene interaction network for CHD4 and the altered adjacent genes. At the same time, the STRING database was employed to generate the protein-protein interaction network of CHD4. Finally, the two interaction networks were compared.

Screening of the co-expressed genes of CHD4

Genes that were positively or negatively co-expressed with

CHD4 were identified through data mining of the TCGA database.

Prediction of upstream regulatory miRNAs of CHD4

To further identify the upstream genes that regulate CHD4, miRNAs that may bind to the 3'UTR of CHD4 mRNA were explored through the TargetScan database.

Gene set enrichment analysis (GSEA)

Online functional analysis of differentially expressed genes was carried out using Metascape. The cellular mechanisms in which CHD4 possibly plays a role in OSCC were investigated via GSEA.

Statistical analysis

The R (version 3.6.3) software was used for statistical analysis and visualization. Kaplan-Meier analysis was used to determine the survival time of patients with OSCC, and the log-rank test was employed to test for statistical significance. Spearman's correlation coefficient was used for correlation analysis. When comparing means, a significant difference was indicated by $P < 0.05$.

Results

CHD4 expression is upregulated in OSCC

We used the TCGA database to identify CHD4 as our target research molecule. The expression of CHD4 was upregulated in the vast majority of tumors in the pan-cancer analysis (*Figure 1A*). Moreover, the expression of CHD4 in matched and unmatched tumor tissues of OSCC is higher than that in adjacent cancer tissues. It is found that AUC is 0.814 by constructing ROC curve, so CHD4 has good prediction research value (*Figure 1B-1D*).

Correlation between CHD4 expression and the prognosis of patients with OSCC

In the TCGA database, we found that patients with OSCC who had reduced CHD4 expression had superior overall survival (OS) and disease-specific survival (DSS) (*Figure 2A*). Further stratified analysis revealed that patients with low CHD4 expression had a better prognosis in patients with T2, N0, M0, Stage II, Stage III, male and female, older

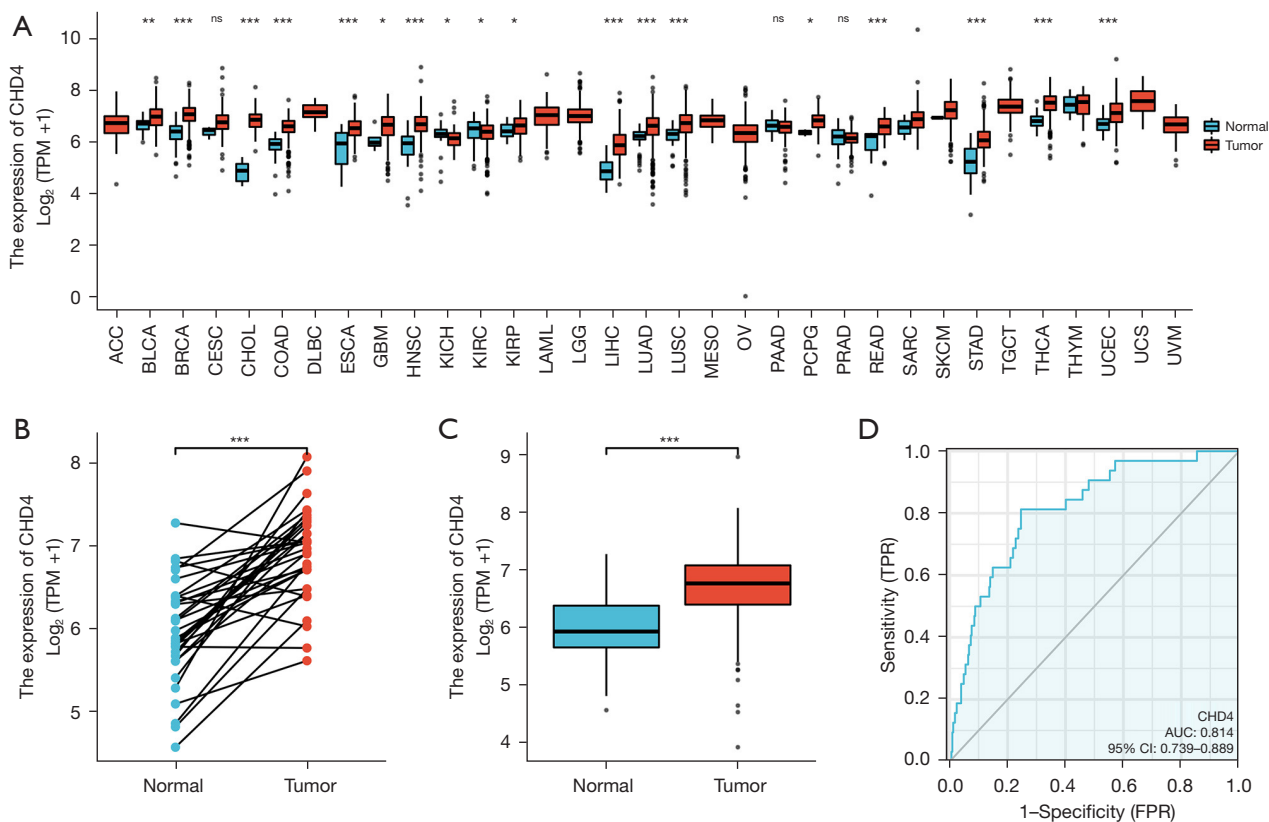


Figure 1 CHD4 expression is upregulated in oral squamous cell carcinoma. (A) The CHD4 expression level in pan-cancer. (B) Expression levels of CHD4 in paracancerous tissues (n=32) and unpaired OSCC tissues (n=329) in The Cancer Genome Atlas database. (C) Expression levels of CHD4 in paracancerous tissues (n=32) and matched OSCC tissues (n=329). (D) Receiver-operating characteristic curve analysis demonstrating the strong capacity of CHD4 to distinguish between tumor and normal tissues. Statistically significant difference: *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$. ns, no significance; TPM, transcripts per million; TPR, true positive rate; AUC, area under the curve; FPR, false positive rate; OSCC, oral squamous cell carcinoma.

than 60 years, G2, G3, and whether or not they had smoked or consumed alcohol in the past (Figure 2B).

The relationships between CHD4 expression and clinical data of patients with OSCC

In comparison with the clinical data of OSCC patients, we can find that patients with higher expression of CHD4 have higher TNM stage, clinical stage and pathological stage, which is consistent with the previous research results (Figure 3).

Construction of a nomogram

The nomogram constructed using the TCGA database and the findings of our multivariate analysis had a C-index

of 0.575 (0.548–0.602) for predicting 1-, 3-, and 5-year survival in patients with OSCC (Figure 4A). The calibration curve of the nomogram also demonstrated that it had some predictive power (Figure 4B).

Identification of genes and protein that interact with CHD4

Using GeneMANIA, we mapped out the gene-gene interaction network of CHD4 and its modified neighboring genes (Figure 5A). Using the STRING database, the protein-protein interaction network of CHD4 was constructed (Figure 5B).

Screening of co-expressed genes of CHD4

The OSCC data in the TCGA database were used to

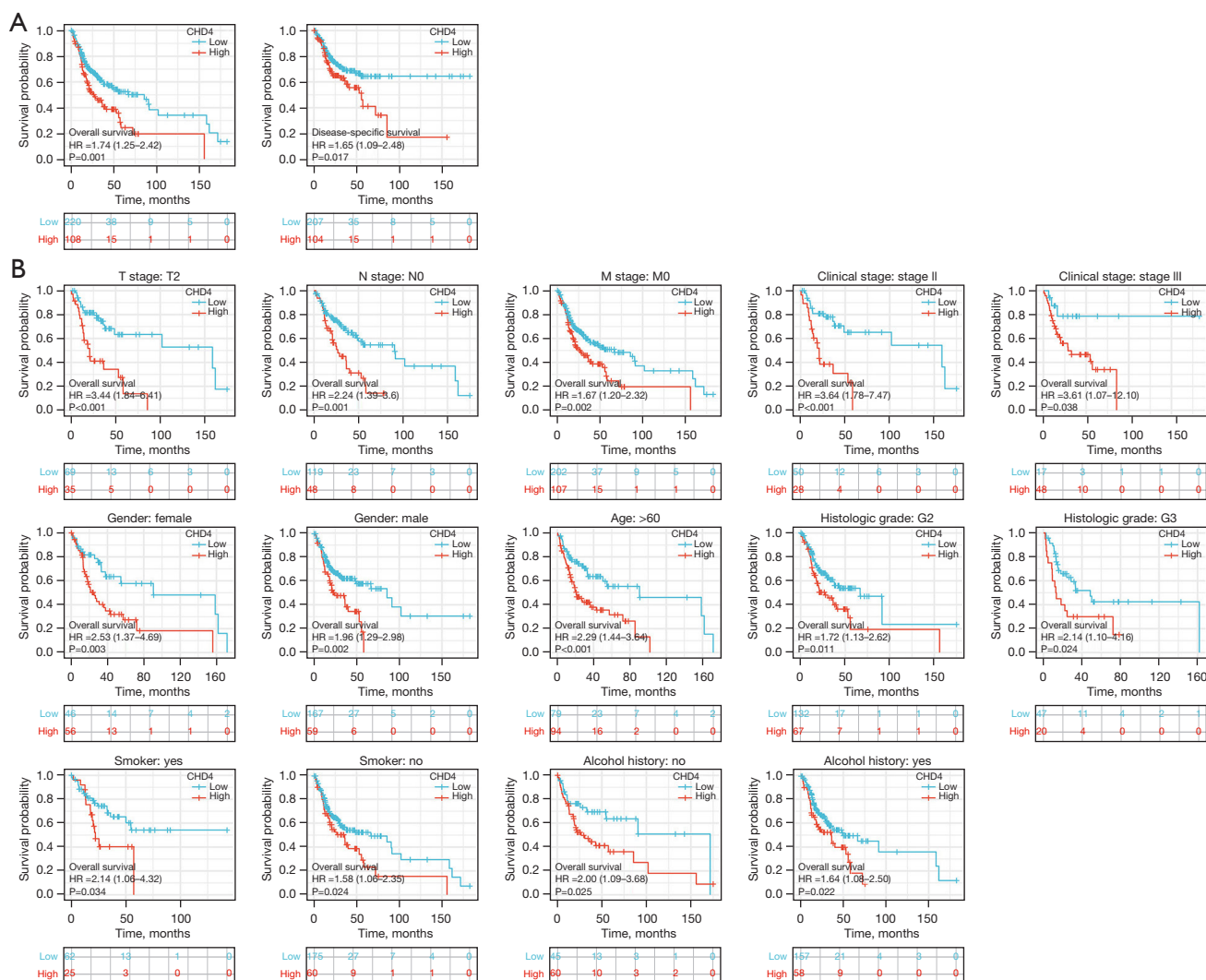


Figure 2 The relationship between CHD4 expression in The Cancer Genome Atlas database and patient prognosis in oral squamous cell carcinoma. (A) The association of CHD4 expression with overall survival and disease-specific survival in patients with OSCC. (B) The association of CHD4 expression with patient prognosis of OSCC in stratified subgroup analysis. Statistically significant difference: P < 0.05. OSCC, oral squamous cell carcinoma.

identify the genes that are positively or negatively co-expressed with CHD4, and heat maps of the top 50 positively (Figure 6A) and negatively (Figure 6B) co-expressed genes were constructed.

Due to the impact gene mutations have on clinical diagnosis and treatment of OSCC, we next investigated the association between CHD4 and OSCC-related gene exons from the TCGA database, including *TP53*, *NOTCH1*, *CASP8*, *PTEN*, *TP63*, *ANXA1*, *CDH1*, *CTNNB1*, *GDF15*,

and *EGFR* (Figure 7).

Predicted upstream regulatory miRNAs of CHD4 in the TargetScan database

To further identify the upstream genes that regulate CHD4 we further investigated the miRNAs that may bind to the 3'UTR of CHD4 mRNA through the TargetScan database. The analysis showed that hsa-miR-194-5p was the most

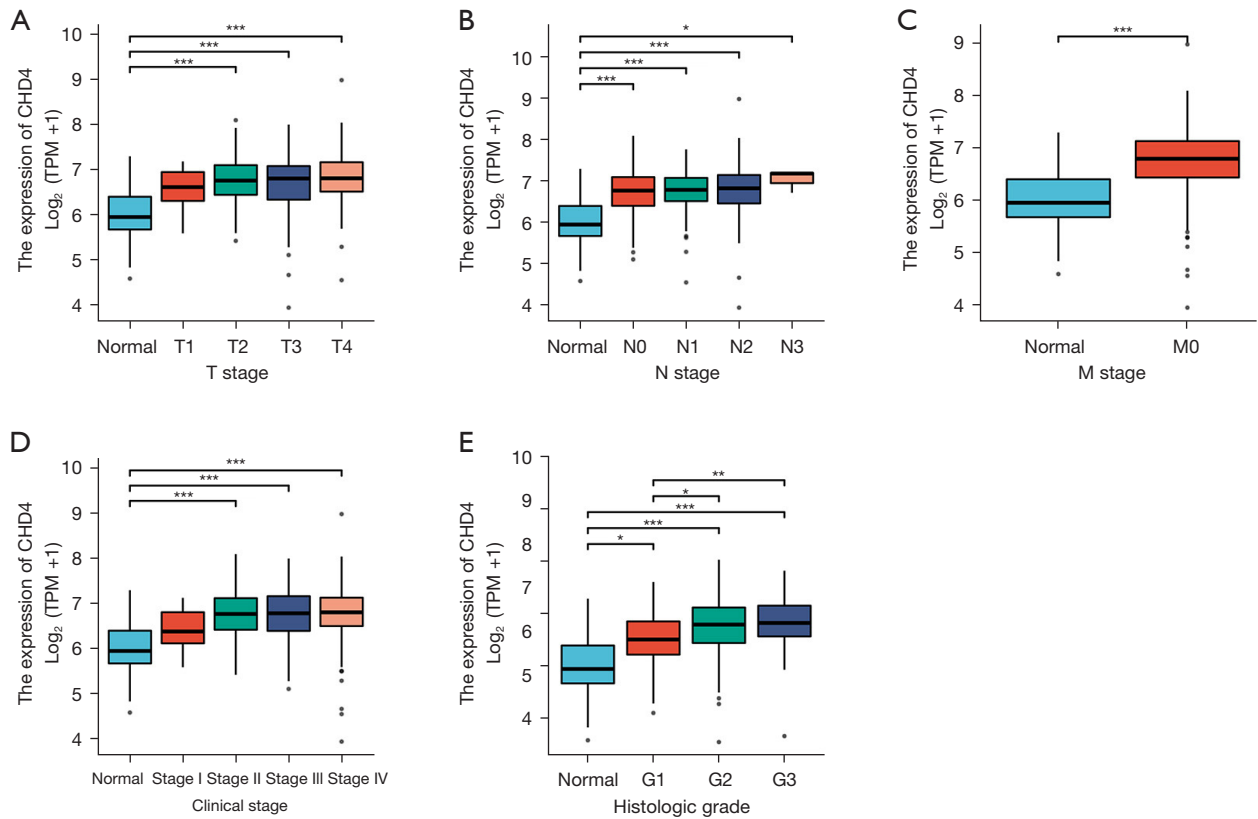


Figure 3 The relationships between CHD4 expression and clinical data of patients with oral squamous cell carcinoma. *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$. TPM, transcripts per million.

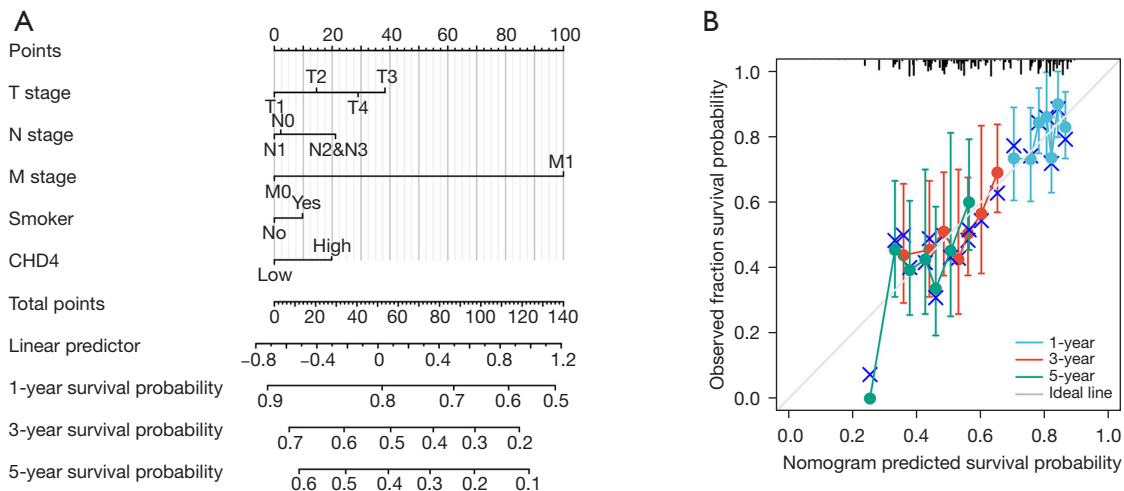


Figure 4 A nomogram to predict the survival of patients with OSCC and its calibration plot. (A) The nomogram to predict the 1-, 3-, and 5-year survival of patients with OSCC, and (B) the calibration plot for the nomogram. OSCC, oral squamous cell carcinoma.

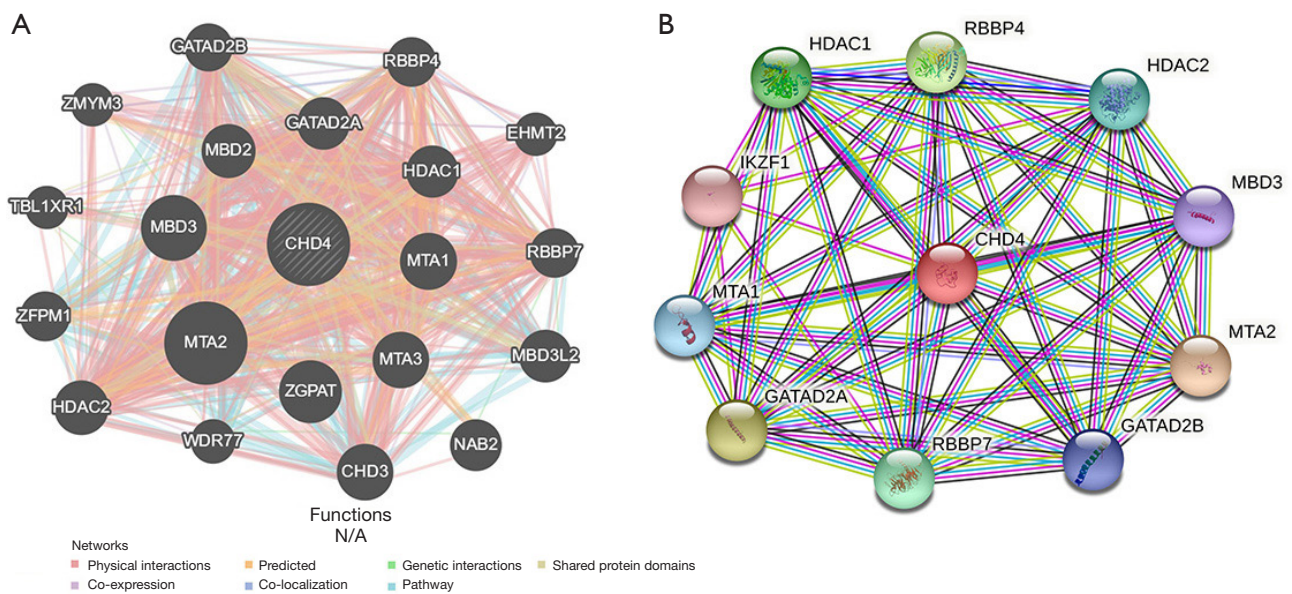


Figure 5 Identification of CHD4-interacting genes and proteins. (A) Gene-gene interaction network of CHD4 in GeneMANIA database. (B) The protein-protein interaction network of CHD4 in the STRING database.

likely upstream regulatory miRNA of CHD4. Details of the predicted miRNAs are shown in *Table 1*.

GSEA analysis results

Kyoto Encyclopedia of Genes and Genomes functional analysis was performed online using Metascape. We found 8 statistically significant possible related pathways: (I) REACTOME PI3K AKT SIGNALING IN CANCER; (II) NABA ECM GLYCOPROTEINS; (III) REACTOME ADHERENS JUNCTIONS INTERACTIONS; (IV) KEGG TGF BETA SIGNALING PATHWAY; (V) KEGG MAPK SIGNALING PATHWAY; (VI) WP CYTOKINES AND INFLAMMATORY RESPONSE; (VII) WP ANGIOGENESIS; (VIII) REACTOME PLATELET CALCIUM HOMEOSTASIS (*Figure 8*).

Discussion

Squamous cell carcinoma, which accounts for over 90% of all oral malignancies, is notorious for its persistence and rapid rate of metastasis (15). At present, although OSCC has made progress in systematic treatment, the survival rate of patients is still very low due to late diagnosis and multiple potential recurrence factors. Biomarkers have evolved into highly effective diagnostic, prognostic, and therapeutic tools

through the use of multi-omics technology (16).

In this study, we employed bioinformatics analysis to investigate the potential roles of CHD4 in the etiology and progression of OSCC. Through analysis of TCGA data, we found that CHD4 expression was elevated in OSCC and that patients with low CHD4 expression had a better prognosis than those with high expression. In our correlation analysis of CHD4 expression with clinical parameters of patients with OSCC, we found that patients with higher CHD4 expression tended to have more advanced TNM, clinical, and pathological stages. The nomogram we constructed based on the multivariate analysis results also adds clinical value to our research findings. We also constructed network maps of genes interacting with CHD4 in the GeneMANIA and STRING databases, as well as a heat map of genes co-expressed with CHD4 in the TCGA database. These results provide some insights for future cytological basic experiments.

With the recurrence and progression of cancer, gene mutations are particularly important in the selection of last-line treatment plans, and many guidelines emphasize the importance of genetic testing in diagnosis and treatment (17). The whole genome exon sequencing results showed that TP53, NOTCH1 and CDKN2A were the most common mutation genes in head and neck squamous cell carcinoma. In addition, research has shown that CASP8,

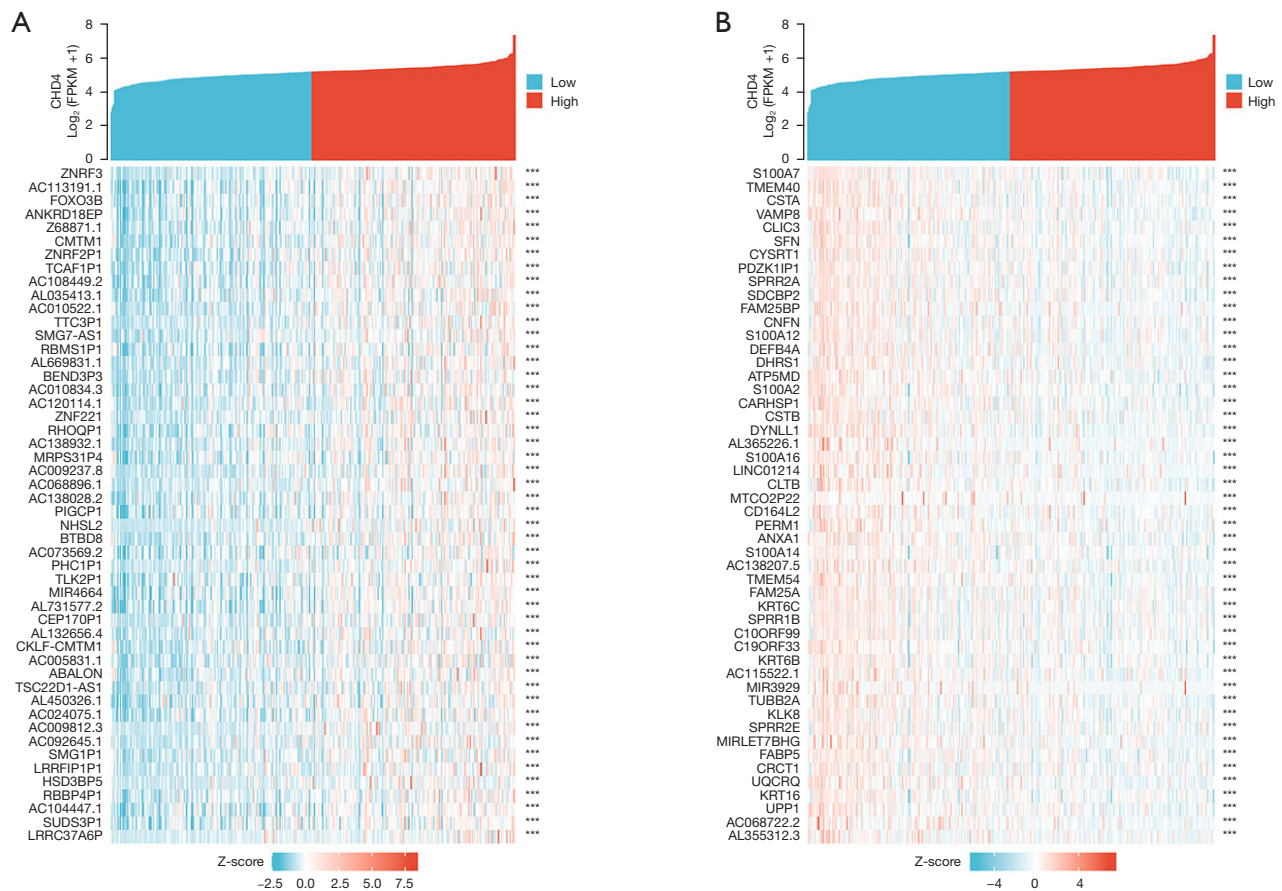


Figure 6 Heat maps of genes co-expressed with CHD4 in oral squamous cell carcinoma. (A) Positively correlated genes co-expressed with CHD4 in OSCC; (B) negatively correlated genes co-expressed with CHD4 in OSCC. FPKM, fragments per kilobase of exon model per million mapped fragments; OSCC, oral squamous cell carcinoma.

PTEN and TP63 have important regulatory functions in the differentiation of squamous cells, and that mutations in these genes are the main driving factors for the occurrence and development of head and neck squamous cell carcinoma (17-19). The expression of each of these genes is essential for the proper development of squamous cells. Head and neck squamous cell carcinoma has many key biomarkers, including ANXA1, CDH1, CTNNB1, and TGFB1, which are expressed as proteins (17). This study focused on the mutation of the above hot spot genes in OSCC. We found that CHD4 had good correlation with TP53, NOTCH1, CASP8, PTEN, TP63, ANXA1, CDH1, CTNNB1, GDF15 and EGFR. We then used the TargetScan database to search for additional relevant miRNAs that could bind to the 3'UTR of CHD4 mRNA. Our analysis determined that hsa-miR-194-5p was the most plausible upstream regulatory miRNA of CHD4. At present, there have been some studies

on this miRNA in tumors, such as PCED1B-AS1, which can induce immunosuppression in hepatocellular carcinoma by double targeting PD-L1 and PD-L2 with spongy hsa-miR-194-5p (20). Zhang *et al.* found that miR-194-5p also mediates GDNF-induced glioma cell proliferation and migration (21). A study on laryngeal cancer showed that hsa_circ_0023028 knockdown inhibits cell migration, proliferation, and invasion (22). Vajen *et al.* reported that miRNA-192-5p inhibits the migration of triple-negative breast cancer cells and directly regulates Rho GTPase-activating protein 19 (23). However, hsa-miR-194-5p has not been the subject of any studies on OSCC.

In the final part of our study, we performed a GSEA analysis and found eight statistically significant potentially relevant pathways: (I) REACTOME PI3K AKT SIGNALING IN CANCER; (II) NABA ECM GLYCOPROTEINS; (III) REACTOME ADHERENS

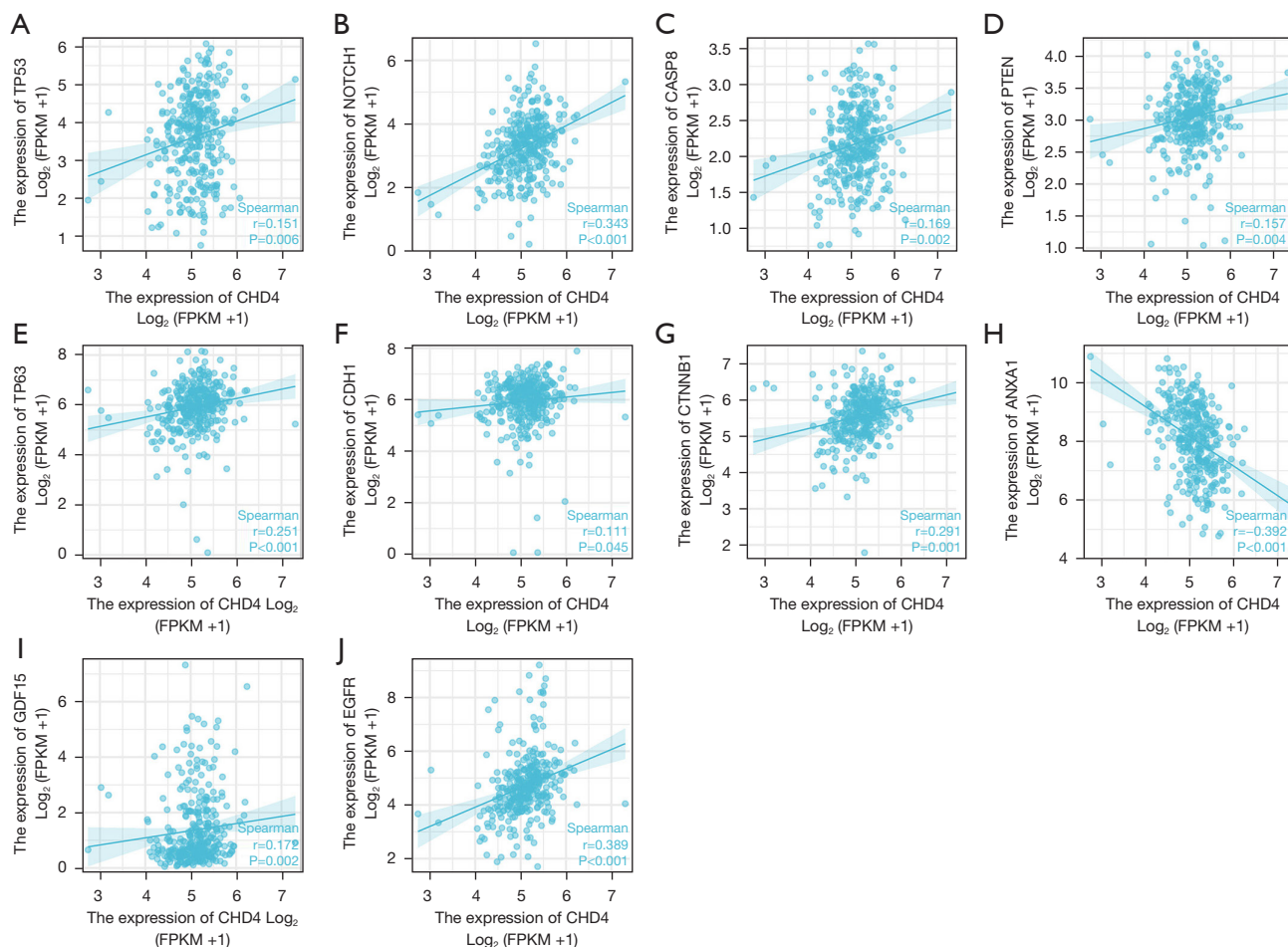


Figure 7 Correlation between CHD4 and oral squamous cell carcinoma-related gene exons. $P<0.05$ indicates a statistically significant difference. FPKM, fragments per kilobase of exon model per million mapped fragments.

Table 1 The best miRNAs predicted to bind to the 3'UTR of CHD4 in the TargetScan database

Position of CHD4 3' UTR	Predicted consequential pairing of target region (top) and miRNA (bottom)	Site type	Context++ score	Context++ score percentile	Weighted context++ score	Conserved branch length	PCT	Predicted relative KD
Hsa-miR-194-5p	5' UCCCCACUGUACGCCUGUUACA; 3' AGGUGUACCUCAACGACAAUGU	8mer	-0.13	83	-0.13	4.030	0.42	-4.539

KD, k-dimensional; PCT, percentage.

JUNCTIONS INTERACTIONS; (IV) KEGG TGF BETA SIGNALING PATHWAY; (V) KEGG MAPK SIGNALING PATHWAY; (VI) WP CYTOKINES AND INFLAMMATORY RESPONSE; (VII) WP ANGIOGENESIS; (VIII) REACTOME PLATELET CALCIUM HOMEOSTASIS.

As a tumor research hotspot, anoikis has recently become

the focus of more scientists. When normal epithelial cells lose touch with the surrounding extracellular matrix, a process known as anoikis occurs and cell death rapidly follows. Anoikis is a special type of programmed cell death that plays a critical role in normal morphogenesis, tissue homeostasis, disease development, and tumor metastasis. When cells lose their attachment to ECM, the pro

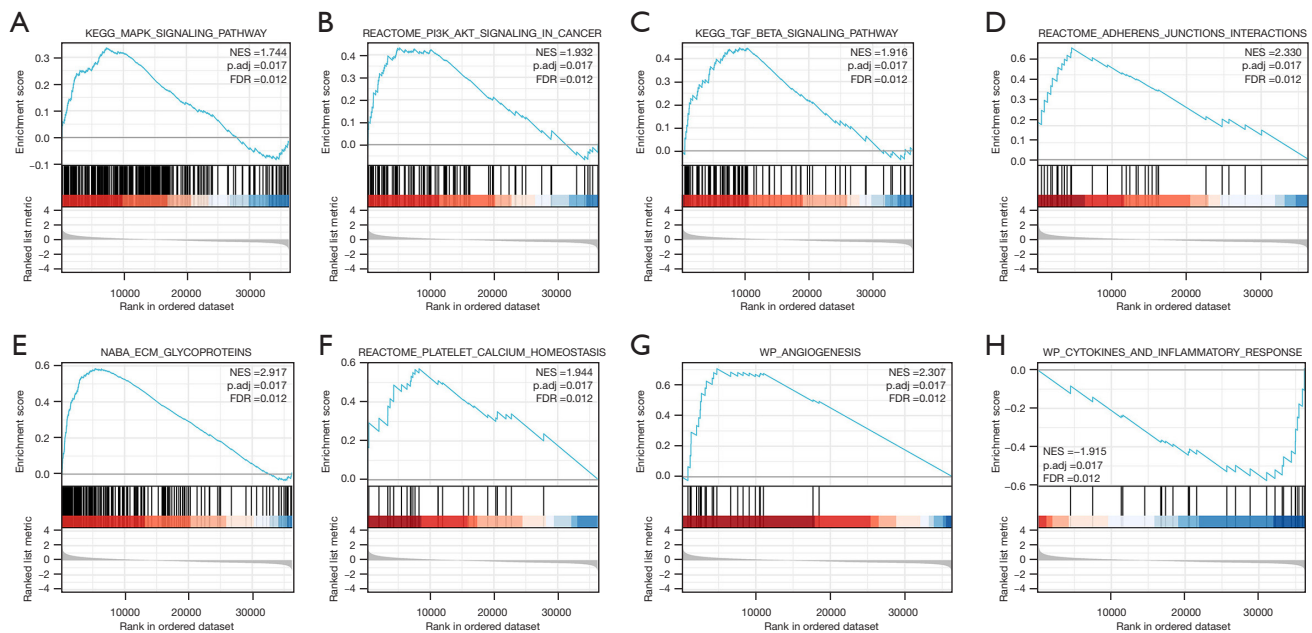


Figure 8 The eight statistically significant Kyoto Encyclopedia of Genes and Genomes pathways. MSigDB utilizes a set of biological process genes called gene set G. $P < 0.05$; NES, normalized enrichment score; FDR, false discovery rate.

apoptotic protein Bmf, which remains on the cytoskeleton, dissociates from dynein light chain 2 and translocates to mitochondria, thus promoting the occurrence of nesting apoptosis (24). However, a common feature of tumor development and growth is the viability of transformed cells under “independent” growth conditions. This resistance to nestless apoptosis has been proved to be related to the loss of homeostasis in the cellular environment, cancer growth and metastasis, and this acquired ability is called the resistance to anoikis apoptosis (25). Cancer cells with anti anoikis apoptosis can spread to distant tissues or organs through the peripheral circulation system and cause cancer metastasis. To study the molecular mechanism of anti anoikis apoptosis will be helpful to explore effective therapies for human malignant tumors.

Combined with the results of our GSEA analysis, we also found that in previous studies, these predicted signal pathways have also shown the correlation with tumor anoikis apoptosis. For example, celecoxib enhances the anticancer effect of cisplatin and induces homing apoptosis in osteosarcoma through PI3K/Akt pathway (26). CEMIP overexpression triggered by AMPK/GSK3 β /beta-catenin cascade promotes the migration and invasion of apoptotic prostate cancer cells by enhancing metabolic reprogramming (27). Small-molecule RGD integrin

antagonists can inhibit cell adhesion and cell migration, and induce anoikis in glioblastoma cells (28). Stereospecific effect of ginsenoside 20-Rg3 on TGF inhibition- β Induces epithelial mesenchymal transformation and inhibits lung cancer migration, invasion and anti nesting apoptosis (29). Silencing of tetramethylthioacetic acid and transthyroxine protein inhibits the metastasis of L-thyroxine in apoptotic prostate cancer cells by regulating MAPK/ERK pathway (27). The loss of p120 catenin induces metastatic progression of breast cancer by inducing the resistance to apoptosis and enhancing the signal transduction of growth factor receptor (30). Anti nesting apoptotic gastric cancer cells pass C/EBP β mediated autocrine and paracrine signals of PDGFB promote angiogenesis and peritoneal metastasis (31). Overexpression of multiple epidermal growth factors such as domain 11 saves the survival of apoptosis through tumor cell platelet interaction in triple negative breast cancer cells (32). Therefore, our research results in this project combined with previous research results, our findings suggest that hsa-miR-194-5p regulates CHD4 via the PI3K/AKT signaling pathway, which in turn increases tumor anoikis resistance and contributes to the poor prognosis of OSCC. These findings provide a more reliable baseline against which to gauge future improvements to cytological and other *in vitro* and *in vivo* experiments.

Conclusions

This study adds to the growing body of data that CHD4 plays an important role in the emergence of OSCC and has the potential to serve as a biomarker for the progression of the disease. Our results provide a potential target for the development of OSCC anti-cancer strategies

Acknowledgments

Funding: None.

Footnote

Reporting Checklist: The authors have completed the TRIPOD reporting checklist. Available at <https://atm.amegroups.com/article/view/10.21037/atm-22-6332/rc>

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <https://atm.amegroups.com/article/view/10.21037/atm-22-6332/coif>). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013).

Open Access Statement: This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

References

- Král D, Pink R, Šašková L, et al. Bone invasion by oral squamous cell carcinoma. *Acta Chir Plast* 2021;63:139-44.
- Madera M, Tirado Amador L, Leal Acosta C. Therapeutic Options in Unresectable Oral Squamous Cell Carcinoma: A Systematic Review. *Cancer Manag Res* 2021;13:6705-19.
- Bray F, Ferlay J, Soerjomataram I, et al. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2018;68:394-424.
- Marsh D, Suchak K, Moutasim KA, et al. Stromal features are predictive of disease mortality in oral cancer patients. *J Pathol* 2011;223:470-81.
- Sun Q, Zhang T, Xiao Q, et al. Procyanidin B2 inhibits angiogenesis and cell growth in oral squamous cell carcinoma cells through the vascular endothelial growth factor (VEGF)/VEGF receptor 2 (VEGFR2) pathway. *Bioengineered* 2022;13:6500-8.
- Zheng M, Jiang YP, Chen W, et al. Snail and Slug collaborate on EMT and tumor metastasis through miR-101-mediated EZH2 axis in oral tongue squamous cell carcinoma. *Oncotarget* 2015;6:6797-810.
- Min F, Liu X, Li Y, et al. Carnosic Acid Suppresses the Development of Oral Squamous Cell Carcinoma via Mitochondrial-Mediated Apoptosis. *Front Oncol* 2021;11:760861.
- Liu L, Liu B, Zhu LL, et al. CYFRA21-1 as a serum tumor marker for follow-up patients with squamous cell lung carcinoma and oropharynx squamous cell carcinoma. *Biomark Med* 2013;7:591-9.
- Schiegnitz E, Kämmerer PW, Koch FP, et al. GDF 15 as an anti-apoptotic, diagnostic and prognostic marker in oral squamous cell carcinoma. *Oral Oncol* 2012;48:608-14.
- Xie S, Shan XF, Yau V, et al. Hyperion imaging system reveals heterogeneous tumor microenvironment of oral squamous cell carcinoma patients at T1N0M0 stage. *Ann Transl Med* 2020;8:1513.
- Wu Z, Dong Y, Wang Y, et al. Clinical application of indocyanine green fluorescence navigation technology to determine the safe margin of advanced oral squamous cell carcinoma. *Gland Surg* 2022;11:352-7.
- Hall JA, Georgel PT. CHD proteins: a diverse family with strong ties. *Biochem Cell Biol* 2007;85:463-76.
- Mills AA. The Chromodomain Helicase DNA-Binding Chromatin Remodelers: Family Traits that Protect from and Promote Cancer. *Cold Spring Harb Perspect Med* 2017;7:a026450.
- Shimono Y, Murakami H, Kawai K, et al. Mi-2 beta associates with BRG1 and RET finger protein at the distinct regions with transcriptional activating and repressing abilities. *J Biol Chem* 2003;278:51638-45.
- Strassen U, Hofauer B, Jacobi C, et al. Management of locoregional recurrence in cutaneous squamous cell carcinoma of the head and neck. *Eur Arch*

- Otorhinolaryngol 2017;274:501-6.
16. Yuan Z, Li WT, Ye XD, et al. Novel functional magnetic resonance imaging biomarkers for assessing response to therapy in hepatocellular carcinoma. *Clin Transl Oncol* 2014;16:599-605.
 17. Nakagaki T, Tamura M, Kobashi K, et al. Targeted next-generation sequencing of 50 cancer-related genes in Japanese patients with oral squamous cell carcinoma. *Tumour Biol* 2018;40:1010428318800180.
 18. Stransky N, Egloff AM, Tward AD, et al. The mutational landscape of head and neck squamous cell carcinoma. *Science* 2011;333:1157-60.
 19. Agrawal N, Frederick MJ, Pickering CR, et al. Exome sequencing of head and neck squamous cell carcinoma reveals inactivating mutations in NOTCH1. *Science* 2011;333:1154-7.
 20. Fan F, Chen K, Lu X, et al. Dual targeting of PD-L1 and PD-L2 by PCED1B-AS1 via sponging hsa-miR-194-5p induces immunosuppression in hepatocellular carcinoma. *Hepatol Int* 2021;15:444-58.
 21. Zhang BL, Dong FL, Guo TW, et al. MiRNAs Mediate GDNF-Induced Proliferation and Migration of Glioma Cells. *Cell Physiol Biochem* 2017;44:1923-38.
 22. Chen X, Su X, Zhu C, et al. Knockdown of hsa_circ_0023028 inhibits cell proliferation, migration, and invasion in laryngeal cancer by sponging miR-194-5p. *Biosci Rep* 2019;39:BSR20190177.
 23. Vajen B, Greiwe L, Schäffer V, et al. MicroRNA-192-5p inhibits migration of triple negative breast cancer cells and directly regulates Rho GTPase activating protein 19. *Genes Chromosomes Cancer* 2021;60:733-42.
 24. Khan SU, Fatima K, Malik F. Understanding the cell survival mechanism of anoikis-resistant cancer cells during different steps of metastasis. *Clin Exp Metastasis* 2022;39:715-26.
 25. Nagaprashantha LD, Vatsyayan R, Lelsani PC, et al. The sensors and regulators of cell-matrix surveillance in anoikis resistance of tumors. *Int J Cancer* 2011;128:743-52.
 26. Liu B, Yan S, Qu L, et al. Celecoxib enhances anticancer effect of cisplatin and induces anoikis in osteosarcoma via PI3K/Akt pathway. *Cancer Cell Int* 2017;17:1.
 27. Zhang P, Song Y, Sun Y, et al. AMPK/GSK3 β / β -catenin cascade-triggered overexpression of CEMIP promotes migration and invasion in anoikis-resistant prostate cancer cells by enhancing metabolic reprogramming. *FASEB J* 2018;32:3924-35.
 28. Russo MA, Paolillo M, Sanchez-Hernandez Y, et al. A small-molecule RGD-integrin antagonist inhibits cell adhesion, cell migration and induces anoikis in glioblastoma cells. *Int J Oncol* 2013;42:83-92.
 29. Kim YJ, Choi WI, Jeon BN, et al. Stereospecific effects of ginsenoside 20-Rg3 inhibits TGF- β 1-induced epithelial-mesenchymal transition and suppresses lung cancer migration, invasion and anoikis resistance. *Toxicology* 2014;322:23-33.
 30. Schackmann RC, Klarenbeek S, Vlug EJ, et al. Loss of p120-catenin induces metastatic progression of breast cancer by inducing anoikis resistance and augmenting growth factor receptor signaling. *Cancer Res* 2013;73:4937-49.
 31. Du S, Yang Z, Lu X, et al. Anoikis resistant gastric cancer cells promote angiogenesis and peritoneal metastasis through C/EBP β -mediated PDGFB autocrine and paracrine signaling. *Oncogene* 2021;40:5764-79.
 32. Huang CP, Tsai YF, Lin YS, et al. Overexpression of multiple epidermal growth factor like domains 11 rescues anoikis survival through tumor cells-platelet interaction in triple negative breast Cancer cells. *Life Sci* 2022;299:120541.

Cite this article as: Li Q, Wang K, Shen Y, Lin C, Miao J, Hu X. Bioinformatics based exploration of hsa-miR-194-5p regulation of CHD4 through PI3K/AKT signal pathway to enhance tumor resistance to apoptosis due to loss of nests and participate in poor prognosis of oral squamous cell carcinoma. *Ann Transl Med* 2023;11(2):107. doi: 10.21037/atm-22-6332