

Experimental study: using the continuous wave Doppler spectrum technique to detect left atrial pressure

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Background: This study sought to investigate the accuracy of estimating left atrial pressure (LAP) using the continuous wave Doppler spectrum of mitral regurgitation.

Methods: Dog models of left atrial hypertension with mitral regurgitation were established with disposable biopsy forceps and the injection of melamine formaldehyde resin microsphere suspension. A total of 40 models of left atrial hypertension with different hemodynamic statuses were established by injecting either esmolol or dobutamine in which the spectrums of mitral regurgitation were clear and the regurgitation velocity exceeded 3.5 m/s. The continuous wave Doppler spectrums of mitral regurgitation were recorded and analyzed to estimate left atrial pressure (LAP_{ECHO}). The mean left atrial pressure (LAP_{C-MEAN}), the isovolumic diastolic left atrial pressure (LAP_{C-MEAN}), were also measured using the catheter method in the same cardiac cycle.

Results: The LAP_{ECHO} (mean \pm standard deviation; 11.77 \pm 4.36 mmHg) was correlated with the LAP_{C-MEAN} (11.51 \pm 4.77 mmHg; r=0.887, P=0.000), but the difference was not statistically significant (P=0.459). The LAP_{ECHO} was correlated with the LAP_{C-IVRT} (12.16 \pm 4.72 mmHg; r=0.883, P=0.000), but the difference was not statistically significant (P=0.271). There was a correlation between the LAP_{C-MEAN} and the LAP_{C-IVRT} (r=0.987, P=0.000), and the difference was statistically significant (P=0.471).

Conclusions: This study suggests that the ultrasound evaluation of LAP correlates well with LAP measured using the gold standard catheter method, and is a simple, convenient, non-invasive method to quantitatively estimate LAP. This method is promising, but further large-scale animal experiments and clinical studies need to be conducted.

Keywords: Left atrial pressure (LAP); continuous wave Doppler spectrum; mitral regurgitation; left atrial pressure measured by ultrasound (LAP_{ECHO}); LAP_C

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Introduction

Left atrial pressure (LAP) is an important hemodynamic parameter of cardiac function (1). At present, LAP is mainly measured using the catheter method, or pulmonary artery wedge pressure is measured rather than LAP. These methods are invasive, and cannot be routinely used in clinical practice. Currently, a variety of non-invasive methods for evaluating LAP, such as the "Doppler + sphygmomanometer" method have been explored. Due to their inaccuracy and complexity, these methods have not been used in clinical practice (2,3).

In the study of the left ventricular relaxation time constant (τ), based on the Weiss exponential equation (P = e^{-t/\tau + B}), the simplified Bernoulli equation ($\Delta P = 4V^2$), and P = ΔP + LAP, Bai (4-6) used mathematical methods to derive "binary linear equations", which can be used to calculate τ , and have been verified by animal experiments (7). The equations have 2 variables; that is, τ and LAP. If τ is eliminated, LAP can be calculated. This method has some advantages. The first is simplicity. It only needs to measure the two intervals of the descending branch of the mitral regurgitation spectrum and substitute them into the above equations. The second is accuracy. The equations are used in the deduction process, thus, the integrity of the theory was preserved (7).

In this study, we created a Beagle dog model of acute mitral regurgitation combined with impaired diastolic function to increase the LAP, and then recorded a continuous wave Doppler spectrum of mitral regurgitation to calculate the LAP. The catheter method was used as the "gold standard" to measure LAP to verify the accuracy of this non-invasive ultrasound LAP measurement method.

We present the following article in accordance with the ARRIVE reporting checklist (available at https://atm. amegroups.com/article/view/10.21037/atm-21-6913/rc).

Methods

Experimental animals

All animal experiments were conducted in the Nongnong (Beijing) Life Science and Technology Company. Experiments were performed under a project license (No. 20180826-115) granted by ethics board of Animal Experimental Center of Nongnong (Beijing) Life Science and Technology Company, in compliance with institutional guidelines for the care and use of animals.

A protocol was prepared before the study without registration. 20 healthy beagle dogs were provided by Nongnong (Beijing) Biotechnology Co., Ltd., aged 2.0–3.6 years (2.8±0.7 years) with a body weight of 13.5–23.0 kg (18.2±2.7 kg). There were 8 females and 12 males. The animals were kept in a constant temperature of 18–22 °C with free access to water and food in separate cages. Surgery was performed after 7 days of adaptive feeding. At the end of the procedure, a rapid intravenous injection of 10% potassium chloride at 0.5 mL per kilogram of the body weight was used to euthanize the dogs under anesthesia.

Main reagents and instruments

The Resona7 color Doppler ultrasound system was purchased from Shenzhen Mindray Biomedical Electronics Co., Ltd. (China). The phased array heart probe, probe frequency 2.5 MHz. MP160 type BIOPAC multi-lead physiological recorder was purchased from American BIOPAC company. An ACM608 animal ventilator was purchased from Beijing Aerospace Changfeng Co., Ltd. (China). A WZS-50F6 dual-channel micro-injection pump was purchased from Zhejiang Smith Medical Instrument Co., Ltd. (China). Intravascular catheters (5F, RH*5SP0061M) were purchased from Terumo Medical Products Co., Ltd of Japan. A A70N25A arterial sheath group and GA35183 guide wire were purchased from Japan Termao Medical Products Co., Ltd. Melamine formaldehyde resin microspheres (average particle size 51.7 µm) were purchased from Wuhan Huake Weike Technology Co., Ltd. (China). The configuration method for melamine microsphere suspension was as follow: an injection of 200 mg of melamine microsphere, mixed with 100 mL of sodium chloride, shaken by ultrasonic shaker, and an injection of lidocaine hydrochloride (5 mL:0.1 mg) purchased from Shanghai Hefeng Pharmaceutical Co., Ltd. the company.

Establishing an animal model of mitral regurgitation combined with increased LAP

The Beagle dogs were fasted for 24 hours before surgery. They were anesthetized by an intravenous injection of Propofol (Sichuan Guorui Salt Industry Co., Ltd., China) (20–40 mg/kg), and the skin was prepared on the neck, chest, and inner limbs. The dogs were placed in the supine position on the operating table, the trachea was intubated,

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the animal ventilator was connected, and the dogs were kept warm (7). Electrocardiograms were attached to the upper and lower limbs with electrode pads (7). Venous access was established in the marginal ear vein and maintained with 0.9% sodium chloride via intravenous injection (1-2 drops/s). After anesthesia, the pressure measurement catheter was inserted into the left ventricle via the right femoral artery. The chest was opened with a middle line incision to expose the heart, after which the pericardium was cut, the pericardium hammock sewn, and the heart subsequently fixed. The pressure measuring catheter was inserted into the left atrium through the left atrial appendage, and the catheter was fixed with a purse. The left common carotid artery was dissected and the distal end of the left common carotid artery was ligated, followed by a puncture of the left common carotid artery with a 16 G trocar. A guide wire was then inserted into the 7F arterial sheath, followed by continuation using disposable biopsy forceps to make a mitral regurgitation model. Under ultrasound guidance: (I) the guide wire entered the left ventricle through the left common carotid artery, the arterial sheath was sent into the left ventricle along the guide wire, and the guide wire was then withdrawn; (II) the front end of the arterial sheath was sent to the mitral valve anterior valve tip when the mitral valve was open, the disposable sterile cystoscope biopsy forceps were delivered through the arterial sheath; (III) the biopsy forceps were opened when they reached the level of the tendon below the edge of the anterior leaflet of the mitral valve, and the tendon was clamped when the mitral valve was opened to cause the tendon to break; (IV) ultrasonic observation of light-medium reflux was defined as the successful establishment of the animal model; (V) if unsuccessful, the above procedure was repeated until the model was successfully established, and the biopsy forceps were then withdrawn. The degree of mitral regurgitation was estimated by the Doppler color reflux area as follow: mild—a color reflux area accounting for <20% of the left atrium area; moderate-a color reflux area accounting for 20-40% of the left atrium area; and severe-a color reflux area accounting for >40% of the left atrium area. After the successful creation of the mitral regurgitation model, under the guidance of ultrasound, the arterial sheath was withdrawn into the aortic sinus, and the left common carotid sheath was moved to shift the front end into the left coronary sinus. Melamine formaldehyde resin microsphere suspension was slowly injected into the left coronary artery sinus with 2 mL of microsphere suspension each time,

every 5–10 min, which led to increased left ventricular enddiastolic pressure (LVEDP). The left ventricular pressure (LVP) corresponding to the starting point of the QRS complex of the electrocardiogram represents the LVEDP, and the increase of LVEDP \geq 5 mmHg indicated the successful construction of acute ischemic left ventricular diastolic dysfunction (7,8). To adjust the hemodynamic state of the LAP by different degrees, a pump injection of esmolol hydrochloride was used to reduce myocardial contractility, or a pump injection of dobutamine hydrochloride was used to improve myocardial contractility; hemodynamic stability was regarded as successful modeling. In the end, each dog adjusted two stable cardiac function states through drugs. A multi-lead physiological recorder was used to monitor the electrocardiogram (ECG), LVP, and LAP in real time.

Continuous wave Doppler spectrum acquisition of mitral regurgitation

Echocardiography was performed on the left atrial hypertension models of different hemodynamic states in which the spectrums of mitral regurgitation were clear, and the regurgitation velocity exceeded 3.5 m/s. The continuous wave Doppler spectrums of mitral regurgitation were saved. Because continuous wave Doppler has high velocity resolution, it can detect very high-speed blood flow.

The following acquisition method was used: echocardiographic examination, adjust the probe position and rotate the probe direction on the apical 4-chamber heart and apical left ventricular long-axis section, color Doppler display the continuous wave Doppler sampling lines parallel to the direction of the reflux beam, obtain the best mitral regurgitation spectrum, set the spectral Doppler scan speed to 200 mm/s, and store a continuous wave Doppler spectrum loop of mitral regurgitation (see *Figure 1A*). The physiological instrument simultaneously recorded and stored the electrocardiogram, LVP curve, and the left atrium pressure curve.

Ultrasonic measurement of LAP and catheter measurement

The ultrasonic measurement of LAP was calculated using the method described below.

Formulas

The following formulas were used:

$$\tau = (t1-t2)/\ln((LAP+16)/(LAP+4))$$
[1]



Figure 1 Continuous wave Doppler spectrum groove edge of mitral regurgitation. (A) Continuous wave Doppler spectrum of mitral regurgitation. (B) Automatic envelope of spectrum in "A" by software. FR, frame rate; DR, dynamic range; WF, wall filtering; SVD, sampling volume depth; PRF, pulse repetition frequency.

[2]

 $\tau = (t1 - t3) / \ln((LAP + 36) / (LAP + 4))$

There are 2 variables, τ and LAP, in the formula. The durations of t1–t2 and t1–t3 of the descending branch of the continuous wave Doppler spectrum of mitral regurgitation were substituted, and the LAP was then calculated (5-7).

t1-t2, and t1-t3 measurements

The stored dynamic images of the mitral regurgitation continuous wave Doppler spectrums were analyzed using Mindray Company's patented software. To automatically identify the spectrum boundary, a smooth envelope was fitted and drawn (see *Figure 1B*); the software automatically determined the time values for t1, t2, and t3 of the velocity on the envelope line at 1, 2, and 3 m/s, and calculated the duration of t1-t2, and t1-t3 (7).

LAP_{ECHO} calculation

The research group wrote the binary linear Eq. [1] and Eq. [2] (see above) into software, input t1–t2, t1–t3, which could automatically calculate LAP_{ECHO} . LAP_{ECHO} was calculated for 3 consecutive cardiac cycles, and the average value was selected for the statistical analysis. The software



Figure 2 Physiological curve recorded by a polyconducting physiological recorder. From top to bottom: the ECG, LVP curve, LAP curve, left ventricular pressure dP/dt curve. LAP_{C-MAX}: 17.914 mmHg; LAP_{C-MIN}: 7.385 mmHg; LAP_{C-MEAN}: 11.146 mmHg; LAP_{C-IVKT}: 12.081 mmHg. ECG, electrocardiogram; LAP, left atrial pressure; LAP_{C-MAX}, the maximum left atrial pressure; LAP_{C-MIN}, the minimum left atrial pressure; LAP_{C-MEAN}, the mean left atrial pressure; LAP_{C-WEAN}, the isovolumic diastolic left atrial pressure.

can be downloaded from: https://sourceforge.net/projects/ tauformula/files/.

The catheter measurement of the LAP (LAP_C) adopts the data analysis software (AcqKnowledge4.2) of the multilead physiological recorder to directly measure the mean LAP (LAP_{C-MEAN}), isovolumic diastolic LAP (LAP_{C-IVRT}), maximum LAP (LAP_{C-MAX}), and the minimum LAP (LAP_{C-MIN}) within a cardiac cycle (see *Figure 2*). The measurement method of LAP during isovolumic diastole (after the aortic valve closure and before the mitral valve opening) is to measure the average value of the LAP, which is recorded as LAP_{C-IVRT} . The LAP_{C-MEAN} , and LAP_{C-IVRT} , etc., were measured by catheterization within 3 consecutive cardiac cycles, and the average value was used for the statistical analysis.

Statistical analysis

SPSS 25.0 software was used for the statistical analysis.



Figure 3 Mitral valve tendon rupture specimen. The two arrows indicate the two broken ends of the tendon cut by the biopsy forceps.

Table 1 LAP measured by	iltrasound and	catheter	(mmHg)
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Parameters (n=40)	Ranges	Mean ± SD
LAP _{ECHO}	4.24–22.93	11.77±4.36
LAP _{C-MEAN}	3.68–22.87	11.51±4.77
LAP _{C-IVRT}	3.94–23.54	12.16±4.72
LAP _{C-MAX}	8.94–25.30	15.70±4.36
LA _{PC-MIN}	1.57–20.81	8.33±4.33

LAP, left atrial pressure; LAP_{ECHO}, left atrial pressure measured by ultrasound; LAP_{C-MEAN}, the mean left atrial pressure; LAP_{C-IVRT}, the isovolumic diastolic left atrial pressure; LAP_{C-MAX}, the maximum left atrial pressure; LAP_{C-MIN}, the minimum left atrial pressure.

The measurement data are expressed as mean \pm standard deviation (SD), and the relationship between the LAP_C and LAP_{ECHO} was assessed by a Pearson correlation analysis. The student's *t*-test was used to compare the difference between 2 values. A P value <0.05 was considered statistically significant.

Results

Animal model construction

The Beagle model of mitral regurgitation was successfully established (see *Figure 3*). After the mitral regurgitation model was successfully established, 2 mL of melamine formaldehyde resin microsphere suspension was injected into the left aortic sinus at a rate of 5 mL/20 s every 5– 10 minutes. The LVEDP was increased by >5 mmHg (with an average increase of 17 mmHg), which represented the successful establishment of the acute ischemic left ventricular diastolic dysfunction model. This process took about 90 minutes. After 20 minutes of continuous observation, 40 models of left atrial hypertension with different hemodynamic states were established by injecting either esmolol or dobutamine.

LAP measured by ultrasound and catheter

Table 1 shows the LAP measured by ultrasound (LAP_{ECHO}), mean left atrial pressure (LAP_{C-MEAN}), isovolumic diastolic left atrial pressure (LAP_{C-IVRT}), maximum left atrial pressure (LAP_{C-MAX}), and minimum left atrial pressure (LAP_{C-MIN}).

Relationship between the LAP_{ECHO} and LAP_{C-MEAN}

In this study, the mitral regurgitation spectrum was taken 3 times using the formula described above. t1, t2, and t3 occurred in the isovolumic diastolic phase of the cardiac physiological cycle. Thus, the LAP calculated by ultrasound corresponded to LAP_{C-IVBT}. Clinically, the average LAP is often used as a parameter to determine the size of the LAP. Thus, the correlations among the LAP_{ECHO} , LAP_{C-} IVRTT, and LAP_{C-MEAN} were analyzed separately. The LAP_{C-} MEAN, which is the "gold standard", was selected to verify the accuracy of the LAP_{ECHO} . The LAP_{ECHO} was between 4.24–22.93 (11.77±4.36) mmHg, and the LAP_{C-MEAN} was between 3.68-22.87 (11.51±4.77) mmHg; there was no statistically significant difference between the 2 groups (P=0.459). The correlation between the LAP_{C-MEAN} and LAP_{ECHO} was statistically significant (r=0.887, P=0.000; see Figure 4). In relation to the Bland-Altman method analysis, the average value of the difference between 40 cases of the LAP_{C-MEAN} and LAP_{ECHO} paired data was \overline{d} =-0.26, the SD of the difference was 2.21, and the 95% consistency limit was $\overline{d} \pm 1.96$ SD =-0.26±1.96×2.21; that is (-4.60, 4.07). The 2.5% (1/40) point was outside the 95% consistency limit, indicating that the 2 methods were consistent (see Figure 5).

Relationship between the LAP_{ECHO} and LAP_{C-IVRT}

The LAP_{ECHO} measured by the ultrasound method was between 4.24–22.93 (11.77 \pm 4.36) mmHg, and the LAP_{C-IVRTS} measured by the catheter method was between 3.94–23.54 (12.16 \pm 4.72) mmHg; the difference between the 2 groups was not statistically significant (P=0.271).

The correlation between the LAP_{C-IVRT} and LAP_{ECHO} was statistically significant (r=0.883, P=0.000; see *Figure 6*). In relation to the Bland-Altman method analysis, the average value of the difference between the 40 cases of LAP_{C-IVRT} and

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Figure 4 The correlation between the LAP_{ECHO} and LAP_{C-MEAN}. LAP, left atrial pressure; LAP_{ECHO}, left atrial pressure measured by ultrasound; LAP_{C-MEAN}, the mean left atrial pressure.



Figure 5 The relationship between the LAP_{ECHO} and LAP_{C-MEAN} using the Bland-Altman method. The abscissa is the average of the LAP_{ECHO} and LAP_{C-MEAN}, the difference between the ordinates; the blue line is the average of the difference between the LAP_{ECHO} and LAP_{C-MEAN} (\overline{d}); the red line is ($\overline{d} \pm 1.96$ SD). LAP, left atrial pressure; LAP_{ECHO}, left atrial pressure measured by ultrasound; LAP_{C-MEAN}, the mean left atrial pressure.



Figure 6 The correlation between the $\mathrm{LAP}_{\text{ECHO}}$ and $\mathrm{LAP}_{\text{C-IVRT}}$ LAP_{ECHO}, left atrial pressure measured by ultrasound; LAP_{C-IVRD} the isovolumic diastolic left atrial pressure.

 LAP_{ECHO} paired data was $\overline{d}=0.39$, the SD of the difference was 2.23, and the 95% consistency limit was $\overline{d} \pm 1.96$ SD =0.39±1.96×2.23; that is, (-3.98, 4.76). The 5% (2/40) point



Figure 7 The relationship between the LAP_{ECHO} and LAP_{C-IVRT} using the Bland-Altman method. The abscissa is the average of the LAP_{ECHO} and LAP_{C-IVED}, the difference between the ordinates; the blue line is the average of the difference between the LAP_{ECHO} and LAP_{C-IVRT} (\overline{d}); the red line is ($\overline{d} \pm 1.96$ SD). LAP_{ECHO}, left atrial pressure measured by ultrasound; LAP_{C-IVRT}, the isovolumic diastolic left atrial pressure.

was outside the 95% consistency limit, and the 2 methods were considered to have good consistency (see Figure 7).

The relationship between the LAP_{C-MEAN} and LAP_{C-IVRT}

In clinical practice, the average LAP is usually used as an indicator for evaluating cardiac function. The LAP calculated by the formula in this study corresponded to the isovolumic diastolic LAP. The LAP_{C-MEAN} was between 3.68–22.87 (11.51±4.77) mmHg, the LAP_{C-IVRT} was between 3.94-23.54 (12.16±4.72) mmHg, and the difference was statistically significant (P=0.000). The analysis between the LAP_{C-MEAN} and LAP_{C-IVRT} showed a significant correlation (r=0.987, P=0.000; see Figure 8). In relation to the Bland-Altman method analysis, the average value of the difference between the paired data of the LAP_{C-MEAN} and LAP_{C-IVRT} was \overline{d} =-0.65, the SD of the difference was 0.76, and the 95% consistency limit ($\overline{d} \pm 1.96$ SD) was -0.65±1.96×0.76; that is, (-2.14, 0.83). The 10% (4/40) point was outside the 95% consistency limit, and the consistency between the 2 methods was considered good (see Figure 9).

Discussion

Following the wide application of ultrasound technology in clinical settings, some scholars have studied the method of using ultrasound technology to evaluate the LAP. "Doppler + sphygmomanometer" is the most commonly used method to study the LAP. In patients with mitral

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Figure 8 The correlation between the LAP_{C-MEAN} and LAP_{C-IVRT} . LAP_{C-MEAN}, the mean left atrial pressure; LAP_{C-IVRT} , the isovolumic diastolic left atrial pressure.



Figure 9 The relationship between the LAP_{C-MEAN} and LAP_{C-IVRT} using the Bland-Altman method. The abscissa is the average of the LAP_{C-MEAN} and LAP_{C-IVRT}, the difference between the ordinates; the blue line is the average of the difference between LAP_{C-MEAN} and LAP_{C-IVRT} (\overline{d}); the red line is ($\overline{d} \pm 1.96$ SD). LAP_{C-MEAN}, the mean left atrial pressure; LAP_{C-IVRT}, the isovolumic diastolic left atrial pressure.

regurgitation, the maximum transvalvular pressure difference (ΔP) of mitral regurgitation is the difference between left ventricular systolic pressure (P) and LAP; thus, the LAP can be calculated as P minus mitral regurgitation pressure difference; that is, LAP = P – ΔP . In patients without left ventricular outflow tract stenosis, P is similar to brachial artery systolic pressure, and can be measured with a sphygmomanometer. The peak velocity of mitral regurgitation measured by continuous Doppler spectrum can be calculated automatically using the following formula: $\Delta P = 4V^2$ (3). However, using the cuff method, measurement errors of brachial artery systolic pressure are common, especially when there is calcification in the brachial artery. This affects the accuracy of LAP measurements.

In the 20th century, many scholars systematically observed and studied the relationship between pulmonary vein blood flow and LAP (9,10), and found that patients with high LAP changed from the systolic pulmonary vein blood flow spectrum to diastolic pulmonary vein blood flow spectrum. Such qualitative evaluation studies cannot measure LAP quantitatively. There are many types of pulmonary vein blood flow spectrums. The spectrum types are easily affected by LAP, left ventricular systolic function, diastolic function, heart rate, age, etc., which affect the accuracy and effectiveness of ultrasound evaluations of LAP. In addition, it is sometimes difficult to collect negative phase waves of atrial contraction by transthoracic ultrasound.

Some scholars have explored the use of mitral valve positive flow spectrum to evaluate LAP, suggesting that there is a certain correlation between them (11). However, the accuracy of this method is poor due to factors such as left ventricular relaxation, ventricular compliance, left ventricular preload, ventricular interaction, pericardial limitation, and left atrial function (12).

Tissue Doppler imaging can quantify myocardial wall motion amplitude, early diastolic mitral annulus tissue Doppler velocity (E'), and the left ventricular relaxation time constant (τ). Some scholars have used the ratio of early diastolic mitral flow velocity (E) to early diastolic mitral annular tissue Doppler velocity (E/E') to evaluate LAP. The calculation formula of LAP is LAP = $1.24 \times (E/e') + 1.9$ (12). However, relevant research results differ, and some scholars believe that it has only a moderate correlation compared to the catheter method, while other scholars say that the correlation is poor or there is no correlation (13-15).

Bai derived a novel method to measure LAP by continuous wave Doppler spectrum (7,16). This method measures the blood flow velocity at 1, 2, and 3 m/s at times t1, t2, and t3 at the descending branch of the mitral regurgitation curve, calculates the duration of t1–t2, and t1–t3, and then estimates the LAP. In the process of formula derivation, 3 points (t1, 1 m/s), (t2, 2 m/s), (t3, 3 m/s) were selected for the calculation. This method of taking integers is convenient for the calculation; the 3 points are always taken in the isovolumic relaxation phase (6).

LAP is composed of systolic and diastolic phases, and in the clinic, average pressure is usually used. There was a significant difference between the LAP_{C-MEAN} and $LAP_{C-IVRTP}$. The isovolumic diastolic phase was closed after the aortic valve was closed, but before the mitral valve was opened; the isovolumic diastolic phase LAP curve was relatively straight, located in the ascending branch of the v wave, close to the v wave peak, and higher than the average pressure of the left

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atrium. There were obvious differences between the LAP_{C-MEAN} and LAP_{C-IVRT} , and the LAP_{IVRT} was slightly larger than the LAP_{C-MEAN} .

In this study, we used animal experiments to verify a new method for noninvasive quantitative measurement of LAP. The method was non-invasive and simple, and had a good correlation with catheter measurements. The method does not require systolic blood pressure measurements, but only needs a clear continuous Doppler spectrum of mitral regurgitation. Based on current advances in computer and software technology, if the ultrasound system is slightly modified and the relevant software of the method is implemented, (t1-t2) and (t1-t3) can be automatically measured, and the LAP can be calculated to achieve "1 key" to "measure" LAP. Continuous wave Doppler is commonly used in echocardiography. If this method can be verified by animal experiments and large-cohort clinical trials, it will enable the real-time quantitative measurement of LAP during conventional echocardiography and has epoch-making significance for the accurate diagnosis and treatment of heart diseases (8).

Limitations

Our study had some limitations. First, the study was an animal study with a small sample size, and only an animal model of acute mitral regurgitation combined with increased LAP was used. The calculation still needs to be verified by large-cohort clinical trials. Second, this method is only applicable when there is mitral regurgitation, and the spectrums of mitral regurgitation should be clear and the regurgitation velocity should exceed 3.5 m/s. Third, when collecting the continuous Doppler spectrum of mitral regurgitation, the position and angle of the probe must be adjusted to ensure that the Doppler sampling lines are parallel and pass through the regurgitation beam in each plane, which is highly dependent on the operator. In addition, a clear mitral regurgitation spectrum must be collected to accurately measure t1, t2, and t3, and to accurately measure LAP.

Conclusions

This study showed the good correlation between the LAP_{ECHO} and LAP_{C} . Further, the new method for detecting LAP by continuous wave Doppler spectrum is simple, convenient, non-invasive, and can be used to quantitatively estimate LAP, which is promising. Further large-scale

animal experiments and clinical studies need to be conducted.

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Footnote

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Company, in compliance with institutional guidelines for the care and use of animals.

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