



Effects of notoginseng leaf triterpenes on small molecule metabolism after cerebral ischemia/reperfusion injury assessed using MALDI-MS imaging

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Background: Notoginseng leaf triterpenes (PNGL) is believed to have neuroprotective effects via the inhibition of inflammatory response and neuronal apoptosis. However, its mechanisms underlying the anti-ischemia/reperfusion (I/R) injury effects on the regulation of small molecule metabolism in rat brain remains unclear. The purpose of this study was thus to explore the mechanisms of PNGL on the regulation of small molecule metabolism in rat brain after I/R injury using matrix-assisted laser desorption ionization-mass spectrometry imaging (MALDI-MSI).

Methods: As a model of *in vivo* cerebral I/R injury, male Sprague-Dawley (SD) rats were established with a middle cerebral artery occlusion/reperfusion (MCAO/R) model after PNGL administration with 40 mg·kg⁻¹ through intraperitoneal injection (i.p.) for 7 days. We assessed the neurological behavior, regional cerebral blood flow (r CBF), neuron injury, and spatial distribution of metabolic small molecules.

Results: Our *in vivo* results suggested that PNGL increased cerebral blood flow and relieved neurological dysfunction. Furthermore, using MALDI-MSI, we demonstrated that PNGL regulated 16 endogenous small molecules implicated in metabolic networks including tricarboxylic acid (TCA) cycle, adenosine triphosphate (ATP) metabolism, malate-aspartate shuttle, metal ions, and antioxidants underwent noticeable changes after reperfusion for 24 h.

Conclusions: PNGL is a novel cerebrovascular agent that can improve cerebral blood flow and attenuate adverse neurological disorders. The mechanisms are closely correlated with relative metabolic pathways, which offers insight into exploring new mechanisms in PNGL for the treatment of cerebral I/R injury.

Keywords: Cerebral I/R injury; notoginseng leaf triterpenes (PNGL); matrix-assisted laser desorption ionization-mass spectrometry imaging (MALDI-MSI); small molecule metabolism

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Introduction

Stroke is a complex disease that affects different brain regions, leading to disruption of abundant neural circuits, and it involves complicated injury cascade responses (1). Of all stroke events, ischemic stroke (caused by decreased blood flow to the brain) accounts for approximately 87% in nature (2). Ischemic stroke causes a complex pathophysiological response including destruction of the neurovascular unit, neuronal death and inflammation (3,4). Rapid supply of cerebral blood flow is the primary target of ischemic stroke treatment and the prerequisite for neuroprotective therapies (5). Except for tissue plasminogen activator (tPA), few pharmacological agents have succeeded in clinical trials. Therefore, the search for promising neuroprotective compounds to restore cerebral blood flow and improve neurological deficits after ischemic stroke to overcome the clinical therapeutic limitations remains an urgent issue.

Currently, matrix-assisted laser desorption/ionization mass spectrometry imaging (MALDI-MSI) is an innovative methodology that enables label-free localization of molecular species in tissue sections mounted on electrically conductive slides (6). By means of unique advantages of label free, molecular specificity, high-throughput analysis, simple sample preparation, and in situ positioning (7), MALDI-MSI plays an irreplaceable role for measuring the spatial distribution of small molecules (8,9), metabolites (10), proteins (11), peptides (12), lipids (13), or drugs (14). Furthermore, recent improvements in MALDI-MSI have been shown to be useful for different kinds of cerebrovascular disease (8,15).

Panax notoginseng, is a traditional Chinese medicinal plant that has a long history of use in China for preventing and treating ischemic cerebrovascular diseases (16). The current research on *Panax notoginseng* mainly focuses on the roots of *Panax notoginseng*. However, the stem and leaf parts of *Panax notoginseng* are often ignored. Notoginseng leaf triterpenes (PNGL) is the total saponins extracted from the stem and leaf of *Panax notoginseng*, which has the functions of regulating blood lipid, sedative and hypnotic, analgesic, anti-inflammatory, and anti-aging (17–20). More importantly, our previous studies have mainly reported the neuroprotective effect of PNGL after ischemic stroke in the inhibition of inflammation and apoptosis (21). However, the mechanism of PNGL on brain metabolic small molecules and their spatial distributions remain unsystematic explored.

The present study is the first to investigate the protective effects of PNGL on cerebral I/R injury correlated with

cerebral blood flow, neurological scores, and histomorphology. MALDI-MSI was used to further explore the anti-cerebral I/R injury effects of PNGL on the metabolism of endogenous substances in relation to glucose metabolism, ATP metabolism, antioxidant molecules, and metal ions. Additionally, chemical fingerprinting data have revealed that the content of total saponins of *Panax notoginseng* stems and leaves are mainly protopanaxadiol-type saponins, and contains almost no protopanaxatriol-type saponins (21). As shown in Figure S1A,B, monomeric saponins contained in protopanaxadiol-type saponins mainly include ginsenoside Rb1, ginsenoside Rc, ginsenoside Rb2, and ginsenoside Rb3. In addition, eleven batches of PNGL samples were detected by the fingerprinting assay (Figure S2).

We present the following article in accordance with the ARRIVE reporting checklist (available at <http://dx.doi.org/10.21037/atm-20-4898>).

Methods

Drugs

PNGL was supplied by Jilin Academy of Chinese Medicine (Jilin, China). A positive drug dl-3-n-butylphthalide (NBP) was purchased from CSPC-NBP Pharmaceutical Co., Ltd (Hebei, China). Before study, they were freshly dissolved in 0.9% normal saline.

Animals

Male Sprague-Dawley (SD) rats (purchased from Beijing Vital River Laboratories, Beijing, China) weighing 250–280 g were used in this study. The study was approved by the Institutional Animal Care and Use Committee of Chinese Academy of Medical Sciences and Peking Union Medical College (SYXK 2017-0020). All rats care and experimental procedures were reported in accordance with the Laboratory Animal Ethics Committee of the Institute of Medicinal Plant Development, Peking Union Medical College and complied with NIH Guidelines for the Care and Use of Laboratory Animals. All rats were randomized into four experimental groups, as followed: sham group, MCAO/R model group, PNGL (40 mg·kg⁻¹) group, and NBP (20 mg·kg⁻¹) group. For drug administration, PNGL, NBP or 0.9% normal saline was exposed to continuous intraperitoneal injection (once per day) for 7 d prior to MCAO surgery. At 24 h after reperfusion, some brain tissues were fixed for histopathology and small molecule

metabolism detection.

Middle cerebral artery occlusion/reperfusion (MCAO/R) surgery and regional cerebral blood flow (r CBF) measurement

Male SD rats were anaesthetized with ketamine (80 mg·kg⁻¹) and xylazine (10 mg·kg⁻¹) via intraperitoneal injections. MCAO/R was proceeded by experimenters who were unaware of the grouping scheme, following a previously described method (22). After MCAO surgery, the wound was disinfected with iodine, and then the wound was sutured with sterile surgical suture to reduce the bleeding. We also injected tramadol (2.5 mg·kg⁻¹) by tail intravenous to relieve the pain caused by the operation. Reperfusion was induced for 24 h by removing the suture. Rats in the sham group were operated using the same method but with no occlusion in middle cerebral artery. The body temperature was maintained at 37±0.5 °C until rats woke up using a heating pad (Sunbeam, USA). The incision was sewed after ischemia and cerebral blood flow in the ischemic core region and peripheral region after reperfusion 24 h was assessed by the laser Doppler flowmeter (moor FLPI-2, UK).

Neurological score

Neurological performances of all animals were performed by two blinded investigators using a 5-point scale following a previously described method (22). The neurological function was scored according to a series of scales from 0 to 4. The highest score represents the most severe neurological deficits.

Histopathology staining

Histopathology staining (24 h postreperfusion) was implemented as reported previously (23). The brain samples were embedded in paraffin and coronally dissected into 5-µm-thick sections. Then paraffin sections were stained with hematoxylin-eosin (H&E) and Nissl staining to reveal histopathological lesions.

MSI

Frozen sections (10 µm) of the brain tissues were taken for MALDI-MSI using a Leica CM1950 cryostat (Leica Microsystems GmbH, Wetzlar, Germany) -20 °C and placed the thawed brain tissue slices on electrically conductive glass slides coated with indium tin oxide (ITO). The MALDI

MSI experiments were implemented using the Autoflex Speed™ MALDI-TOF/TOF MS (Bruker Daltonics, Bremen, Germany) as described reported previously (8,24).

Data and statistical analysis

Experimental data are expressed as the means ± standard deviation (SD) and were statistically evaluated using SPSS17 software (IBM Corporation, New York, NY, USA). Differences between the groups were analyzed by one-way ANOVA and Tukey's post hoc test (*t*-test), and *P*<0.05 was considered statistically significant.

Results

PNGL increased r CBF and reduced neurologic deficits

As shown in *Figure 1A,B*, the r CBF in different groups was evaluated using a laser Doppler flowmeter system. The results revealed that the administration of PNGL and NBP for 7 days resulted in a significant increase in r CBF at 24 h after reperfusion versus the model group. Meanwhile, a visible amelioration in neurologic deficit was observed in PNGL administration for 7 days (*Figure 1C*). In addition, NBP did not exhibit significant differences on the neurological deficit scores compared with the PNGL.

PNGL inhibited neuronal loss

As shown in *Figure 1D,E*, H&E-stained slides of brain sections from each group were detected under a light microscope. Untreated rats have many neurons present in the pyknotic nuclei and weak staining, whereas MCAO/R rats have pale nuclei in the cerebral cortex regions. In contrast, administration of PNGL for 7 days prior to MCAO notably reduced the pyknotic nuclei in the cerebral cortex. Analogously, as shown in *Figure 1F,G*, Nissl staining from each group were also detected under a light microscope. MCAO/R rats have many neurons present in the shrunken phenotype and weak staining in the cerebral cortex. However, compared with the model group, PNGL administration for 7 days prior to MCAO exhibited strong staining in the cerebral cortex. Additionally, NBP did not exhibit significant differences on neuronal loss compared with the PNGL.

PNGL decreased the interruption of TCA cycle

The results of *Figure 2A,B* demonstrated that the MCAO/R

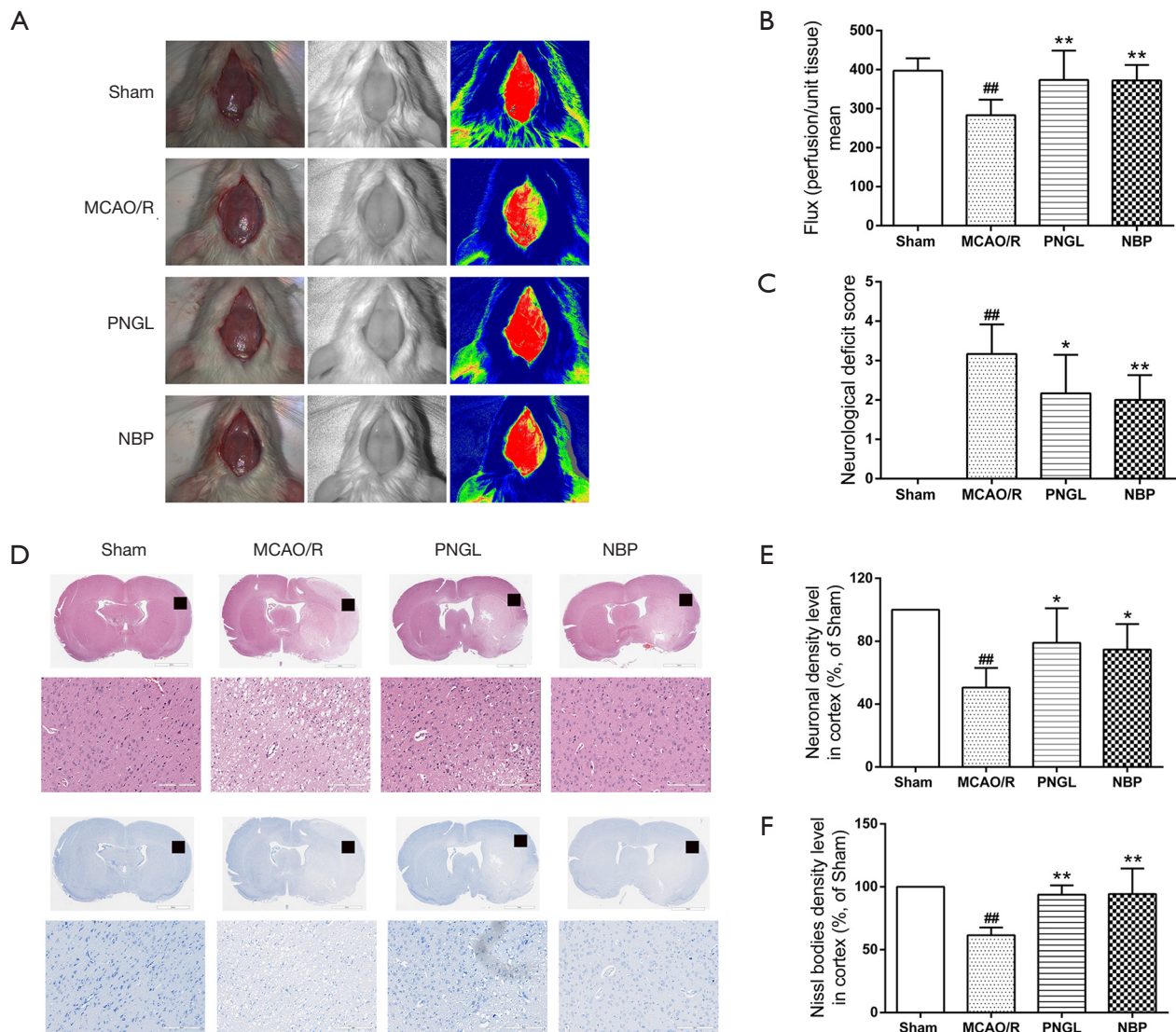


Figure 1 PNGL improves r CBF and decreases neuronal loss. (A) The images of r CBF in the cortex region. The magnitude of r CBF is expressed by different colors, and blue represents the most severe ischemia. (B) Flux mean value of r CBF in different groups, $n=5-6$ per group. (C) Neurological deficit scores are assessed in rats with MCAO 2 h then reperfusion 24 h, $n=6$ per group. (D) Pathological sections stained with H&E in the cortex region for each group. (E) Quantitative analysis in the cortex region in all groups of H&E staining, $n=4$ rats per group, scale bar =200 μm . (F) Pathological sections stained with Nissl in the cortex region for each group. (G) Quantitative analysis in the cortex region in all groups of Nissl staining, $n=4$ rats per group, scale bar =200 μm . Data are expressed as the mean \pm SD. ^{##} $P<0.01$ vs. sham group; ^{*} $P<0.05$, ^{**} $P<0.01$ vs. MCAO/R group. PNGL, notoginseng leaf triterpenes; r CBF, regional cerebral blood flow; MCAO, middle cerebral artery occlusion; H&E, hematoxylin-eosin.

R model group remarkably increased the content of glucose (Figure 2A) and citric acid (Figure 2B) in rats. However, PNGL can significantly decreased the content of glucose and citric acid in both the striatum and cerebral cortex. Moreover, NBP did not exhibit significant differences on the content of glucose and citric acid

compared with the PNGL.

PNGL increased the glutamine content and improved the malate-aspartate shuttle

MALDI-MSI maps of glutamine, aspartate and

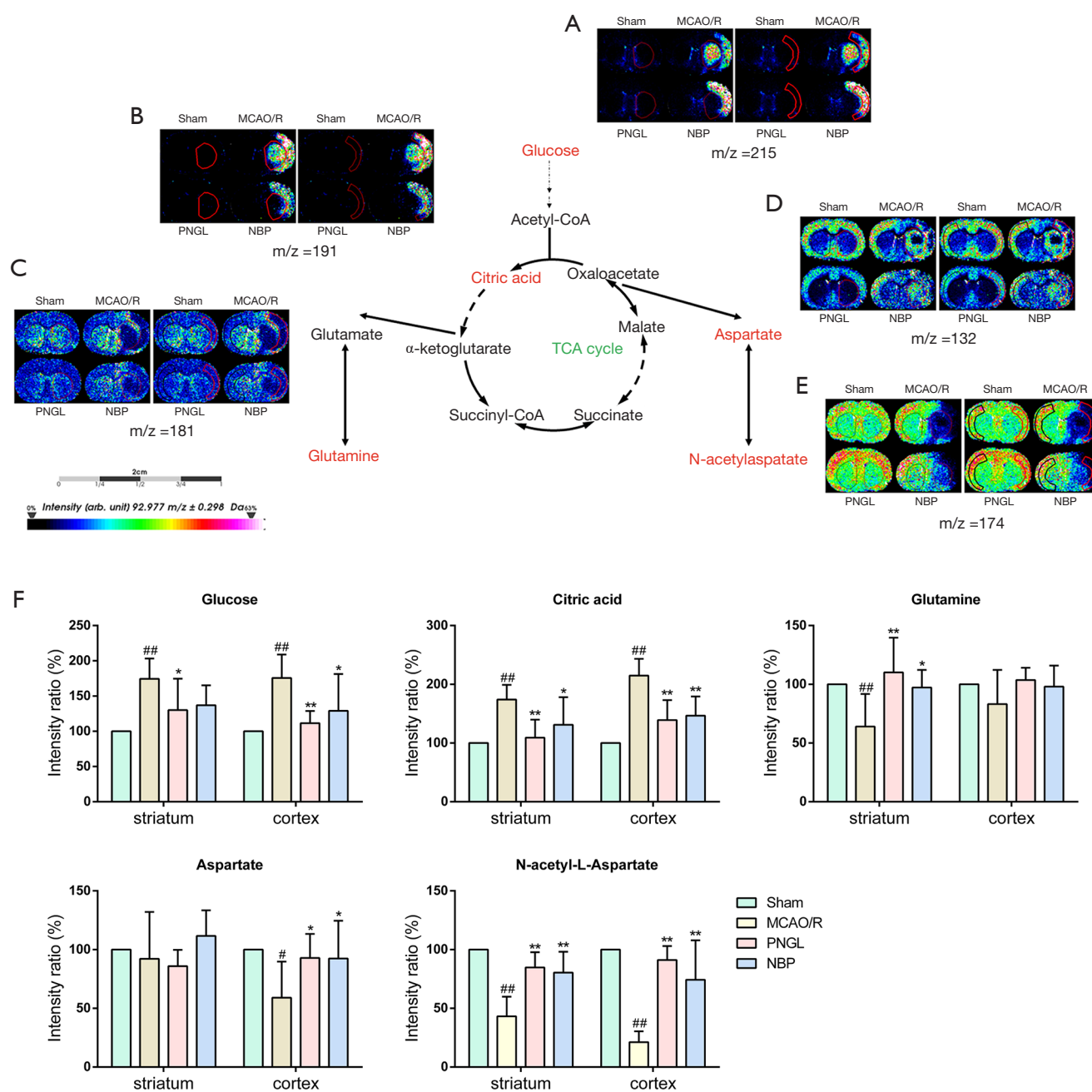


Figure 2 PNLG decreases the interruption of TCA cycle and improves the malate-aspartate shuttle. Ion image distributions of (A) glucose, (B) citric acid, (C) glutamine, (D) aspartate, and (E) N-acetylaspartate. (F) Quantitative data of glucose, citric acid, glutamine, aspartate and N-acetylaspartate in the striatum and cerebral cortex. Data are expressed as the mean \pm SD ($n=5$), scale bar = 2 cm. ^{##} $P < 0.01$ vs. sham group; ^{*} $P < 0.05$, ^{**} $P < 0.01$ vs. MCAO/R group. PNLG, notoginseng leaf triterpenes; MCAO/R, middle cerebral artery occlusion/reperfusion; MALDI MSI, matrix-assisted laser desorption ionization-mass spectrometry imaging.

N-acetylaspartate (NAA) were observed as shown in *Figure 2C,D,E*, respectively. The results revealed that the MCAO/R model group decreased the content of glutamine in the striatum region. However, the content of glutamine was increased after PNGL administration (*Figure 2C*). Our results also revealed that the aspartate content decreased in the ischemic region while PNGL obviously increased the aspartate level in the cerebral cortex (*Figure 2D*). The distribution of NAA shown in *Figure 2E* demonstrated that the content of NAA decreased in the ischemic region. PNGL administration for 7 days increased the level of NAA both in the striatum and cerebral cortex. However, the other regions in rat brain were not significantly changes in aspartate. In addition, NBP did not exhibit significant differences on the changes of glutamine, aspartate and NAA compared with the PNGL.

PNGL increased ATP metabolism

As shown in *Figure 3*, the MCAO/R rats showed low levels of GMP (A), AMP (B) and ADP (C) and the consequently decreased content of other related substances including adenosine (D) and inosine (E) in the ischemic region, whereas the MCAO/R rats showed high levels in the infarct area. However, we found that PNGL administration significantly increased the levels of AMP, ADP, adenosine, and inosine in both the striatum and cerebral cortex. The level of GMP in the cerebral cortex was relatively higher than that in the striatum. In addition, PNGL administration for 7 days can significantly decrease xanthine content in both the striatum and cerebral cortex region. Moreover, NBP did not exhibit significant differences on the changes of ATP-related metabolites compared with the PNGL.

PNGL increased the content of antioxidants

As shown in *Figure 4A,B,C*, MCAO/R model group decreased the levels of glutathione (A), ascorbic acid (AA) (B) and taurine (C) in the striatum and cerebral cortex regions. However, the amount of glutathione, AA and taurine were significantly increased in both the striatum and cerebral cortex after PNGL administration. Moreover, NBP did not exhibit significant differences on the content of antioxidants compared with the PNGL.

PNGL maintained the homeostasis of Na⁺ and K⁺

We also evaluated the content of Na⁺ and K⁺ using 1,

5-DAN hydrochloride as a matrix. *Figure 4* showed that MCAO/R model group increased Na⁺ level, whereas the level of K⁺ remarkably decreased both in the striatum and cerebral cortex regions compared with the sham group. However, PNGL administration for 7 days significantly reduced the Na⁺ level and increased K⁺ level.

Discussion

Insufficient supply of oxygen and glucose caused by middle cerebral artery occlusion causes sodium-potassium pump imbalance and leads to membrane depolarization, which is mainly reflected in the destruction of TCA cycle and the blocking of ATP synthesis and metabolism (7). Finally, neuronal death occurred due to excitotoxicity, mitochondrial dysfunction, immune response, and acidotoxicity caused by membrane depolarization (5).

PNGL is the total saponins extracted from the stems and leaves of *Panax notoginseng*. Many reports have demonstrated that PNGL exerts a neuroprotective effect in alleviating neurological disorders through the suppression of inflammation, apoptosis and autophagy (21,25). PNGL was found to counteract aberrant autophagy and apoptosis in hippocampal neurons of mice with cognitive impairment induced by sleep deprivation (25). Importantly, our previous study has shown that the neuroprotective effects of PNGL against cerebral I/R injury via inhibiting inflammatory response and neuronal apoptosis (21). In this work, we found that PNGL had a neuroprotective effect on cerebral I/R injury by improving the cerebral blood flow, neurological function, and the levels of small molecule metabolism during the acute phase after ischemic stroke.

Glucose is the most important and basic energy-supplying substance of various organisms, and the major energy source in the human brain and nervous system. Citric acid is the main mediator of the TCA cycle, and it has been reported to be concentrated in the striatum and cerebral cortex of ischemic areas (7). Our data showed that the contents of glucose and citric acid had significant increase in the right cerebral cortex and striatum compared with the sham group. Importantly, PNGL administration can significantly decreased this abnormal accumulation of glucose and citric acid in both the striatum and cerebral cortex.

Glutamine is mainly located in astrocytes and is essential for maintaining nitrogen homeostasis and ammonia detoxification in the brain. In addition, glutamine can also ensure the effective circulation of the neurotransmitters

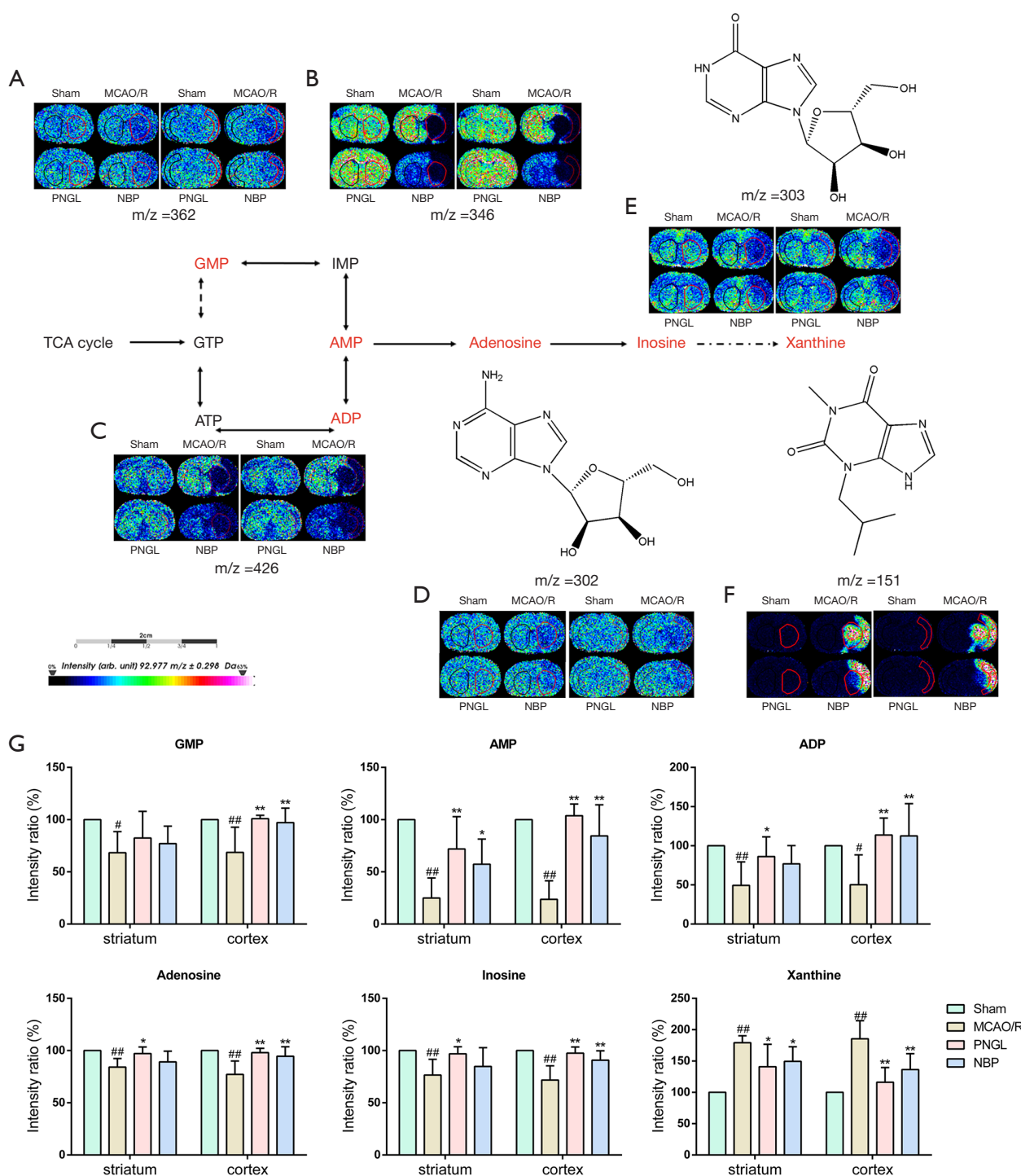


Figure 3 PNGL increases ATP metabolism. Ion image distributions of (A) GMP, (B) AMP, (C) ADP, (D) adenosine, (E) inosine, (F) xanthine. (G) Quantitative data of GMP, AMP, ADP, adenosine, inosine and xanthine in the striatum and cerebral cortex. Data are expressed as the mean ± SD (n=5), scale bar =2 cm. #P<0.05, ##P<0.01 vs. sham group; *P<0.05, **P<0.01 vs. MCAO/R group. PNGL, notoginseng leaf triterpenes; MCAO/R, middle cerebral artery occlusion/reperfusion; MALDI MSI, matrix-assisted laser desorption ionization-mass spectrometry imaging; ATP, adenosine triphosphate; GMP, guanosine monophosphate; AMP, adenosine monophosphate; ADP, adenosine diphosphate.

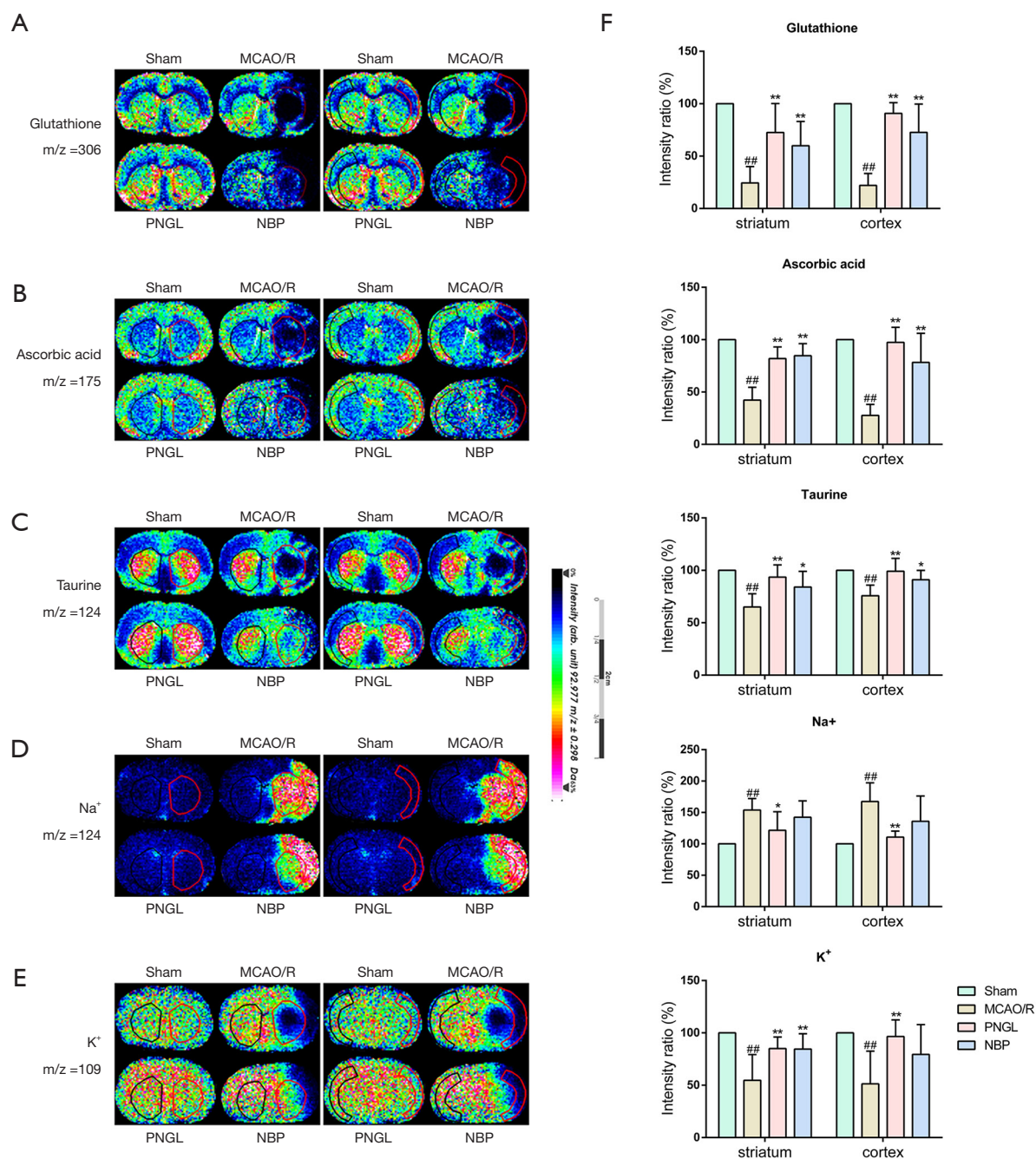


Figure 4 PNGL increases the content of antioxidants and maintains the homeostasis of Na⁺ and K⁺. Ion image distributions of (A) glutathione, (B) ascorbic acid, (C) taurine, (D) Na⁺, (E) K⁺. (F) Quantitative data of glutathione, ascorbic acid, taurine, Na⁺ and K⁺ in the striatum and cerebral cortex. Data are expressed as the mean ± SD (n=5), scale bar = 2 cm. ##P<0.01 vs. sham group; *P<0.05, **P<0.01 vs. MCAO/R group. PNGL, notoginseng leaf triterpenes; MCAO/R, middle cerebral artery occlusion/reperfusion; MALDI MSI, matrix-assisted laser desorption ionization-mass spectrometry imaging.

between neurons and astrocytes (26). Our results showed that the content of glutamine had significant reduction in the ischemic area, whereas the content of glutamine clearly increased after PNGL administration, especially in the striatum. In addition, L-aspartate (L-Asp) exchanges glutamate and protons through mitochondrial aspartic acid/glutamate carriers and plays an important role in the malate-aspartate shuttle (27). NAA has a higher concentration in normal brain tissue, but that was significantly reduced under pathological conditions such as neuronal damage (28). Our results also showed that the contents of aspartate and NAA decreased in the ischemic hemisphere. However, PNGL administration significantly increased the contents of aspartate and NAA.

Mitochondria generates ATP by oxidative phosphorylation to become a powerhouse for eukaryotic cells (29). ATP can be catabolized to ADP, AMP, adenosine, inosine, hypoxanthine, xanthine, and eventually to uric acid following two catabolic routes known as the “IMP pathway” and the “adenosine pathway” (30). The decreased cerebral blood flow restricted oxygen delivery in the MCAO/R-treated brain, which interrupted mitochondrial oxidative phosphorylation, resulting in metabolic disturbances of ATP synthesis (24). Previous studies have reported that levels of ATP, AMP, ADP, adenosine, hypoxanthine, and creatine were changed after MCAO challenge (8,31). In our study, we visualized the content changes and spatial distribution of 6 metabolites involved in ATP metabolism in MCAO/R-treated rats by MALDI-MSI, as presented in *Figure 3*. The MSI results also showed that PNGL obviously increased the ADP, GMP, AMP, adenosine, and inosine levels and consequently decreased the xanthine content in the ischemic hemisphere, suggesting that the disturbance of ATP catabolism was reversed by PNGL (7). These results indicated that the distribution and content of energy-related metabolites are affected by ischemia, mainly manifesting in the striatum and cerebral cortex.

ROS are mainly generated in the ischemic penumbra particularly after reperfusion (32), which triggers inflammatory responses, consequently increasing blood brain barrier (BBB) permeability and resulting in neuronal death (33). Glutathione (GSH), AA and taurine have the function of scavenging ROS and are considered to be an important antioxidants in the brain (15). As shown in *Figure 4A,B,C*, the content of GSH, AA, and taurine were remarkably declined in the ischemic regions, especially in the striatum and cerebral cortex. However, administration with PNGL increased the level of GSH, AA and taurine

both in the striatum and cerebral cortex.

The membrane potential necessary for neuronal activity depends on the formation of normal Na^+ and K^+ concentration gradients inside and outside the cell membrane. Focal ischemia leads to abnormal mitochondrial function, which induces ATP depletion and ultimately inhibits Na^+/K^+ -ATPase (34). Previous research has shown that neuronal necrosis after permanent occlusion of the middle cerebral artery are mainly due to the destruction of ion influx and efflux caused by the abnormal function of Na^+/K^+ -ATPase (35,36). In our research, *Figure 4D,E* shows MCAO/R injury increased the content of Na^+ , and reduced the content of K^+ in the striatum and cerebral cortex. These results are consistent with previous research reports, indicating that ion homeostasis was disturbed under ischemic conditions (7). Furthermore, PNGL increased the K^+ content and decreased the Na^+ content. One of the mechanisms is that PNGL restores the dysfunction of Na^+/K^+ -ATPase, which normally maintains the ion influx and efflux, and finally improves neuronal necrosis after cerebral artery occlusion.

Our results also demonstrated that brain region-specific differences are affected by PNGL in neurological function between small molecule metabolism isolated from the striatum and cerebral cortex of MCAO/R-treated rats. Thus, small molecule metabolism from the two brain regions can be regulated by different mechanisms and are therefore also likely to be differently affected by PNGL or ischemia reperfusion injury. We also showed functional differences between striatal and cortical small molecule metabolism. Regarding both ATP metabolism and antioxidants, the striatal content exhibited no difference compared with the cortical content. Meanwhile, the two brain regions compared here differed in glutamine and aspartate. Specifically, glutamine is distributed in the striatum region, and aspartate is distributed in the cerebral cortex. The obvious differences are most likely relevant to the differences in physiological function between the regions. The presented differences stress the importance of studying small molecule metabolisms from different brain regions separately, especially when exploring ischemic diseases that affect the striatum and/or the cerebral cortex.

In summary, using MALDI-MSI with assistance of 1,5-diaminonaphthalene hydrochloride to visualize small molecule metabolite systems will improve our understanding of the content and unique distribution in which small molecule metabolite act during cerebral ischemia. *Figure 5* shows the effects of PNGL on 16 altered

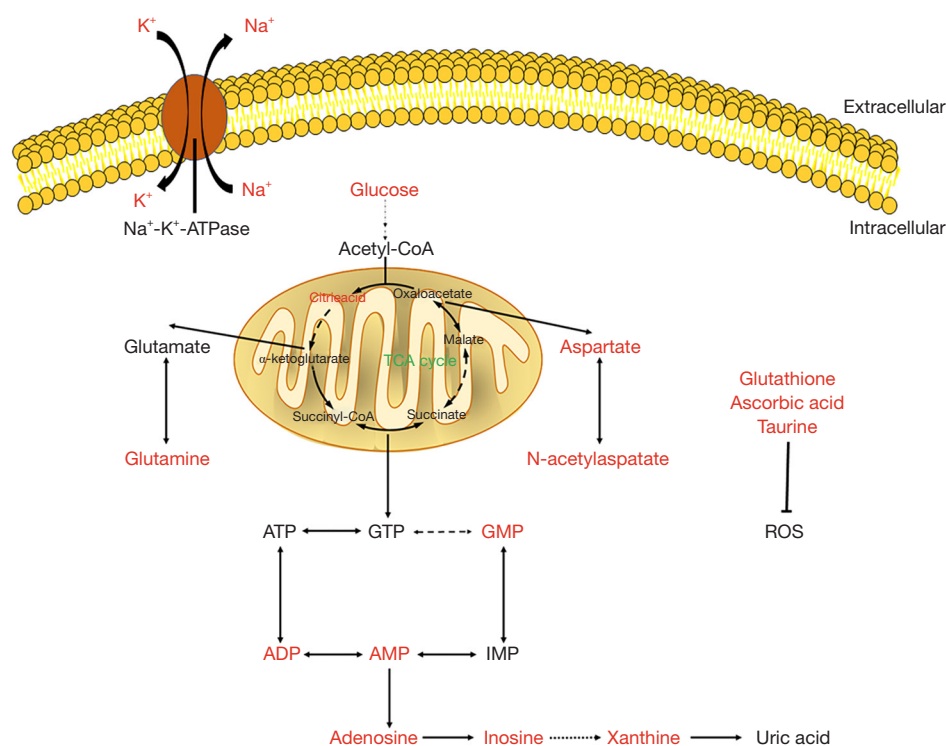


Figure 5 PNGL exerts a neuroprotective effect by regulating the abnormal small molecule metabolism in the early acute phase with MCAO rats, which is specifically reflected in the reduced abnormal accumulation of glucose and citric acid, increased ATP metabolism rate, augmented rate of the malate aspartate shuttle, increased antioxidants and maintained ionic homeostasis. PNGL, notoginseng leaf triterpenes; MCAO, middle cerebral artery occlusion; ATP, adenosine triphosphate.

endogenous small molecule metabolism. The results show that PNGL exerts a neuroprotective effect by reducing the activation of glial cells and rescuing the abnormal metabolism of small molecule in the early acute phase with MCAO rats, which is specifically reflected in the reduced abnormal accumulation of glucose and citric acid, increased ATP metabolism rate, augmented rate of the malate aspartate shuttle, increased antioxidants and maintained ionic homeostasis. Our approach offers critical insight into exploring new mechanisms for the PNGL treatment of cerebral ischemic/reperfusion injury.

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Footnote

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Data Sharing Statement: Available at <http://dx.doi.org/10.21037/atm-20-4898>

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uniform disclosure form (available at <http://dx.doi.org/10.21037/atm-20-4898>). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was approved by the Institutional Animal Care and Use Committee of Chinese Academy of Medical Sciences and Peking Union Medical College (SYXK 2017-0020). All rats care and experimental procedures were reported in accordance with the Laboratory Animal Ethics Committee of the Institute of Medicinal Plant Development, Peking Union Medical College and complied with NIH Guidelines for the Care and Use of Laboratory Animals.

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References

- George PM, Steinberg GK. Novel Stroke Therapeutics: Unraveling Stroke Pathophysiology and Its Impact on Clinical Treatments. *Neuron* 2015;87:297-309.
- Minnerup J, Sutherland BA, Buchan AM, et al. Neuroprotection for stroke: current status and future perspectives. *Int J Mol Sci* 2012;13:11753-72.
- Feigin VL, Forouzanfar MH, Krishnamurthi R, et al. Global and regional burden of stroke during 1990-2010: findings from the Global Burden of Disease Study 2010. *Lancet* 2014;383:245-54.
- Bennett DA, Krishnamurthi RV, Barker-Collo S, et al. The global burden of ischemic stroke: findings of the GBD 2010 study. *Glob Heart* 2014;9:107-12.
- Zhou Z, Lu J, Liu WW, et al. Advances in stroke pharmacology. *Pharmacol Ther* 2018;191:23-42.
- Caprioli RM, Farmer TB, Gile J. Molecular imaging of biological samples: localization of peptides and proteins using MALDI-TOF MS. *Anal Chem* 1997;69:4751-60.
- Liu H, Chen R, Wang J, et al. 1,5-Diaminonaphthalene hydrochloride assisted laser desorption/ionization mass spectrometry imaging of small molecules in tissues following focal cerebral ischemia. *Anal Chem* 2014;86:10114-21.
- Tian F, Liu R, Fan C, et al. Effects of Thymoquinone on Small-Molecule Metabolites in a Rat Model of Cerebral Ischemia Reperfusion Injury Assessed using MALDI-MSI. *Metabolites* 2020;10:27.
- Nilsson A, Fehniger TE, Gustavsson L, et al. Fine mapping the spatial distribution and concentration of unlabeled drugs within tissue micro-compartments using imaging mass spectrometry. *PLoS One* 2010;5:e11411.
- Xu J, Zhang Z, Liu R, et al. Function of complement factor H and imaging of small molecules by MALDI-MSI in a methamphetamine behavioral sensitization model. *Behav Brain Res* 2019;364:233-44.
- Chaurand P, Norris JL, Cornett DS, et al. New developments in profiling and imaging of proteins from tissue sections by MALDI mass spectrometry. *J Proteome Res* 2006;5:2889-900.
- Clemis EJ, Smith DS, Camenzind AG, et al. Quantitation of spatially-localized proteins in tissue samples using MALDI-MRM imaging. *Anal Chem* 2012;84:3514-22.
- Cornett DS, Reyzer ML, Chaurand P, et al. MALDI imaging mass spectrometry: molecular snapshots of biochemical systems. *Nat Methods* 2007;4:828-33.
- Oppenheimer SR, Wehr AY. Imaging mass spectrometry in drug discovery and development. *Bioanalysis* 2015;7:2609-10.
- Li W, Liu H, Jiang H, et al. (S)-Oxiracetam is the Active Ingredient in Oxiracetam that Alleviates the Cognitive Impairment Induced by Chronic Cerebral Hypoperfusion in Rats. *Sci Rep* 2017;7:10052.
- Leung AW, Mo ZX, Zheng YS. Reduction of cellular damage induced by cerebral ischemia in rats. *Neurochem Res* 1991;16:687-92.
- Yao CL, Pan HQ, Wang H, et al. Global profiling combined with predicted metabolites screening for discovery of natural compounds: Characterization of ginsenosides in the leaves of *Panax notoginseng* as a case study. *J Chromatogr A* 2018;1538:34-44.
- Fu JH, Li XZ, Shang XH, et al. Protective effects of saponins of stem and leaf of *Panax notoginseng* on acute myocardial ischemia in anaesthetic dogs. *Zhongguo Zhong Yao Za Zhi* 2006;31:62-5.
- Chen ZH, Wang DC, Li HL, et al. Hemodynamic effects of san chi (*Panax notoginseng*) root, leaf, flower and saponins on anesthetized dogs. *Yao Xue Xue Bao*

- 1983;18:818-22.
20. Liu XY, Wang S, Li CJ, et al. Dammarane-type saponins from the leaves of *Panax notoginseng* and their neuroprotective effects on damaged SH-SY5Y cells. *Phytochemistry* 2018;145:10-7.
 21. Xie W, Zhu T, Dong X, et al. HMGB1-triggered inflammation inhibition of notoginseng leaf triterpenes against cerebral ischemia and reperfusion injury via MAPK and NF- κ B signaling pathways. *Biomolecules* 2019;9:512.
 22. Meng X, Wang M, Wang X, et al. Suppression of NADPH oxidase- and mitochondrion-derived superoxide by Notoginsenoside R1 protects against cerebral ischemia-reperfusion injury through estrogen receptor-dependent activation of Akt/Nrf2 pathways. *Free Radic Res* 2014;48:823-38.
 23. Wang Z, Huang Y, Cheng Y, et al. Endoplasmic reticulum stress-induced neuronal inflammatory response and apoptosis likely plays a key role in the development of diabetic encephalopathy. *Oncotarget* 2016;7:78455-72.
 24. Liu RZ, Fan CX, Zhang ZL, et al. Effects of DL-3-n-butylphthalide on Cerebral Ischemia Infarction in Rat Model by Mass Spectrometry Imaging. *Int J Mol Sci* 2017;18:2451.
 25. Cao Y, Yang Y, Wu H, et al. Stem-leaf saponins from *Panax notoginseng* counteract aberrant autophagy and apoptosis in hippocampal neurons of mice with cognitive impairment induced by sleep deprivation. *J Ginseng Res* 2020;44:442-52.
 26. Jeitner TM, Battaile K, Cooper AJ. Critical Evaluation of the Changes in Glutamine Synthetase Activity in Models of Cerebral Stroke. *Neurochem Res* 2015;40:2544-56.
 27. Altinok O, Poggio JL, Stein DE, et al. Malate-aspartate shuttle promotes l-lactate oxidation in mitochondria. *J Cell Physiol* 2020;235:2569-81.
 28. Moffett JR, Ross B, Arun P, et al. N-Acetylaspartate in the CNS: from neurodiagnostics to neurobiology. *Prog Neurobiol* 2007;81:89-131.
 29. Dimroth P, von Ballmoos C. ATP synthesis by decarboxylation phosphorylation. *Results Probl Cell Differ* 2008;45:153-84.
 30. Barsotti C, Ipata PL. Metabolic regulation of ATP breakdown and of adenosine production in rat brain extracts. *Int J Biochem Cell Biol* 2004;36:2214-25.
 31. Folbergrova J, Zhao Q, Katsura K, et al. N-tert-butyl-alpha-phenylnitron improves recovery of brain energy state in rats following transient focal ischemia. *Proc Natl Acad Sci U S A* 1995;92:5057-61.
 32. Sherif IO, Al-Shaalan NH. Vildagliptin Attenuates Hepatic Ischemia/Reperfusion Injury via the TLR4/NF- κ B Signaling Pathway. *Oxid Med Cell Longev* 2018;2018:3509091.
 33. Szydlowska K, Tymianski M. Calcium, ischemia and excitotoxicity. *Cell Calcium* 2010;47:122-9.
 34. Cross HR, Radda GK, Clarke K. The role of Na⁺/K⁺ ATPase activity during low flow ischemia in preventing myocardial injury: a ³¹P, ²³Na and ⁸⁷Rb NMR spectroscopic study. *Magn Reson Med* 1995;34:673-85.
 35. Nagafuji T, Koide T, Takato M. Neurochemical correlates of selective neuronal loss following cerebral ischemia: role of decreased Na⁺,K⁺-ATPase activity. *Brain Res* 1992;571:265-71.
 36. Yang GY, Chen SF, Kinouchi H, et al. Edema, cation content, and ATPase activity after middle cerebral artery occlusion in rats. *Stroke* 1992;23:1331-6.

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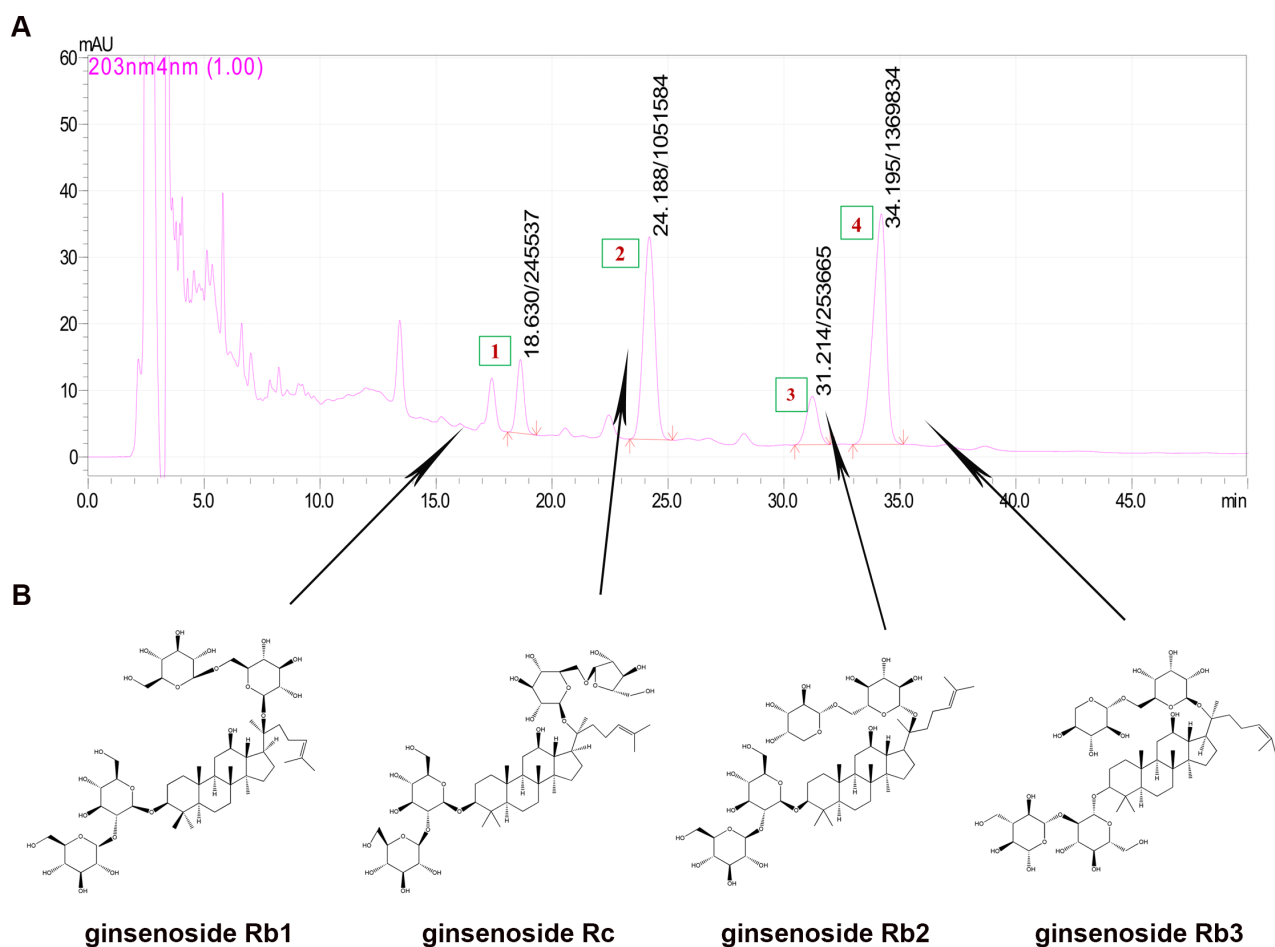


Figure S1 The chromatograms and chemical components of PNGL detected by HPLC. (A) The chromatograms of PNGL samples. (B) Representative chemical component structures, primarily contains ginsenoside Rb1, ginsenoside Rc, ginsenoside Rb2, ginsenoside Rb3. PNGL, notoginseng leaf triterpenes; HPLC, high-performance liquid chromatography.

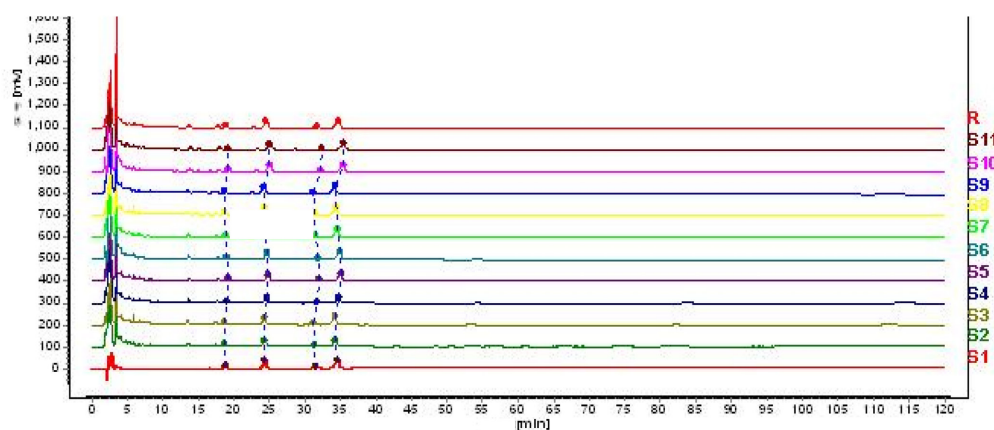


Figure S2 The chemical fingerprinting of PNGL detected by the HPLC. The chemical fingerprinting from eleven batches of PNGL samples. PNGL, notoginseng leaf triterpenes; HPLC, high-performance liquid chromatography.