Development and validation of a novel miRNA classifier as a prognostic signature for stage II/III colorectal cancer

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Background: The TNM staging remains the gold standard for determining the prognosis of patients with colorectal cancer (CRC), which is inadequate at identifying the subset of high-risk stage II and III patients that have a high potential of developing tumor recurrence and may experience death. Emerging evidence indicates that not only microRNAs (miRNAs) play important functional role in CRC development but may serve as important disease biomarkers. In this study we aimed to develop a miRNA-based classifier as a prognostic signature for improving the clinical outcome of patients with stage II/III CRC.

Methods: We performed a systematic and comprehensive discovery step to identify differentially expressed miRNAs in CRC. We subsequently determined the prognostic relevance of these miRNAs in stage II/III patients using qRT-PCR and developed a miRNA-based classifier for predicting disease-free survival (DFS) in a clinical cohort (n=186).

Results: Based upon miRNA expression profiling studies, we identified a panel of 10 miRNAs which are consistently differentially expressed in CRC *vs.* normal tissues. By using cox proportional hazard models, we then developed 6-miRNA-classifier (miR-183, -20a, -21, -195, -139 and -20a) to predict prognosis in clinical cohort, that had significantly superior predictive performance compared to other clinicopathological factors, and could successfully identify high-risk stage II and III CRC patients with poor prognosis [hazard ratio (HR) =2.16; P=0.0048]. In a multivariate analysis, this miRNA-based classifier emerged as an independent prognostic signature for poor DFS.

Conclusions: Our miRNA-based classifier is a reliable predictive tool for determining prognosis in patents with stage II/III CRC, and might be able to identify high-risk patients that are candidates for more targeted personalized clinical management and surveillance.

Keywords: MicroRNA (miRNA); colorectal cancer (CRC); biomarker; prognosis

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Introduction

Colorectal cancer (CRC) is currently the third most common cancer worldwide, with more than one million new cases diagnosed annually. The outcomes of CRC patients in early and late stages are drastically different, with the 5-year survival rates of ~93% for stage I disease and a dismal 8% for stage IV patients. Although 60% of CRC patients with (stages I–III) present with a resectable disease at the time of diagnosis, approximately 40–50% of such patients who undergo curative surgery or another 20–30% that are post-surgically treated with adjuvant chemotherapy, eventually relapse and experience a metastatic disease and

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eventual death (1-3). This clinical challenge highlights the limitation that the current golden standard of Tumor, Node, Metastasis (TNM)-based classification is inadequate at identifying the risk for tumor recurrence, which leads to potential under or over-treatment of a subset of patients with CRC.

At present, post-surgery, 5-fluorouracil (5FU)-based chemotherapy remains the standard of care treatment for some high-risk patients with stage II disease, and all patients with stage III CRC, as it helps improve survival rates by 10-20% (4,5). For stage II patients who present with specific high-risk clinical features, including advanced T stage, low differentiation grade, tumor perforation and few examined lymph nodes, are generally offered 5FUbased adjuvant treatment. Among these, ~20% of stage II patients that are deemed low-risk clinically, experience tumor relapse. On the other hand, for stage III patients, 30-40% of patients do not show any evidence for tumor recurrence in 5 years even when left untreated, while ~40% patients that receive adjuvant treatment still experience tumor recurrence and eventually die, highlighting the need for more intensive chemotherapy or the potential use of novel targeted therapies (6,7). Taken together, these data underscore the need for identification of novel and robust prognostic biomarkers that can better guide treatment decisions in CRC patients with stage II and III disease.

Although in the recent years several studies have reported potential gene-expression based prognostic biomarkers for stage II/III patients, their adoption and routine use in the clinics have been hampered due to the need for high specimen quality and the lack of consensus and difficulties with analytical approaches. In this regard, microRNAs (miRNAs) have recently emerged as promising substrates for development of prognostic biomarkers in cancers, including CRC. MiRNAs are short (18-22 nt in length) and evolutionarily conserved non-coding RNAs. Compared to mRNAs or proteins, miRNAs are relatively immune to degradation by RNAses, and hence be readily detected and accurately quantified in a variety of clinical specimens including fresh frozen and formalin-fixed paraffinembedded (FFPE) tissues. Additionally, miRNAs have emerged as key frontiers in gene regulation due to their ability to regulate a broad range of biological processes in various human diseases, particularly cancer⁸. We and others have previously highlighted that specific miRNAs may contribute to CRC pathogenesis, and many of these may serve as biomarkers for diagnosis, prognosis and metastasisprediction in CRC patients (8-11).

However, since the clinical usefulness of miRNAs in predicting the prognosis of stage II/III CRC patients remains unclear, we envisaged the present study to address this important gap in knowledge. Accordingly, we performed a systematic and comprehensive identification of CRC-specific miRNAs that are differentially expressed (DE) in stage II/III CRCs, followed by determining their combinatorial efficiency in predicting disease free survival by analyzing their expression in multiple, independent cohorts of patients with CRC.

We present the following article in accordance with the STROBE reporting checklist (available at http://dx.doi. org/10.21037/atm-20-1751).

Methods

Patients and tumor specimens

Clinical specimens analyzed for the miRNA classifier (n=186) were obtained from patients enrolled between 2008 and 2012 at the Shanghai Tenth People's Hospital, Shanghai, China. Patients were staged according to the American Joint Committee on Cancer (AJCC) staging guidelines. Detailed patient information is listed in Table 1. All CRC patients were followed up for survival for at least up to 5 years after surgery. Patients treated with radiotherapy or chemotherapy before surgery were excluded from this study. Personal or family history of polyposis or Lynch syndrome, personal history of inflammatory bowel disease, R1 or R2 resections (microscopic or macroscopic neoplastic involvement of surgical margins, respectively), and cases with lack of available FFPE tissues were also excluded from this study. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013) and was approved by the institutional review boards of Shanghai Tenth People's Hospital, School of Medicine, Tongji University (ID: KN84-01). Informed consent was taken from all the patients.

Biomarker screening phase

We performed an initial biomarker discovery phase wherein we performed an extensive published literature survey for all miRNA expression profiling studies published between 2006 and 2014. We included studies that exclusively performed direct comparison for miRNA expression profiles between normal and CRC tissues. MiRNAs were ranked according to several criteria as described previously (12):

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Variables –	Clinical cohort					
	Number of patients	Low risk	High risk	Р		
Age, years				0.079		
>71	94	53	41			
≤71	92	40	52			
Gender				0.086		
Female	96	49	47			
Male	90	44	46			
Tumor location				0.003**		
Proximal	50	16	34			
Distal	136	77	59			
Tumor size				0.380		
Small	94	44	50			
Large	92	49	43			
Lymph node metastasis				0.092		
Negative	118	60	58			
Positive	68	33	35			
Histological type				0.057		
Well/moderate	158	109	49			
Poor	27	18	9			
Serum CEA				0.380		
Low	84	42	42			
Hiah	84	46	38			

1.

Pearson chi-squared testing was used to compare the correlation between triple-miRNA based classifier and clinical variables. **, P<0.01. CEA, carcinoembryonic antigen.

(I) each miRNA was consistently reported as DE; (II) the direction of expression change (up- or down-regulation) was consistent across all studies; (III) the frequency of miRNA expression alteration was consistent and reported in multiple studies.

Quantitative MiRNA expression analysis

High

The miRNA expression analysis was performed using QuantStudio6 Flex Real-Time PCR System (Applied Biosystems, Foster City, CA, USA). All miRNA TaqMan probes were purchased from Ambion (Austin, TX, USA). The qRT-PCR assays were conducted using TaqMan MicroRNA Reverse Transcription Kit and TaqMan Universal PCR Master Mix kit (Applied Biosystems, Foster City, CA, USA) according to manufacturer's instructions. The relative expression of miRNA was determined by $2^{-\Delta\Delta Ct}$ method using miR-16 as a normalizer, as described previously (9,13-15).

Statistical analysis

All statistical analyses were performed using Medcalc version 12.3, SPSS version 13.0 or GraphPad Prism version 6.0. We conducted receiver operating characteristic (ROC) curves and calculated the area under the ROC curves (AUC) to evaluate the predictive power of candidate miRNAs for prognosticating CRC patients. For the disease-free

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survival (DFS) analysis, we defined the probability that patients remained free of tumor recurrence or death as the first event. Data were analyzed from the date of surgery to the time of the first event or the date on which data were censored, according to the Kaplan-Meier method, and the curves were compared using the log-rank test. To develop a miRNA panel and determining patient survival, we used Cox's proportional hazard regression models and obtained a risk score derived from this prediction model. We categorized patients into high-risk and low-risk group based on median cutoff value. Furthermore, we calculated estimate hazard ratios (HRs) for each miRNA, clinic-pathological variables and combination model, based on univariate and multivariate Cox proportional hazard regression models. All data were expressed as mean \pm standard deviation (SD). We compared two groups using the two-sided χ^2 test for categorical variables. All P values were two-sided, and those less than 0.05 were considered statistically significant.

Results

Systematic discovery and identification of potential miRNAs for prognosis prediction in stage II and III CRC patients

During the past decade, several miRNAs with a prognostic potential for CRC patients have been identified, but majority of them have failed to validate across different studies. In order to avoid bias in selection of prognostic candidate miRNAs reported in previous studies, we initially performed a comprehensive and systematic literature review to identify most frequently and consistently reported miRNAs that are DE between CRC and normal tissues (Figure 1). Performing an exhaustive search of miRNA profiling studies, we identified a panel of 60 up-regulated and 41 down-regulated miRNAs that showed consistent data and were reported in at least 3 independent studies (Table S1). In order to narrow down this list further, we thereafter selected DE-miRNAs consistently reported in ≥10 studies and gathered 10 miRNAs (miR-20a, -31, -183, -182, -21, -17, -145, -139, -195 and -215) which were significantly DE in CRCs, implicating their important role in the development of this disease, and their potential relevance in determining the clinical outcome of stage II/III CRC patients.

Development of a prognostic miRNA classifier to predict survival in stage II/III patients

We subsequently enrolled a clinical cohort of 186 patients

to determine the optimal miRNA combinations for survival prediction. Accordingly, we measured expression level of each miRNA in CRC tissues, and the expression data was normalized by Z score transformation, allowing the comparison of different cohort independent of the original signal intensities. Cox proportional hazard models were used to build a prognostic classifier. Of note, we adopted the back-step elimination algorithm to exclude non-significant confounders and thereafter identified miR-183, miR-21, miR-20a, miR-139 and miR-195 was the optimal combination for survival prediction. We then derived a formula to calculate the risk score for their risk of disease recurrence for every patient based on their individual six miRNA expression levels, where the risk score = $(-1.2681 \times \text{miR}-139) + (-0.8916 \times \text{miR}-145)$ + (0.7084 × miR-183) + (1.4509 × miR-195) + (0.9662 × miR- $20a) + (-1.5493 \times miR-21).$

Performance evaluation of the miRNA-classifier in a clinical cobort of stage II/III CRC patients

We performed ROC analysis to evaluate the prediction accuracy of individual miRNAs and 6-miRNA classifier between DFS and recurrence/death. As shown in *Table 2*, our 6-miRNA-classifier (AUC: 0.705) significantly improved prediction ability of individual miRNA (AUC range, 0.530–0.643). Furthermore, compared to known clinicopathological risk factors, such as poor differentiation, lymph node metastasis, and tumor location, our newly developed miRNA-based classifier revealed the highest predictive accuracy (AUC =0.715; *Figure 1B*).

When we assessed the distribution of each patient's survival status and risk scores generated by this miRNAbased classifier, patients with lower risk scores showed better outcomes vs. those with higher risk scores (Figure 1C), highlighting its high predictive accuracy (Figure 2A). Based on cutoff value (the median value of all patients' risk scores), we divided patients into high-risk group and low-risk group and noted that high-risk group had worse prognosis compared to patients in the low-risk group (HR =2.16, P=0.0048; Figure 2B). Furthermore, we observed high risk scores have strong tendency in association with proximal tumor (P=0.003), lymph node metastasis (P=0.092) and poor differentiation (P=0.057, Table 1). In the univariate analysis, this 6-miRNA-classifier emerged as the strongest predictor of DFS (HR =2.1604, P=0.0059) compared to other clinicopathological variables such as serum CEA (HR =1.8134, P=0.0384), lymph node metastasis (HR =1.5021, P=0.1227) and tumor differentiation (HR =1.0408,



Figure 1 Overview of the study design and receiver operating characteristic (ROC) curves for the comparison of the prognostic accuracy of miRNA-classifier and clinicopathological variables in clinical cohort. (A) The overview of the study design. We performed a systematic and comprehensive discovery step to identify differentially expressed miRNAs in colorectal cancer (CRC). We subsequently determined the prognostic relevance of these miRNAs in stage II/III patients using qRT-PCR and developed a miRNA-based classifier for predicting disease-free survival (DFS) in a clinical cohort (n=186), which was later validated in an independent cohort (n=192). (B) The ROC analysis was used for the discrimination between disease free and recurrence or death cases. (C) The distribution of each patient's risk scores and survival status (recurrence or death). AUC, area under curve; DE, differentially expressed.

P=0.9125). Consistently, multivariate analysis revealed that this 6-miRNA classifier was an independent prognostic factor in stage II/III CRC patients (HR =2.5727, P=0.0015, *Table 3*).

When stratified by tumor stage, this miRNA-classifier still demonstrated clinically and statistically significant predictive power. As depicted in *Figure 2B,C*, the AUC of miRNA-classifier is 0.73 and 0.67 in stage II and stage III respectively. In consistent, stage II patients with higher *vs.* lower risk scores had poor prognosis (HR =2.09, P=0.049). When we analyzed the subset of stage III patients separately, the 6-miRNA-classifier also showed to be a highly predictive prognostic indicator, wherein patients in the high-risk group were more likely to have a poor outcome

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Table 2 The area under a ROC curve (AUC) of individual miRNAs and 6-miRNA classifier for disease free survival analysis in the clinical validation cohort

MIDNA	Clinical cohort			
	AUC	SE	95% CI	
miR-139	0.643	0.0465	0.570–0.712	
miR-145	0.585	0.0490	0.511–0.657	
miR-183	0.625	0.0437	0.551–0.695	
miR-195	0.575	0.0502	0.501–0.647	
miR-20a	0.626	0.0447	0.553–0.696	
miR-21	0.530	0.0456	0.455–0.603	
miR-17	0.608	0.0460	0.533–0.678	
miR-182	0.633	0.0438	0.560–0.703	
miR-215	0.542	0.0473	0.467–0.615	
miR-31	0.559	0.0460	0.485–0.632	
6-miRNA classifier	0.705	0.0429	0.634–0.770	

ROC, receiver operating characteristic; SE, standard error; CI, confidence interval.

vs. those with low-risk (HR =2.26, P=0.041; Figure 2D). Notably, the high-risk stage II group yielded similar survival curves as stage III patients (HR =0.91, P=0.7463; Figure 2E), suggesting our classifier is able to identify high risk stage II group which has same prognosis as stage III group. Collectively, these results indicate that our newly developed 6-miRNA-classifier could successfully segregate high vs. low-risk patients with stage II/III disease. Which highlighting that our 6-miRNA based classifier is indeed a promising and reliable prognostic tool for identifying high-risk stage II and stage III patients, which has important implications for their clinical management.

Discussion

In this study, we have firstly performed a systematic discovery step, followed by development and validation of a novel prognostic tool based on a miRNA-classifier aimed at improving the predictive potential for the clinical outcomes of stage II/III CRC patients following surgery. Based upon a logical discovery, clinical validation steps, we provide data that our triple-miRNA based classifier was able to successfully discriminate high *vs.* low-risk CRC patients with a better predictive performance compared to the currently used TNM classification based clinicopathological variables used for determining therapeutic decision-making in stage II/III patients with CRC.

Although several studies have recently suggested that assessment of gene or protein expression changes may be used for prognostication of stage II/III CRC patients, methodological standardization including tissue handling, RNA or DNA processing, and lack of stability of these analytes have hampered the adoption of these biomarkers in routine clinical settings. In contrast, measurement of expression alterations of miRNAs offers several distinct advantages for their clinical use as biomarkers as these short non-coding RNA genes are highly stable in a variety of clinical specimens, have important functional role in regulating gene expression of key cancer-related genes, and their expression can be very accurately measured using simple PCR-based analytical tools. In view of these salient features of miRNAs, in this study we aimed to develop a miRNA-based predictive model for improved prognostication of stage II/III CRC patients.

In order to identify prognosis-related miRNAs for stage II/III CRC patients, we first selected robustly and DE miRNAs between cancer and normal tissues. We hypothesized that aberrant expression of these miRNAs may directly correlate with prognosis in stage II/III patients. Based upon a discovery step involving systematic literature review for miRNA expression profiling studies, we identified several DE miRNAs such as miR-145 (16-18), miR-21 (19,20), miR-17 (21) and miR-20a (22). We subsequently measured expression level of 10 candidate miRNAs, which



Figure 2 The prediction performance of 6-miRNA-classifier in the clinical cohort. (A) The predictive power of 6-miRNA-classifier was demonstrated in stage II/III patients by ROC analysis. (B,C,D,E) All the patients were divided into low and high-risk group based on risk scores calculated from 6-miRNA-classifier. The Kaplan-Meier analysis was used to estimate the prognosis of low and high-risk group in stage II and stage III. *, P<0.05; **, P<0.01. HR, hazard ratio; ROC, receiver operating characteristic; AUC, area under curve; DFS, disease-free survival.

were reported to be differentially altered in colorectal *vs.* normal tissues at least 10 or more studies. We thereafter derived predictive models by using Cox's regression model,

and identified a 6-miRNA-classifier consisting of miR-183, miR-145, miR-20a, miR-21, miR-195 and miR-139, which was significantly superior in its prognostic accuracy

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	Univariate analysis			Multivariate analysis		
	HR	95% CI	Р	HR	95% CI	Р
Age (>71 <i>vs.</i> ≤71 years)	1.6310	0.9636-2.7604	0.0684	-	-	-
Gender (female vs. male)	0.8628	0.5139–1.4484	0.5765	-	-	-
Tumor location (proximal vs. distal)	1.3471	0.7782-2.3316	0.2872	-	_	-
Tumor size (large <i>vs.</i> small)	1.0200	0.9000-1.1560	0.7568	-	_	-
Lymph node metastasis (pos vs. neg)	1.5021	0.8961-2.5179	0.1227	-	_	-
Differentiation (poor vs. well/mod)	1.0408	0.5104-2.1224	0.9125	-	_	-
Serum CEA (high vs. low)	1.8134	1.0324–3.1853	0.0384*	1.9074	1.0854–3.3519	0.0248*
6-miRNA-classifier (high vs. low)	2.1604	1.2479–3.7401	0.0059**	2.5727	1.4333–4.6179	0.0015**

Table 3 Univariate and multivariate association for the 6-miRNA-classifier and other clinicopathological characteristics with disease-free survival

*, P<0.05; **, P<0.01. CEA, carcinoembryonic antigen; HR, hazard ratio; CI, confidence interval.

compared to the expression of individual miRNAs.

The biological function of these identified miRNAs selected for our classifier has been investigated previously. MiR-183 is a member of miR-183 cluster, which is comprised of miR-183, -182 and 96. The miR-183 family is reported to be highly expressed in CRC and exerts its oncogenic activity via inhibition of several tumor suppressor genes (23). Furthermore, miR-183 was reported to be associated with poor prognosis in CRC patients (24). MiR-195, is a tumor suppressor, since its overexpression results in the inhibition of proliferation and metastasis in various cancers (25-28). MiR-195 is downregulated in CRC and its reduced expression associates with poor prognosis (29,30). MiR-139 was shown to be down-regulated in a stagedependent manner in CRC, and regulates the expression of several oncogenes such as NOTCH1 (31), IGF1R (32,33), MAPK, NF-KB, and STAT3 (34). MiR-145 (35,36) and miR-21 (37) are well-known miRNAs which function as tumor suppressor and onco-miR in CRC. MiR-20a was also reported to promote tumor development through suppression of GABBR1 (38). Considering the functional role as well as the clinical significance of these miRNAs in the development of cancer, it is rational to evaluate their expression in a miR-classifier for predicting prognosis of CRC patients.

With regards to potential limitations, our current study is retrospective in nature, and our results must be validated in future, prospective, multi-center clinical trials. In addition, some of the clinical parameters such as vascular invasion or number of analyzed lymph nodes were not consistently recorded or evaluated in our retrospective cohorts, which may be easier to address in a future well-defined patient cohort.

In conclusion, we provide compelling evidence that our newly developed miRNA-based prognostic classifier tool can effectively stratify patients with stage II/III CRCs into high and low risk groups based upon clinical outcomes, thereby adding significant prognostic value to the currently used clinicopathological risk factors used for such purposes. If validated in future studies, such a miRNA classifier potentially offers tremendous clinical value in directing personalized treatment regimens and clinical management of patients with stage II/III CRC.

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Table S1 Differential expressed miRNAs reported in at least 3 expression profiling studies (up: n=60, down: n=41)

miRNA		Accession number	Mature sequence (5'→3')	Studies [suppl. ref. (23,39-58)]	Expression
Name	Symbol	(miRBase)			·
hsa-miR-20a-5p	miR-20a	MIMAT0000075	uaaagugcuuauagugcagguag	12 (39,40,42-45,47,49,54-57)	Up
hsa-miR-183-5p	miB-183	MIMAT0000261		12(39,40,42,43,46-49,51,53,54,57) 12(39,43,46-49,51,54,57)	Up
hsa-miR-182-5p	miR-182	MIMAT0000259	uuuggcaaugguagaacucacacu	12 (39-43,46-49,51,54)	Up
has-miR-21-5p	miR-21	MIMAT0000076	uagcuuaucagacugauguuga	11 (39,40,43,45-47,49,51,54,55,57)	Up
hsa-miR-17-5p	miR-17	MIMAT0000070	caaagugcuuacagugcagguag	10 (39,40,42,43,45,47,49,51,54,55)	Up
hsa-miR-224-5p	miR-224	MIMAT0000281	caagucacuagugguuccguu	9 (39,40,42,44,46,49,51,56,57)	Up
hsa-miR-106a-5p	miR-106a	MIMAT0000103	aaaagugcuuacagugcagguag	9 (39,40,43,47,49,51,55-57)	Up
hsa-miR-19a-3p	miR-19a	MIMAT0004490	aguuuugcauaguugcacuaca	9 (39,40,43,45,47,49,51,54,57)	Up
hsa-miR-92a-3p	miR-92a	MIMAT0000092	uauugcacuugucccggccugu	8 (40,42,45,47,51,55-57)	Up
nsa-miR-90-5p hsa-miR-203a-3p	miR-96	MIMAT000095		8 (39,40,43,44,46,47,50,57)	Up
hsa-miR-135b-5p	miR-205a	MIMAT0000258		8 (43 46 47 50 52 53 56 57)	Up
hsa-miR-18a-5p	miR-18a	MIMAT0000072	uaaggugcaucuagugcagauag	8 (39,42,43,46,48,49,51,56)	Up
hsa-miR-93-5p	miR-93	MIMAT0000093	caaagugcuguucgugcagguag	7 (39,41,43,47,48,54,55)	Up
hsa-miR-95-3p	miR-95	MIMAT0026473	исааиаааидисидиидааии	7 (39,40,42,53,55-57)	Up
hsa-miR-106b-5p	miR-106b	MIMAT0000680	uaaagugcugacagugcagau	7 (39,41,45,47,48,55,56)	Up
hsa-miR-522-3p	miR-522	MIMAT0002868	aaaaugguucccuuuagagugu	6 (43,44,47,48-50)	Up
hsa-miR-1246	miR-1246	MIMAT0005898	aauggauuuuuggagcagg	6 (44,45,48,49,51,53)	Up
hsa-miR-130b-3p	miR-130b	MIMAT0000691	cagugcaaugaugaaagggcau	6 (39,40,46,48,51,54)	Up
hsa-miR-148a-3p	miR-148a	MIMAT0001412		6 (23,40,43,44,54,57) 6 (41,42,47,49,51)	Up
hsa-miR-17-3p	miR-17*	MIMAT0001412	acuacaauaaaaacacuuduaa	5 (40 43 47 49 57)	Up
hsa-miR-183-3p	miR-183*	MIMAT0004560	gugaauuaccgaagggccauaa	5 (44,45,47,52,53)	Up
hsa-miR-221-3p	miR-221	MIMAT0000278	agcuacauugucugcuggguuuc	5 (23,40,45,54,56)	Up
hsa-miR-503-5p	miR-503	MIMAT0002874	uagcagcgggaacaguucugcag	5 (44,46,48,49,51)	Up
hsa-miR-181b-5p	miR-181b	MIMAT0000257	aacauucauugcugucggugggu	5 (39,40,45,55,57)	Up
hsa-miR-34a-5p	miR-34a	MIMAT0000255	uggcagugucuuagcugguugu	5 (39-41,55,57)	Up
hsa-miR-15a-5p	miR-15a	MIMAT0000068	uagcagcacauaaugguuugug	5 (23,40,43,54,57)	Up
has-miR-21-3p	miR-21*	MIMAT0004494	caacaccagucgaugggcugu	4 (44,48,49,51)	Up
hsa-miR-25-3p	miR-25	MIMAT0000081	cauugcacuugucucggucuga	4 (39,40,43,49)	Up
hsa-miR-210-3p	miR-210	MIMAT0000267	cugugcgugugacagcggcuga	4 (40,48,55,57)	Up
hsa-miR-223-3p	miR-223	MIMAT0000280	ugucaguuugucaaauacccca	4 (43,47,55,56)	Up
hsa-miR-429	miR-429	MIMAT0001536	uaauacugucugguaaaaccgu	4 (44,45,49,51)	Up
hsa-miR-584-5p	miR-584	MIMAT0003249	uuaugguuugccugggacugag	4 (46-48,50)	Up
hsa-miR-1247-5p	miR-1247	MIMAT0005899	acccgucccguucguccccgga	4 (41,44,50,53)	Up
hsa-miR-29a-3p	miR-29a	MIMAT0000086	uagcaccaucugaaaucgguua	4 (39,40,42,57)	Up
hsa-miR-338-3p	miR-338	MIMAT0000763		4 (23,40,44,55)	Up
hsa-miR-135a-5p	miR-135a	MIMAT0000428		4 (23,40,55,57)	Up
hsa-miR-20b-5p	miR-20b	MIMAT0000232	caaaquqcucauaquqcaqquaq	3 (23 45 49)	Up
hsa-miR-98-5p	miR-98	MIMAT0000096	uqaqquaquaaquuquauuquu	3 (40.43.54)	Up
hsa-miR-105-5p	miR-105	MIMAT0000096	ugagguaguaguuguauuguu	3 (40,44,50)	Up
hsa-miR-128-3p	miR-128	MIMAT0000424	ucacagugaaccggucucuuu	3 (23,40,48)	Up
hsa-miR-182-3p	miR-182	MIMAT0000260	ugguucuagacuugccaacua	3 (40,53,57)	Up
hsa-miR-185-5p	miR-185	MIMAT0000455	uggagagaaaggcaguuccuga	3 (23,47,55)	Up
has-miR-198	miR-198	MIMAT0000228	gguccagagggggagauagguuc	3 (44,47,53)	Up
hsa-miR-222-3p	miR-222	MIMAT0000279	agcuacaucuggcuacugggu	3 (40,55,56)	Up
hsa-miR-320a	miR-320a	MIMAT0000510	aaaagcuggguugagagggcga	3 (23,40,47)	Up
hsa-miR-339-5p	miR-339	MIMAT0000764	ucccuguccuccaggagcucacg	3 (40,43,51)	Up
hsa-miR-346	miR-346	MIMAT0000773	ugucugcccgcaugccugccucu	3 (47,53,55)	Up
has-mir-421	mir-421	MIMAT0003339	aucaacagacauuaauugggcgc	3 (43,48,49)	Up
hsa-miR-424-3p	miR-424	MIMAT0004749	caaaacgugaggcgcugcuau	3 (41,44,49)	Up
hsa-miR-452-5p	miR-452	MIMAT0001635	aacuguuugcagaggaaacuga	3 (43,46,47)	Up
hsa-miR-645	miR-645	MIMAT0003315	ucuaggcugguacugcuga	3 (23,47,53)	Up
hsa-miR-663b	miR-663b	MIMAT0005867	gguggcccggccgugccugagg	3 (44,48,53)	Up
has-miR-142-3p	miR-142"	MIMAT0000434		3 (40,42,54)	Up
hsa-miR-29b-3p	miR-29b	MIMAT0000100		3 (39,43,57)	Un
hsa-miR-196b-5p	miR-196b	MIMAT0001080	uagguagguagguagguagg	3 (42.43.51)	Up
hsa-miR-145-5p	miR-145	MIMAT0000437	guccaguuuucccaggaaucccu	14 (39,40,42,43,45-47,49-53,57,58)	Down
hsa-miR-139-5p	miR-139	MIMAT0000250	ucuacagugcacgugucuccagu	13 (39,40,43,44,46,48-54,57)	Down
hsa-miR-195-5p	miR-195	MIMAT0000461	uagcagcacagaaauauuggc	11 (39,40,44,46-53)	Down
hsa-miR-215-5p	miR-215	MIMAT0000272	augaccuaugaauugacagac	10 (44-49,52,54,55,58)	Down
hsa-miR-378a-5p	miR-378a	MIMAT0000731	cuccugacuccagguccugugu	9 (43,44,46,47,49-53)	Down
hsa-miR-422a	miR-422a	MIMAT0001339	acuggacuuagggucagaaggc	9 (39,44,46,48-52,54)	Down
hsa-miR-143-3p	miR-143	MIMAT0000435	ugagaugaagcacuguagcuc	8 (39,42,47,49,51-53,21)	Down
hsa-miR-30a-5p	miR-30a	MIMAT0000087	uguaaacauccucgacuggaag	8 (39,44,46-49,53,21)	Down
hsa-miR-30a-3p	miR-30a*	MIMAT0000088	cuuucagucggauguuugcagc	8 (39,40,44,49,52,53,55,57)	Down
hsa-miR-1-3p	miR-1	MIMAT0000416	uggaauguaaagaaguauguau	7 (39,46,47,50,53-55)	Down
hsa-miR-149-5p	miR-149			(40,44,49,51,53,54,57)	Down
nsa-miH-138-5p	тін-138		agcugguguugugaaucaggccg	(44,45,47,49,51,53)	Down
наэ-шіп-342-3р hsa-miP-100 5~	miR-100			r (44,47,48,01-03,00) 6 (47-49 52 52 57)	Down
hsa-miR-150-5p	miR-150	MIMAT0000451	UCUCCCAACCCUUduaccadud	6 (41,43,44,47-49)	Down
hsa-miR-497-5p	miR-497	MIMAT0002820	cagcagcacacuauaauuuau	6 (42,44,46,48,49.53)	Down
hsa-miR-10b-5p	miR-10b	MIMAT0000254	uacccuguagaaccgaauuuquq	6 (39,44,47,49,51,53)	Down
hsa-miR-125a-5p	miR-125a	MIMAT0000443	ucccugagacccuuuaaccuguga	6 (39,41,43,47,49,58)	Down
hsa-miR-30c-5p	miR-30c	MIMAT0000244	uguaaacauccuacacucucagc	6 (39,47,51-53,57)	Down
hsa-miR-133a-3p	miR-133a	MIMAT0000427	uuugguccccuucaaccagcug	6 (39,47,49,53,54,57)	Down
hsa-miR-375	miR-375	MIMAT0000728	uuuguucguucggcucgcguga	5 (43,47,49,52,54)	Down
hsa-miR-125b-5p	miR-125b	MIMAT0000423	ucccugagacccuaacuuguga	5 (39,43,47,53,58)	Down
hsa-miR-133b	miR-133b	MIMAT0000770	uuugguccccuucaaccagcua	5 (23,43,47,53,57)	Down
hsa-miR-99a-5p	miR-99a	MIMAT0000097	aacccguagauccgaucuugug	5 (44,49,51-53)	Down
hsa-miR-124-3p	miR-124	MIMAT0000422	uaaggcacgcggugaaugcc	4 (44,47,53,57)	Down
hsa-miR-137	miR-137	MIMAT0000429	uuauugcuuaagaauacgcguag	4 (46,53,57,58)	Down
hsa-miR-186-5p	miR-186	MIMAT0000456	caaagaauucuccuuuugggcu	4 (41,47,52,53)	Down
hsa-miR-147b	miR-147b		gugugcggaaaugcuucugcua	4 (43,46,47,53)	Down
hsa-miR-26a-5p	miR-26a		uucaaguaauccaggauaggcu	4 (47,52,53,58)	Down
nsa-miR-29c-3p	miR-290		uagcaccauuugaaaucgguua	4 (43,45,47,53)	Down
наз-шін-140-3р has-miP-29-25	miR-22			4 (44,43,31,33) A (AA AQ 51 5A)	Down
haə-miR-486-5ρ	miR-486	MIMAT0002177	uccuguacugageugeeegaa	न (नन,नव,वन,वन) 4 (50-53)	Down
hsa-miR-204-5p	miR-204	MIMAT0002177		4 (53 54 57 58)	Down
hsa-miR-454-5p	miR-454		accoualicaatiauuduotieude	3 (23 48 53)	Down
hsa-miR-101-30	miR-101	MIMAT000009	Uacaguacuguaguaguaga	3 (47,52,53)	Down
hsa-miR-192-5p	miR-192	MIMAT0000222	cugaccuaugaauugacagacc	3 (47,52,55)	Down
hsa-miR-383-5p	miR-383	MIMAT0000738	agaucagaaggugauuguqqcu	3 (44,50,53)	Down
hsa-miR-585-3p	miR-585	MIMAT0003250	ugggcguaucuguaugcua	3 (46,48,53)	Down
hsa-miR-141-3p	miR-141	MIMAT0000432	uaacacugucugguaaagaugg	3 (23,47,53)	Down
hsa-miR-187-3p	miR-187	MIMAT0000262	ncanancnnanannacsaccaa	3 (47,53,57)	Down







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