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Blocking the 'MIDAS' touch of Enterococcus faecalis

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Introduction

The treatment and prevention of hospital-acquired infections represents a major challenge to patient care. It is estimated that approximately 1.7 million hospital-acquired infections occur annually in the United States alone, accounting for almost 100,000 deaths and \$5-10 billion in associated health care costs (1,2). Catheter-associated urinary tract infections (CAUTI) account for up to 40% of all hospital-acquired infections (3), and thus constitute the greatest burden of acquired disease in the nosocomial environment. The risk of bacteriuria in catheterized patients is estimated to be 5-10% per day (4), and most patients with an indwelling urinary catheter for 30 days or longer develop bacteriuria (5). Multidrug resistance (MDR) is now frequently associated with pathogens that cause CAUTI, leading to increased rates of treatment failure with standard antibiotic therapies and a rise in the use of second and third-line agents. A wide-range of Gram-positive and Gramnegative organisms can cause CAUTI, including Escherichia coli, Klebsiella pneumoniae, Proteus mirabilis, Staphylococcus spp. and Enterococcus spp. (6). Enterococcus spp., in particular Enterococcus faecalis (E. faecalis), cause up to 25% of all CAUTIs and are increasingly associated with MDR in the nosocomial setting (7-9). Here we review the exciting new findings reported by Flores-Mireles and co-workers, who have defined the mechanism of growth and biofilm formation by E. faecalis in human urine, and translated this new knowledge into the development of a vaccine that can prevent CAUTI in a mouse infection model (10).

The MIDAS motif of Ebp is important for adherence of *E. faecalis* to fibrinogen

E. faecalis produce a heteropolymeric extracellular hair-

like fimbrial structure called the endocarditis- and biofilmassociated pilus (Ebp) (11). Ebp is composed of three subunits; a major repeating subunit that comprises the bulk of the organelle (EbpC), a minor subunit that forms the base of the structure (EbpB) and a tip-located adhesin (EbpA). E. faecalis mutants that do not express the EbpA adhesin, or contain a mutation in the metal ion-dependent adhesion site (MIDAS) motif of EbpA, are attenuated in a mouse CAUTI model (12). Using a series of in vitro biofilm assays, Flores-Mireles et al. showed that the N-terminal domain of EbpA (EbpA^{NTD}), where the MIDAS motif is located, is important for mediating biofilm formation, which is dependent on adherence to the extracellular matrix (ECM) protein fibrinogen (10). Fibrinogen-dependent biofilm formation was also ablated following mutation of the EbpA MIDAS motif, demonstrating that this motif is critical for structural conformation of EbpA $^{\rm NTD}$ and coordinating E. faecalis adhesion to and colonization of surfaces (Figure 1).

Catheter implantation invokes fibrinogen release in the mouse bladder

What is the source of fibrinogen during infection, and how is this related to catheter implantation? Flores-Mireles *et al.* investigated the presence of fibrinogen in the lumen of the mouse bladder post-catheterization (10). Accumulation of fibrinogen was detected within 1 hour of catheterization, and increased in a time-dependent manner. By 24 hours, significant accumulation of fibrinogen was observed on the bladder epithelium and on the catheter surface, demonstrating that fibrinogen accumulation represents a major component of the inflammatory response to catheterization. Infection of catheterized mice revealed that

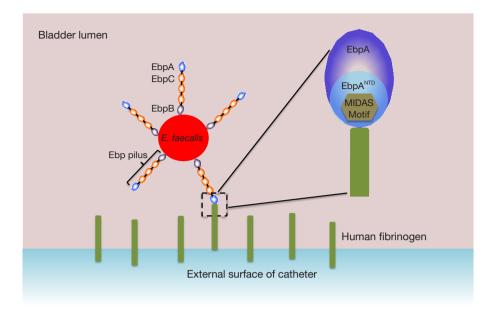


Figure 1 Schematic representation of *E. faecalis* endocarditis- and biofilm-associated pilus (Ebp) mediated binding to released and deposited fibrinogen on an implanted catheter surface in the mouse bladder. The E. faecalis Ebp is an extracellular organelle composed of the structural proteins EbpA, EbpB and EbpC. The N-terminal domain of the tip-located EbpA adhesin (EbpA^{NTD}) contains a MIDAS motif. The MIDAS motif is critical for the structural conformation of the EbpA^{NTD} and for mediating the interaction between EbpA^{NTD} and fibrinogen. *E. faecalis*, *Enterococcus faecalis*, MIDAS, metal ion-dependent adhesion site.

wild-type *E. faecalis*, but not mutants deficient in functional EbpA, adhered to the bladder epithelium and catheter surface. Thus, adherence to remodeled bladder and catheter surfaces coated with host-released fibrinogen via EbpA defines the molecular mechanism of biofilm formation by *E. faecalis*.

Fibrinogen: more than just a sticky surface

EbpA-mediated adherence of *E. faecalis* to fibrinogen only partly explains the ability of this pathogen to cause CAUTI. Intriguingly, *E. faecalis* did not grow in human urine collected from healthy individuals. However, supplementation of human urine with fibrinogen supported growth and the formation of catheter-associated biofilms. In the assays performed by Flores-Mireles *et al.*, an *E. faecalis* mutant containing a non-functional MIDAS motif in EbpA was unable to form a biofilm in fibrinogen-supplemented human urine (10). Thus, it appears that catheter implantation induces fibrinogen release, and this serves as both an essential growth nutrient and a receptor target for specific adhesion by *E. faecalis*. More broadly, urinary fibrin/fibrinogen degradation products were reported to be elevated in patients with chronic pyelonephritis (13) as well

as during UTI with frank hematuria in children (14). Thus, the role of fibrinogen and its breakdown products in forms of UTI beyond CAUTI may be underappreciated given its complex role in other infectious diseases. Fibrinogen may help to restrain bacterial growth by promoting clearance by host immune cells in some circumstances, but may be detrimental when excessive coagulation promotes bacterial adhesion, survival and dissemination (15). One would also assume that pathogens such as *E. faecalis* must secrete proteases that can efficiently break down fibrinogen in order to enable its uptake as a nutrient source.

Vaccination with EbpA can prevent CAUTI

The identification of a critical interaction between the EbpA^{NTD} and fibrinogen for *E. faecalis* adherence and biofilm formation suggests that blocking this interaction might represent a novel strategy for the prevention of CAUTI. Flores-Mireles *et al.* tested this hypothesis by vaccinating mice with EbpA and then challenging these mice with *E. faecalis* following catheterization (10). The EbpA vaccine was administered to mice intramuscularly with two booster doses at 4-week intervals. This induced a robust and long-lasting response, demonstrating the

immunogenicity of EbpA as a vaccine antigen. Vaccinated and control mice were then catheterized and challenged with *E. faecalis*. Excitingly, this strategy resulted in a 4-log reduction in bacterial burdens associated with both the bladder and catheter in the vaccinated mice. Immunization of mice with other Ebp components—i.e., EbpB and EbpC, or the C-terminal domain of EbpA did not provide protection from catheter-associated infection. Thus, vaccination with EbpA, specifically EbpA^{NTD}, can generate a protective immune response by targeting a disease defining EbpA-fibrinogen interaction. This represents a viable alternative strategy for the prevention of CAUTI.

Conclusions and outlook

The increasing incidence of CAUTI caused by multidrug resistant pathogens in the nosocomial setting represents a major challenge to patient health. E. faecalis is one of the most significant causes of device-related infections, and resistance to last-line antibiotics such as vancomycin by this pathogen is of major concern. The study by Flores-Mireles et al. utilized detailed molecular knowledge of a novel hostpathogen interaction, in this case between the EbpA adhesin N-terminal domain and fibrinogen, to develop a new antivirulence targeted vaccine (10). The work paves the way for more detailed molecular and structural analysis of this key interaction, and the identification of the specific metal ion(s) involved in the activation of the MIDAS motif for providing structural conformation to the $\operatorname{EbpA}^{\operatorname{NTD}}$. There are important parallels between their approach and other methods currently being developed to prevent UTIs caused by Gram-negative pathogens. Fimbriae-mediated adherence is critical to enable colonization of the urinary tract by all uropathogens. In the case of Gram-negative pathogens such as uropathogenic *E. coli*, vaccines targeting fimbrial adhesins (16-18), as well as novel therapeutics that block adhesinreceptor interactions (19-21), have been used successfully in animal models (22). These approaches offer a viable alternative to antibiotic treatment. The next step requires the assessment of efficacy in carefully designed clinical trials. If inhibitor molecules could be developed against the EbpA^{NTD} of *E. faecalis* in the same way that mannosides can target the FimH adhesin of uropathogenic E. coli, one could envision a whole new anti-virulence treatment and prevention program for CAUTI. It may also be possible to develop multivalent vaccines composed of a range of adhesins from different uropathogens. Finally, preventing the induction of fibrinogen release, or even other ECM

proteins if relevant, may also represent a strategy to limit bacterial growth and adherence in the catheterized urinary tract.

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