Intra-abdominal infection in acute pancreatitis in eastern China: microbiological features and a prediction model

Cheng Zhu^{1#}, Sheng Zhang^{2#}, Han Zhong³, Zhichun Gu³, Yuening Kang⁴, Chun Pan⁵, Zhijun Xu⁶, Erzhen Chen¹, Yuetian Yu⁷, Qian Wang⁸, Enqiang Mao¹

¹Department of Emergency, Ruijin Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China; ²Department of Critical Care Medicine, Ruijin Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China; ³Department of Pharmacy, Ren Ji Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China; ⁴Department of Rheumatology, Ren Ji Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China; ⁵Department of Critical Care Medicine, Zhongda Hospital, School of Medicine, Southeast University, Nanjing, China; ⁶Department of Critical Care Medicine, The Second Affiliated Hospital, Zhejiang University School of Medicine, Hangzhou, China; ⁷Department of Critical Care Medicine, Ren Ji Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China; ⁸Department of Emergency Internal Medicine, Shuguang Hospital Affiliated to Shanghai University of Traditional Chinese Medicine, Shanghai, China

Contributions: (I) Conception and design: C Zhu, Y Yu; (II) Administrative support: E Chen; (III) Provision of study materials or patients: C Pan, Y Yu, S Zhang; (IV) Collection and assembly of data: C Zhu, Y Yu, Z Xu; (V) Data analysis and interpretation: H Zhong, Y Yu, Z Gu; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

[#]These authors contributed equally to this work.

Correspondence to: Yuetian Yu. Department of Critical Care Medicine, Ren Ji Hospital, Shanghai Jiao Tong University School of Medicine, 145, Middle Shandong Road, Shanghai 200001, China. Email: fishyyt@sina.com; Enqiang Mao. Department of Emergency, Ruijin Hospital, Shanghai Jiao Tong University School of Medicine, 197, the Second Ruijin Road, Shanghai 200025, China. Email: maoeq@yeah.net; Qian Wang. Department of Emergency Internal Medicine, Shuguang Hospital Affiliated to Shanghai University of Traditional Chinese Medicine, 528, Zhangheng Road, Shanghai 201203, China. Email: docqianwang@126.com.

Background: This study aimed to investigate the microbiol distribution of intra-abdominal infection in patients with acute pancreatitis, and to develop a reliable prediction model to guide the use of antibiotics.

Methods: Inpatient with acute pancreatitis between January 2015 and June 2020 were enrolled in the study. Participants were divided into the intra-abdominal infection group and non-infection group. Isolated pathogens and antibiotic susceptibility were documented. Characteristics parameters, laboratory results, and outcomes were also compared. Least absolute shrinkage and selection operator (LASSO) regression model was used to select the risk factors associated with intra-abdominal infection in patients with acute pancreatitis. Logistic regression analysis, random forest model, and artificial neural network were also used to validate the performance of the selected predictors in intra-abdominal infection prediction. A novel nomogram based on selected predictors was established to provide individualized risk of developing intra-abdominal infection in patients with acute pancreatitis.

Results: A total amount of 711 participants were enrolled in the study, and of these, 182 (25.6%) had intra-abdominal infection. Of the 247 isolated pathogens, 45 (18.2%) were multidrug-resistant bacteria, and antibiotic susceptibility was lower than that of China Antimicrobial Surveillance Network 2020. The LASSO method identified 5 independent predictors [intra-abdominal pressure (IAP), acute physiology and chronic health evaluation II (APACHE II), computed tomography severity index (CTSI), the severity of pancreatitis, and intensive care unit (ICU) admission] of intra-abdominal infection, which were validated by three different models. The area under the curve was >0.95 for all 5 predictors. A clinically useful nomogram based on these predictors was successfully established.

Conclusions: Multidrug-resistant bacteria were quite common in intra-abdominal infection. IAP, APACHE II, CTSI, the severity of pancreatitis, and ICU admission were identified as risk factors and the new nomogram based on these could help clinicians estimate the risk of intra-abdominal infection and optimize antimicrobial prescription for acute pancreatitis patients.

Keywords: Acute pancreatitis; intra-abdominal infection; microbiology; prediction model

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Introduction

Acute pancreatitis is an inflammatory condition of the pancreas that is also known to involve the peri-pancreatic tissues (1). It has various underlying etiologies, such as acute cholecystitis, hyperlipidemia, and excessive alcohol consumption (1). Secondary intra-abdominal infection of acute pancreatitis is one of the leading complications that has been of great concern in recent years. Epidemiological studies have shown that infection of the pancreas and peripancreatic necrosis occurs in about 20-40% of patients with severe acute pancreatitis, which can cause life-threatening organ dysfunction and lead to poor prognosis (2). In a meta-analysis of 6,970 patients, it was indicated that the mortality rate could be as high as 35.2% in acute pancreatitis with intraabdominal infection and organ failure (3). Therefore, the early identification of these infectious diseases is a major challenge in managing patients with acute pancreatitis, especially in severe cases.

Prophylactic antibiotics in acute pancreatitis was once believed to reduce the incidence of infection-related complications (4,5). However, the effects of treatment are still controversial due to the heterogeneity of enrolled patients in clinical trials and different antimicrobial applications (6). The abuse of antibiotics will lead to gut microflora dysbiosis and increased medical expenses (7). Therefore, it is important to identify those at high risk of developing an infection and to recognize microbiological features after infection onset. It was reported that some of the host factors like diabetes and gallstones were once recognized as the risk factors for secondary intra-abdominal infection (1,2). However, no comprehensive systematic analysis with large sample size has been performed and the valuable information provided to clinical practice is limited. Thus, the present study was performed to establish a reliable clinical prediction model by least absolute shrinkage and selection operator (LASSO) regression and was further validated in the logistic regression model, random forest model, and artificial neural network model, respectively. With the valuable nomogram established, we hope it could optimize the antimicrobial prescription for

acute pancreatitis patients with suspected intra-abdominal infection.

We present the following article in accordance with the TRIPOD reporting checklist (available at http://dx.doi. org/10.21037/atm-21-399).

Methods

The retrospective clinical study was conducted at 6 pancreatitis diagnosis and treatment centers in eastern China (Ruijin Hospital, Ren Ji Hospital, Zhongda Hospital, Sir Run Run Shaw Hospital, The Second Zhejiang Hospital, and Shu Guang Hospital). The analyzed data were obtained from electronic medical records of each hospital between January 2015 and June 2020 by 6 attending physicians. The Ethics Committee of Shanghai Jiaotong University School of Medicine approved the study (No. 2019-RES-082), and the study was conducted in accordance with the declaration of Helsinki (as revised in 2013). Written informed consent was obtained from all patients.

Study population

Inpatients who were diagnosed with acute pancreatitis and aged 18–80 years were included in the study. The exclusion criteria were as follows: (I) was diagnosed with an extraabdominal infectious disease during hospital stay; (II) was pregnancy; and (III) had incomplete medical history.

Extra-abdominal infectious diseases include bloodstream infections, pneumonia (hospital-acquired pneumonia or ventilator-acquired pneumonia), urinary tract infections, and skin and soft tissue infections. All diagnostic criteria of the infection were based on the practice guideline of the European Society of Clinical Microbiology and Infectious Diseases (8).

Disease definitions

Based on the guidelines of the World Society of Emergency Surgery 2019 (1), acute pancreatitis can be diagnosed by any 2 of the following 3 criteria: (I) acute epigastric pain, which

can radiate to the back or waist; (II) the level of amylase and/or lipase in blood samples is at least 3 times higher than the normal limit; and (III) enhanced computed tomography (CT) or magnetic resonance imaging (MRI) was performed, and typical pancreatic lesions can be detected.

Secondary intra-abdominal infection is suspected in patients with acute pancreatitis when the 1 of the following criteria is met: (I) new-onset fever (body temperature >38.5 °C) or persistent fever; (II) the level of inflammatory biomarkers [leukocyte count, neutrophil count, procalcitonin (PCT), and C-reactive protein (CRP)] is increased; and (III) clinical symptoms of continuous deterioration and organ dysfunction are detected. The diagnostic methods consist of the following: (I) gas configurations in the necrotic pancreatic tissue can be detected via enhanced CT (Figure S1); and (II) positive culture results are acquired from the fine-needle aspiration (FNA) sample (1,2).

Identification of strains and antimicrobial susceptibility testing operation

CT-guided FNA was performed to obtain drainage fluid from peri-pancreatic tissues (Figure S2). To identify pathogens, samples were gram and fluorescence stained and were also amplified by bacteria culture. Identification of pathogens was also conducted by the VITEK-2 compact automated microbiology system (BioMerienx, France, Version 1.7), and the results of the antimicrobial susceptibility testing were determined based on the standards of the Clinical and Laboratory Standards Institute (CLSI). Information on the pathogens that caused intraabdominal infection was obtained from each research center's microbiological database.

Only the first isolated pathogen from the drainage fluid samples was tested in our study. Multidrug-resistant (MDR) strain was defined as acquired non-susceptibility to at least 1 agent in 3 or more antimicrobial categories (9). China Antimicrobial Surveillance Network data were obtained from the internet (http://www.chinets.com) to compare antimicrobial susceptibility.

Clinical assessment and data collection

Clinical medical information of the hospitalized acute pancreatitis patients included in the present study was obtained from the electronic medical record management system of each hospital. The clinical characteristics of the enrolled patients were as follows: (I) basic information, including age, sex, comorbidity (hypertension, diabetes), and body mass index; (II) etiology of pancreatitis; (III) severity of the disease [Acute Physiology and Chronic Health Evaluation II (APACHE II) score, CT severity index (CTSI)]; (IV) intra-abdominal pressure (IAP); (V) biomarkers of inflammation (PCT, CRP, and white blood cell counts); (VI) change in coagulation index; and (VII) change in clinical biochemical index on days 1, 3, 7, and 14 and the prognosis (intra-abdominal infection, dead).

Statistical analysis

Categorical variables were presented as counts (n) and percentages (%), and compared using χ^2 -test or Fisher's exact test. Normally distributed variables were compared using Student's *t*-test, and were expressed as means (standard deviations), whereas non-normally distributed data was compared using Wilcoxon rank sum test and reported as medians (interquartile range). Kaplan-Meier plot and logrank test were used for the survival analysis.

To identify risk factors for intra-abdominal infection in patients with acute pancreatitis, we applied a series of artificial intelligence algorithms. The LASSO regression model with the "lambda.1se" criterion was used for the selection of variables. This analytic strategy enables us to exclude non-essential variables, retaining a subset of the most important variables for outcome prediction. The predictive performance of the selected variables was further validated in the logistic regression model, random forest model, and artificial neural network model, respectively. Model discrimination and calibration were assessed by receiver-operating characteristic (ROC) curves and calibration curves, respectively. In addition, ROC curve parameters, including sensitivity, specificity, correctly classified rate, positive likelihood ratio, and negative likelihood ratio, were also computed for each model. To reduce the risk of overfitting, the whole cohort was randomly divided into the training cohort and validation cohort with a ratio of 6:4. In each model, the training cohort was used for model development and the validation cohort was used for the unbiased assessment of model performance. Finally, a novel nomogram based on the selected predictors was established to provide individualized risk of developing intra-abdominal infection in patients with acute pancreatitis. Statistical analyses were performed using R (University of Auckland, New Zealand, Version 3.6.2), SPSS (IBM, USA, Version 21.0), and GraphPad Prism



Figure 1 Flow chart of the study.

(GraphPad Software, USA, Version 7.0). A 2-sided P value <0.05 was considered statistically significant (10,11).

Results

Study population

A total of 874 hospitalized patients who were diagnosed with acute pancreatitis were initially screened, and of these, 163 were excluded due to factors, such as extra-abdominal infection and pregnancy. Finally, 711 patients were included in the study and 182 (25.6%) of these were diagnosed with intra-abdominal infection (*Figure 1*).

Isolated bacteria and antimicrobial susceptibility

In total, 247 pathogens from 182 FNA samples were isolated. Of these, 146 (59.1%) were gram-negative

bacteria, 47 (19%) were gram-positive bacteria, while the other 32 were fungi. *Enterobacteriaceae* accounted for the main gram-negative bacteria. Forty-five (18.2%) pathogens were identified as MDR bacteria (*Table 1*). Antimicrobial susceptibility of the 4 main gram-negative bacteria (*Escherichia coli, Klebsiella pneumoniae, Pseudomonas aeruginosa*, and *Acinetobacter baumannii*) is shown in *Figure 2*, and a relatively high incidence of resistance to carbapenem and 3rd-generation cephalosporin was indicated. For gram-positive bacteria, vancomycin, linezolid, teicoplanin, and tigecycline were considered the best choices for antimicrobial susceptibility (close to 100%) (Figure S3).

Clinical features and outcomes of patients with or without intra-abdominal infection

The clinical features of patients with or without intraabdominal infection are listed in *Table 2*. It was indicated

 Table 1 Microbiology distribution of the 247 pathogens from 182

 patients with intra-abdominal infection

Pathogens	Isolated, n (%)
Total microorganisms	247 (100.0)
Gram-negative bacteria	146 (59.1)
Escherichia coli	47 (19.1)
Klebsiella pneumoniae	42 (17.1)
Pseudomonas aeruginosa	26 (10.5)
Acinetobacter baumannii	14 (5.7)
Enterobacter cloacae	5 (2.0)
Proteus mirabilis	5 (2.0)
Serratia marcescens	4 (1.6)
Other	3 (1.2)
Gram-positive bacteria	47 (19.0)
Enterococcus faecalis	15 (6.1)
Enterococcus faecium	12 (4.9)
Staphylococcus aureus	10 (4.0)
Coagulase-negative staphylococci	7 (2.8)
Other	3 (1.2)
Fungi	32 (12.9)
Candida albicans	22 (8.9)
Non-Candida albicans	10 (4.0)
Anaerobic bacteria	22 (9.0)
Bacteroides fragilis	17 (6.9)
Other	5 (2.0)
MDR bacteria	45 (100.0)
MDR Gram-negative bacteria	35 (77.7)
ESBL-producing Escherichia coli	12 (26.7)
ESBL-producing Klebsiella pneumoniae	10 (22.2)
MDR Pseudomonas aeruginosa	7 (15.6)
MDR Acinetobacter baumannii	6 (13.2)
MDR Gram-positive bacteria	10 (22.3)
Methicillin-resistant Staphylococcus aureus	4 (8.9)
Methicillin-resistant CoNS	4 (8.9)
Vancomycin-resistant Enterococci	2 (4.5)

CoNS, coagulase-negative staphylococci; ESBL, extended spectrum β -lactamase; MDR, multidrug resistant.

that more acute pancreatitis patients with intra-abdominal infection were admitted to intensive care unit (ICU). A high CTSI, APACHE II score, and IAP were found in the intraabdominal infection group, as well as more patients with diabetes and excessive alcohol consumption (*Table 2*). The laboratory results showed a higher serum PCT level and neutrophil count in patients with intra-abdominal infection (Tables S1-S4).

It was also noted that high mortality (9.9% vs. 0.8%) and long hospital stay (40.5 vs. 16 days) were associated with acute pancreatitis patients diagnosed with intra-abdominal infection (P<0.01) (*Figure 3*).

Identification and validation predictive factors for intraabdominal infection in patients with acute pancreatitis

Variable selection using the LASSO regression model

The whole cohort was randomly divided into the training cohort and validation cohort with a ratio of 6:4. The demographic and clinical data of patients in the training and validation groups are summarized in *Table 3* and Table S5. Most of the included variables and outcomes were well balanced between the two groups. Using the LASSO model with the "lambda.1se" criterion, 5 variables (intra-abdominal pressure, APACHE II score, CTSI, ICU admission, and severity grade) were retained for subsequent model construction and intra-abdominal infection prediction (*Figure 4*).

Development, validation, and assessment of models for predicting intra-abdominal infection

Three predictive models were built by incorporating the five selected variables. The logistic regression model (*Figure 5*), random forest model (*Figure 6*), and artificial neural network (*Figure 7*) were used. ROC curve parameters at the optimal cut-off point according to different models were compared (*Table 4*). It was indicated that all of the areas under the curve were >0.95 and had high sensitivity and specificity in both the training set and validation set (all >93%). In addition, calibration curves indicated good agreement between predicted probability and observed probability for intra-abdominal infection in both the training set and validation set.

Estimating the probability of intra-abdominal infection using the nomogram

For the development of the nomogram, we incorporated the five selected predictors as prognostic features: intra-

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Figure 2 Antimicrobial susceptibility of the four main gram-negative bacteria. (A) *Escherichia coli*; (B) *Klebsiella pneumoniae*; (C) *Pseudomonas aeruginosa*; (D) *Acinetobacter baumannii*.

abdominal pressure, APACHE II score, CTSI, ICU admission, and severity grade. This novel 5 predictor-based nomogram can be utilized to predict individualized risk of intra-abdominal infection in patients with acute pancreatitis (*Figure 8*).

Discussion

Data of the 711 patients with acute pancreatitis were

analyzed in our study and indicated that gram-negative bacteria were still the main cause of intra-abdominal infection with high antimicrobial resistance trends. A new nomogram model that consisted of five risk factors (IAP, ICU admission, APACHE II score, CTSI, and severity of pancreatitis) was built and verified, and revealed a high predictive value.

Gut is the largest microecological reservoir of the human body which is mainly filled with *Enterococcus*, *Enterobacter*

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Table 2 Demographics and	clinical character	istics of patients	with acute pa	ncreatitis
() I				

Variables	Total (n=711)	Non-IAI group (n=529)	IAI group (n=182)	P value
Sex, n (%)				0.31
Female	242 (34.0)	174 (32.9)	68 (37.4)	
Male	469 (66.0)	355 (67.1)	114 (62.6)	
Age, median (IQR)	49 (37, 62)	50 (38, 62)	48 (37, 59)	0.3
Severity of pancreatitis, n (%)				<0.01
AP	294 (41.4)	291 (55)	3 (1.6)	
MSAP	188 (26.4)	179 (33.8)	9 (4.9)	
SAP	229 (32.2)	59 (11.2)	170 (93.4)	
Cause of pancreatitis, n (%)				<0.01
Hyperlipidemic pancreatitis	323 (45.4)	237 (44.8)	86 (47.3)	
Biliary pancreatitis	207 (29.1)	114 (21.6)	93 (51.1)	
Other	181 (25.5)	178 (33.6)	3 (1.6)	
Alcohol consumption, n (%)				<0.01
No	632 (88.9)	493 (93.2)	139 (76.4)	
Yes	79 (11.1)	36 (6.8)	43 (23.6)	
Smoking, n (%)				0.07
No	467 (65.7)	358 (67.7)	109 (59.9)	
Yes	244 (34.3)	171 (32.3)	73 (40.1)	
Diabetes, n (%)				<0.01
No	487 (68.5)	417 (78.8)	70 (38.5)	
Yes	224 (31.5)	112 (21.2)	112 (61.5)	
Hypertension, n (%)				0.75
No	511 (71.9)	378 (71.5)	133 (73.1)	
Yes	200 (28.1)	151 (28.5)	49 (26.9)	
Admission, n (%)				<0.01
Emergency Department	447 (62.9)	423 (80.0)	24 (13.2)	
ICU	264 (37.1)	106 (20.0)	158 (86.8)	
CTSI, median (IQR)	3.0 (4, 5)	4.0 (2, 4)	5.0 (5, 7)	<0.01
BMI, median (IQR)	25.1 (23.4, 26.8)	25.1 (23.4, 26.5)	25.2 (22.8, 27.6)	0.67
APACHE II, median (IQR)	7 (6, 8)	6 (5, 7)	10 (7, 13)	<0.01
Drainage, n (%)				<0.01
No	641 (90.2)	529 (100.0)	112 (61.5)	
Yes	70 (9.8)	0 (0)	70 (38.5)	
Operation, n (%)				<0.01
No	695 (97.7)	529 (100.0)	166 (91.2)	
Yes	16 (2.3)	0 (0)	16 (8.8)	

Table 2 (continued)

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Table 2 (continued)

Variables	Total (n=711)	Non-IAI group (n=529)	IAI group (n=182)	P value
CRRT, n (%)				<0.01
No	702 (98.7)	528 (99.8)	174 (95.6)	
Yes	9 (1.3)	1 (0.2)	8 (4.4)	
IAP, median (IQR)	14 (12, 18)	12 (11, 16)	19 (17, 22.8)	<0.01
Initiation of enteral nutrition, n (%)				<0.01
Day 0	10 (1.4)	5 (0.9)	5 (2.7)	
Day 1	12 (1.7)	11 (2.1)	1 (0.5)	
Day 2	81 (11.4)	77 (14.6)	4 (2.2)	
Day 3	330 (46.4)	284 (53.7)	46 (25.3)	
Day 4	213 (30.0)	122 (23.1)	91 (50.0)	
Day 5	57 (8.0)	29 (5.5)	28 (15.4)	
Day 6	8 (1.1)	1 (0.2)	7 (3.8)	
LOS, median (IQR)	20 (12, 31)	16 (10, 22)	40.5 (36, 51.8)	<0.01
Prognosis, n (%)				<0.01
Survival	689 (96.9)	525 (99.2)	164 (90.1)	
Non-survival	22 (3.1)	4 (0.8)	18 (9.9)	

AP, acute pancreatitis; APACHE II, Acute Physiology and Chronic Health Evaluation II; BMI, body mass index; CRRT, continuous renal replacement therapy; CTSI, computed tomography severity index; IAI, intra-abdominal infection; IAP, intra-abdominal pressure; ICU, intensive care unit; IQR, interquartile range; LOS, length of stay; MSAP, mild-severe acute pancreatitis; SAP, severe acute pancreatitis.



Figure 3 Ninety-day survival curves of hospitalized patients with acute pancreatitis (intra-abdominal infection *vs.* non intra-abdominal infection).

Table 3 Demographic and clinical characteristics of patients in training and validation groups

Variables	Total (n=711)	Validation group (n=294)	Train group (n=417)	P value
Sex, n (%)				0.02
Female	242 (34.0)	85 (28.9)	157 (37.6)	
Male	469 (66.0)	209 (71.1)	260 (62.4)	
Age, median (IQR)	49 (38, 62)	50 (37, 63)	49 (38, 60)	0.65
Severity of pancreatitis, n (%)				0.99
AP	294 (41.4)	122 (41.5)	172 (41.2)	
MSAP	188 (26.4)	78 (26.5)	110 (26.4)	
SAP	229 (32.2)	94 (32.0)	135 (32.4)	
Cause of pancreatitis, n (%)				0.38
Hyperlipidemic pancreatitis	323 (45.4)	125 (42.5)	198 (47.5)	
Biliary pancreatitis	207 (29.1)	88 (29.9)	119 (28.5)	
Other	181 (25.5)	81 (27.6)	100 (24)	
Alcohol consumption, n (%)				0.6
No	632 (88.9)	264 (89.8)	368 (88.2)	
Yes	79 (11.1)	30 (10.2)	49 (11.8)	
Smoking, n (%)				0.56
No	467 (65.7)	189 (64.3)	278 (66.7)	
Yes	244 (34.3)	105 (35.7)	139 (33.3)	
Diabetes, n (%)				0.98
No	487 (68.5)	202 (68.7)	285 (68.3)	
Yes	224 (31.5)	92 (31.3)	132 (31.7)	
Hypertension, n (%)				0.76
No	511 (71.9)	209 (71.1)	302 (72.4)	
Yes	200 (28.1)	85 (28.9)	115 (27.6)	
Admission, n (%)				0.29
Emergency Department	447 (62.9)	192 (65.3)	255 (61.2)	
ICU	264 (37.1)	102 (34.7)	162 (38.8)	
CTSI, median (IQR)	3 (4.0, 5.0)	3 (4.0, 5.0)	3 (4.0, 5.0)	0.33
BMI, median (IQR)	25.1 (23.4, 26.8)	25.3 (23.3, 27)	25.1 (23.4, 26.8)	0.71
APACHE II, median (IQR)	7 (6, 8)	7 (5, 8)	7 (6, 8)	0.33
Drainage, n (%)				0.38
No	641 (90.2)	269 (91.5)	372 (89.2)	
Yes	70 (9.8)	25 (8.5)	45 (10.8)	
Operation, n (%)				0.57
No	695 (97.7)	289 (98.3)	406 (97.4)	
Yes	16 (2.3)	5 (1.7)	11 (2.6)	

Table 3 (continued)

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Table 3 (continued)

Variables	Total (n=711)	Validation group (n=294)	Train group (n=417)	P value
CRRT, n (%)				1
No	702 (98.7)	290 (98.6)	412 (98.8)	
Yes	9 (1.3)	4 (1.4)	5 (1.2)	
IAP, median (IQR)	14 (12, 18)	14 (12, 18)	14 (12, 18)	0.89
Initiation of enteral nutrition, n (%)				0.52
Day 0	10 (1.4)	6 (2.0)	4 (1.0)	
Day 1	12 (1.7)	4 (1.4)	8 (1.9)	
Day 2	81 (11.4)	36 (12.2)	45 (10.8)	
Day 3	330 (46.4)	145 (49.3)	185 (44.4)	
Day 4	213 (30.0)	80 (27.2)	133 (31.9)	
Day 5	57 (8.0)	20 (6.8)	37 (8.9)	
Day 6	8 (1.1)	3 (1.0)	5 (1.2)	
LOS, median (IQR)	20 (12, 31)	20 (11, 30)	21 (12, 32)	0.47
IAI, n (%)				0.51
No	529 (74.4)	223 (75.9)	306 (73.4)	
Yes	182 (25.6)	71 (24.1)	111 (26.6)	
Prognosis, n (%)				1
Survival	689 (96.9)	285 (96.9)	404 (96.9)	
Non-survival	22 (3.1)	9 (3.1)	13 (3.1)	

AP, acute pancreatitis; APACHE II, Acute Physiology and Chronic Health Evaluation II; BMI, body mass index; CRRT, continuous renal replacement therapy; CTSI, computed tomography severity index; IAI, intra-abdominal infection; IAP, intra-abdominal pressure; ICU, intensive care unit; IQR, interquartile range; LOS, length of stay; MSAP, mild-severe acute pancreatitis; SAP, severe acute pancreatitis.

and *Bacteroides fragilis* (1,12). As an inflammatory disease, the permeability of intestinal mucosa could be increased by neutrophil-derived elastase, which could also destroy the tissue, increase the vascular permeability, and promotes the microecological imbalance. Thus, during the period of acute pancreatitis, the integrity of mucosa could also be destroyed. Therefore, bacteria translocation was easily recognized from the intestine to abdomen, and *Enterococcus* or *Enterobacter* could be isolated from the FNA samples (12). What's more, high concentrations of inflammatory mediators like IL-1 and IL-6 created during the cause of acute pancreatitis could lead to the obstruction of lymphatic reflux which is a "helping hand" to bacteria translocation (13).

Most abdominal infections secondary to acute pancreatitis should be classified as hospital-acquired infections. For patients with severe acute pancreatitis, especially for those who need to be admitted to the ICU, isolated pathogens were often MDR, which might relate to the previous antimicrobial exposure in the ICU (13,14). For gram-negative bacteria, the β -lactam/ β -lactamase inhibitor complex was a better choice than carbapenem. For gramnegative bacteria, vancomycin, linezolid, and teicoplanin were found to have high susceptibility.

The APACHE II score can be used to comprehensively evaluate the acute and chronic physiological status of patients (15). The acute physiology score includes 12 physiological measurements, such as body temperature and respiratory rate, that clearly reflects the inflammatory state (16). Chronic health evaluation can assess the immune condition of the host, plays an important role in the antiinflammatory response. Therefore, more patients with a high APACHE II score could be recognized in the intraabdominal infection group (10 *vs.* 6) in our study, and more patients with a higher score were admitted to the ICU



Figure 4 Selection of risk factors of intra-abdominal infection using the LASSO logistic regression algorithm. (A) LASSO coefficient profiles of the 43 candidate variables. Vertical line was plotted at the given lambda, selected by 10-fold cross-validation with minimum classification error plus 1 standard error, respectively. For the optimal lambda that gives minimum classification error plus 1 standard error, 5 features with a non-0 coefficient were selected. (B) Penalization coefficient lambda in the LASSO model was tuned using 10-fold cross-validation and the "lambda.1se" criterion. Area under the curve (AUC) metrics (y-axis) were plotted against log(lambda) (bottom x-axis). Top x-axis indicates the number of predictors for the given log(lambda). Red dots indicate average AUC for each model at the given lambda, and vertical bars through the red dots show the upper and lower values of the AUC according to the 10-fold cross-validation. Vertical black lines define the optimal lambda that gives the minimum classification error plus 1 standard error.



Figure 5 Performance of the logistic regression algorithm in intra-abdominal infection prediction. (A,B) Receiver-operating characteristic curves; (C,D) calibration curves.



Figure 6 Development and assessment of the random forest algorithm in intra-abdominal infection prediction. (A) Relationship between out-of-bag error and number of trees. In total, 114 trees are selected to establish a random forest model; (B) feature importance; (C,D) receiver-operating characteristic curves; (E,F) calibration curves.

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Figure 7 Development and assessment of the artificial neural network algorithm in intra-abdominal infection prediction. (A) Structure of artificial neural network; (B) feature importance; (C,D) receiver-operating characteristic curves; (E,F) calibration curves.

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Table 4 Receiver-operating	characteristic curves at t	the optimal cut-off	point according to	o different models
			porte according to	

Models	AUC	Sensitivity	Specificity	Correctly classified	LR+	LR-
Training set						
Logistic model	0.99	0.91	0.97	0.95	27.84	0.09
Random forest model	0.98	0.93	0.95	0.94	18.93	0.08
Artificial neural network model	0.99	0.96	0.96	0.96	22.48	0.05
Validation set						
Logistic model	0.98	0.93	0.95	0.95	18.85	0.07
Random forest model	0.98	0.94	0.93	0.94	14.03	0.06
Artificial neural network model	0.98	0.93	0.95	0.95	18.85	0.07

AUC, area under the curve; LR, likelihood ratio.



Figure 8 Nomogram to predict intra-abdominal infection was developed using the LASSO model selected predictors (intra-abdominal pressure, APACHE II score, CTSI, ICU admission, and severity grade were identified as risk factors). ***, P<0.001. APACHE II, Acute Physiology and Chronic Health Evaluation II; CTSI, CT severity index; ICU, Intensive care unit.

(86.8% *vs.* 20%), which was proved to be a risk factor for secondary intra-abdominal infection.

Many of the 711 included patients with acute pancreatitis could not be treated with injections alone, but also required

drainage of peritoneal effusion (38.5%), operation (8.8%), and continuous renal replacement therapy (4.4%), especially those admitted to the ICU. Surgery and minimally invasive treatment can destroy the integrity of skin and mucosa

which pose a hidden danger for secondary infection (17,18).

CT outperforms all other scores, not only for diagnostic accuracy but also for demonstrating the extent of the disease, as well as for guiding interventional procedures. Moreover, contrast-enhanced CT is the gold standard modality in the imaging evaluation of patients with acute pancreatitis (19). In 1990 Balthazar *et al.* established the CT severity index (CTSI) (20). This system grades pancreatic inflammation and necrosis, and is able to predict the morbidity and mortality of acute pancreatitis. Therefore, it was still found to be an independent risk factor for intraabdominal infection, as well as IAP.

A prediction model with 5 main risk factors was built during our 6-year study, and a high predictive value for intra-abdominal infection was confirmed. However, there are many limitations. First, prophylactic antimicrobial application was not further analyzed. Therefore, the relationship between antimicrobial resistance and its consumption could not be established. Second, no indicators related to immune status were analyzed, such as the human leukocyte antigen D-related expression or the CD4+Th/CD8+Th cell ratio, which was reported to be a risk factor for MDR infection in patients with severe acute pancreatitis (21-23). Third, although the sample size of the present study was adequate, the mortality was relatively low, which led to the uncompleted subgroup analysis. Further prospective trials with large sample size are needed in the future to obtain a higher grade of evidence.

Conclusions

Gram-negative bacteria, especially *Enterobacteriaceae*, are the main cause of intra-abdominal infection. Therefore, more attention should be paid to MDR pathogens. A new nomogram model that consisted of 5 risk factors (IAP, ICU admission, APACHE II score, CTSI, and severity of pancreatitis) was built for intra-abdominal infection prediction for patients with acute pancreatitis. Multicenter prospective validation cohorts should be used in subsequent studies to achieve a higher level of evidence.

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Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was conducted in accordance with the declaration of Helsinki (as revised in 2013). The Ethics Committee of Shanghai Jiaotong University School of Medicine approved the study (No. 2019-RES-082), and written informed consent was obtained from all patients.

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References

- Leppäniemi A, Tolonen M, Tarasconi A, et al. 2019 WSES guidelines for the management of severe acute pancreatitis. World J Emerg Surg 2019;14:27.
- Banks PA, Bollen TL, Dervenis C, et al. Classification of acute pancreatitis--2012: revision of the Atlanta classification and definitions by international consensus. Gut 2013;62:102-11.
- Werge M, Novovic S, Schmidt PN, et al. Infection increases mortality in necrotizing pancreatitis: A systematic review and meta-analysis. Pancreatology 2016;16:698-707.

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- Bassi C, Larvin M, Villatoro E. Antibiotic therapy for prophylaxis against infection of pancreatic necrosis in acute pancreatitis. Cochrane Database Syst Rev 2003;(4):CD002941.
- Villatoro E, Bassi C, Larvin M. Antibiotic therapy for prophylaxis against infection of pancreatic necrosis in acute pancreatitis. Cochrane Database Syst Rev 2006;(4):CD002941.
- Villatoro E, Mulla M, Larvin M. Antibiotic therapy for prophylaxis against infection of pancreatic necrosis in acute pancreatitis. Cochrane Database Syst Rev 2010;(5):CD002941.
- Cresci GA, Bawden E. Gut Microbiome: What We Do and Don't Know. Nutr Clin Pract 2015;30: 734-46.
- Tacconelli E, Cataldo MA, Dancer SJ, et al. ESCMID guidelines for the management of the infection control measures to reduce transmission of multidrug-resistant Gram-negative bacteria in hospitalized patients. Clin Microbiol Infect 2014;20 Suppl 1:1-55.
- Magiorakos AP, Srinivasan A, Carey RB, et al. Multidrugresistant, extensively drug-resistant and pandrug-resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. Clin Microbiol Infect 2012;18:268-81.
- Fernández-Delgado M, Sirsat MS, Cernadas E, et al. An extensive experimental survey of regression methods. Neural Netw 2019;111:11-34.
- 11. Alhamzawi R, Ali HTM. The Bayesian adaptive lasso regression. Math Biosci 2018;303:75-82.
- 12. Lu JD, Cao F, Ding YX, et al. Timing, distribution, and microbiology of infectious complications after necrotizing pancreatitis. World J Gastroenterol 2019;25:5162-73.
- Li Y, Shen H, Zhu C, et al. Carbapenem-Resistant Klebsiella pneumoniae Infections among ICU Admission Patients in Central China: Prevalence and Prediction Model. Biomed Res Int 2019;2019:9767313.
- 14. Zheng B, Dai Y, Liu Y, et al. Molecular Epidemiology and Risk Factors of Carbapenem-Resistant Klebsiella

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pneumoniae Infections in Eastern China. Front Microbiol 2017;8:1061.

- Knaus WA, Draper EA, Wagner DP, et al. APACHE II: a severity of disease classification system. Crit Care Med 1985;13:818-29.
- Kumar S, Gattani SC, Baheti AH, et al. Comparison of the Performance of APACHE II, SOFA, and mNUTRIC Scoring Systems in Critically Ill Patients: A 2-year Crosssectional Study. Indian J Crit Care Med 2020;24:1057-61.
- Yu Y, Shen H, Zhu C, et al. Infections Caused by Extended-Spectrum beta-Lactamase Producing Escherichia Coli in Systemic Lupus Erythematosus Patients: Prevalence, Risk Factors, and Predictive Model. Biomed Res Int 2018;2018:8296720.
- Zhang Z, Smischney NJ, Zhang H, et al. AME evidence series 001-The Society for Translational Medicine: clinical practice guidelines for diagnosis and early identification of sepsis in the hospital. J Thorac Dis 2016;8:2654-65.
- Balthazar EJ. Acute pancreatitis: assessment of severity with clinical and CT evaluation.Radiology 2002;223:603-13
- Balthazar EJ, Robinson DL, Megibow AJ, et al. Acute pancreatitis: value of CT in establishing prognosis. Radiology 1990;174:331-6.
- Yu ZX, Chen XC, Zhang BY, et al. Association between HLA-DR Expression and Multidrug-resistant Infection in Patients with Severe Acute Pancreatitis. Curr Med Sci 2018;38:449-54.
- Ferrian S, Ross M, Conradie F, et al. Frequency of Circulating CD4(+)Ki67(+)HLA-DR(-) T Regulatory Cells Prior to Treatment for Multidrug Resistant Tuberculosis Can Differentiate the Severity of Disease and Predict Time to Culture Conversion. Front Immunol 2018;9:2438.
- 23. Shen H, Yu Y, Chen SM, et al. Dectin-1 Facilitates IL-18 Production for the Generation of Protective Antibodies Against Candida albicans. Front Microbiol 2020;11:1648.

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Figure S1 Enhanced computed tomography of acute pancreatitis with infection. (A-F) Gas configurations in the necrotic pancreatic tissue can be detected (green arrows).



Figure S2 Computed tomography-guided fine-needle aspiration was performed to obtain drainage fluid from peri-pancreatic tissues. (A,B,C) Left para-colonic sulcus drainage (prone position); (D,E,F) right para-colonic sulcus drainage (supine position).



Figure S3 Antimicrobial susceptibility of gram-positive bacteria. Teicoplanin, vancomycin, linezolid, and tigecycline are the best choice for Gram-positive bacteria.

Table S1 Laboratory results of patients with acute pancreatitis (day 1)

Variables	Total (n=711)	Non-IAI group (n=529)	IAI group (n=182)	P value
Cr, median (IQR)	62 (54, 72)	62 (54, 70)	61.5 (51, 77.8)	0.44
Hct, median (IQR)	0.4 (0.3, 0.4)	0.4 (0.3, 0.4)	0.4 (0.3, 0.4)	0.14
WBC, median (IQR)	10.2 (7.4, 13.8)	9.9 (6.8, 13.1)	11.4 (8.8, 15.8)	<0.01
PNEUT, median (IQR)	82.8 (74.5, 87.7)	81.3 (71.6, 86.9)	86.2 (81.8, 89.3)	<0.01
Neutrophils, median (IQR)	8.4 (5.5, 11.8)	7.9 (5.1, 11)	9.7 (7.3, 13.7)	<0.01
TLC, median (IQR)	1.1 (0.8, 1.5)	1.2 (0.9, 1.6)	1 (0.7, 1.3)	<0.01
PLYMPH, median (IQR)	10.8 (7.2, 17.4)	12 (8, 19.6)	8.6 (5.7, 11.7)	<0.01
Lymphocytes, median (IQR)	1.1 (0.8, 1.5)	1.2 (0.9, 1.7)	1 (0.7, 1.3)	<0.01
EOS, median (IQR)	0 (0, 0.1)	0.1 (0, 0.1)	0 (0, 0)	<0.01
PEO, median (IQR)	0.4 (0.1, 1.3)	0.6 (0.1, 1.6)	0.1 (0, 0.4)	<0.01
Hb, median (IQR)	132 (117, 145)	133 (118, 145)	126.5 (112, 146)	0.18
PLT, median (IQR)	181 (139, 224)	180 (141, 221)	185 (138, 234.8)	0.31
Lactates, median (IQR)	1.7 (1.3, 2.2)	1.6 (1.3, 2.1)	1.9 (1.4, 2.6)	<0.01
PT, median (IQR)	13.2 (12.4, 14.2)	13 (12.3, 13.9)	13.9 (13.1, 14.8)	<0.01
APTT, median (IQR)	30.4 (27.8, 33.6)	30.1 (27.7, 33.1)	31.9 (28.2, 36)	<0.01
TT, median (IQR)	17.3 (16.2, 18.8)	17.3 (16.2, 18.8)	17.2 (16.1, 19)	0.99
Fg, median (IQR)	4.8 (3.2, 6.3)	4.5 (3, 6)	5.4 (3.9, 6.6)	<0.01
FDP, median (IQR)	8.5 (4, 17.3)	6.4 (3.1, 13.8)	15.2 (9.2, 25.1)	<0.01
DD, median (IQR)	2.4 (1, 5.2)	1.7 (0.8, 4.1)	4.5 (2.6, 8.2)	<0.01
TG, median (IQR)	2 (1.1, 4.5)	2 (1.1, 4.3)	2.2 (1.1, 5.4)	0.11
TC, median (IQR)	4.5 (3.3, 6)	4.6 (3.4, 6.1)	4.3 (2.9, 6)	0.07
LP, median (IQR)	0.1 (0, 0.2)	0.1 (0, 0.2)	0.1 (0, 0.2)	0.12
ApoA, mean ± SD	0.9±0.3	1±0.3	0.8±0.3	<0.01
ApoB, median (IQR)	0.8 (0.6, 1)	0.8 (0.6, 1.1)	0.7 (0.6, 1)	0.03
ApoE, median (IQR)	4.8 (3.7, 7.2)	4.8 (3.7, 7.2)	4.8 (3.9, 8)	0.36
LDLc, median (IQR)	2.1 (1.5, 3.1)	2.3 (1.6, 3.2)	1.7 (1.1, 2.5)	<0.01
HDLc, median (IQR)	0.9 (0.6, 1.1)	0.9 (0.7, 1.1)	0.7 (0.5, 0.9)	<0.01
FFA, median (IQR)	0.7 (0.6, 0.9)	0.7 (0.6, 0.9)	0.7 (0.6, 0.9)	0.28
Na, median (IQR)	138 (136, 140)	138 (136, 140)	137 (134, 140)	<0.01
K, median (IQR)	3.8 (3.5, 4.1)	3.8 (3.5, 4.1)	3.8 (3.5, 4.2)	0.52
CO2CP, median (IQR)	23.4 (20.4, 26)	23.6 (21.2, 25.9)	22.3 (18.9, 26)	0.02
P, median (IQR)	0.8 (0.6, 1)	0.9 (0.7, 1.1)	0.7 (0.5, 1)	<0.01
Ca, median (IQR)	2.1 (2, 2.2)	2.1 (2, 2.2)	2 (1.8, 2.1)	<0.01
LDH, median (IQR)	278 (200.2, 388.8)	274 (193.8, 382)	289 (228.5, 408.5)	0.04
CK, median (IQR)	74 (45, 106)	75 (46, 105)	69.5 (38.2, 121)	0.41
MYO, median (IQR)	30 (17.5, 36)	29.4 (16.1, 31.6)	30 (21.6, 62.6)	<0.01
Tnl, median (IQR)	0 (0, 0.5)	0 (0, 0.5)	0 (0, 0.5)	<0.01
PCT, median (IQR)	0.3 (0.1, 1.1)	0.2 (0, 0.7)	0.8 (0.3, 2.1)	<0.01
CRP, median (IQR)	79 (50.2, 135)	80 (54, 135)	75 (43, 142)	0.24
AG, median (IQR)	1.2 (1, 1.5)	1.2 (1, 1.5)	1.2 (1, 1.4)	0.74
ALT, median (IQR)	26 (15, 46)	24 (15, 44)	31.5 (17.2, 56)	<0.01
Alb, median (IQR)	37 (33, 40)	37 (33, 40)	36 (33, 40)	0.56
AST, median (IQR)	35 (24.8, 53.2)	35 (24, 52)	36 (26, 63)	0.03
TBIL, median (IQR)	17.2 (11.8, 24.9)	16.8 (11.7, 24.6)	18.1 (12.4, 27.6)	0.19
ALP, median (IQR)	73 (55, 104)	71 (53.5, 97.5)	82 (60, 122.5)	<0.01
DBIL, median (IQR)	3.7 (2.1, 6.8)	3.6 (2.1, 6.3)	4.1 (2.2, 9.3)	0.03

AG, albumin-to-globulin ratio; Alb, albumin; ALP, alkaline phosphatase; ALT, alanine transaminase; ApoA, apolipoprotein A; ApoB, apolipoprotein B; ApoE, apolipoprotein E; APTT, activated partial thromboplastin time; AST, aspartate transaminase; Ca, calcium; CK, creatine kinase; CO2CP; carbon dioxide combining power; Cr, creatinine; CRP, C-reactive protein; DBIL, direct bilirubin; DD, D-dimer; EOS, eosinophil; FDP, fibrin degradation products; FFA, free fatty acids; Fg, fibrinogen; Hb, hemoglobin; Hct, hematocrit; IQR, interquartile range; K, potassium; HDLc, high-density lipoprotein cholesterol; LDH, lactate dehydrogenase; LDLc, low-density lipoprotein cholesterol; LP, lipoprotein; MYO, myoglobin; Na, sodium; P, phosphorus; PCT, procalcitonin; PEO, percentage of eosinophils; PLT, platelets; PLYMPH, percentage of lymphocytes; PNEUT, percentage of neutrophils; PT, prothrombin time; SD, standard deviation; TBIL, total bilirubin; TC, total cholesterol; TG, total glyceride; Tnl, troponin; TLC, total lymphocyte count; TT, thrombin time; WBC, white blood cells.

Table S2 Laboratory	results of	patients wit	h acute pancr	eatitis (day 3)
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Variables	Total (n=711)	Non-IAI group (n=529)	IAI group (n=182)	P value
Cr, median (IQR)	62 (52, 71)	62 (53, 71)	59 (48, 71)	0.18
Hct, median (IQR)	0.4 (0.3, 0.4)	0.4 (0.4, 0.4)	0.4 (0.3, 0.4)	<0.01
WBC, median (IQR)	8.8 (5.9, 13)	8.2 (5.4, 12.2)	10.8 (7.7, 14.7)	<0.01
PNEUT, median (IQR)	79.6 (67.5, 85.8)	76 (63.2, 84.9)	84.8 (79.9, 88.1)	<0.01
Neutrophils, median (IQR)	7 (4.2, 10.3)	6.2 (3.8, 9.5)	9 (6.2, 12.6)	<0.01
TLC, median (IQR)	1.2 (0.8, 1.6)	1.3 (1, 1.7)	1 (0.7, 1.3)	<0.01
PLYMPH, median (IQR)	12.8 (8.1, 21.7)	15.3 (8.9, 24.7)	9.4 (6.1, 12.4)	<0.01
Lymphocytes, median (IQR)	1.1 (0.8, 1.6)	1.2 (0.8, 1.7)	0.9 (0.7, 1.3)	<0.01
EOS, median (IQR)	0.1 (0, 0.2)	0.1 (0, 0.2)	0.1 (0, 0.2)	0.41
PEO, median (IQR)	0.5 (0.1, 2)	0.5 (0.1, 1.7)	0.6 (0.1, 2.1)	0.67
Hb, median (IQR)	128 (114, 142)	131 (118, 143)	120 (110, 134.8)	<0.01
PLT, median (IQR)	185 (145, 241.5)	187 (148.2, 238.8)	178.5 (139.5, 252)	0.46
Lactates, median (IQR)	1.6 (1.4, 2.1)	1.6 (1.4, 2)	1.8 (1.4, 2.2)	0.16
PT, median (IQR)	13.4 (12.7, 14.3)	13.3 (12.6, 14.2)	13.7 (13, 14.9)	<0.01
APTT, median (IQR)	30.9 (28.2, 34.5)	30.6 (28, 33.9)	31.9 (28.9, 35.7)	<0.01
TT, median (IQR)	17.5 (16.2, 19)	17.6 (16.4, 19.1)	17.3 (15.8, 18.6)	0.01
Fg, median (IQR)	4.8 (3.6, 6.3)	4.7 (3.5, 6.2)	5.3 (4, 6.7)	<0.01
FDP, median (IQR)	12.2 (6.1, 21.4)	10.1 (4.7, 18.4)	17.4 (11.9, 26.6)	<0.01
DD, median (IQR)	3.8 (1.7, 6.9)	3 (1.3, 5.9)	5.9 (3.5, 8.3)	<0.01
Na, median (IQR)	138 (136, 141)	138 (136, 140)	138 (135.5, 142)	0.65
K, median (IQR)	3.8 (3.6, 4.1)	3.8 (3.6, 4.1)	3.7 (3.5, 4.1)	0.25
CO2CP, median (IQR)	24.1 (21.6, 26.9)	24 (22, 26.2)	24.5 (21, 27.9)	0.36
P, median (IQR)	0.9 (0.6, 1.1)	1 (0.7, 1.1)	0.7 (0.5, 1)	<0.01
Ca, median (IQR)	2.2 (2, 2.3)	2.2 (2.1, 2.3)	2 (1.9, 2.2)	<0.01
LDH, median (IQR)	265 (188, 329.2)	264 (186, 327)	274 (207.5, 361.5)	0.1
CK, median (IQR)	64 (43, 86)	64 (44, 86)	59 (37, 89.5)	0.37
PCT, median (IQR)	0.2 (0, 1.2)	0.2 (0, 0.9)	0.2 (0, 1.4)	0.6
CRP, median (IQR)	74 (52, 100.5)	74 (49, 100.2)	74 (54, 100.5)	0.42
AG, median (IQR)	1.2 (1, 1.4)	1.2 (1, 1.4)	1.1 (0.9, 1.3)	<0.01
AST, median (IQR)	25 (18, 36)	25 (18, 38)	24 (18, 34)	0.46
Alb, median (IQR)	35 (31, 38)	35 (32, 39)	32 (29, 36)	<0.01
ALT, median (IQR)	21 (14, 38)	21 (14, 37)	20 (14, 39)	0.7
TBIL, median (IQR)	19 (13.6, 26.8)	18.3 (13.3, 25.3)	20.3 (15.1, 30.2)	<0.01
ALP, median (IQR)	64 (51.8, 84)	65 (53, 85)	62 (49, 83.5)	0.21
DBIL, median (IQR)	3.8 (2.3, 6.1)	3.6 (2.3, 5.5)	5 (2.7, 8)	<.01
AFP, median (IQR)	1.9 (1.4, 2.8)	2 (1.4, 2.9)	1.6 (1.2, 2.6)	0.01
CEA, median (IQR)	1.9 (1.3, 2.6)	1.9 (1.3, 2.6)	1.8 (1.3, 2.6)	0.78
IL-6, median (IQR)	24.6 (16.6, 34.8)	23.8 (15.3, 32.9)	32.9 (22.4, 46.3)	<0.01
IL-8, median (IQR)	125.5 (84, 190.3)	114.5 (78.2, 176.3)	190.4 (109.6, 245.9)	<0.01
IL-10, median (IQR)	44.5 (2.4, 67.4)	32.6 (2.4, 57.3)	67.8 (46.4, 87.3)	<0.01

AG, albumin-to-globulin ratio; Alb, albumin; ALP, alkaline phosphatase; ALT, alanine transaminase; ApoA, apolipoprotein A; ApoB, apolipoprotein B; ApoE, apolipoprotein E; APTT, activated partial thromboplastin time; AST, aspartate transaminase; Ca, calcium; CK, creatine kinase; CO2CP, carbon dioxide combining power; Cr, creatinine; CRP, C-reactive protein; DBIL, direct bilirubin; DD, D-dimer; EOS, eosinophils; FDP, fibrin degradation products; FFA, free fatty acids; Fg, fibrinogen; Hb, hemoglobin; Hct, hematocrit; IQR, interquartile range; K, potassium; HDLc, high-density lipoprotein cholesterol; LDH, lactate dehydrogenase; LDLc, low-density lipoprotein cholesterol; LP, lipoprotein; MYO, myoglobin; Na, sodium; P, phosphorus; PCT, procalcitonin; PEO, percentage of eosinophils; PLT, platelets; PLYMPH, percentage of lymphocytes; PNEUT, percentage of neutrophils; PT, prothrombin time; TBIL, total bilirubin; TC, total cholesterol; TG, total glyceride; Tnl, troponin; TLC, total lymphocyte count; TT, thrombin time; WBC, white blood cells.

Table S3 Laboratory results of patients with acute pancreatitis (day 7)

Variables	Total (n=711)	Non-IAI group (n=529)	IAI group (n=182)	P value
Cr, median (IQR)	64 (54, 76)	65 (55, 76)	58 (46.5, 72)	<0.01
Hct, median (IQR)	0.4 (0.3, 0.4)	0.4 (0.3, 0.4)	0.4 (0.3, 0.4)	<0.01
WBC, median (IQR)	8.4 (6, 11.9)	7.8 (5.7, 10.7)	10.2 (7.5, 13.2)	<0.01
PNEUT, median (IQR)	78.3 (65.9, 85)	74.6 (62.4, 83.6)	83.9 (78.5, 87.7)	<0.01
Neutrophils, median (IQR)	6.5 (4.1, 9.9)	5.8 (3.6, 8.8)	8.7 (6.1, 12)	<0.01
TLC, median (IQR)	1.2 (0.9, 1.6)	1.3 (0.9, 1.8)	1 (0.7, 1.3)	<0.01
PLYMPH, median (IQR)	13.3 (8.4, 23.6)	15.9 (9.5, 27.3)	9.2 (6.5, 13.6)	<0.01
Lymphocytes, median (IQR)	1.2 (0.8, 1.6)	1.3 (0.8, 1.8)	1 (0.7, 1.3)	<0.01
EOS, median (IQR)	0.2 (0.1, 0.2)	0.2 (0.1, 0.2)	0.2 (0.1, 0.2)	0.2
PEO, median (IQR)	1.5 (0.9, 2.5)	1.5 (0.9, 2.4)	1.5 (0.9, 2.9)	0.27
Hb, median (IQR)	125 (112, 138)	128 (114.2, 141)	117 (107, 130)	<0.01
PLT, median (IQR)	193 (148.5, 255)	197 (151.8, 255.2)	181 (138.5, 253)	0.3
Lactates, median (IQR)	1.8 (1.5, 2.4)	1.8 (1.5, 2.3)	1.9 (1.6, 2.5)	0.14
PT, median (IQR)	13.3 (12.6, 14.2)	13.2 (12.6, 13.9)	13.6 (12.8, 14.6)	<0.01
APTT, median (IQR)	30.5 (28.1, 33.8)	30.4 (28.1, 33.3)	31 (28.3, 34)	0.48
TT, median (IQR)	17.5 (16.2, 18.8)	17.6 (16.3, 18.9)	17.3 (15.8, 18.8)	0.05
Fg, median (IQR)	4.8 (3.6, 6.2)	4.7 (3.5, 6.2)	5 (4.2, 6.4)	<0.01
FDP, median (IQR)	13.7 (6.7, 21.7)	11.4 (5.1, 18.4)	19.2 (13.4, 28.7)	<0.01
DD, median (IQR)	4.3 (1.9, 7)	3.5 (1.4, 6.4)	6.3 (4, 8.7)	<0.01
Na, median (IQR)	139 (136, 141)	139 (136, 141)	139 (136, 142)	0.6
K, median (IQR)	3.8 (3.5, 4.1)	3.8 (3.5, 4.1)	3.8 (3.5, 4.1)	0.45
CO2CP, median (IQR)	28 (25.6, 30.1)	27.7 (25.4, 29.7)	28.4 (26, 31.3)	<0.01
P, median (IQR)	0.9 (0.7, 1.1)	1 (0.7, 1.2)	0.8 (0.6, 1)	<0.01
Ca, median (IQR)	2.2 (2, 2.3)	2.2 (2.1, 2.3)	2.1 (1.9, 2.2)	<0.01
LDH, median (IQR)	260 (190, 300)	258 (186, 297)	264 (196, 324)	0.27
CK, median (IQR)	82 (38, 136)	84 (42.8, 135.8)	77 (31, 145)	0.43
PCT, median (IQR)	0.2 (0, 1.2)	0.2 (0, 1.2)	0.2 (0, 1.2)	0.63
CRP, median (IQR)	74 (53, 97)	75 (54, 97)	70 (49, 97)	0.8
AG, median (IQR)	1.2 (1.1, 1.5)	1.2 (1.1, 1.5)	1.3 (1.1, 1.4)	0.88
ALT, median (IQR)	34 (23, 53)	34 (23, 53)	33 (22, 52)	0.38
Alb, median (IQR)	38 (34, 42)	37 (34, 42)	38 (34, 41)	0.53
AST, median (IQR)	23 (18, 35)	23 (18, 37)	23 (17.8, 33)	0.5
TBIL, median (IQR)	14.1 (10.5, 19.3)	14.3 (10.6, 19.9)	13.4 (10.3, 18.9)	0.19
ALP, median (IQR)	69 (56, 89.2)	69 (56, 88)	70 (56, 93)	0.79
DBIL, median (IQR)	2.7 (1.6, 5.1)	2.7 (1.6, 5.1)	2.6 (1.5, 4.9)	0.53

AG, albumin-to-globulin ratio; Alb, albumin; ALP, alkaline phosphatase; ALT, alanine transaminase; ApoA, apolipoprotein A; ApoB, apolipoprotein B; ApoE, apolipoprotein E; APTT, activated partial thromboplastin time; AST, aspartate transaminase; Ca, calcium; CK, creatine kinase; CO2CP; carbon dioxide combining power; Cr, creatinine; CRP, C-reactive protein; DBIL, direct bilirubin; DD, D-dimer; EOS, eosinophils; FDP, fibrin degradation products; FFA, free fatty acids; Fg, fibrinogen; Hb, hemoglobin; Hct, hematocrit; K, potassium; HDLc, high-density lipoprotein cholesterol; IQR, interquartile range; LDH, lactate dehydrogenase; LDLc, low-density lipoprotein cholesterol; LP, lipoprotein; MYO, myoglobin; Na, sodium; P, phosphorus; PCT, procalcitonin; PEO, percentage of eosinophils; PLT, platelets; PLYMPH, percentage of lymphocytes; PNEUT, percentage of neutrophils; PT, prothrombin time; TBIL, total bilirubin; TC, total cholesterol; TG, total glyceride; Tnl, troponin; TLC, total lymphocyte count; TT, thrombin time; WBC, white blood cells.

Table S4 Laboratory results of patients with acute pancreatitis (day 14)

Variables	Total (n=711)	Non-IAI group (n=529)	IAI group (n=182)	P value
Cr, median (IQR)	65 (53, 77)	67 (56, 78)	60 (48.2, 73)	<0.01
Hct, median (IQR)	0.4 (0.3, 0.4)	0.4 (0.3, 0.4)	0.3 (0.3, 0.4)	<0.01
WBC, median (IQR)	8.5 (5.8, 11.9)	8.6 (6, 11.7)	8.3 (5.6, 12)	0.41
PNEUT, median (IQR)	78.2 (66.8, 84.5)	73.8 (60.8, 83.3)	81.6 (75.9, 86.5)	<0.01
Neutrophils, median (IQR)	6.9 (4.1, 10)	5.6 (3.6, 9.2)	8.1 (5.7, 10.9)	<0.01
TLC, median (IQR)	1.2 (0.9, 1.7)	1.3 (1, 1.7)	1.1 (0.8, 1.4)	<0.01
PLYMPH, median (IQR)	13.3 (9, 22.9)	15.6 (9.7, 27.1)	10.6 (8, 15.4)	<0.01
Lymphocytes, median (IQR)	1.1 (0.7, 2.1)	1.3 (0.8, 2.6)	0.9 (0.6, 1.5)	<0.01
EOS, median (IQR)	0.2 (0.1, 0.3)	0.2 (0.1, 0.3)	0.2 (0.1, 0.3)	0.19
PEO, median (IQR)	1.9 (1.1, 2.7)	1.9 (1.2, 2.6)	1.8 (0.9, 3.1)	0.74
Hb, mean ± SD	122.7±20.4	125.4±20.3	117.6±19.7	<0.01
PLT, median (IQR)	201 (156, 264.5)	204 (155.8, 259.8)	198 (156, 266.5)	0.76
Lactates, median (IQR)	1.9 (1.5, 2.4)	1.8 (1.5, 2.4)	1.9 (1.4, 2.4)	0.44
PT, median (IQR)	13.3 (12.5, 14.1)	13.2 (12.5, 14)	13.4 (12.7, 14.4)	0.02
APTT, median (IQR)	29.8 (27.4, 33.1)	29.8 (27.6, 32.7)	29.8 (27.4, 33.7)	0.58
TT, median (IQR)	17.6 (16.4, 18.8)	17.7 (16.4, 18.8)	17.5 (16.3, 19.1)	0.71
Fg, median (IQR)	4.6 (3.5, 5.9)	4.6 (3.3, 5.9)	4.7 (3.8, 5.9)	0.21
FDP, median (IQR)	14.7 (6.4, 22.4)	10.6 (4.7, 18.4)	18.6 (12.4, 28)	<0.01
DD, median (IQR)	4.5 (2.1, 7.3)	3.5 (1.4, 5.9)	6.2 (3.6, 8.7)	<0.01
Na, median (IQR)	139 (137, 142)	139 (137, 141.2)	139 (137, 142)	0.5
K, median (IQR)	3.8 (3.6, 4.2)	3.8 (3.6, 4.1)	3.8 (3.5, 4.2)	0.36
CO2CP, mean±SD	28.9 ± 4.4	28 ± 4.5	29.7 ± 4.2	<0.01
P, median (IQR)	1 (0.7, 1.2)	1.1 (0.7, 1.2)	0.8 (0.6, 1.1)	<0.01
Ca, median (IQR)	2.2 (2.1, 2.4)	2.3 (2.1, 2.4)	2.1 (2, 2.3)	<0.01
LDH, median (IQR)	216 (164, 270.5)	215 (158, 266)	217 (165.2, 274.8)	0.4
CK, median (IQR)	57 (38, 95.2)	56 (38, 96)	58 (38, 93.5)	0.88
PCT, median (IQR)	0.2 (0, 0.7)	0.2 (0, 0.6)	0.3 (0.1, 1)	<0.01
CRP, median (IQR)	32 (22, 46)	31 (21, 45)	35 (25, 48)	0.03
AG, median (IQR)	1.2 (1.1, 1.5)	1.3 (1.1, 1.5)	1.2 (1, 1.4)	0.08
ALT, median (IQR)	21 (14, 39)	23 (15, 40)	17 (13, 32)	<0.01
ALB, median (IQR)	38 (34, 41)	38 (34, 42)	37.5 (34, 41)	0.11
AST, median (IQR)	24 (18, 34)	24 (18, 34)	24 (17, 34.5)	0.91
TBIL, median (IQR)	14.6 (10.5, 22.1)	14.3 (10.5, 22.1)	15.1 (10.4, 21.2)	0.88
ALP, median (IQR)	68 (53, 86.2)	68.5 (53, 87)	65.5 (52, 85.2)	0.5
DBIL, median (IQR)	3.1 (1.8, 5.3)	3 (1.8, 5.1)	3.2 (1.7, 5.6)	0.68

AG, albumin-to-globulin ratio; Alb, albumin; ALP, alkaline phosphatase; ALT, alanine transaminase; ApoA, apolipoprotein A; ApoB, apolipoprotein B; ApoE, apolipoprotein E; APTT, activated partial thromboplastin time; AST, aspartate transaminase; Ca, calcium; CK, creatine kinase; CO2CP, carbon dioxide combining power; Cr, creatinine; CRP, C-reactive protein; DBIL, direct bilirubin; DD, D-dimer; EOS, eosinophils; FDP, fibrin degradation products; FFA, free fatty acids; Fg, fibrinogen; Hb, hemoglobin; Hct, hematocrit; K, potassium; HDLc, high-density lipoprotein cholesterol; LDH, lactate dehydrogenase; IQR, interquartile range; LDLc, low-density lipoprotein cholesterol; LP, lipoprotein; MYO, myoglobin; Na, sodium; P, phosphorus; PCT, procalcitonin; PEO, percentage of eosinophils; FLT, platelets; PLYMPH, percentage of lymphocyte; PNEUT, percentage of neutrophils; PT, prothrombin time; SD, standard deviation; TBIL, total bilirubin; TC, total @CNAPEsteroff_TG_state_b_glwgetide; T_NIItroppenest_G_v_G_total lymphocyte count; TT, thrombin time; WBC, white blogQd GetBs_/10.21037/atm-21-399

Table S5 Laboratory results of patients in training and validation groups

Variables	Total (n=711)	Validation group (n=294)	Train group (n=417)	P value
Cr, median (IQR)	62 (54, 72)	63 (55, 75)	60 (53, 70)	0.04
Hct, median (IQR)	0.4 (0.3, 0.4)	0.4 (0.3, 0.4)	0.4 (0.3, 0.4)	0.28
WBC, median (IQR)	10.2 (7.4, 13.8)	10.3 (7.3, 14.2)	10.2 (7.5, 13.6)	0.54
PNEUT, median (IQR)	82.8 (74.5, 87.7)	82.3 (74.5, 87.5)	83.3 (74.8, 87.7)	0.58
Neutrophils, median (IQR)	8.4 (5.5, 11.8)	8.7 (5.4, 12.1)	8.3 (5.6, 11.6)	0.61
TLC, median (IQR)	1.1 (0.8, 1.5)	1.1 (0.9, 1.5)	1.1 (0.8, 1.6)	0.69
PLYMPH, median (IQR)	10.8 (7.2, 17.4)	11.1 (7, 17.3)	10.5 (7.4, 17.6)	0.88
Lymphocytes, median (IQR)	1.1 (0.8, 1.5)	1.1 (0.9, 1.5)	1.1 (0.8, 1.6)	0.67
EOS, median (IQR)	0 (0, 0.1)	0 (0, 0.1)	0 (0, 0.1)	0.52
PEO, median (IQR)	0.4 (0.1, 1.3)	0.4 (0.1, 1.5)	0.3 (0.1, 1.2)	0.65
Hb, median (IQR)	132 (117, 145)	134.5 (118, 147)	130 (116, 144)	0.02
PLT, median (IQR)	181 (139, 224)	181 (140.2, 220.8)	181 (139, 227)	0.98
Lactates, median (IQR)	1.7 (1.3, 2.2)	1.7 (1.3, 2.2)	1.6 (1.3, 2.1)	0.17
PT, median (IQR)	13.2 (12.4, 14.2)	13.3 (12.4, 14.2)	13.2 (12.5, 14.2)	0.52
APTT, median (IQR)	30.4 (27.8, 33.6)	30.4 (27.8, 34.1)	30.4 (27.9, 33.5)	0.75
TT, median (IQR)	17.3 (16.2, 18.8)	17.5 (16.3, 19)	17.1 (16.1, 18.8)	0.12
Fg, median (IQR)	4.8 (3.2, 6.3)	4.8 (3.1, 6.3)	4.8 (3.4, 6.3)	0.4
FDP, median (IQR)	8.5 (4, 17.3)	7.2 (3.9, 17.3)	9.6 (4.3, 17.2)	0.17
DD, median (IQR)	2.4 (1, 5.2)	2.1 (1, 5)	2.9 (1.1, 5.3)	0.14
TG, median (IQR)	2 (1.1, 4.5)	1.9 (0.9, 4.2)	2 (1.1, 4.9)	0.08
TC, median (IQR)	4.5 (3.3, 6)	4.5 (3.3, 6)	4.6 (3.3, 6.1)	0.87
LP, median (IQR)	0.1 (0, 0.2)	0.1 (0.1, 0.2)	0.1 (0, 0.2)	0.55
ApoA, mean±SD	0.9 ± 0.3	0.9 ± 0.3	0.9 ± 0.3	0.91
ApoB, median (IQR)	0.8 (0.6, 1)	0.8 (0.6, 1)	0.8 (0.6, 1)	0.74
ApoE, median (IQR)	4.8 (3.7, 7.2)	4.6 (3.5, 6.6)	5 (3.9, 7.7)	<0.01
LDLc, median (IQR)	2.1 (1.5, 3.1)	2.2 (1.5, 3.1)	2.1 (1.5, 3)	0.64
HDLc, median (IQR)	0.9 (0.6, 1.1)	0.9 (0.7, 1.1)	0.8 (0.6, 1.1)	0.26
FFA, median (IQR)	0.7 (0.6, 0.9)	0.7 (0.5, 0.9)	0.7 (0.6, 0.9)	0.12
Na, median (IQR)	138 (136, 140)	138 (136, 140)	138 (135, 141)	0.62
K, median (IQR)	3.8 (3.5, 4.1)	3.8 (3.5, 4.1)	3.8 (3.5, 4.1)	0.27
CO2CP, median (IQR)	23.4 (20.4, 26)	23.6 (20.9, 25.9)	23.3 (20, 26)	0.64
P, median (IQR)	0.8 (0.6, 1)	0.8 (0.6, 1)	0.8 (0.6, 1)	0.79
Ca, median (IQR)	2.1 (2, 2.2)	2.1 (2, 2.2)	2.1 (2, 2.2)	0.24
LDH, median (IQR)	278 (200.2, 388.8)	282 (189, 396)	275 (212, 386)	0.68
CK, median (IQR)	74 (45, 106)	70 (43, 104.8)	75 (47, 112)	0.2
MYO, median (IQR)	30 (17.5, 36)	30 (18.1, 39.7)	30 (16.5, 34.4)	0.25
Tnl, median (IQR)	0 (0, 0.5)	0 (0, 0.5)	0 (0, 0.5)	0.34
PCT, median (IQR)	0.3 (0.1, 1.1)	0.3 (0.1, 1.1)	0.3 (0.1, 1)	0.58
CRP, median (IQR)	79 (50.2, 135)	79 (50, 135)	79 (53, 135)	0.84
AG, median (IQR)	1.2 (1, 1.5)	1.2 (1, 1.5)	1.2 (1, 1.4)	0.34
ALT, median (IQR)	26 (15, 46)	24 (15, 46)	27 (15, 46.5)	0.93
Alb, median (IQR)	37 (33, 40)	37 (33, 40)	36 (33, 39)	0.55
AST, median (IQR)	35 (24.8, 53.2)	36 (26, 54)	34 (23.5, 53)	0.13
TBIL, median (IQR)	17.2 (11.8, 24.9)	17.6 (11.9, 25.6)	16.8 (11.6, 24.4)	0.3
ALP, median (IQR)	73 (55, 104)	75 (57, 108)	72 (54, 99.5)	0.15
DBIL, median (IQR) © Annals of Translational Medicine. A	3.7 (2.1, 6.8) ll rights reserved.	3.8 (2, 7.1)	.3.7 (2.2, 6.8) http://dx.doi.org/10.2	0.9 1037/atm-21-399

AG, albumin-to-globulin ratio; Alb, albumin; ALP, alkaline phosphatase; ALT, alanine transaminase; ApoA, apolipoprotein A; ApoB, apolipoprotein B; ApoE, apolipoprotein E; APTT, activated partial thromboplastin time; AST, aspartate transaminase; Ca, calcium; CK, creatine kinase; CO2CP, carbon dioxide combining power; Cr, creatinine; CRP, C-reactive protein; DBIL, direct bilirubin; DD, D-dimer; EOS, eosinophils; FDP, fibrin degradation products; FFA, free fatty acids; Fg, fibrinogen; Hb, hemoglobin; Hct, hematocrit; K, potassium; HDLc, high-density lipoprotein cholesterol; IQR, interquartile range; LDH, lactate dehydrogenase; LDLc, low-density lipoprotein cholesterol; LP, lipoprotein; MYO, myoglobin; Na, sodium; P, phosphorus; PCT, procalcitonin; PEO, percentage of eosinophils; PLT, platelets; PLYMPH, percentage of lymphocytes; PNEUT, percentage of neutrophils; PT, prothrombin time; TBIL, total bilirubin; TC, total cholesterol; TG, total glyceride; Tnl, troponin; TLC, total lymphocyte count; TT, thrombin time; WBC, white blood cells.