



Genetic variants in Chinese patients with sporadic dilated cardiomyopathy: a cross-sectional study

Cheng Shen^{1,2}, Lei Xu¹, Xiaoning Sun³, Aijun Sun^{1,4}, Junbo Ge^{1,4}

¹Department of Cardiology, Zhongshan Hospital, Fudan University, Shanghai Institute of Cardiovascular Diseases, Shanghai, China; ²Department of Cardiology, Affiliated Hospital of Jining Medical University, Jining Key Laboratory for Diagnosis and Treatment of Cardiovascular Diseases, Jining, China; ³Department of Cardiovascular Surgery, Zhongshan Hospital, Fudan University, Shanghai Institute of Cardiovascular Diseases, Shanghai, China; ⁴Institutes of Biomedical Sciences, Fudan University, Shanghai, China

Contributions: (I) Conception and design: L Xu, X Sun; (II) Administrative support: A Sun, J Ge; (III) Provision of study materials or patients: C Shen, L Xu, X Sun; (IV) Collection and assembly of data: C Shen, L Xu; (V) Data analysis and interpretation: C Shen, L Xu, X Sun; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

Correspondence to: Dr. Lei Xu; Dr. Xiaoning Sun. Fenglin Road 180, Shanghai, China. Email: bri3stone@163.com; sun.xiaoning@zs-hospital.sh.cn.

Background: Multiple genes have been associated with familial dilated cardiomyopathy (DCM). However, the role of genetic factors in sporadic DCM (SDCM) remains unclear. Therefore, we studied the genetic variations in Chinese patients with SDCM.

Methods: Sixty-six unrelated Chinese patients (mean age 49.1±17.0 years; 71% male) diagnosed with SDCM were enrolled. The clinical history and genomic DNA of the cohort were collected and examined. The exons of 24 genes closely associated with familial DCM (*ABCC9*, *ACTC1*, *ACTN2*, *DES*, *LAMA4*, *LDB3*, *LMNA*, *MYBPC3*, *MYH6*, *MYH7*, *MYPN*, *PLN*, *PSEN1*, *PSEN2*, *RBM20*, *SCN5A*, *SGCD*, *TAZ*, *TCAP*, *TMPO*, *TNNI3*, *TNNT2*, *TPM1*, and *VCL*) were sequenced using targeted next-generation sequencing method. All called nonsynonymous variants and their occurrence frequencies were compared against population data from public databases. And the nonsynonymous variants were also evaluated for pathogenicity by PolyPhen 2 (PP2) and Sorts Intolerant From Tolerant (SIFT) algorithms.

Results: Eighty-five nonsynonymous variants were detected in 17 genes. The variants and their occurrence frequencies in the patients were compared against population data from the 1000 Genomes and NHLBI (National Heart, Lung, and Blood Institute) Go Exome Sequencing Project. Forty-nine nonsynonymous variants had occurrence frequencies that were significantly higher in the study patients than in the general population, indicating that they have the potential to increase the risk of DCM. The risk variants were distributed in 40 (61%) patients, among whom 25 carried a single variant, while the remaining patients carried multiple (2 to 4) variants. Risk variants occurred more frequently in *MYBPC3* (14% of the patients), *SCN5A* (14%), *MYH7* (12%), *MYPN* (9%), and *LDB3* (8%), as verified by Poisson distribution analysis, which were considered “the five risky genes”.

Conclusions: We found that genetic variants with potential risk for DCM were commonly present in SDCM patients, indicating that genetic factors contribute to the pathogenesis, and (probably) the onset, of DCM in these patients.

Keywords: Sporadic dilated cardiomyopathy (SDCM); mutations; risky genes

Submitted Nov 25, 2021. Accepted for publication Jan 05, 2022.

doi: 10.21037/atm-21-6774

View this article at: <https://dx.doi.org/10.21037/atm-21-6774>

Introduction

Dilated cardiomyopathy (DCM) is characterized by left ventricular (LV) dilatation and impaired systolic function, and is a leading cause of heart failure (1). Idiopathic DCM is defined when no other discernible causes, such as ischemia, valvular disease, and myocarditis, are present. Epidemiological studies have shown that a proportion of idiopathic DCM cases are familial (2). Familial DCM (FDCM) is primarily transmitted through an autosomal dominant pattern, while other patterns, including X-chromosomal, autosomal recessive, and mitochondrial transmission, are much less common or rare. To date, an increasing number of individual genes have been associated with inheritance in familial DCM cases (1,2).

However, more idiopathic DCM cases are sporadic, and the etiology of sporadic DCM (SDCM) remains largely unknown. SDCM patients are diagnosed with DCM manifestation but no family history, which are common in clinical practice. So far, compared to the information obtained from a large number of studies on FDCM cases, much less information is available regarding the role of genetic factors, such as rare genetic variations, on the pathogenesis of SDCM. Previously, a few studies on mutations of several individual genes implicated the role of genetic factors in the development of SDCM (3-6). Several cohorts, mainly containing Caucasian patients with FDCM or a combination of FDCM and SDCM, were also investigated for mutations in a relatively small number of selected candidate genes, including *MYH7*, *TNNT2*, *SCN5A*, *CSRP3*, and *LBD* (7-9). The consideration of genetic variants is important when evaluating the pathogenicity of a genomic variant. The genetic variants were identified using the genome analysis toolkit and the genetic databases. A systematic assessment of correlation between more candidate genes and SDCM cases would help to determine whether SDCM has genetic influences, and if so, whether SDCM shares the same or similar sets of genetic risk factors that are associated with FDCM, and hence, whether these two forms of DCM share similar or major pathogenetic paths. Li *et al.* reported some genetic variants in 24 SDCM patients recently, while the sample size is small (10). We aimed to expand the subjective scale and study the characteristics of genetic variants in 66 unrelated Chinese patients with SDCM using a next-generation sequencing technique targeting on 24 genes that have been previously reported to be associated with DCM primarily based on familial cases (11,12). We present the following article in accordance with the STROBE reporting

checklist (available at <https://atm.amegroups.com/article/view/10.21037/atm-21-6774/rc>).

Methods

Subjects

All procedures performed in this study involving human participants were in accordance with the Declaration of Helsinki (as revised in 2013). This cross-sectional clinical investigation was conducted in compliance with the guidelines for genetic research in the protocol approved by the Ethics Committees of Zhongshan Hospital (No. 2006-87). All participants signed a written informed consent. All participants in this study were unrelated individuals who were Han Chinese living in eastern China. Patients diagnosed with idiopathic DCM were hospitalized and recruited into the study from the Cardiology Department of Zhongshan Hospital. The inclusion criteria for the SDCM cohort were consistent with the guidelines described by the AHA Scientific Statement and position statement of the ESC working group (13,14). The exclusion criteria included peripartum cardiomyopathy and secondary dilated cardiomyopathies caused by ischaemic heart diseases, hypertension, valvular diseases, endocrine disorders (such as diabetic cardiomyopathy and hyperthyroid cardiomyopathy), inflammation, toxicity, and stress. All patients in this study reported no family history of DCM based on their recollection. Further clinical and genetic screenings were not conducted.

Clinical data collection

Clinical information available from the study subjects included date of birth, gender, vital status, clinical diagnosis, age at diagnosis (age at genetic testing was recorded if the age at diagnosis was not available), family history of DCM and other cardiovascular or muscular diseases, cardiovascular history (including myocardial infarction, hypertension, myocarditis, and drug/toxin exposure), and cardiac structure and function [including maximal LV wall thickness, LV ejection fraction (LVEF), and LV dimensions].

Genomic DNA (deoxyribonucleic acid) sequencing and data analysis

Genomic DNA was isolated from the peripheral blood of the participants. From each sample, 5 µg of genomic DNA was dissolved in 50 µL of water and fragmented to a size

of 100–300 bp, as judged by agarose gel electrophoresis. The patients' DNA samples were screened using a next-generation sequencing technique for point mutation variants in exons of the 24 genes that were reported to be associated with DCM primarily based on FDCM cases. The following genes were included: *ABCC9* (ATP-sensitive potassium channel regulatory subunit SUR2A), *ACTC1* (cardiac actin), *ACTN2* (α -actinin-2), *DES* (desmin), *LAMA4* (laminin a-4), *LDB3* (cypher), *LMNA* (lamin A/C), *MYBPC3* (myosin-binding protein C), *MYH6* (α -myosin heavy chain), *MYH7* (β -myosin heavy chain), *MYPN* (myopalladin), *PLN* (phospholamban), *PSEN1* (presenilin 1), *PSEN2* (presenilin 2), *RBM20* (RNA binding protein 20), *SCN5A* (cardiac sodium channel), *SGCD* (δ -sarcoglycan), *TAZ* (tafazzin), *TCAP* (telethonin), *TMPO* (thymopoietin), *TNNI3* (cardiac troponin I), *TNNT2* (cardiac troponin T), *TPM1* (tropomyosin α -1 chain), and *VCL* (metavinculin). Next-generation sequencing was performed at the sequencing platform of the Shanghai Institute of Cardiovascular Diseases.

Variants were called by aligning the raw sequence data to the human GRCh37 reference genome with manual verification. Novel variants were defined as those variants that were not found in the SNP Database (dbSNP) build 137 of the National Center for Biotechnology Information, and that had not been reported at the completion of the study. All called nonsynonymous variants of the genes in the DCM cohort and their occurrence frequencies were compared against population data from public databases, including the 1000 Genomes Project (<http://www.1000genomes.org>), and NHLBI (National Heart, Lung, and Blood Institute) Go Exome Sequencing Project (<http://evs.gs.washington.edu/EVS/>). The sequencing data of the 197 Chinese adults deposited in the 1000 Genomes Project database were used as the Chinese (ethnic-matched) control group in this study. Sequence and occurrence frequency data from other populations in this database, and those from African American and European American populations in the NHLBI Go Exome Sequencing Project database were combined together and used as the control group of the non-Chinese populations.

The relationship between the nonsynonymous variants and DCM was evaluated by comparing the occurrence frequencies of the DCM cohort and the control populations. Details of the comparison are described in the Results section below. The nonsynonymous variants were also evaluated for pathogenicity by PolyPhen 2 (PP2)

and Sorts Intolerant From Tolerant (SIFT) algorithms. PP2 scores were obtained using HumVar model software (<http://genetics.bwh.harvard.edu/pph2/index.shtml>). PP2 scores ranged from 0 to 1, with three levels of pathogenic potential, i.e., probably damaging, possibly damaging, and benign). SIFT scores were also obtained using a software tool (http://sift-dna.org/www/Extended_SIFT_chr_coords_submit.html). The SIFT scores range from 0 to 1, and were divided into damaging or tolerated levels with 0.05 as the threshold value.

Statistical analysis

Normally distributed continuous data were presented as mean \pm standard deviation (SD) and compared using the Student *t*-test. However, if the normality test (Shapiro-Wilk) and/or equal variance test failed, the Mann-Whitney rank-sum test was used to compare the continuous data. Categorical variables were presented as frequencies and analyzed by Fisher's exact test using the Simple Interactive Statistical Analysis web-based software (SISA Binomial, Southampton, UK). In all statistical tests, $P < 0.05$ was used to determine whether differences were statistically significant. To evaluate the occurrence frequencies of genetic variants, a model of simple Poisson process was used. We assumed that the number of variants occurring in a given gene approximately follows the Poisson distribution, $p(k) = e^{-\lambda} \lambda^k / k!$. $P < 0.05$ indicated a significantly high occurrence frequency. Poisson distribution and *P* values were computed using a web-based tool (<http://www.vassarstats.net/poissonfit.html#down>). The same method was also used to evaluate the significance of the prevalence of genetic variants.

Results

Study cohort

The SDCM cohort included 66 patients (47 males). The average age at diagnosis of patients in this cohort was 49.1 ± 17.0 years. All enrolled patients had symptoms of heart failure at the time of enrollment, with a majority of them in the New York Heart Association's (NYHA) classes III and IV. Echocardiographic findings showed a mean LV end-diastolic diameter (LVEDD) of 69 ± 9 mm, a mean LV end-systolic diameter (LVESD) of 57 ± 10 mm, and a mean LVEF of $34\% \pm 10\%$.

Nonsynonymous variants of DCM-associated genes in the SDCM cohort

A total of 85 nonsynonymous variants, mostly single-nucleotide mutations, were detected by sequencing in 17 out of the 24 DCM-related genes in the DCM cohort (listed in *Table 1*). In this cohort, nonsynonymous variants were absent in seven DCM-related genes. Fifty-five (65%) of the called nonsynonymous variants were either registered in the databases we searched or were reported previously (we denoted these variants as ‘known variants’). The remaining 30 (35%) of the called nonsynonymous variants have not yet been recorded anywhere else, and were therefore denoted herein as ‘novel variants’. All novel variants were rare frequency mutations (i.e., allele frequency <0.5% in the combined control populations). Meanwhile, among the 55 known variants, 19 (35%) had higher allele frequencies (>0.5%) in the combined control populations.

Occurrence frequencies of the nonsynonymous variants

To determine the possible relationship between the nonsynonymous variants and DCM in SDCM patients, we first compared the occurrence frequency (allele frequency) of the variants identified in the DCM cohort of this study with those of the control populations. We used two control populations, i.e., Chinese and non-Chinese controls, as described in the Methods section above. In addition, we also combined the Chinese and non-Chinese controls to form the general control population. The comparison indicated that the variants could be divided into three groups with statistically separable patterns.

The allele frequencies of variants in the first pattern group in the SDCM cohort were significantly higher than those in the general control population, but were statistically similar between the Chinese and non-Chinese control populations. These statistical features suggest that the variants falling in this pattern group exhibit a potential risk for SDCM (herein, we denoted these variants as ‘risk variants’). There were 49 variants (58% of the total nonsynonymous variants called) in this risky group. Most risk variants were not present in the control populations, except for six variants that had very low allele frequencies in the control populations. All risk variants were heterozygous mutations, except for a patient who carried a homozygous E334K mutation. It is noteworthy that all 30 novel variants were risk variants.

The allele frequencies of the variants in the second

pattern group were statistically different between the Chinese and non-Chinese populations, but were statistically similar between the SDCM group and the Chinese control group. These results suggest that variants with these statistical features (30 variants, 35% of the total variants called) exhibit specific occurrence in the Chinese population, but are not likely to indicate an increased risk of SDCM. We found that the allele frequencies of variants that did not fall in the previous two pattern groups in the SDCM cohort were statistically similar to those in the general control population, indicating that these six variants (7% of the total variants called) were shared globally and do not exhibit a specific risk of SDCM. We denoted variants in the second and third groups as ‘low-risk variants’. *Table 1* displays the statistical data that are summarized here.

The risk variants were distributed in 16 DCM-associated genes, which accounted for 94% of the genes on which nonsynonymous variants were found, and 67% of the total number of genes in our study. They were distributed in 40 (61%) patients, among whom 25 carried a single variant, 11 carried two variants, two carried three variants, and one carried four variants. As shown in *Table 2*, the prevalence of risk variants of each gene ranged from 1 to 9 among the 66 patients. *MYBPC3* and *SCN5A* exhibited the highest prevalence (9/66, 14%). Poisson distribution analysis revealed that *MYBPC3*, *SCN5A*, *MYH7*, *MYPN*, and *LDB3* had significantly higher prevalence (14%, 14%, 12%, 9%, and 8%, respectively) than the other genes. They had a combined prevalence of 31/66 (47%) in the SDCM cohort when multiple risk variants of these genes in a patient were counted as a single occurrence. *MYBPC3*, *SCN5A*, *MYH7*, and *LDB3* also housed more risk variants than the other genes, and the Poisson distribution analysis identified that they had a significantly high probability to house risk variants. These results suggested that these genes (which we denoted as ‘risky genes’) were closely related to SDCM.

Pathogenic potential of the nonsynonymous variants

Seventeen known variants, 11 risky variants, and six low-risk variants have been previously found to increase susceptibility to some cardiac diseases, including Brugada syndrome, LV noncompaction, hypertrophic cardiomyopathy, and distal myopathy (*Table 1*). However, none of the variants were directly related to DCM, except for A1180V and R1193Q of *SCN5A*, which were reported in the DCM cases (26,30).

To obtain further information on the pathogenic potential of the nonsynonymous variants of the DCM-

Table 1 Nonsynonymous variants of the DCM-associated genes found in the sporadic DCM cohort and their occurrence frequency compared with control populations

Gene	dbSNP ID	Change in AA	Number of DCM patients	DCM (D) allele counts	Chinese (C) allele counts	All Chinese (AC) allele counts	Non-Chinese (NC) allele counts	All reference population (A) allele counts	P (D-C)	P (D-NC)	P (C-NC)	P (D-A)	P (AC-NC)	Pattern	Disease reported
<i>ABCC9</i>	rs149319186	K976I	1	1/132	2/394	3/526	0/14,796	2/15,190	1.0000	0.0088	0.0007	0.0000*	0.0000*	2	
<i>ABCC9</i>	R1197C		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*	0.0086*	1	
<i>ACTN2</i>	K96R		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*	0.0086*	1	
<i>ACTN2</i>	M316T		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*	0.0086*	1	
<i>ACTN2</i>	rs80257412	D475N	6	6/132	35/394	41/526	20/14,796	55/15,190	0.1334	0.0000	0.0000	0.0000*	0.0000*	2	
<i>DES</i>	R78L		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*	0.0086*	1	
<i>LAMA4</i>	A41V		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*	0.0086*	1	
<i>LAMA4</i>	C91S		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*	0.0086*	1	
<i>LAMA4</i>	rs71543223	A283D	54	108/132	343/394	451/526	1,444/1,790	1,787/2,184	0.1507	0.8195	0.0024	0.0083*	0.0083*	2	
<i>LAMA4</i>	rs1050348	Y498H	65	107/132	309/394	416/526	9,713/1,4796	10,022/15,190	0.6209	0.0001	0.0000	0.0000*	0.0000*	2	
<i>LAMA4</i>	rs2032567	G1117S	66	116/132	338/394	454/526	11,172/14,796	11,510/15,190	0.6609	0.0007	0.0000	0.0000*	0.0000*	2	
<i>LAMA4</i>	rs1050349	P1119R	34	41/132	144/394	185/526	3,252/14,796	3,396/15,190	0.2924	0.0151	0.0000	0.0000*	0.0000*	2	
<i>LAMA4</i>	rs70940811	V1315I	1	1/132	6/394	7/526	4/14,796	10/15,190	0.6860	0.0434	0.0000	0.0000*	0.0000*	2	
<i>LAMA4</i>	G1356R		2	2/132	0/394	2/526	0/14,796	0/15,190	0.0626	0.0001	1.0000	0.0001*	0.0001*	1	
<i>LAMA4</i>	rs201094782	Y1391H	1	1/132	1/394	2/526	0/14,796	1/15,190	0.4393	0.0000	0.0259	0.0012*	0.0012*	2	
<i>LAMA4</i>	rs3734292	V1815I	11	12/132	31/394	43/526	15/14,796	46/15,190	0.7136	0.0000	0.0000	0.0000*	0.0000*	2	
<i>LDB3</i>	T28K		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*	0.0086*	1	
<i>LDB3</i>	rs3740343	V55I	7	7/132	38/394	45/526	23/14,796	61/15,190	0.1053	0.0000	0.0000	0.0000*	0.0000*	2	LYNC (15)
<i>LDB3</i>	E139K		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*	0.0086*	1	
<i>LDB3</i>	rs45521338	R218C	1	1/132	2/394	3/526	1/14,796	3/15,190	1.0000	0.0176	0.0020	0.0002*	0.0002*	2	
<i>LDB3</i>	S330P		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*	0.0086*	1	
<i>LDB3</i>	rs113817827	V426I	1	1/132	2/394	3/526	2/14,796	4/15,190	1.0000	0.0263	0.0001	0.0004*	0.0004*	2	
<i>LDB3</i>	M456R		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*	0.0086*	1	
<i>LDB3</i>	rs145983824	P498L	1	1/132	0/394	1/526	3/14,796	3/15,190	0.2510	0.0349	1.0000	0.0340*	0.0340*	1	
<i>LMNA</i>	R220H		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*	0.0086*	1	
<i>MYBPC3</i>	rs3729989	S236G	2	2/132	13/394	15/526	1,475/14,348	1,488/14,742	0.3769	0.0001	0.0000	0.0000*	0.0000*	2	HCM (16)
<i>MYBPC3</i>	E334K		4	5/132	0/394	5/526	0/14,796	0/15,190	0.0009	0.0000	1.0000	0.0000*	0.0000*	1	HCM (17)
<i>MYBPC3</i>	R409G		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*	0.0086*	1	

Table 1 (continued)

Table 1 (continued)

Gene	dbSNP ID	Change in AA	Number of DCM patients	DCM (D) allele counts	Chinese (C) allele counts	All Chinese (AC) allele counts	Non-Chinese (NC) allele counts	All reference population (A) allele counts	P (D-C)	P (D-NC)	P (C-NC)	P (D-A)	P (AC-NC)	Pattern	Disease reported
<i>MYBPC3</i>	G416S		1	1/132	0/394	1/526	3/14,466	3/14,860	0.2510	0.0357	1.0000	0.0348*		1	HCM (18)
<i>MYBPC3</i>	P459fs		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*		1	HCM (19)
<i>MYBPC3</i>	R835L		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*		1	
<i>MYBPC3</i>	R895H		1	1/132	0/394	1/526	4/14,078	4/14,472	0.2510	0.0456	0.1045	0.0444*		1	
<i>MYH6</i>	rs28711516	G56R	1	1/132	10/394	11/526	1,155/14,796	1,165/15,190	0.3060	0.0005	0.0001		0.0000*	2	
<i>MYH6</i>	rs365990	V1101A	21	21/132	68/394	89/526	6,644/14,796	6,712/15,190	0.7894	0.0000	0.0000		0.0000*	2	
<i>MYH6</i>	rs28730771	A1130T	6	6/132	27/394	33/526	168/1,790	195/2,184	0.2990	0.0600	0.1188	0.1087		3	
<i>MYH6</i>	rs34935550	E1295Q	2	2/132	10/394	12/526	43/14,796	53/15,190	0.7389	0.0600	0.0000		0.0000*	2	
<i>MYH6</i>	rs45574136	Q1593L	2	2/132	11/394	13/526	327/14,796	338/15,190	0.5328	1.0000	0.3877	1.0000		3	
<i>MYH6</i>	rs61742476	V1613A	1	1/132	11/394	12/526	248/14,796	259/15,190	0.3106	0.7288	0.1092	0.7299		3	
<i>MYH6</i>	K1860R		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*		1	
<i>MYH7</i>	rs121913653	T441M	1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*		1	Distal myopathy (20)
<i>MYH7</i>	rs121913625	R453C	1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*		1	HCM (21)
<i>MYH7</i>	I736T		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*		1	HCM (22)
<i>MYH7</i>	rs121913628	E924K	1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*		1	HCM (18)
<i>MYH7</i>	LS1139LD		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*		1	
<i>MYH7</i>	R1250W		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*		1	LVNC (23)
<i>MYH7</i>	G1520R		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*		1	
<i>MYH7</i>	R1897H		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*		1	
<i>MYPN</i>	R482K		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*		1	
<i>MYPN</i>	rs10823148	F628L	23	27/132	67/394	94/526	5,819/14,796	5,886/15,190	0.3617	0.0000	0.0000		0.0000*	2	
<i>MYPN</i>	rs10997975	S691N	25	31/132	86/394	117/526	5,801/14,796	5,887/15,190	0.3617	0.0002	0.0000		0.0000*	2	
<i>MYPN</i>	rs7916821	S707N	22	27/132	67/394	94/526	5,788/14,796	5,855/15,190	0.3617	0.0000	0.0000		0.0000*	2	
<i>MYPN</i>	rs3814182	S803R	36	46/132	134/394	180/526	7,714/14,796	7,848/15,190	0.9156	0.0001	0.0000		0.0000*	2	
<i>MYPN</i>	rs181848049	G847V	1	1/132	2/394	3/526	0/14,796	2/15,190	1.0000	0.0088	0.0007		0.0000*	2	
<i>MYPN</i>	rs151282801	R1042C	1	1/132	0/394	1/526	3/14,796	3/15,190	0.2510	0.0349	1.0000	0.0340*		1	
<i>MYPN</i>	rs7079481	P1135T	29	33/132	82/394	115/526	5,983/14,796	6,065/15,190	0.3312	0.0002	0.0000		0.0000*	2	

Table 1 (continued)

Table 1 (continued)

Gene	dbSNP ID	Change in AA	Number of DCM patients	DCM (D) allele counts	Chinese (C) allele counts	All Chinese (AC) allele counts	Non-Chinese (NC) allele counts	All reference population (A) allele counts	P (D-C)	P (D-NC)	P (C-NC)	P (D-A)	P (AC-NC)	Pattern	Disease reported
<i>MYPN</i>	rs138813730	L1161I	3	3/132	0/394	3/526	12/14,796	12/15,190	0.0155	0.0003	1.0000	0.0003*		1	
<i>MYPN</i>	rs199585352	S1296T	1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*		1	
<i>RBM20</i>	G40W		1	1/132	0/394	1/526	0/6,356	0/6,750	0.2510	0.0204	1.0000	0.0192*		1	
<i>RBM20</i>	P48delinsPP		1	1/132	0/394	1/526	0/6,356	0/6,750	0.2510	0.0204	1.0000	0.0192*		1	
<i>RBM20</i>	rs143785916	R641Q	1	1/132	3/394	4/526	2/6,356	5/6,750	1.0000	0.0598	0.0018		0.0000*	2	
<i>RBM20</i>	rs138926584	R673Q	1	1/132	0/394	1/526	7/6,356	7/6,750	0.2510	0.1517	1.0000	0.1436		3	
<i>RBM20</i>	rs1417635	W768S	66	132/132	394/394	526/526	6,125/6,127	6,519/6,521	1.0000	1.0000	1.0000	1.0000		3	
<i>RBM20</i>	rs188054898	Q856X	1	1/132	0/394	1/526	0/6,356	0/6,750	0.2510	0.0204	1.0000	0.0192*		1	
<i>RBM20</i>	R1057Q		4	4/132	11/394	15/526	5/6,356	16/6,750	1.0000	0.0001	0.0000		0.0000*	2	
<i>RBM20</i>	R1182H		1	1/132	0/394	1/526	0/6,356	0/6,750	0.2510	0.0204	1.0000	0.0192*		1	
<i>RBM20</i>	rs942077	E1223Q	59	101/132	315/394	416/526	4719/6,356	5,034/6,750	0.3902	0.6154	0.0122		0.0142*	2	
<i>SCN5A</i>	rs199473071	R225Q	3	3/132	0/394	3/526	0/14,118	0/14,512	0.0155	0.0000	1.0000	0.0000*		1	
<i>SCN5A</i>	rs199473561	A226V	1	1/132	0/394	1/526	0/14,118	0/14,512	0.2510	0.0093	1.0000	0.0090*		1	BS (16)
<i>SCN5A</i>	rs1805124	H558R	16	16/132	38/394	54/526	3,520/14,410	3,558/14,804	0.4110	0.0007	0.0000		0.0000*	2	AF, LQTS (24)
<i>SCN5A</i>	rs45600438	R668C	1	1/132	0/394	1/526	0/14,264	0/14,658	0.2510	0.0089	1.0000	0.0086*		1	
<i>SCN5A</i>	R659W		1	1/132	0/394	1/526	0/14,784	0/15,178	0.2510	0.0093	1.0000	0.0090*		1	LQTS (25)
<i>SCN5A</i>	rs1805125	P1090L	3	3/132	9/394	12/526	4/14,236	13/14,630	1.0000	0.0000	0.0000		0.0000*	2	LQTS (25)
<i>SCN5A</i>	rs41310765	A1180V	1	1/132	1/1,314	2/1,446	0/14,794	1/15,188	0.4393	0.0088	0.0816	0.0162*		1	AF, DCM (26)
<i>SCN5A</i>	rs41261344	R1193Q	8	10/132	20/394	30/526	17/14,796	37/15,190	0.2832	0.0000	0.0000		0.0000*	2	BS, LQT, CCD, DCM (27)
<i>SCN5A</i>	rs199473251	I1448N	1	1/132	0/394	1/526	0/14,122	0/14,516	0.2510	0.0092	1.0000	0.0089*		1	
<i>SCN5A</i>	P1619T		1	1/132	0/394	1/526	0/14,796	0/15,190	0.2510	0.0088	1.0000	0.0086*		1	
<i>SGCD</i>	C34G		1	1/132	0/394	1/526	0/13,987	0/14,381	0.2510	0.0094	1.0000	0.0091*		1	
<i>SGCD</i>	P253fs		1	1/132	0/394	1/526	0/13,890	0/14,284	0.2510	0.0094	1.0000	0.0092*		1	
<i>SGCD</i>	Q283R		1	1/132	0/394	1/526	0/14,022	0/14,416	0.2510	0.0093	1.0000	0.0091*		1	
<i>TAZ</i>	H111N		1	1/132	0/394	1/526	0/12,353	0/12,747	0.2510	0.0106	1.0000	0.0103*		1	
<i>TNNI2</i>	R151Q		2	2/132	0/394	2/526	0/14,310	0/14,704	0.0626	0.0001	1.0000	0.0001*		1	
<i>TNNI2</i>	rs3730238	K260R	4	4/132	14/394	18/526	889/14,796	903/15,190	1.0000	0.1946	0.0400		0.1452	3	HCM (28)

Table 1 (continued)

Table 1 (continued)

Gene	dbSNP ID	Change in AA	Number of DCM patients	DCM (D) allele counts	Chinese (C) allele counts	All Chinese (AC) allele counts	Non-Chinese (NC) allele counts	All reference population (A) allele counts	P (D-C)	P (D-NC)	P (C-NC)	P (D-A)	P (AC-NC)	Pattern	Disease reported
<i>[†]TPM1</i>		D34E	1	1/132	0/394	1/526	0/14,308	0/14,702	0.2510	0.0091	1.0000	0.0089*		1	
<i>[‡]VCL</i>	rs144683137	M209L	1	1/132	1/394	2/526	0/14,796	1/15,190	0.4393	0.0088	0.0259		0.0012*	2	
<i>[‡]VCL</i>	rs201528612	P398S	1	1/132	1/394	2/526	0/14,796	1/15,190	0.4393	0.0088	0.0259		0.0012*	2	

DCM patients: the patients with sporadic DCM enrolled in this study; Chinese: the 197 Chinese adults in the 1000 Genomes Project (phase 1), for A1180V of SCN5A samples from additional 460 unrelated healthy Chinese (29) were included; all Chinese: DCM patients + Chinese; non-Chinese: a combination of populations excluding the Chinese population in the 1000 Genomes Project (phase 1) and American populations in NHLBI Go Exome Sequencing Project; all control populations: Chinese + non-Chinese, referred as the general control population in the text. Patterns of the grouped variants: [†], pattern 1, having potential risk of sporadic DCM: when P(C-NC) ≥ 0.05; a variant was judged to be in this pattern group if P(D-A) < 0.05. [‡], pattern 2, being Chinese-specific without risk of sporadic DCM: when P(C-NC) < 0.05 and P(D-C) ≥ 0.05; groups D and C were pooled together to form group AC, and a variant was judged to exhibit this pattern if P(AC-NC) < 0.05. Pattern 3, being shared globally without risk of sporadic DCM: a variant that falls neither in pattern 1 nor in pattern 2. *, P < 0.05. AA, amino acid; DCM, dilated cardiomyopathy; LVNC, left ventricular noncompaction; HCM, hypertrophic cardiomyopathy; AF, atrial fibrillation; LQTS, long QT syndrome; BS, Brugada syndrome.

associated genes in the SDCM cohort, we performed an analysis of the nonsynonymous variants using the protein function prediction algorithms, PP2 and SIFT (Table S1). In total, 62 variants could be predicted by both algorithms (73% of the total nonsynonymous variants), which also included one non-sense and two frame-shift variants that could be assumed to cause damaging consequences in the protein function (31). Twenty-six risk variants (53% of the total risk variants) were predicted to be damaging, and eight risk variants (16% of the total risk variants) were predicted to be tolerated. On the other hand, 23 low-risk variants (63% of the total low-risk variants) were predicted to be tolerated, and five low-risk variants (14% of the total low-risk variants) were predicted to be damaging. Fisher's exact test indicated a significant difference (P=0.000) in the predicted results between the risky and non-risky variant groups. Therefore, the protein function prediction algorithms confirmed that majority of the risk variants exhibit pathogenic potential, whereas the majority of low-risk variants were predicted to be harmless based on the protein structure-function relationship. We noticed that in the risky genes of SDCM, *SCN5A* housed the most risk variants that were predicted to be damaging.

Comparison of clinical symptoms between the patients with and without risk variants

Given the determination of the risk variants, we further explored whether there were differences in clinical symptoms between the patients with and without risk variants, and compared the symptoms between these two groups of patients (as shown in Tables S2,S3). The results indicated that patients with risk variants did not manifest any symptoms or abnormalities that were significantly different from those seen in the patients without risk variants.

Discussion

This study was specifically designed to screen genes known to be associated with FDCM in SDCM patients. To our knowledge, similar studies have not yet been reported in the literature. In this study, we investigated a cohort of 66 unrelated Chinese patients with diagnosed SDCM. We performed mutational screening of 24 genes known to be associated with FDCM using a next-generation sequencing technique. A major finding of this study was that the at-risk genotypes are common (61%) in SDCM patients. The

Table 2 Genes hosting variants with potential risk of sporadic DCM in the DCM cohort being studied

Gene	Prevalence ^a of variants with risk of sporadic DCM	P value ^c	Prevalence ^a of variants	Number of variants with risk of sporadic DCM	P value ^d	Number of total variants
<i>MYBPC3</i>	9 (13.6%) ^b	0.000*	11	6	0.010*	7
<i>SCN5A</i>	9 (13.6%)	0.000*	36	7	0.002*	10
<i>MYH7</i>	8 (12.1%)	0.001*	8	8	0.001*	8
<i>MYPN</i>	6 (9.0%)	0.011*	142	4	0.140	10
<i>LDB3</i>	5 (7.6%)	0.039*	14	5	0.034*	8
<i>RBM20</i>	4 (6.1%)	0.114	135	4	0.140	9
<i>LAMA4</i>	4 (6.1%)	0.114	236	3	0.261	10
<i>SGCD</i>	3 (4.5%)	0.277	3	3	0.261	3
<i>ACTN2</i>	2 (3.0%)	0.546	8	2	0.528	3
<i>TNNT2</i>	2 (3.0%)	0.546	6	1	0.830	2
<i>MYH6</i>	1 (1.5%)	0.840	34	1	0.830	2
<i>ABCC9</i>	1 (1.5%)	0.840	2	1	0.830	7
<i>DES</i>	1 (1.5%)	0.840	1	1	0.830	1
<i>LMNA</i>	1 (1.5%)	0.840	1	1	0.830	1
<i>TPM1</i>	1 (1.5%)	0.840	1	1	0.830	1
<i>TAZ</i>	1 (1.5%)	0.840	1	1	0.830	1
<i>VCL</i>	0 (0%)	1.000	2	0	1.000	2
<i>ACTC1</i>	0 (0%)		0	0		0
<i>PLN</i>	0 (0%)		0	0		0
<i>PSEN1</i>	0 (0%)		0	0		0
<i>PSEN2</i>	0 (0%)		0	0		0
<i>TCAP</i>	0 (0%)		0	0		0
<i>TMPO</i>	0 (0%)		0	0		0
<i>TNNI3</i>	0 (0%)		0	0		0

^a, prevalence was designated as the sum of patients carrying a variant (either homozygous or heterozygous) in each gene. When prevalence was calculated, variants co-occurring in a gene were counted separately. Likewise, variants co-occurring in a patient were also counted separately. ^b, prevalence in this column was also expressed as a percentage of the total patients in the DCM cohort. ^{c,d}, P values were computed using Poisson distribution as described in the Methods section. The means of the fitted Poisson distribution (λ) were 0.97 and 0.95, respectively. Only those genes that contained nonsynonymous variants were included in the computation. *, P<0.05.

second major finding was that *MYBPC3* and *SCN5A* were found to be the most prevalent risky genes for SDCM. The five significantly risky genes out of the 24 genes, namely *MYBPC3*, *SCN5A*, *MYH7*, *MYPN*, and *LDB3*, had a combined prevalence of 47% in the patient cohort, which accounted for 78% of the prevalence of the total risk variants. In addition, 30 novel variants with a potential to increase the risk of DCM were identified in this study.

Comparison of the prevalence of risk variants with other studies

The prevalence of 61% for total risk variants observed in this study was higher than that reported in previous studies that included both FDCM and SDCM. In a study by Millat *et al.*, a prevalence of 19% was observed in 105 DCM patients (8). Another study by Hershberger *et al.* was 11.5% in 313 DCM

patients, although the study cohorts of these studies contained both FDCM and SDCM patients (32). We consider that these differences are partly due to the fact that our study included more genes than these previous studies. For example, only six genes (*MYH7*, *TNNT2*, *SCN5A*, *TCAP*, *LDB3*, and *CSRP3*) were screened in the 313-case study, which excluded two genes with high prevalence (*MYBPC3* and *MYPN*) found in our study. Given the fact that there are more DCM-associated genes, the actual prevalence of genetic risk variants in SDCM patients would be expected to be even higher. Indeed, a later study from Hershberger *et al.* including additional genes further expanded the prevalence (up to 27%) of the total variants that are likely cause DCM (7). The second factor contributing to this difference is the criteria used to define risk variants. In this study, we used a statistics-based criterion that compares prevalence; a variant was considered to be risky when its prevalence in the DCM cohort was significantly higher than that in the general population (on the condition that the prevalence of this variant in the Chinese population was similar to that in the general population). However, more complicated criteria, which possibly depend on familial cases to a greater extent, were used previously. For example, in the 313-case study, a variant was considered disease-causing if it caused a change in a conserved amino acid, a frame-shift, premature truncation, a mis-splicing event, and also segregated with the disease in multiple affected individuals or was identified in multiple unrelated probands, or had previously been reported to be associated with DCM (32). We believe that the criteria used in the present study are more appropriate for detection of novel risk variants, especially in sporadic cases where segregation of multiple occurrence is unlikely. In the risk variants identified by this criterion, 51% were predicted to be functionally damaging by both the PP2 and SIFT algorithms (in addition to the frame-shift and non-sense variants), and there were significantly more predicted damaging variants in the risky group than in the non-risky group, with a P value of 0.000. These facts confirm the effectiveness of the criteria used in our study. With the rapid expansion of population genomic databases, the accuracy of identification of risk variants using statistics-based criteria should be rapidly increased.

Although our study reported a high prevalence of risk variants, the major fraction of the prevalence was attributed to the variants hosted in a few genes. The five risky genes with the highest prevalence (*MYBPC3*, *SCN5A*, *MYH7*, *MYPN*, and *LDB3*) were distributed in 47% of the total SDCM patients and 78% of patients with risk variants. This

distribution overlaps with the five genes with the highest prevalence (*LMNA*, *MYBPC3*, *MYH7*, *MYH6*, and *TNNT2*) previously obtained from a mixed cohort mainly containing Caucasian patients with FDCM and SDCM. Determination of the genes with the highest prevalence of risky genes may assist in clinical practice by narrowing down the genes to be screened and saving the cost of diagnostic tests. Although the 24 candidates are not the whole targeted genes of SDCM, the results of the study showed the genetic variants from the candidate genes, suggesting the genetic variants are common in DCM even though the patients with no familial history. Besides the environment factors, the genetic factor plays an important role in the pathogenesis of SDCM. The findings of the research indicate the genetic screening is valuable for DCM patients in clinical practice. In addition, our study used next-generation sequencing technology rather than chain termination to determine the DNA sequence, which is faster and more cost-efficient (29). The accurate sequencing results in an array of multiple genes obtained in our study confirm that this high-throughput sequencing technology facilitates diagnostic classification and can improve risk stratification in affected patients.

It is important to note that in this study, the DCM patients were diagnosed as sporadic cases primarily based on the patients' recollection of their family history, which is sometimes inaccurate, and was not verified by medical examination. Therefore, the DCM cases in this study can only be considered as "apparently" sporadic.

Are genetic factors a risk for DCM in sporadic cases?

The role of genetic factors in the pathogenesis of SDCM remains largely unknown. Previous population studies mainly from Caucasian population showed that the prevalence of disease-causing mutations of some DCM-associated genes is similar for SDCM and FDCM (7,30). Our study conducted in a different ethnic group, namely Chinese, are therefore helpful to further map out the role of genetic factors in SDCM.

Mutational screening studies, such as the present study, provide clear evidence to show that genetic variation has a relation to SDCM. However, compared to the familial cases, it is more difficult to determine the contribution of genetic factors in sporadic cases that do not involve family history, i.e., cases where the disease is not apparently hereditary. Several hypothetical interpretations could be offered. Firstly, it appears that each individual risky variant only

modestly increases the risk, which is not sufficient to result in the onset of the disease by itself. Therefore, those who carry multiple risk variants are likely to develop the disease. Indeed, 14 of the 40 (35%) patients with risk variants in our study were found to carry multiple risk variants. We cannot exclude the possibility of patients with single risk variants concurrently carrying risk variants of DCM-associated genes, either known or unknown at present, that are not included in our study.

In addition to genetic factors, environmental factors may play a significant role in the expression of pathogenic mechanisms. Our previous study on a segregated DCM family with A1180V of *SCN5A* (which encodes the cardiac sodium channel) demonstrated that the functional phenotype of this genetic variation was specifically aggravated at high heart rates (26). The results of that study suggested that the risk of DCM of this variation increases in those carriers with physical activity and lifestyles that increase the average daily heart rate. Notably, A1180V of *SCN5A* was found to be a risky variant in the present study.

Is there a specific phenotype that differentiates SDCM patients with risk variants from those without?

A unique result of this study is the report of summarized symptoms and the statistical comparison of the symptoms between patients with and without risk variants (shown in Tables S2,S3). Our data clearly demonstrated that there were no differences in the profile of DCM-related symptoms between these two patient groups. DCM was the only phenotype for individuals who carried risk variants reported in our study. No other clear genotype-phenotype correlations could be concluded from the data. A clear unique phenotype-genotype relationship does not appear to be present for individual genes either. For example, many DCM patients with variants in *LMNA* and *SCN5A* were reported to suffer from conduction system diseases (26,33); however, in our study cohort, only one patient had first-degree atrioventricular block, whereas other patients carrying risk variants in *LMNA* and *SCN5A* were not diagnosed with any conduction system diseases. Also, genes listed among the highest prevalence encode proteins (myosin-binding protein C, cardiac sodium channel, β -myosin heavy chain, myopalladin, and cypher), which have markedly different functions, implicate the heterogeneity of the phenotype-genotype relation. In addition, we noticed that patients carrying multiple variants did not exhibit aggravated symptoms (see Table S2),

indicating an absence of additive effects for risk variants on the symptoms of the disease. Certainly, we do not exclude the possibility that some individual variants may not directly cause DCM. However in these cases, DCM is not primary, which is not the concern of this study. Taken together, our data supports the hypothesis that the presence of a risky genotype in SDCM only imposes an increased risk for the onset of DCM, but does not affect the progress of DCM after the onset of the disease. However, the current study does not contain sufficient information regarding the mechanism through which risk variants lead to the onset of SDCM, which was not the purpose of this study and requires further investigation.

Conclusions

In summary, this study suggests that at-risk genomic variants are a major pathogenic factor of SDCM, and *MYBPC3*, *SCN5A*, *MYH7*, *MYPN*, and *LDB3* are the major genes hosting the at-risk genomic variants. This study also identifies a number of novel variants that are possibly associated with SDCM. These results not only expand the spectrum of DCM genetics, but also provide new information that helps to provide insights into the pathogenesis of SDCM.

Acknowledgments

Funding: This study was supported by the Surface Project of National Natural Science Foundation of China (82070242 to Lei Xu); and the National Science Fund for Distinguished Young Scholars (81725002 to Aijun Sun); and the Innovation Program of Shanghai Municipal Education Commission to Aijun Sun.

Footnote

Reporting Checklist: The authors have completed the STROBE reporting checklist. Available at <https://atm.amegroups.com/article/view/10.21037/atm-21-6774/rc>

Data Sharing Statement: Available at <https://atm.amegroups.com/article/view/10.21037/atm-21-6774/dss>

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <https://atm.amegroups.com/article/view/10.21037/atm-21-6774/coif>). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. All procedures performed in this study involving human participants were in accordance with the Declaration of Helsinki (as revised in 2013). This cross-sectional clinical investigation was conducted in compliance with the guidelines for genetic research in the protocol approved by the Ethics Committees of Zhongshan Hospital (No. 2006-87). All participants signed a written informed consent.

Open Access Statement: This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

References

- Rosenbaum AN, Agre KE, Pereira NL. Genetics of dilated cardiomyopathy: practical implications for heart failure management. *Nat Rev Cardiol* 2020;17:286-97.
- Peters S, Johnson R, Birch S, et al. Familial Dilated Cardiomyopathy. *Heart Lung Circ* 2020;29:566-74.
- Xu JH, Gu JY, Guo YH, et al. Prevalence and Spectrum of NKX2-5 Mutations Associated With Sporadic Adult-Onset Dilated Cardiomyopathy. *Int Heart J* 2017;58:521-9.
- Asselbergs FW, Sammani A, Elliott P, et al. Differences between familial and sporadic dilated cardiomyopathy: ESC EORP Cardiomyopathy & Myocarditis registry. *ESC Heart Fail* 2021;8:95-105.
- Li J, Liu WD, Yang ZL, et al. Prevalence and spectrum of GATA4 mutations associated with sporadic dilated cardiomyopathy. *Gene* 2014;548:174-81.
- Zhou W, Zhao L, Jiang JQ, et al. A novel TBX5 loss-of-function mutation associated with sporadic dilated cardiomyopathy. *Int J Mol Med* 2015;36:282-8.
- Hershberger RE, Norton N, Morales A, et al. Coding sequence rare variants identified in MYBPC3, MYH6, TPM1, TNNC1, and TNNI3 from 312 patients with familial or idiopathic dilated cardiomyopathy. *Circ Cardiovasc Genet* 2010;3:155-61.
- Millat G, Bouvagnet P, Chevalier P, et al. Clinical and mutational spectrum in a cohort of 105 unrelated patients with dilated cardiomyopathy. *Eur J Med Genet* 2011;54:e570-5.
- Li D, Morales A, Gonzalez-Quintana J, et al. Identification of novel mutations in RBM20 in patients with dilated cardiomyopathy. *Clin Transl Sci* 2010;3:90-7.
- Li M, Xia S, Xu L, et al. Genetic analysis using targeted next-generation sequencing of sporadic Chinese patients with idiopathic dilated cardiomyopathy. *J Transl Med* 2021;19:189.
- Hershberger RE, Morales A, Siegfried JD. Clinical and genetic issues in dilated cardiomyopathy: a review for genetics professionals. *Genet Med* 2010;12:655-67.
- Hershberger RE, Siegfried JD. Update 2011: clinical and genetic issues in familial dilated cardiomyopathy. *J Am Coll Cardiol* 2011;57:1641-9.
- Bozkurt B, Colvin M, Cook J, et al. Current Diagnostic and Treatment Strategies for Specific Dilated Cardiomyopathies: A Scientific Statement From the American Heart Association. *Circulation* 2016;134:e579-646.
- Pinto YM, Elliott PM, Arbustini E, et al. Proposal for a revised definition of dilated cardiomyopathy, hypokinetic non-dilated cardiomyopathy, and its implications for clinical practice: a position statement of the ESC working group on myocardial and pericardial diseases. *Eur Heart J* 2016;37:1850-8.
- Xing Y, Ichida F, Matsuoka T, et al. Genetic analysis in patients with left ventricular noncompaction and evidence for genetic heterogeneity. *Mol Genet Metab* 2006;88:71-7.
- Van Driest SL, Vasile VC, Ommen SR, et al. Myosin binding protein C mutations and compound heterozygosity in hypertrophic cardiomyopathy. *J Am Coll Cardiol* 2004;44:1903-10.
- Bahrudin U, Morisaki H, Morisaki T, et al. Ubiquitin-proteasome system impairment caused by a missense cardiac myosin-binding protein C mutation and associated with cardiac dysfunction in hypertrophic cardiomyopathy. *J Mol Biol* 2008;384:896-907.
- Song L, Zou Y, Wang J, et al. Mutations profile in Chinese patients with hypertrophic cardiomyopathy. *Clin Chim Acta* 2005;351:209-16.
- Lin J, Zheng DD, Tao Q, et al. Two novel mutations of the MYBPC3 gene identified in Chinese families with hypertrophic cardiomyopathy. *Can J Cardiol* 2010;26:518-22.
- Tajsharghi H, Oldfors A. Myosinopathies: pathology and mechanisms. *Acta Neuropathol* 2013;125:3-18.

21. Alcalai R, Seidman JG, Seidman CE. Genetic basis of hypertrophic cardiomyopathy: from bench to the clinics. *J Cardiovasc Electrophysiol* 2008;19:104-10.
 22. Laredo R, Monserrat L, Hermida-Prieto M, et al. Beta-myosin heavy-chain gene mutations in patients with hypertrophic cardiomyopathy. *Rev Esp Cardiol* 2006;59:1008-18.
 23. Dellefave LM, Pytel P, Mewborn S, et al. Sarcomere mutations in cardiomyopathy with left ventricular hypertrabeculation. *Circ Cardiovasc Genet* 2009;2:442-9.
 24. Viswanathan PC, Benson DW, Balsler JR. A common SCN5A polymorphism modulates the biophysical effects of an SCN5A mutation. *J Clin Invest* 2003;111:341-6.
 25. Chiu SN, Wu MH, Su MJ, et al. Coexisting mutations/polymorphisms of the long QT syndrome genes in patients with repaired Tetralogy of Fallot are associated with the risks of life-threatening events. *Hum Genet* 2012;131:1295-304.
 26. Ge J, Sun A, Paajanen V, et al. Molecular and clinical characterization of a novel SCN5A mutation associated with atrioventricular block and dilated cardiomyopathy. *Circ Arrhythm Electrophysiol* 2008;1:83-92.
 27. Hwang HW, Chen JJ, Lin YJ, et al. R1193Q of SCN5A, a Brugada and long QT mutation, is a common polymorphism in Han Chinese. *J Med Genet* 2005;42:e7; author reply e8.
 28. Jáchymová M, Muravská A, Paleček T, et al. Genetic variation screening of TNNT2 gene in a cohort of patients with hypertrophic and dilated cardiomyopathy. *Physiol Res* 2012;61:169-75.
 29. Gu W, Miller S, Chiu CY. Clinical Metagenomic Next-Generation Sequencing for Pathogen Detection. *Annu Rev Pathol* 2019;14:319-38.
 30. Kwon HW, Lee SY, Kwon BS, et al. Long QT syndrome and dilated cardiomyopathy with SCN5A p.R1193Q polymorphism: cardioverter-defibrillator implantation at 27 months. *Pacing Clin Electrophysiol* 2012;35:e243-6.
 31. Lakdawala NK, Funke BH, Baxter S, et al. Genetic testing for dilated cardiomyopathy in clinical practice. *J Card Fail* 2012;18:296-303.
 32. Hershberger RE, Parks SB, Kushner JD, et al. Coding sequence mutations identified in MYH7, TNNT2, SCN5A, CSRP3, LBD3, and TCAP from 313 patients with familial or idiopathic dilated cardiomyopathy. *Clin Transl Sci* 2008;1:21-6.
 33. Kawakami H, Ogimoto A, Tokunaga N, et al. A Novel Truncating LMNA Mutation in Patients with Cardiac Conduction Disorders and Dilated Cardiomyopathy. *Int Heart J* 2018;59:531-41.
- (English Language Editor: A. Kassem)

Cite this article as: Shen C, Xu L, Sun X, Sun A, Ge J. Genetic variants in Chinese patients with sporadic dilated cardiomyopathy: a cross-sectional study. *Ann Transl Med* 2022;10(3):129. doi: 10.21037/atm-21-6774

Table S1 Pathogenic potential of the nonsynonymous variants of the DCM-associated genes in the sporadic DCM cohort

Gene	dbSNP ID	Change in AA	Pattern	PP2 Prediction (score)	Chrs.	Position (B37)	cDNA substitution	Orientation	SIFT Prediction (score)	Predicted damaging ^a	Predicted tolerated ^b
ABCC9	rs149319186	K976I	2	Benign (0.36)	12	21998706	A/T	-	Damaging (0.04)		
ABCC9		R1197C	1	Probably damaging (0.93)	12	21981972	C/T	-	Damaging (0.04)	X	
ACTN2		K96R	1	Probably damaging (0.98)	1	236882239	A/G	+	Tolerated (0.06)		
ACTN2		M316T	1	Probably damaging (0.96)	1	236902672	T/C	+	Tolerated (0.12)		
ACTN2	rs80257412	D475N	2	Probably damaging (0.99)	1	236910983	G/A	+	Tolerated (0.24)		
DES		R78L	1	Benign (0.00)	2	220283417	G/T	+	Tolerated (0.54)		X
LAMA4		A41V	1	Benign (0.00)	6	112575231	C/T	-	Tolerated (0.15)		X
LAMA4		C91S	1	Probably damaging (1.00)	6	112537595	T/A	-	Damaging (0.00)	X	
LAMA4	rs71543223	A283D	2	Benign (0.00)	6	112508769	CA/AC	-			
LAMA4	rs1050348	Y498H	2	Benign (0.00)	6	112493872	T/C	-	Tolerated (0.34)		X
LAMA4	rs2032567	G1117S	2	Benign (0.00)	6	112457390	G/A	-	Tolerated (1.00)		X
LAMA4	rs1050349	P1119R	2	Probably damaging (0.99)	6	112457383	C/G	-	Tolerated (0.13)		
LAMA4	rs70940811	V1315I	2	Benign (0.00)	6	112452195	G/A	-	Tolerated (0.43)		X
LAMA4		G1356R	1	Probably damaging (1.00)	6	112451145	G/C	-	Damaging (0.00)	X	
LAMA4	rs201094782	Y1391H	2	Probably damaging (1.00)	6	112450240	T/C	-	Tolerated (0.19)		
LAMA4	rs3734292	V1815I	2	Probably damaging (0.98)	6	112430669	G/A	-	Tolerated (0.18)		
LDB3		T28K	1	Probably damaging (1.00)	10	88428531	CA/AG	+			
LDB3	rs3740343	V55I	2	Benign (0.02)	10	88439193	G/A	+	Tolerated (0.42)		X
LDB3		E139K	1		10	88446896	G/A	+	Tolerated (0.79)		
LDB3	rs45521338	R218C	2	Benign (0.05)	10	88451756	C/T	+	Tolerated (0.07)		X
LDB3		S330P	1	Benign (0.20)	10	88476170	T/C	+	Tolerated (0.26)		X
LDB3	rs113817827	V426I	2	Benign (0.15)	10	88476458	G/A	+	Tolerated (0.28)		X
LDB3		M456R	1	Probably damaging (0.97)	10	88477741	T/G	+	Damaging (0.00)	X	
LDB3	rs145983824	P498L	1	Probably damaging (1.00)	10	88477867	C/T	+	Damaging (0.00)	X	
LMNA		R220H	1	Benign (0.37)	1	156104615	G/A	+	Damaging (0.01)		
MYBPC3	rs3729989	S236G	2	Benign (0.00)	11	47370041	A/G	-	Tolerated (1.00)		X
MYBPC3		E334K	1	Benign (0.33)	11	47367848	G/A	-	Damaging (0.00)		

Table S1 (continued)

Table S1 (continued)

Gene	dbSNP ID	Change in AA	Pattern	PP2 Prediction (score)	Chrs.	Position (B37)	cDNA substitution	Orientation	SIFT Prediction (score)	Predicted damaging ^a	Predicted tolerated ^b
MYBPC3	rs113276889	R409G	1	Benign (0.10)	11	47364698	A/G	-	Damaging (0.00)		
MYBPC3	G416S	G416S	1	Probably damaging (0.98)	11	47364677	G/A	-	Damaging (0.01)	X	
MYBPC3	P459fs	P459fs	1		11	47364464	delC	-		X	
MYBPC3	rs147359039	R835L	1	Probably damaging (0.93)	11	47359039	GC/TT	-			
MYBPC3	R895H	R895H	1	Benign (0.41)	11	47357481	G/A	-	Tolerated (0.19)		X
MYH6	rs28711516	G56R	2	Probably damaging (0.95)	14	23876267	G/A	-	Damaging (0.00)	X	
MYH6	rs365990	V1101A	2	Benign (0.00)	14	23861811	T/C	-	Tolerated (1.00)		X
MYH6	rs28730771	A1130T	3	Possibly damaging (0.71)	14	23859610	G/A	-	Tolerated (0.17)		
MYH6	rs34935550	E1295Q	2	Probably damaging (0.95)	14	23858697	G/C	-	Damaging (0.01)	X	
MYH6	rs45574136	Q1593L	3	Benign (0.00)	14	23855705	A/T	-	Tolerated (0.14)		X
MYH6	rs61742476	V1613A	3	Benign (0.00)	14	23855645	T/C	-	Tolerated (1.00)		X
MYH6	K1860R	K1860R	1	Benign (0.00)	14	23852515	AA/GG	-			
MYH7	rs121913653	T441M	1	Benign (0.02)	14	23898249	C/T	-	Tolerated (0.07)		X
MYH7	rs121913625	R453C	1	Probably damaging (1.00)	14	23898214	C/T	-	Damaging (0.00)	X	
MYH7	I736T	I736T	1	Probably damaging (1.00)	14	23894983	T/C	-	Tolerated (0.13)		
MYH7	rs121913628	E924K	1	Probably damaging (1.00)	14	23893268	G/A	-	Damaging (0.02)	X	
MYH7	LS1139LD	LS1139LD	1	Benign (0.44)	14	23889361	GTC/AGA	-			
MYH7	R1250W	R1250W	1	Probably damaging (0.99)	14	23888797	C/T	-	Damaging (0.02)	X	
MYH7	G1520R	G1520R	1	Benign (0.07)	14	23886163	G/A	-	Damaging (0.03)		
MYH7	R1897H	R1897H	1	Probably damaging (1.00)	14	23883068	G/A	-	Damaging (0.00)	X	
MYPN	R482K	R482K	1	Probably damaging (0.99)	10	69918370	G/A	+	Tolerated (0.12)		
MYPN	rs10823148	F628L	2	Benign (0.00)	10	69926334	C/G	+	Tolerated (0.67)		X
MYPN	rs10997975	S691N	2	Benign (0.00)	10	69933921	G/A	+	Tolerated (0.66)		X
MYPN	rs7916821	S707N	2	Benign (0.03)	10	69933969	G/A	+	Tolerated (0.39)		X
MYPN	rs3814182	S803R	2	Benign (0.12)	10	69934258	C/G	+	Tolerated (0.46)		X
MYPN	rs181848049	G847V	2	Probably damaging (1.00)	10	69934389	G/T	+	Damaging (0.01)	X	
MYPN	rs151282801	R1042C	1	Probably damaging (1.00)	10	69955255	C/T	+	Damaging (0.00)	X	

Table S1 (continued)

Table S1 (continued)

Gene	dbSNP ID	Change in AA	Pattern	PP2 Prediction (score)	Chrs.	Position (B37)	cDNA substitution	Orientation	SIFT Prediction (score)	Predicted damaging ^a	Predicted tolerated ^b
MYPN	rs7079481	P1135T	2	Probably damaging (1.00)	10	69959242	C/A	+	Damaging (0.00)	X	
MYPN	rs138313730	L1161I	1	Probably damaging (1.00)	10	69959320	C/A	+	Damaging (0.00)	X	
MYPN	rs199585352	S1296T	1	Probably damaging (0.99)	10	69970135	T/A	+	Damaging (0.02)	X	
RBM20	G40W		1	Probably damaging (1.00)	10	112404330	G/T	+	Damaging (0.00)	X	
RBM20	P48delinsPP		1		10	112404356	insGCC	+			
RBM20	rs143785916	R641Q	2	Probably damaging (0.92)	10	112572077	G/A	+	Tolerated (0.08)		
RBM20	rs138926584	R673Q	3	Probably damaging (1.00)	10	112572173	G/A	+	Damaging (0.02)	X	
RBM20	rs1417635	W768S	3	Benign (0.00)	10	112572458	G/C	+	Tolerated (0.80)		X
RBM20	Q856X		1		10	112579845	C/T	+		X	
RBM20	rs188054898	R1057Q	2	Benign (0.00)	10	112581547	G/A	+	Tolerated (0.85)		X
RBM20	R1182H		1	Probably damaging (1.00)	10	112590912	G/A	+	Damaging (0.02)	X	
RBM20	rs942077	E1223Q	2	Benign (0.32)	10	112595719	G/C	+	Tolerated (0.12)		X
SCN5A	rs199473071	R225Q	1	Possibly damaging (0.88)	3	38655263	G/A	-	Damaging (0.00)	X	
SCN5A	rs199473561	A226V	1	Probably damaging (0.95)	3	38655260	C/T	-	Damaging (0.03)	X	
SCN5A	rs1805124	H558R	2	Benign (0.00)	3	38645420	A/G	-	Tolerated (1.00)		X
SCN5A	rs45600438	R568C	1	Possibly damaging (0.64)	3	38645391	C/T	-	Damaging (0.01)	X	
SCN5A	R659W		1	Probably damaging (0.98)	3	38640457	C/T	-	Damaging (0.00)	X	
SCN5A	rs1805125	P1090L	2	Benign (0.03)	3	38620946	C/T	-	Tolerated (0.65)		X
SCN5A	rs41310765	A1180V	1	Benign (0.02)	3	38616915	C/T	-	Tolerated (0.33)		X
SCN5A	rs41261344	R1193Q	2	Benign (0.01)	3	38616876	G/A	-	Tolerated (0.12)		X
SCN5A	rs199473251	I1448N	1	Possibly damaging (0.76)	3	38598026	T/A	-	Damaging (0.00)	X	
SCN5A	P1619T		1	Probably damaging (0.93)	3	38593008	C/A	-	Damaging (0.00)	X	
SGCD	C34G		1	Probably damaging (1.00)	5	155771595	T/G	+	Damaging (0.01)	X	
SGCD	P253fs		1		5	156186287	insC	+		X	
SGCD	Q283R		1	Probably damaging (0.97)	5	156186376	A/G	+	Tolerated (0.10)		
TAZ	H111N		1	Benign (0.08)	X	153641865	C/A	+	Tolerated (0.29)		X

Table S1 (continued)

Table S1 (continued)

Gene	dbSNP ID	Change in AA	Pattern	PP2 Prediction (score)	Chrs.	Position (B37)	cDNA substitution	Orientation	SIFT Prediction (score)	Predicted damaging ^a	Predicted tolerated ^b
TNNI72		R151Q	1	Probably damaging (1.00)	1	201333463	G/A	-	Damaging (0.04)	X	
TNNI72	rs3730238	K260R	3	Benign (0.01)	1	201330429	A/G	-	Tolerated (0.27)		X
TPM1		D34E	1	Benign (0.00)	15	63335130	C/A	+	Tolerated (1.00)		X
VCL	rs144683137	M209L	2	Benign (0.01)	10	75834503	A/T	+	Tolerated (1.00)		X
VCL	rs201528612	P398S	2	Benign (0.29)	10	75849796	C/T	+	Tolerated (0.09)		X

AA, Amino Acid; Chrs., chromosome; PP2, PolyPhen 2; PP2 scores range from 0 to 1, with three levels of pathogenic potential (probably damaging, possibly damaging, and benign); PP2 scores were obtained by using a web-based tool, HumVar model, at <http://genetics.bwh.harvard.edu/pph2/index.shtml>. SIFT, Sorts Intolerant From Tolerant; SIFT scores range from 0 to 1 (damaging or tolerated) with 0.05 as the threshold value; SIFT scores were obtained by using a web-based tool at http://sift.dna.org/www/Extended_SIFT_chr_coords_submit.html. a, b: A variant is predicted to be damaging when both PP2 and SIFT scores fall in the damaging ranges, while it is predicted to be tolerated when both scores are in the benign or tolerated ranges.

Table S2 Comparison of symptoms between the sporadic DCM patients with and without risk variants

Patient ID	Gender (female=0 male=1)	Age (years)	Drinking (no=0 yes=1)	Smoking (no=0 yes=1)	Hypertension (no=0 yes=1)	Diabetes (no=0 yes=1)	Symptoms to diagnosis (months)	Diagnosis to inclusion (months)	Arrhythmias ¹	NYHA class	Number of risky variants
DCM patients without risk variants											
1398	1	17	0	0	0	0	(N.A.)	180	N	4	0
1400	1	22	0	1	0	0	4	0	3	3	0
1403	1	40	0	0	0	0	(N.A.)	48	1; 3; 4	3	0
1408	0	56	0	0	1	1	6	0	N	4	0
1409	1	61	0	1	0	0	(N.A.)	240	3; 4	3	0
1412	0	60	0	0	0	0	304	120	3; 5	3	0
1421	0	66	0	0	0	1	(N.A.)	96	N	3	0
1424	1	36	(N.A.)	(N.A.)	0	0	(N.A.)	NA	N	4	0
1426	0	62	0	0	0	0	240	0	7	3	0
1432	1	58	0	1	1	0	6	0	3; 4	4	0
1433	1	36	0	1	0	0	2	0	N	2	0
1437	0	42	0	1	0	0	49	11	N	4	0
1446	1	59	1	1	0	1	0	26	N	4	0
1447	0	46	0	0	1	0	0	22	N	1	0
1448	1	49	1	0	0	0	0	11	N	0	0
1454	1	48	0	0	0	0	60	36	N	1	0
1455	1	28	0	0	0	0	0	0	N	3	0
1459	1	51	0	0	0	0	0	60	N	4	0
1462	1	63	1	0	0	1	0	0	N	3	0
1463	1	41	0	1	1	1	7	7	N	4	0
1468	1	77	0	0	0	0	5	5	N	2	0
1471	1	65	1	0	0	0	60	0	N	4	0
1472	1	20	0	0	0	0	0	2	N	2	0
1473	1	81	0	0	0	0	0	60	N	3	0
1475	1	40	0	0	0	0	12	0	N	2	0
1479	1	45	0	0	0	0	0	84	N	4	0
Mean ± SD		48.8±16.6					36.0±81.5	40.3±62.0		3±1	
DCM patients with risk variants											
1401	1	36	1	1	0	0	12	24	0	3	2
1402	1	38	0	0	0	0	(N.A.)	36	4	4	1
1404	1	43	0	1	1	0	(N.A.)	36	N	3	2
1405	0	51	0	0	0	0	1	0	N	3	2
1406	0	51	0	0	0	0	(N.A.)	96	7	4	1
1410	1	77	0	1	1	0	1	0	N	4	4
1411	1	65	0	0	0	1	1	0	5	3	2
1414	1	52	0	0	0	0	0	9	3	3	1

Table S2 (continued)

Table S2(continued)

Patient ID	Gender (female=0 male=1)	Age (years)	Drinking (no=0 yes=1)	Smoking (no=0 yes=1)	Hypertension (no=0 yes=1)	Diabetes (no=0 yes=1)	Symptoms to diagnosis (months)	Diagnosis to inclusion (months)	Arrhythmias ¹	NYHA class	Number of risky variants
1416	1	26	0	1	0	0	1	0	N	3	1
1417	0	51	0	0	0	0	(N.A.)	60	3; 4	4	2
1418	1	60	1	1	0	0	(N.A.)	48	7	3	2
1419	0	55	0	0	0	0	7	1	3	3	3
1428	1	36	0	0	1	0	1	0	N	3	1
1429	1	34	0	1	1	0	2	0	N	1	1
1430	1	61	(N.A.)	(N.A.)	0	0	(N.A.)	(N.A.)	N	3	1
1431	0	16	0	0	0	0	9	0	N	3	2
1435	1	82	0	1	0	0	24	0	5; 7	2	1
1436	0	62	0	0	0	0	31	5	1; 3	2	1
1438	1	19	0	1	0	0	0	0	11	3	1
1439	0	17	0	0	0	0	(N.A.)	(N.A.)	N		1
1440	1	53	0	0	0	0	60	0	N	3	1
1442	1	58	1	1	0	1	11	60	N	4	2
1443	1	50	1	0	0	1	0	0	N	4	1
1444	1	50	1	1	0	0	0	0	N	1	1
1449	1	47	0	0	0	0	0	33	N	3	1
1451	0	63	1	1	0	0	0	5	N	4	2
1452	0	62	0	1	0	0	7	57	N	1	2
1453	0	36	1	0	1	0	0	0	N	4	1
1456	1	69	0	0	0	0	0	18	N	4	1
1457	1	54	0	0	0	0	19	18	N	4	1
1461	1	66	0	0	0	0	2	18	N	4	1
1464	0	51	0	0	0	0	0	12	N	3	1
1465	1	56	1	1	1	0	26	0	N	3	1
1466	1	42	1	1	1	0	2	0	N	2	3
1467	1	48	0	0	1	0	2	2	N	4	1
1469	1	71	0	0	0	0	10	26	N	3	1
1470	1	36	1	1	0	0	1	0	N	4	1
1474	0	46	0	0	1	0	0	36	N	3	1
1478	1	29	0	0	0	0	0	6	N	3	1
1480	0	64	0	0	0	0	0	24	N	3	2
Mean ± SD		49.6±15.9					7.0±12.7	16.6±23.0		3±1	
P values ²	0.579	0.854	0.537	0.431	0.543	0.247	(0.602)	(0.258)	0.778	(0.812)	

1: Arrhythmias: atrial fibrillation or flutter=0; atrial premature beat=1; atrial tachycardia=2; ventricular premature beat=3; ventricular tachycardia=4; first-degree AV block=5; third-degree AV block=6; complete left or right bundle branch block=7. 2: Compared with the corresponding parameter of the patients without risky variants. P values in the parentheses were obtained from the Mann-Whitney rank sum test. N, no symptom; N.A., data not available; SD, standard deviation.

Table S3 Comparison of echocardiographic parameters between the sporadic DCM patients with and without risk variants

Patient ID	ARD (mm)	LVEDD (mm)	LVESD (mm)	LAD (mm)	IVST (mm)	LVPWT (mm)	FS (%)	LVEF (%)	RWT
DCM patients without risk variants									
1398	22	73	68	51	8	8	12	25	0.110
1400	23	84	76	50	8	6	15	30	0.071
1403	33	81	71	50	8	7	13	27	0.086
1408	35	60	51	56	10	10	25	50	0.167
1409	35	69	63	42	9	8	10	20	0.116
1412	30	85	78	46	8	8	13	26	0.094
1421	31	64	51	48	7	9	22	45	0.141
1424	37	76	67	67	9	8	18	36	0.105
1426	33	66	57	36	9	9	16	32	0.136
1432	41	72	56	45	10	9	16	32	0.125
1433	37	68	59	40	10	9	14	28	0.132
1437	25	75	65	51	8	8	13	25	0.107
1446	36	84	61	44	10	10	13	26	0.119
1447	30	65	53	37	8	8	20	40	0.123
1448	37	55	40	40	10	10	29	55	0.182
1454	23	54	45	49	8	8	18	35	0.148
1455	32	58	40	34	10	9	18	36	0.155
1459	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)
1462	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)
1463	33	44	37	39	10	10	31	59	0.256
1468	38	63	41	44	14	12	19	38	0.273
1471	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)
1472	31	64	44	35	9	10	25	52	0.286
1473	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)
1475	36	84	64	50	11	12	10	20	0.240
1479	30	71	64	49	9	8	12	24	0.163
Mean ± SD	32±5	69±11	57±12	46±8	9±1	8±1	17±6	35±11	0.152±0.061
DCM patients with risk variants									
1401	29	54	45	63	8	11	17	32	0.204
1402	26	74	56	66	7	7	17	34	0.095
1404	34	82	70	60	9	10	15	29	0.122
1405	30	59	42	45	8	8	29	56	0.136
1406	29	72	61	47	6	6	17	34	0.083
1410	34	74	55	52	12	13	20	39	0.176
1411	36	75	64	48	10	10	14	28	0.133
1414	35	64	56	52	10	10	20	40	0.156

Table S3 (continued)

Table S3 (continued)

Patient ID	ARD (mm)	LVEDD (mm)	LVESD (mm)	LAD (mm)	IVST (mm)	LVPWT (mm)	FS (%)	LVEF (%)	RWT
1416	30	67	58	52	7	7	12	24	0.104
1417	26	74	62	54	10	10	15	30	0.135
1418	39	81	71	53	12	11	16	33	0.136
1419	32	59	44	39	9	8	24	48	0.136
1428	27	62	43	42	9	10	21	42	0.161
1429	33	74	62	50	7	8	15	30	0.108
1430	35	88	73	58	9	10	15	30	0.114
1431	25	70	60	38	7	7	13	26	0.100
1435	34	66	47	36	10	10	25	50	0.152
1436	30	61	43	48	10	9	23	45	0.148
1438	22	57	48	31	5	9	16	31	0.158
1439	24	67	54	42	9	8	17	35	0.119
1440	30	65	52	50	9	9	18	36	0.138
1442	30	75	60	48	8	8	9	17	0.107
1443	32	65	58	40	8	8	10	21	0.123
1444	28	78	69	44	8	8	12	24	0.103
1449	25	76	63	52	10	10	15	31	0.132
1451	33	68	61	39	8	9	12	23	0.132
1452	30	75	64	68	11	11	14	29	0.147
1453	31	72	59	57	8	8	17	34	0.111
1456	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)
1457	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	16	32	
1461	37	59	43	39	7	9	18	35	0.231
1464	34	63	51	43	8	8	19	38	0.186
1465	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	13	26	
1466	34	75	58	68	10	11	15	29	0.162
1467	27	79	68	50	8	9	14	28	0.180
1469	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)
1470	34	54	38	41	10	10	32	60	0.244
1474	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)
1478	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)	(N.A.)
1480	30	77	65	53	8	8	14	28	0.151
Mean ± SD	31±4	69±8	57±9	49±9	9±2	9±1	17±5	34±9	0.142±0.037
P values	0.246	0.825	0.916	0.150	0.195	0.666	0.742	0.703	0.452

Compared with the corresponding parameter of the patients without risk variants; SD, standard deviation. ARD, aortic root diameter; FS, shortening fraction; LAD, left atrial diameter; LVEDD, left ventricular end-diastolic diameter; LVEF, left ventricular ejection fraction; LVESD, left ventricular end-systolic diameter; LVPWT, left ventricle post wall thickness; IVST, interventricular septal thickness; N, no symptom; N.A., data not available; RWT, relative wall thickness.