NOTE: Please save this file locally before filling in the table, DO NOT work on the file within your internet browser as changes will not be saved. Adobe Acrobat Reader (available free here) is recommended for completion.

The ARRIVE guidelines 2.0: author checklist

The ARRIVE Essential 10

These items are the basic minimum to include in a manuscript. Without this information, readers and reviewers cannot assess the reliability of the findings.

Item		Recommendation	Section/line number, or reason for not reporting
Study design	1	For each experiment, provide brief details of study design including: a. The groups being compared, including control groups. If no control group has been used, the rationale should be stated. b. The experimental unit (e.g. a single animal, litter, or cage of animals).	Methods/page 6,line 125-129. a single rat is regarded as an experimental unit.
Sample size	2	a. Specify the exact number of experimental units allocated to each group, and the total number in each experiment. Also indicate the total number of animals used. b. Explain how the sample size was decided. Provide details of any *a priori* sample size calculation, if done.	Methods/page 6,line 123-129. et in a natural plateau environment. Therefore, the original purpose is to collect basic evidence on the changes in the structure and function of the right ventricle in rats in the plateau natural environment.
Inclusion and exclusion criteria	3	a. Describe any criteria used for including and excluding animals (or experimental units) during the experiment, and data points during the analysis. Specify if these criteria were established *a priori.* If no criteria were set, state this explicitly. b. For each experimental group, report any animals, experimental units or data points not included in the analysis and explain why. If there were no exclusions, state so. c. For each analysis, report the exact value of *n* in each experimental group.	Inclusion criteria: male SD rats, rats raised in different environments to 18 weeks of age. Exclusion criteria: death during feeding, unqualified image quality after cardiac MRI scan. there were no exclusions Methods/page 6,line 125-129.
Randomisation	4	a. State whether randomisation was used to allocate experimental units to control and treatment groups. If done, provide the method used to generate the randomisation sequence. b. Describe the strategy used to minimise potential confounders such as the order of treatments and measurements, or animal/cage location. If confounders were not controlled, state this explicitly.	Random numbers were generated using the standard = RAND() function in Microsoft Excel. Methods/page 6,line 129-131.
Blinding	5	Describe who was aware of the group allocation at the different stages of the experiment (during the allocation, the conduct of the experiment, the outcome assessment, and the data analysis).	Group personnel do not participate in data measurement.
Outcome measures	6	a. Clearly define all outcome measures assessed (e.g. cell death, molecular markers, or behavioural changes). b. For hypothesis-testing studies, specify the primary outcome measure, i.e. the outcome measure that was used to determine the sample size.	Methods/page 7,line 160-162. This experiment is a cohort controlled study, and hypothesis testing study is not applicable
Statistical methods	7	a. Provide details of the statistical methods used for each analysis, including software used. b. Describe any methods used to assess whether the data met the assumptions of the statistical approach, and what was done if the assumptions were not met.	Methods/page 8,line 182-187. Model assumptions were checked using the Shapiro-Wilk normality test and Levene's Test for homogeneity of variance.
Experimental animals	8	a. Provide species-appropriate details of the animals used, including species, strain and substrain, sex, age or developmental stage, and, if relevant, weight. b. Provide further relevant information on the provenance of animals, health/immune status, genetic modification status, genotype, and any previous procedures.	Methods/page 6, line 123-124. Methods/page 6, line 123-124.
Experimental procedures	9	For each experimental group, including controls, describe the procedures in enough detail to allow others to replicate them, including: a. What was done, how it was done and what was used. b. When and how often. c. Where (including detail of any acclimatisation periods). d. Why (provide rationale for procedures).	Methods/page 6,line 122-182. Methods/page 6,line 123-129. Methods/page 6,line 123-129. concentration, and better stability It is an internationally recognized method of anesthesia with many advantages such as small effects on the physiological indicators and vital signs of animals.
Results	10	For each experiment conducted, including independent replications, report: a. Summary/descriptive statistics for each experimental group, with a measure of variability where applicable (e.g. mean and SD, or median and range). b. If applicable, the effect size with a confidence interval.	Results:table 1 and table 2. The size of the confidence interval is 95%.

The Recommended Set

These items complement the Essential 10 and add important context to the study. Reporting the items in both sets represents best practice.

Item		Recommendation	Section/line number, or reason for not reporting
Abstract	11	Provide an accurate summary of the research objectives, animal species, strain and sex, key methods, principal findings, and study conclusions.	ıar function could not be fully recovered.
Background	12	a. Include sufficient scientific background to understand the rationale and context for the study, and explain the experimental approach. b. Explain how the animal species and model used address the scientific objectives and, where appropriate, the relevance to human biology.	Introduction/page 3, line 74-77 and page 4,line 78-120. Advantages and limitations/page 17, line 377-382.
Objectives	13	Clearly describe the research question, research objectives and, where appropriate, specific hypotheses being tested.	ı after returning to normoxic environment.
Ethical statement	14	Provide the name of the ethical review committee or equivalent that has approved the use of animals in this study, and any relevant licence or protocol numbers (if applicable). If ethical approval was not sought or granted, provide a justification.	Footnote/Ethical statement
Housing and husbandry	15	Provide details of housing and husbandry conditions, including any environmental enrichment.	Methods/page 6,line 130-133.
Animal care and monitoring	16	a. Describe any interventions or steps taken in the experimental protocols to reduce pain, suffering and distress. b. Report any expected or unexpected adverse events. c. Describe the humane endpoints established for the study, the signs that were monitored and the frequency of monitoring. If the study did not have humane endpoints, state this.	The rats were placed in a sealed glass container and anesthetized with 2-3% isoflurane premixed in oxygen on a MATRXVIP 3000 small animal anesthesia machine (Midmark Co.,USA).Methods/page 6,line 138-140. In the high altitude group, two rats are found to have inflammation in their lungs during the CMR scan. However, no adverse events are observed during the study period. The research team and veterinary staff monitor the animals twice a day. The health of the rats was monitored according to their body weight (twice a week), food and water intake, activity and fur condition, and the litter was changed twice a week
Interpretation/ scientific implications	17	a. Interpret the results, taking into account the study objectives and hypotheses, current theory and other relevant studies in the literature. b. Comment on the study limitations including potential sources of bias, limitations of the animal model, and imprecision associated with the results.	Discussion/page 11-17,line 249-366. Advantages and limitations/page 17,line 383-385 and .page 18,line 386-387.
Generalisability/ translation	18	Comment on whether, and how, the findings of this study are likely to generalise to other species or experimental conditions, including any relevance to human biology (where appropriate).	The results of this research have not yet been promoted, but they can provide references for related animal experiments and research on plateau heart disease in the future.
Protocol registration	19	Provide a statement indicating whether a protocol (including the research question, key design features, and analysis plan) was prepared before the study, and if and where this protocol was registered.	A protocol was prepared before the study without registration.
Data access	20	Provide a statement describing if and where study data are available.	Data Sharing Statement
Declaration of interests	21	a. Declare any potential conflicts of interest, including financial and non-financial. If none exist, this should be stated. b. List all funding sources (including grant identifier) and the role of the funder(s) in the design, analysis and reporting of the study.	The authors declare that they have no competing interests in this section. nd Technology Basic Research Project (NO. 2021-ZJ-732) and Qinghai province "Kunlun Elite High-end Innovative and Entrepreneurial Talents" program to cultivate leading talents. Project No. : Youth Talent Word (2021) No. 13.

Article information: https://dx.doi.org/10.21037/atm-22-5991

*As the checklist was provided upon initial submission, the page number/line number reported may be changed due to copyediting and may not be referable in the published version. In this case, the section may be used as an alternative reference.