

Chinese herbal component, Praeruptorin E, enhances anti-asthma efficacy and prevents toxicity of aminophylline by targeting the NF- κ B/PXR/CYP3A4 pathway

Ronghua Xu^{1,2#^}, Huiming Deng^{1,2#^}, Lianfang Gan^{1#}, Lifan Zhong¹, Yanxi Deng¹, Qianru Wang¹, Chuanzhu Lv^{3,4}, Ling Huang^{1,4,5^}

¹Hainan Province Key Laboratory for Drug Preclinical Study of Pharmacology and Toxicology Research, Hainan Medical University, Haikou, China; ²Department of Gastrointestinal Surgery, Huazhong University of Science and Technology Union Shenzhen Hospital, Shenzhen, China; ³Key Laboratory of Emergency and Trauma of Ministry of Education, Hainan Medical University, Haikou, China; ⁴Research Unit of Island Emergency Medicine, Chinese Academy of Medical Sciences, Hainan Medical University, Haikou, China; ⁵School of Hainan Provincial Drug Safety Evaluation Research Center, Hainan Medical University, Haikou, China

Contributions: (I) Conception and design: L Huang, R Xu, H Deng; (II) Administrative support: L Huang, R Xu; (III) Provision of study materials or patients: R Xu, H Deng; (IV) Collection and assembly of data: H Deng, L Gan, L Zhong, Y Deng, Q Wang; (V) Data analysis and interpretation: H Deng, L Gan; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

[#]These authors contributed equally to this work.

Correspondence to: Ling Huang. Hainan Province Key Laboratory for Drug Preclinical Study of Pharmacology and Toxicology Research, Hainan Medical University, No. 3 Xueyuan Road, Haikou 571199, China; Research Unit of Island Emergency Medicine, Chinese Academy of Medical Sciences, Hainan Medical University, No. 3 Xueyuan Road, Haikou 571199, China; School of Hainan Provincial Drug Safety Evaluation Research Center, Hainan Medical University, No. 3 Xueyuan Road, Haikou 571199, China. Email: puer6@163.com; Chuanzhu Lv. Key Laboratory of Emergency and Trauma of Ministry of Education, Hainan Medical University, No. 3 Xueyuan Road, Haikou 571199, China; Research Unit of Island Emergency Medicine, Chinese Academy of Medical Sciences, Hainan Medical University, No. 3 Xueyuan Road, Haikou 571199, China. Email: lvchuanzhu677@126.com.

Background: Aminophylline is widely used for the treatment of asthma, but the therapeutic dose is very close to the toxic dose, which makes this drug prone to accumulation poisoning. In the present study, we explored whether the Chinese herbal component, Praeruptorin E (PE), enhances anti-asthma efficacy and prevents the toxicity of aminophylline.

Methods: First, an ovalbumin (OVA)-induced mouse model of asthma, immunohistochemistry, pathological staining, and bronchoalveolar lavage fluid (BALF) were used to detect the lung condition of asthmatic mice. The content of Th2 cytokines in serum was measured by enzyme-linked immunosorbent assay (ELISA), and the expression of related proteins was detected by Western blotting and immunofluorescence. Concentrations of theophylline and its metabolites in rat serum were analyzed by liquid chromatography-tandem mass spectrometry (LC-MS/MS). siRNA transfection and chromatin immunoprecipitation (ChIP) were used to investigate the mechanism of PE.

Results: PE was found to synergize with aminophylline to reduce the infiltration of inflammatory cells, collagen deposition, and mucus hyperplasia in the lungs of asthmatic mice. It inhibited the expression of Th2 cytokines, interleukin (IL)-4, IL-5, and IL-13; promoted lung tissue repair; and reduced the toxic effect of aminophylline on the heart. Moreover, LC-MS/MS analysis showed that PE reduced the plasma concentration of the parent theophylline and its metabolite 1,3-dimethyluric acid (1,3-DMU). PE facilitated aminophylline's suppression of nuclear factor- κ B (NF- κ B), and increased the expression of the xenobiotic nuclear receptor pregnane X receptor (PXR) and its primary target gene, *CYP3A11* [this is the mouse

[^] ORCID: Ronghua Xu, 0000-0002-5745-8915; Huiming Deng, 0000-0002-1409-6926; Ling Huang, 0000-0002-5173-689X.

homolog of cytochrome P450 3A (*CYP3A*) in the asthmatic mouse liver and in the L-02 human fetal hepatocyte cell culture model. In addition, the ChIP assay revealed that PE attenuated the binding of *NF-κB* to the promoter region of the *PXR* gene and prevented the suppression of *PXR* gene expression by *NF-κB*.

Conclusions: PE has a dual function in enhancing the immune regulation and anti-inflammatory effects of theophylline, as well as preventing theophylline toxicity by targeting the *NF-κB*/*PXR*/*CYP3A4* axis. PE is a promising herbal medicine that will benefit asthmatics taking theophylline.

Keywords: Praeruptorin E (PE); aminophylline; nuclear factor-κB (*NF-κB*); pregnane X receptor (*PXR*); asthma

Submitted Nov 30, 2021. Accepted for publication Feb 18, 2022.

doi: 10.21037/atm-22-386

View this article at: <https://dx.doi.org/10.21037/atm-22-386>

Introduction

Asthma is a chronic inflammatory disease of the lungs, characterized by airway obstruction and high reactivity. There are about 300 million people with asthma worldwide, and about 250,000 asthma-related deaths occur every year (1,2). Th2 cytokines [such as interleukin (IL)-4, IL-5, and IL-13] play a key role in the pathogenesis of allergic asthma. These cytokines promote eosinophil infiltration, the release of immunoglobulin E (IgE), airway hyperreactivity, and mucus secretion, leading to the development of allergic asthma (3-5). Currently, anti-airway allergic inflammatory drugs and bronchodilators are the main asthma therapies (6). Aminophylline has been used for the treatment of asthma for many years, producing smooth muscle relaxation and immune regulation, and having anti-inflammatory activity (7). Although aminophylline has a good therapeutic effect on asthma symptoms, in a clinical study, it was found that 21% (9/43) of patients died from aminophylline poisoning, of which 4 patients had toxic levels of theophylline in their serum (8). Another study reported that treatment with aminophylline can lead to cardiac toxicity, such as arrhythmias, chest tightness, and elevated myocardial enzymes (9). One report demonstrated that monitoring plasma concentration levels of theophylline during administration could improve safety (10). However, the concentration of theophylline in the blood cannot be monitored continuously due to the limitations of clinical measurements. Therefore, there is an urgent need to find compatible drugs that can reduce the toxicity of aminophylline and enhance its efficacy in clinical practice.

Nuclear factor-κB (*NF-κB*) is a multipotent transcription factor that has been shown to induce asthma both in mouse models and asthmatic patients (11). Inflammatory factors, such as IL-1β, IL-4, and IL-5, which are regulated

by *NF-κB*, can promote the development of asthma, whereas cytokines regulated by *NF-κB* can promote the expression of *NF-κB* in asthma (12,13). *NF-κB* activation is also closely associated with airway tissue remodeling (fibrosis of surrounding airway tissues, hyperplasia and hypertrophy of smooth muscle cells, and myxogenesis) (14). When asthmatics have airway inflammation and hyperresponsiveness, *NF-κB* and its inflammatory factors can reduce the metabolic clearance activity of theophylline and increase the incidence of theophylline accumulation toxicity, which is more likely to occur in children and elderly patients (15,16). *NF-κB*, a key factor in the inflammatory process, regulates the gene expression of cytochrome P450 3A (*CYP3A*) through the pregnane X receptor (*PXR*), thereby affecting drug metabolism (17). *PXR* activation can upregulate the expression of the metabolic enzyme *CYP3A*, promote drug metabolism, and inhibit the activity of *NF-κB* and the expression of *NF-κB* target genes (18,19). Therefore, inhibition of the *NF-κB* pathway is not only a synergistic anti-asthma effect, but also an effective strategy to reduce the risk of aminophylline poisoning.

Peucedanum praeruptorum Dunn is a perennial herb of the *Peucedanum* genus in the *Apiaceae* family. It has a variety of active ingredients that have antihypertensive, antiosteoporosis, and antitumor biological effects (20-22). It can also inhibit *NF-κB* activity to produce anti-inflammatory, anti-asthmatic, and antitussive phlegm effects (23). Our previous results indicated that the active components of *Peucedanum praeruptorum* Dunn could upregulate the expression and functional activity of *CYP3A4* through the *PXR* pathway (24-26). In the present study, an ovalbumin (OVA)-induced asthma model in mice and *in vitro* experiments showed that Praeruptorin E (PE) could inhibit the expression of *NF-κB* to enhance the

effect of aminophylline treatment in asthma. In addition, the inhibition of NF- κ B by PE promoted the expression of PXR and enhanced the functional activity of the enzyme *CYP3A11* and reduced the accumulation of theophylline and 1,3-dimethyluric acid (1,3-DMU) in asthmatic mice. These findings indicate that PE has a dual function in enhancing the immune regulation and anti-inflammatory effects of theophylline, as well as preventing theophylline toxicity by targeting the NF- κ B/PXR/CYP3A4 axis. Our findings are expected to reduce the incidence of adverse reactions to aminophylline in clinical treatment of asthma.

We present the following article in accordance with the ARRIVE reporting checklist (available at <https://atm.amegroups.com/article/view/10.21037/atm-22-386/rc>).

Methods

Animals

All experiments were approved by the Institutional Animal Care and Treatment Committee of Hainan Medical University (Ratified on October 16, 2017). All procedures involving animals were performed in accordance with the Guidelines for the Humane Treatment of Laboratory Animals (Ministry of Science and Technology of the People's Republic of China; policy No. 2006398). BALB/c male mice (weighing 18–22 g, drug efficacy study) and male Sprague-Dawley (SD) male rats (weighing 160–180 g, metabolic analysis) were purchased from Shanghai SLAC Laboratory Animal Co., Ltd. (Shanghai, China). All animals were bred under the specific pathogen-free in the Hainan Provincial Drug Safety Evaluation Research Center [animal license No. SYXK [Qiong] 2016-0013]. The mice were given standard rodent chow and secondary purified water. The feed environment was a 12-h light/dark cycle, temperature 22–25 °C, and humidity 40–70%. The animals were adapted to the experimental environment for 7 days.

Grouping and the asthma model

The experimental design and dosage of PE (24) are shown in *Figure 1A, 1B*. Briefly, for the asthma model, BALB/c mice and SD rats were intraperitoneally sensitized with 10 μ g or 10 mg of OVA (Sigma-Aldrich) and 20 or 100 mg Al(OH)₃ gel on days 8 and 15. From days 15 to 25, the rats were challenged on 10 consecutive days with the administration of 1 mg/mL or 2% OVA inhalation, 30 min/day. The

normal control group received the same volume of saline subcutaneously (days 8 and 15) and inhalation (days 15–25).

Chemicals and reagents

PE (purity \geq 98.0%) was purchased from Chengdu purechem-standard (Chengdu, China), the PE structure is shown in *Figure 2A*; aminophylline was obtained from Sigma-Aldrich (Louis, MO, USA); human liver L-02 cells were obtained from the Shanghai Cell Bank of the Chinese Academy of Sciences; RPMI-1640 medium and fetal bovine serum (FBS) were purchased from Gibco (Carlsbad, CA, USA); polyvinylidene difluoride (PVDF) membrane was obtained from Millipore (Billerica, CA, USA). The enhanced chemiluminescence (ECL) detection system was from Beyotime Biotechnology (Shanghai, China).

Analysis of bronchoalveolar lavage fluid (BALF)

After anesthetizing the BALB/c mice with chloral hydrate (Sigma-Aldrich), injected intraperitoneally, the mice were humanely killed, the left lungs were lavaged 3 times with cold phosphate-buffered saline (PBS), and BALF was harvested. A total of 1 mL of fresh BALF was taken and centrifuged at 1,500 \times g for 10 min at 4 °C; 10 μ L of BALF pellets were dropped on a glass slide to prepare a smear and then fixed in methanol for 30 s. The BALF smear was immersed in Giemsa (Sigma-Aldrich) staining solution for 20 s at room temperature, and washed with PBS for 1 min. Sections were then immersed in 1% acetic acid solution for 30 s after rinsing 3 times with PBS. Immune cells were then counted under a light microscope at \times 400 magnification.

Enzyme-linked immunosorbent assay (ELISA)

BALF and serum were collected from the left lung or retro-orbital plexus. Concentrations of IL-4, IL-5, IL-13, IgE, NF- κ B p65, lactate dehydrogenase (LDH), hydroxybutyrate dehydrogenase (HBD), creatine kinase (CK)-MB, CK, and aspartate aminotransferase (AST) were measured using ELISA kits (R&D Systems, Minneapolis, MN, USA), according to the manufacturer's instructions.

Electrocardiogram (ECG) collection

After anesthesia, the limbs of mice were connected to the electrodes of the Powerlab multichannel physiological

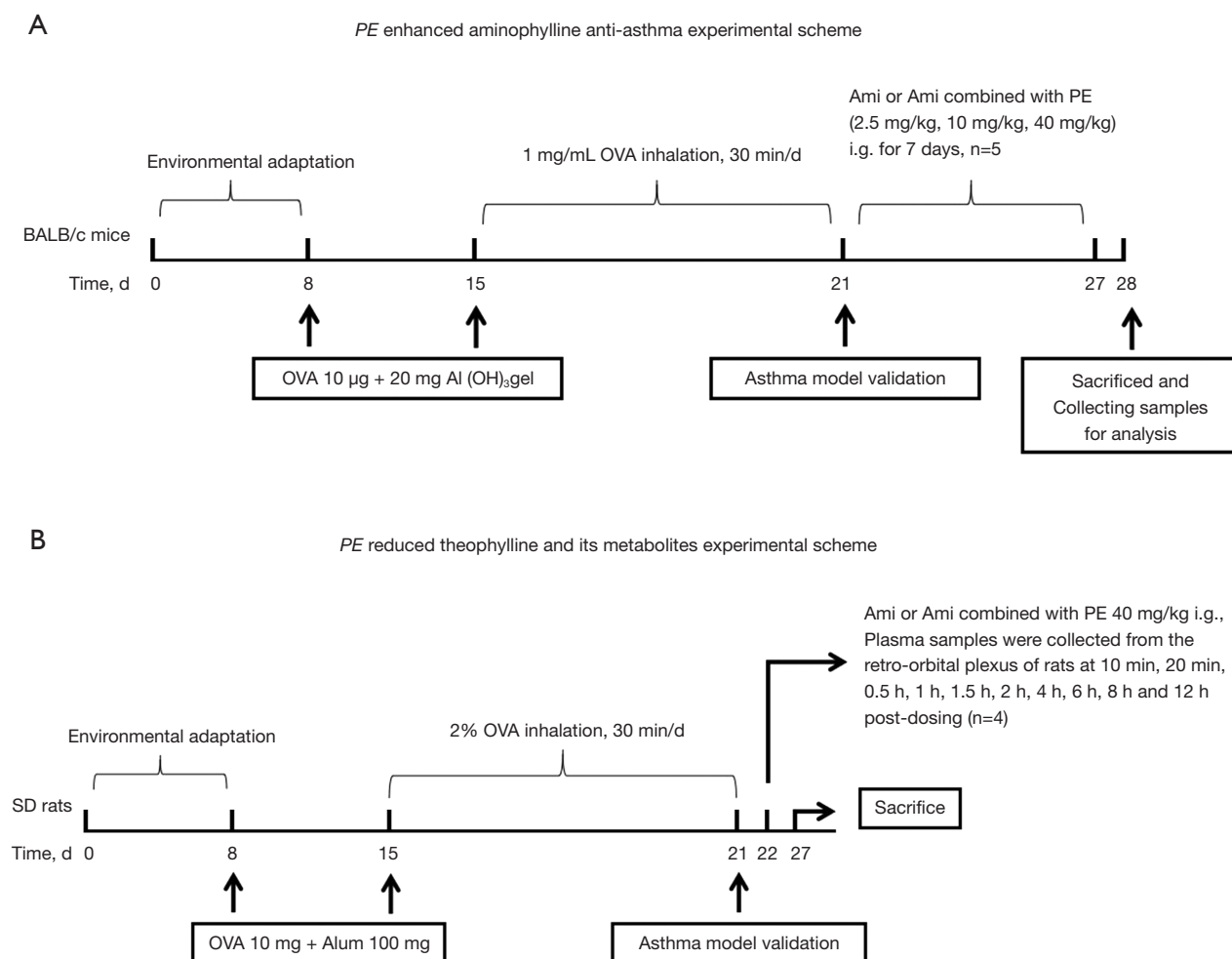


Figure 1 Schematic diagram of the experimental design. PE, Praeruptorin E; OVA, ovalbumin.

recorder (9020P; Nihon Kohden, Tokyo, Japan). Recording was started after display of a satisfactory lead II ECG. A continuously recorded 300-s ECG segment was printed to determine mouse heart rate and to analyze the ECG signal.

Histopathological analysis

Briefly, the right upper lobe lung and trachea were fixed in 4% formaldehyde for 4 days. Each tissue was sliced into 4-µm sections and stained with hematoxylin and eosin (H&E). Periodic acid-Schiff (PAS) and Masson's trichrome (M-T) (Sigma-Aldrich) stains were used to detect the infiltration of inflammatory cells, acidic mucus chromophores, and collagen deposition based on the manufacturer's protocols. Lung and peri-bronchiole

inflammation was evaluated using a score of 0–4 (0, no cells; 1, a few cells; 2, a ring of cells 1 cell deep; 3, a ring of cells 2–4 cells deep; 4, a ring of cells >4 cells deep). Image-Pro Plus software (Media Cybernetics, Maryland, USA) was used to analyze of PAS and M-T stained specimens, as previously described (27).

Immunohistochemical analysis

The right lower lobe lung was fixed in 4% formaldehyde for 4 days. Each tissue was sliced into 4-µm sections, de-waxed, and peroxidase inhibited. Specimens were incubated with the primary antibody at 4 °C overnight. Subsequently, they were washed with PBS 3 times and incubated with appropriate secondary antibodies (PowerVision Two-

Step Histostaining Reagent, PV6001; Beijing Zhong Shan-Golden Bridge Biological Technology, Beijing, China) for 15 min at 37 °C. The samples were stained with 3,3N-Diaminobenzidine Tetrahydrochloride (DBA) Horseradish Peroxidase Color Development Kit (P0203; Beyotime Biotechnology) and counterstained with hematoxylin (Sigma-Aldrich). The progress of the reaction was observed with the aid of a microscope (IX73; Olympus, Tokyo, Japan).

Liquid chromatography-tandem mass spectrometry (LC-MS/MS)

The instruments used for LC-MS/MS were the Agilent 1290 ultra-high performance LC system and Agilent 6410B triple quadrupole mass spectrometer (Thermo Scientific, Waltham, MA, USA).

Method of LC

A Capcell Pak MGII-C18 (2 mm × 100 mm, 3 μm) (Shiseido, Tokyo, Japan) was used as the chromatographic column. Mobile phase A used pure water and mobile phase B used methanol, both containing 0.5% acetic acid. The gradient elution procedure was as follows: 0 min → mobile phase B 10%, 0.5 min → mobile phase B 30%, 1.0 min → mobile phase B 90%, 2 min → mobile phase B 90%, 2.3 min → mobile phase B 93%, 2.5 min → mobile phase B 10%, and 0.5 min → mobile phase B 30%. Other conditions included column temperature of 25 °C; injection volume of 1 μL, flow rate of 0.3 mL/min and running time of 4 min. Under the above conditions, the baseline separation was realized between the targeting analyte and the internal standard.

MS method

The electrospray ion source positive ion detection and multireaction monitoring methods were used for mass spectrometry detection. The mass spectrum parameters were capillary voltage +4,000 V with capillary temperature of 350 °C. The drying gas flow rate was 10 L/min and atomized gas at 30 psi. The specific detection method including ion pair (m/z), fragmentor, and collision energy (CE) for each analyte were as follows:

Theophylline: ion pair 179.2→164.1 m/z, fragmentor 104 V, CE 16 V;

1,3-DMU: ion pair 195.1→180.1 m/z, fragmentor 119 V, CE 12 V;

1-Methyluric acid (1-MU): ion pair 181.0→138.1 m/z, fragmentor 129 V, CE 12 V.

Standard curve

A total of 50 μL blank rat plasma was mixed with a series of working solutions, including 5 μL theophylline (0.5, 1, 2, 5, 10, 20, 50, 100, 200, 500, 1,000 μg/mL), 1-MU, 1,3-DMU (0.05, 0.1, 0.2, 0.5, 1, 2, 5, 10, 20, 50, 100 μg/mL), and 5 μL internal standard solution (1 μg/mL toluene sulfonylurea, aqueous solution). After adding 150 μL acetonitrile and vortexing for 10 min, the solution was centrifuged at 14,000 ×g and 4 °C for 10 min. Subsequently, 1 μL supernatant was injected for the LC-MS/MS analysis.

Quality control of sample

A total of 50 μL blank rat plasma was mixed with a series of working solutions, including 5 μL theophylline (1, 100, 800 μg/mL), 1-MU (0.1, 10, 80 μg/mL), 1,3-DMU (1, 10, 80 μg/mL), and 5 μL internal standard solution (1 μg/mL toluene sulfonylurea, aqueous solution). After adding 150 μL acetonitrile and vortexing for 10 min, the solution was centrifuged at 14,000 g and 4 °C for 10 min. Subsequently, 1 μL supernatant was injected for LC-MS/MS analysis.

Experimental animal sample

Fifty μL plasma was mixed with 5 μL water and 5 μL internal standard solution (1 μg/mL toluene sulfonylurea, aqueous solution). After mixing, 150 μL acetonitrile was added and vortexed for 10 min, followed by centrifugation at 14,000 ×g and 4 °C for 10 min. Subsequently, 1 μL supernatant was injected for the LC-MS/MS analysis.

Cell culture and transfection

L-02 cells were cultured in RPMI-1640 containing supplements of FBS (10%), penicillin (100 U/mL), and streptomycin (100 μg/mL) (Gibco) in a humidified sterile incubator at 37 °C, with gaseous 5% CO₂ added to the atmosphere.

siRNA-PXR was designed and synthesized by Gene Pharma (Shanghai, China). The transfection of siRNA-PXR was carried out using Lipofectamine 2000 transfection reagent (Gibco), according to the manufacturer's protocol. The L-02 cells were infected for 72 h, then the stably transfected cells were treated with PE and lipopolysaccharide (LPS) (Sigma-Aldrich) for 24 h.

Reverse transcription polymerase chain reaction (RT-PCR)

Total RNA was extracted from L-02 cell lung and liver tissue using TransZol Reagent (Generay, 1703G01,

Shanghai, China). Reverse transcription was performed using HiScript-II Q RT SuperMix for qPCR (Vazyme, 7E092G6). Qualitative reverse transcription PCR (qRT-PCR) was performed using ChamQ SYBR Color qPCR Master Mix (Vazyme, 7E092G6, Nanjing, China) and a CFX Connect Real-Time PCR System instrument (Bio-Rad, Hercules, CA, USA), with the supporting analysis software, Bio-Rad CFX Manager (Bio-Rad, Hercules, CA, USA). The cycling conditions were as follows: 95 °C for 30 s, followed by 35 cycles at 95 °C for 10 s, 60 °C for 30 s, 60 °C for 30 s plate read. The sequences of the primer sets (Takara Bio, Talian, China) used for this analysis were as follows: *NF-κB*: 5'-AGGATTTCGATTCCGCTACG-3' (forward) and 5'-CTCACAGTGCTTGCCCACC-3' (reverse), *PXR*: 5'-ATGGACGCTCAGATGCAAAC-3' (forward) and 5'-GGAGGGAGGTTGGTAGTTCC-3' (reverse), *CYP3A4*: 5'-CAATAAGGCACCACCCACCTAT-3' (forward) and 5'-AGGTTTGCCTTTCTCTGCC-3' (reverse), *CYP3A11*: 5'-GGCAAGCCTGTTACTATGAAAG-3' (forward) and 5'-ACTGAGAAGAGCAAAGGATCA A-3' (reverse), *CYP1A2*: 5'-ATCCTGGAGATCTACCGA TACA-3' (forward) and 5'-TATGTAGATACAGCGCTCC TTG-3' (reverse), *CYP2E1*: 5'-CCAACTCTGGACTCCC TTTTAT-3' (forward) and 5'-ACGCCTTGAAATAGTCA CTGTA-3' (reverse), and *actin*: 5'-CCCATCTATGAGGG TTACGC-3' (forward) and 5'-TTCTTGCTGAATCT TTCAGGGAG-3' (reverse). The results were analyzed using a $2^{-\Delta\Delta C_t}$ relative quantification method, and each sample was tested in triplicate with 5 wells per group.

Western blotting

Lung and liver tissues and L-02 cells were lysed in RIPA buffer, and a BCA kit (Beyotime Biotechnology) was used to quantify the level of protein in a sample. Protein bands were separated by polyacrylamide gels electrophoresis (SDS-PAGE, 8–14%) and then repositioned on PVDF membranes. After appropriate blockade, PVDF membranes were incubated with the following primary antibodies: NF-κB p65 (1:1,000) (Cell Signaling Technology, Boston, MA, USA); PXR; CYP3A1, CYP1A2, and CYP2E1 (1:1,000) (Abcam, Cambridge, MA, USA); GAPDH (1:1,000) (MAB5465, MultiSciences, Hangzhou, China); and goat antirabbit IgG (1:5,000) (70-GAR007, MultiSciences), then incubated at room temperature with secondary antibodies for 1 h. ECL chemicals (Beyotime Biotechnology) were used to examine the target proteins. An image analyzer

system (ChemiDoc XRS+; Bio-Rad, Hercules, CA, USA) was used to quantify the results.

Chromatin immunoprecipitation (ChIP)

ChIP assays were conducted using a SimpleChIP Plus Enzymatic Chromatin IP Kit (Cell Signaling Technology), according to the manufacturer's instructions. Briefly, chromatin from cross-linked L-02 cells was sheared by sonication and incubated overnight with specific antibodies (NF-κB; Cell Signaling Technology) followed by proteinase K decrosslinking and purified DNA. Precipitated DNA and input DNA were analyzed using qualitative PCR (qPCR), and the results were presented as normalization to the input DNA. The results were analyzed and normalized by the C_t value obtained by qPCR, %Input = $2^{-\Delta C_t}$ (normalized ChIP) × 100%. The primers for PXR were as follows: 5'-TACATAAAATACCTAGAATA-3' (forward) and 5'-ATTTAAATGGTTAATTTTAT-3' (reverse).

Molecular docking

We searched for the structural formula of PE (ID:013078) in TCMSP (<https://www.tcmsp-e.com/>). The structure of the NF-κB p65 protein (ID:6QHL) was downloaded from the crystal structure data of PDB (<https://www.pdbus.org/>). AutoDock software (version 1.5.6, Scripps Research Institute, San Diego, CA, USA) was used to eliminate the water molecules of the target protein, and to add non-polar hydrogen and calculate the Gasteiger charge. The ligand structure was optimized in an MM2 force field by using Chem3D software (CambridgeSoft, MA, USA). AutoDock Vina 1.1.2 software was used for docking and Visual Studio software (2016, Microsoft, Redmond, USA) was used to visualize the docking results and select the conformation with the lowest interaction energy.

Statistical analysis

The experimental results are expressed as the mean ± standard error of the mean, and the differences between groups were analyzed by one-way analysis of variance and Student's *t*-test using GraphPad Prism 8 (GraphPad Software, San Diego, CA, USA). $P < 0.05$ indicated that the difference was statistically significant. All *in vitro* data are representative of three independent experiments.

Results

PE enhances the inhibitory effect of aminophylline on OVA-induced inflammation

Giemsa and H&E staining was used to investigate whether PE could enhance the therapeutic effect of aminophylline on airway inflammation induced by OVA in mice. As shown in *Figure 2B*, the total number of inflammatory cells, eosinophils, monocytes, neutrophils, and lymphocytes in the OVA group was significantly higher than in the control group. In contrast, the total number of inflammatory cells and eosinophils in the aminophylline group was lower than in the OVA group, while the aminophylline + PE group exhibited even lower numbers than the aminophylline-alone group. The results revealed that PE could enhance aminophylline efficacy and reduce inflammatory effects in the trachea.

The H&E staining results showed that the lung tissue structure was normal without any pathological changes in the control group. A large number of inflammatory cells was observed in the OVA group, mainly eosinophils and neutrophils. The alveolar walls were significantly thickened, most of the alveolar cavities were shrunk or even collapsed, and some of the alveoli were overextended. The aminophylline group exhibited some inflammatory cell infiltration, a small number of eosinophils, and local thickening of the alveolar walls. The number of inflammatory cells in the aminophylline + PE group was significantly lower than in the aminophylline group, and only a small number of eosinophils were found. The local alveolar wall was slightly thickened and some alveolar structures were normal, with a few alveolar dilatations. The least severe pathological changes were observed in the aminophylline + 40 mg/kg PE group. At the same time, the pulmonary pathological immune score of the aminophylline + PE group was lower than for the aminophylline group (*Figure 2C, 2D*).

In the OVA group, the bronchial mucosa was loose and exhibited edema, epithelial cilia were adherent and prostrate, and the pseudo-lamellar structure had disappeared. The submucosal tissue was infiltrated with numerous inflammatory cells, including eosinophils, neutrophils, and monocytes. In the aminophylline group, the mucosal epithelium exhibited edema, part of the mucosal epithelium was exfoliated, while part of the mucosal epithelium was relatively normal with obvious cilium structures intact. Many inflammatory cells were observed in the submucosa, and part of the airway mucosa

and submucosa was exfoliated. In the aminophylline + PE group, the degree of mucosal epithelial edema was lower than in the aminophylline group. The infiltration of submucosal inflammatory cells was lower than in the aminophylline group with minimal, if any, eosinophils present. In addition, the bronchial pathological immune score of the aminophylline + PE group was lower than that of the aminophylline group (*Figure 2E, 2F*).

In addition to asthma mouse lung and bronchus inflammation manifestations, PAS and M-T stains were also used to observe mucus and collagen deposition in lung tissue (27). The results revealed that OVA group mouse lungs displayed remarkably increased airway acidic mucus secretion and collagen accumulation. Mucus secretion and collagen accumulation in asthmatic mice lung tissues was improved in the aminophylline group. In contrast, the mucus secretion and collagen deposition in the aminophylline + PE group was less than in the aminophylline treatment group (*Figure 3A-3D*). These results indicated that PE can enhance the effect of aminophylline by reducing inflammation, mucus secretion, and collagen cell fibrosis in asthmatic mouse lungs.

PE promotes aminophylline to reduce OVA-induced serum NF- κ B, IgE, and Th2 cytokine concentrations

Th2 cytokines, IgE, and NF- κ B play key roles in the pathogenesis of allergic asthma. Therefore, the concentrations of Th2 cytokines in BALF and serum concentrations of IgE and NF- κ B p65 were measured to investigate the synergism of PE with aminophylline in reducing airway inflammation. *Figure 4A-4E* shows that the concentrations of IL-4, IL-5, and IL-13 in BALF and serum IgE and NF- κ B p65 in the OVA group were significantly higher than in the control group. Treatment with aminophylline significantly reduced IL-4, IL-5, and IL-13 in BALF and serum IgE and NF- κ B p65 concentrations, while the addition of PE during aminophylline treatment markedly reduced these concentrations. To investigate further the expression of NF- κ B p65 in the lungs of asthmatic mice, we performed immunofluorescence analysis and found that the expression of NF- κ B p65 in asthmatic mice was significantly lower in the aminophylline + PE group than in the aminophylline alone group (*Figure 4F, 4G*). To further demonstrate that PE had potential in binding to the NF- κ B p65 and then inhibiting NF- κ B p65, we performed molecular docking. The binding modes of PE with the NF- κ B p65 complex are illustrated in *Figure 4H-4J*,

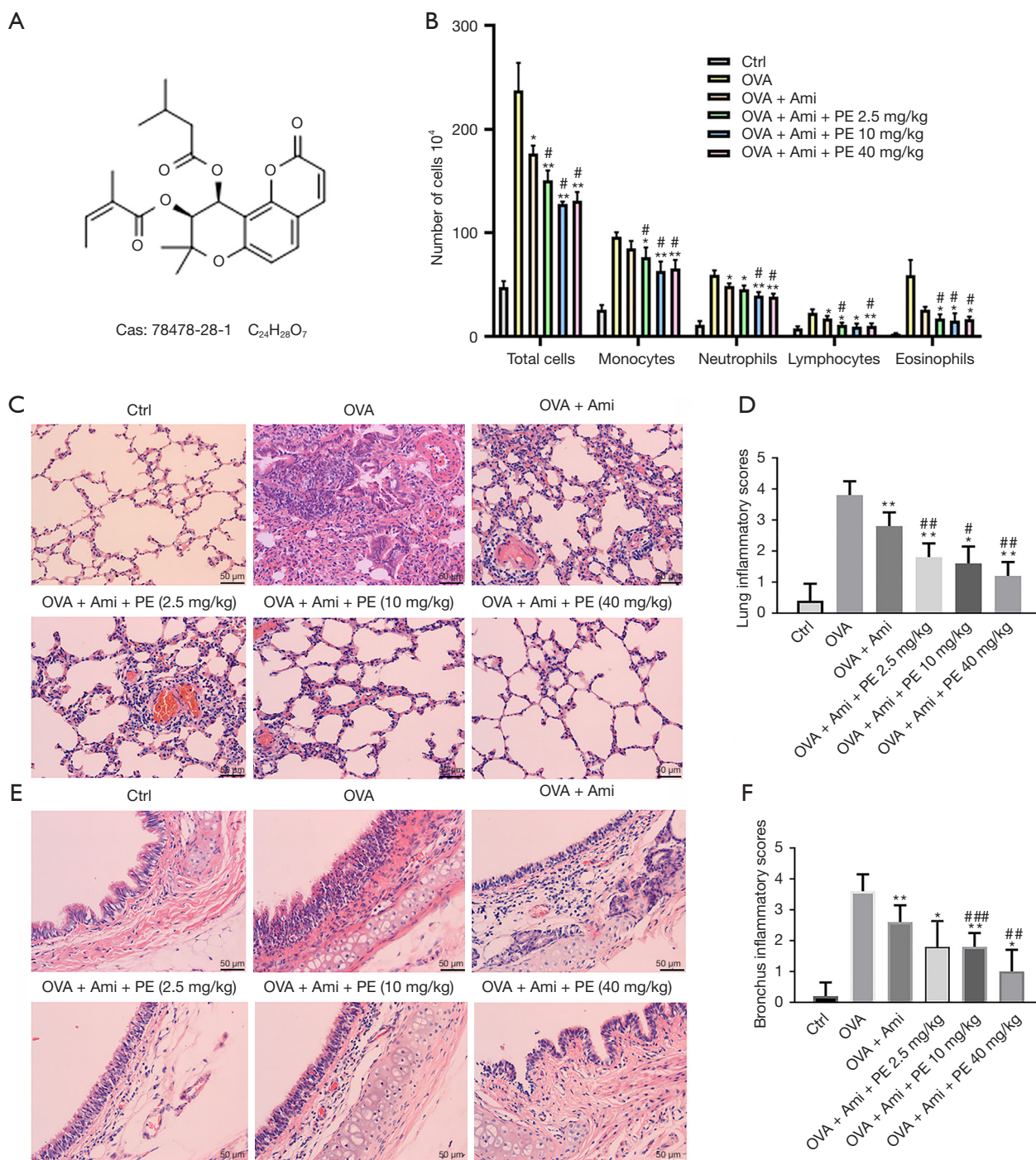


Figure 2 PE cooperates with Ami to reduce lung inflammation in asthmatic mice. (A) Chemical structure of PE. (B) Total number of inflammatory cells, monocytes, neutrophils, eosinophils, and lymphocytes in BALF. (C,D) H&E staining and the pathological immune score of lung tissue in asthmatic mice. (E,F) H&E staining and the pathological immune score of bronchi tissue in asthmatic mice. OVA + Ami group, OVA + Ami + PE 2.5 mg/kg group, OVA + Ami + PE 10 mg/kg group and OVA + Ami + PE 40 mg/kg group compared with the OVA group, * $P < 0.05$, ** $P < 0.01$; OVA + Ami + PE 2.5 mg/kg group, OVA + Ami + PE 10 mg/kg group and OVA + Ami + PE 40 mg/kg group compared with the OVA + Ami group, # $P < 0.05$, ## $P < 0.01$, ### $P < 0.001$. $n = 5$. PE, Praeruptorin E; Ami, aminophylline; BALF, bronchoalveolar lavage fluid; H&E, hematoxylin and eosin; ctrl, control; OVA, ovalbumin.

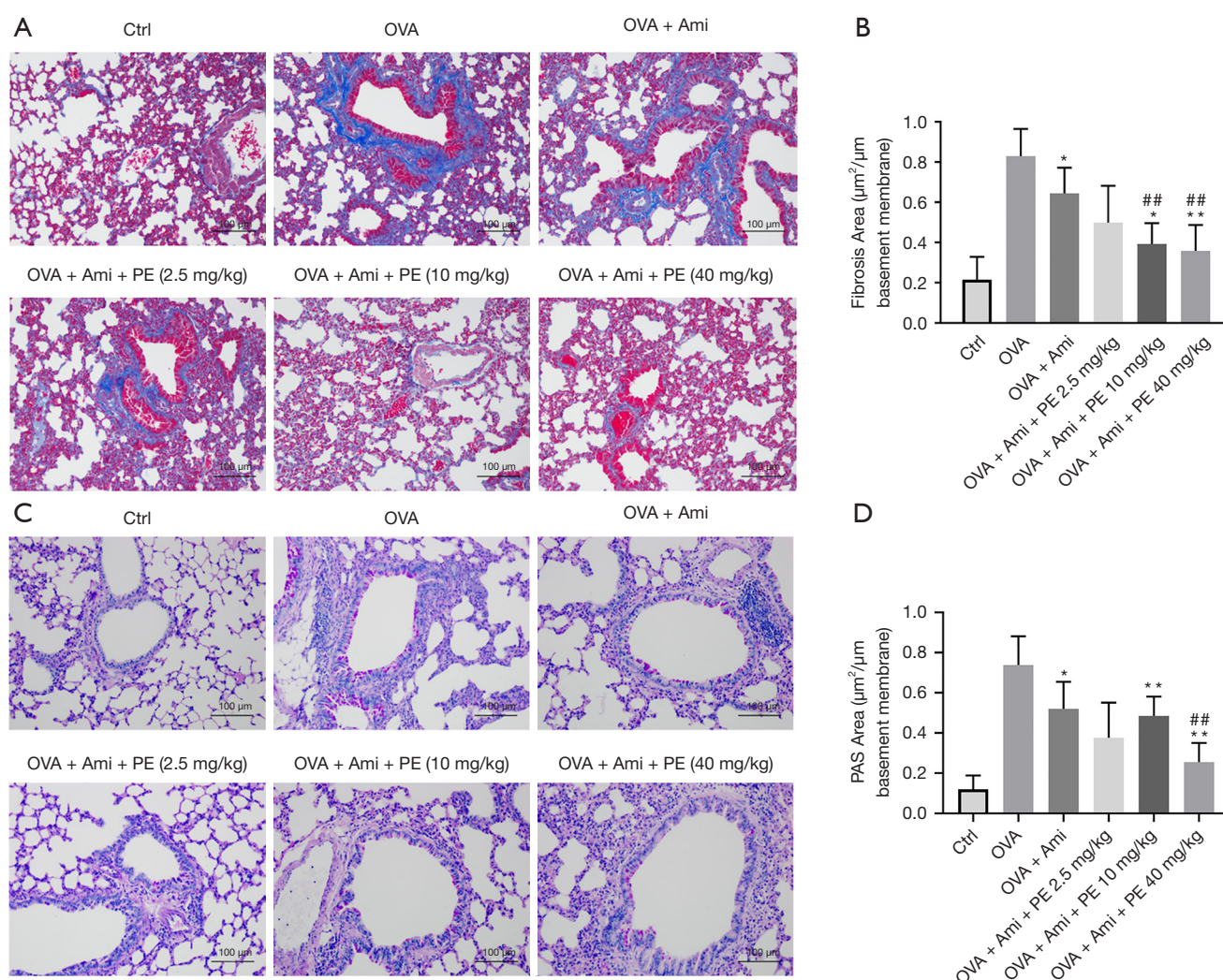
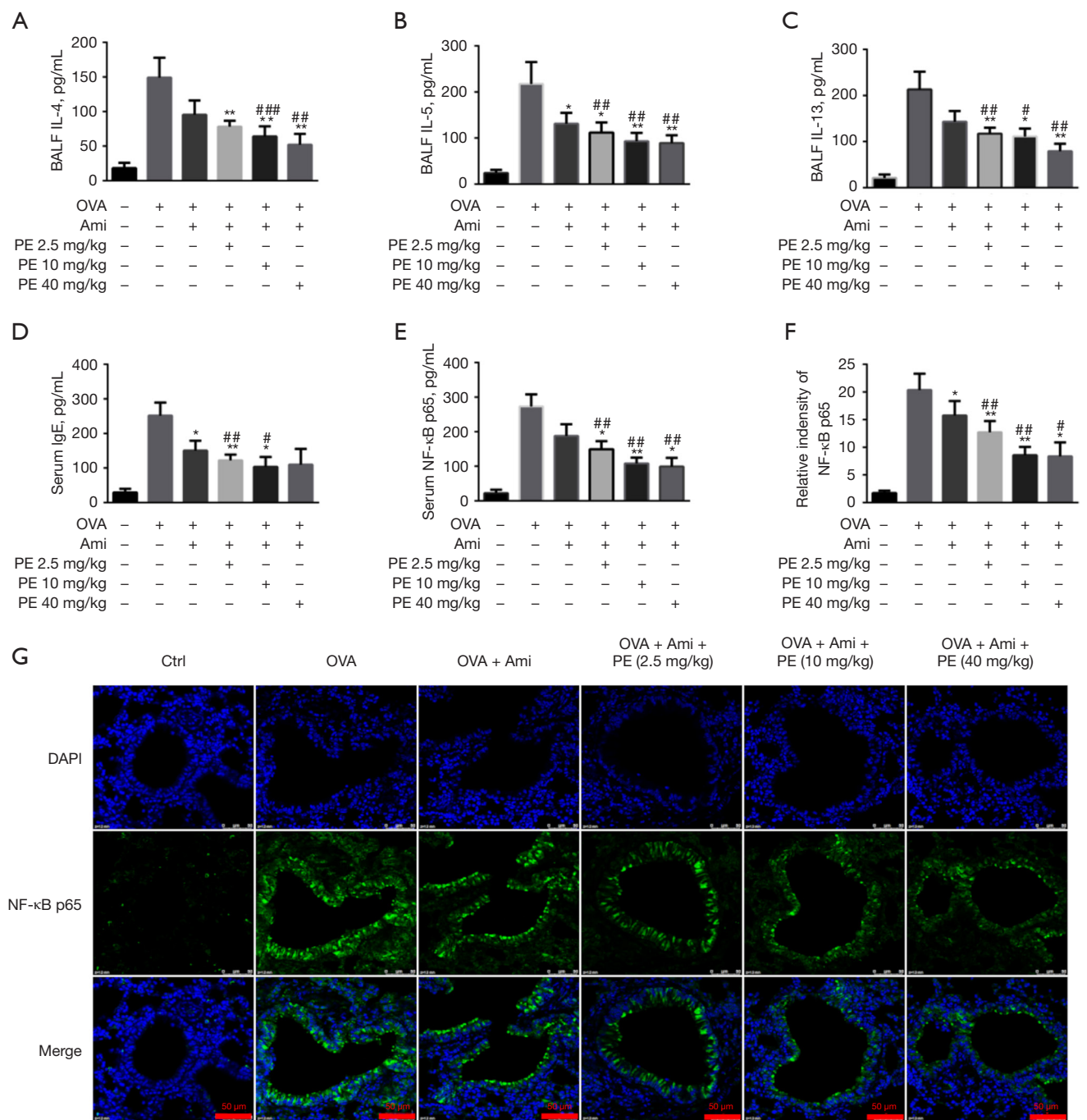


Figure 3 PE enhanced the effect of Ami on the reduction of mucus secretion and collagen deposition in asthmatic mice lungs. (A,B) M-T staining and fibrosis area in the bronchial basement membrane determined by Image-Pro Plus software analysis. (C,D) PAS staining and PAS-positive areas in the bronchial basement membrane. OVA + Ami group, OVA + Ami + PE 2.5 mg/kg group, OVA + Ami + PE 10 mg/kg group and OVA + Ami + PE 40 mg/kg group compared with the OVA group, * $P < 0.05$, ** $P < 0.01$; OVA + Ami + PE 2.5 mg/kg group, OVA + Ami + PE 10 mg/kg group and OVA + Ami + PE 40 mg/kg group compared with the OVA + Ami group, ### $P < 0.01$. $n = 5$. PE, Praeruptorin E; Ami, aminophylline; M-T, Masson's trichrome; PAS, periodic acid-Schiff; ctrl, control; OVA, ovalbumin.

the free energy of binding was -6.71 kcal/mol. PE forms Van der Waals force with Arg¹³¹, Gly¹³⁰, Leu²²², Asn⁴², Ser¹³³, Phe¹¹⁹, and a carbon-hydrogen bond with Gly¹⁷¹ and Lys¹²² of the P chain. Moreover, PE forms π -alkylation with Pro¹²⁹, Ile²¹⁹, Pro¹⁶⁷, Arg¹³², Ile¹²⁸ of the α -chain, and Leu²¹⁸, Ile¹⁶⁸ of the A-chain of NF- κ B p65. These findings indicate that PE could enhance aminophylline efficacy to reduce the level of Th2 cytokines, IgE, and NF- κ B p65 in asthmatic mice.

PE ameliorates aminophylline cardiotoxicity and reduces the accumulation of aminophylline metabolites in vivo

Cardiotoxicity is a common adverse reaction to aminophylline, manifested as arrhythmia and hypofunction of the heart (9,28). Therefore, we investigated whether PE could reduce the cardiotoxicity of aminophylline at a concentration of 450 mg/kg, which is 6 times the usual dose of aminophylline that induces cardiotoxicity. The



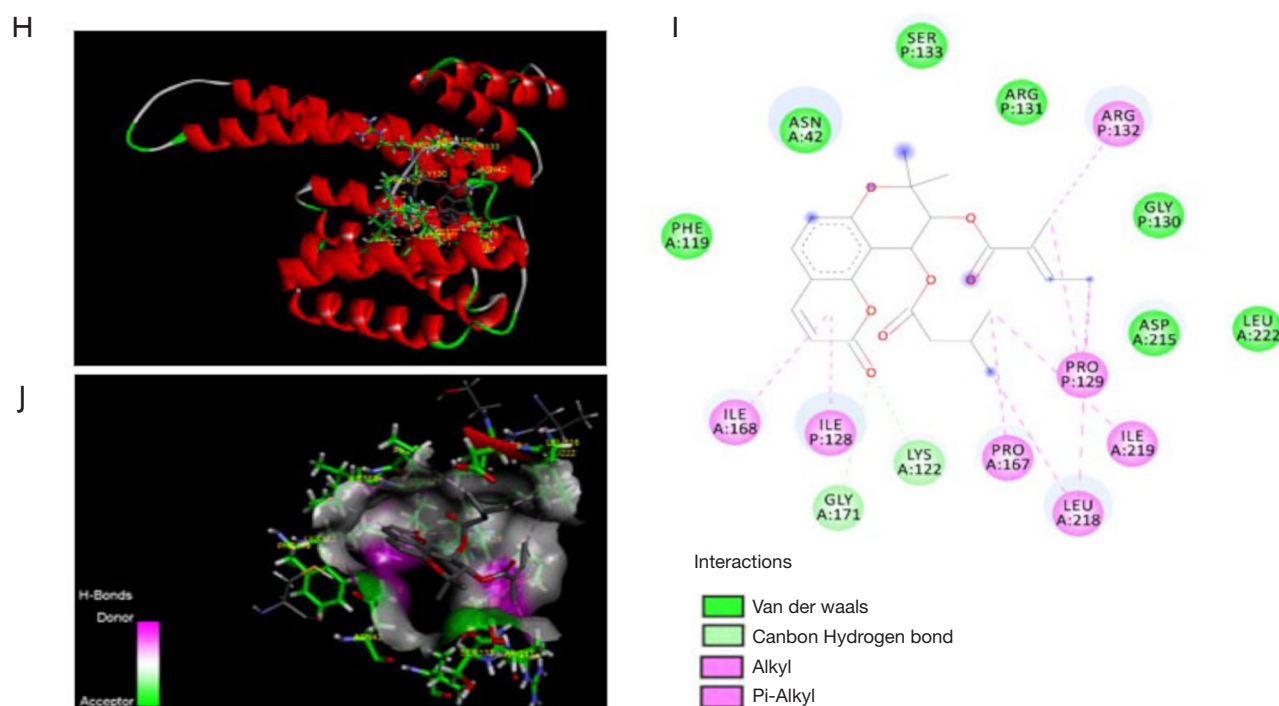


Figure 4 PE facilitates aminophylline inhibition of inflammation factors in asthmatic mice. (A-C) Concentrations of IL-4, IL-5, and IL-13 in BALF. (D,E) Concentrations of IgE and NF-κB p65 in serum. (F,G) Expression levels of NF-κB p65 in the lungs and bronchi detected by immunofluorescence assay. Magnification $\times 400$, green: NF-κB p65, blue: nucleus. OVA + Ami group, OVA + Ami + PE 2.5 mg/kg group, OVA + Ami + PE 10 mg/kg group and OVA + Ami + PE 40 mg/kg group compared with the OVA group, * $P < 0.05$, ** $P < 0.01$; OVA + Ami + PE 2.5 mg/kg group, OVA + Ami + PE 10 mg/kg group and OVA + Ami + PE 40 mg/kg group compared with the OVA + Ami group, # $P < 0.05$, ## $P < 0.01$, ### $P < 0.001$. $n = 5$. (H-J) Molecular docking PE into NF-κB p65, the 3D diagram shows the binding conformation of PE (green) with NF-κB p65 (red). The 2D diagrams show the docking model of PE in the active site of NF-κB p65. PE, Praeruptorin E; IL, interleukin; BALF, bronchoalveolar lavage fluid; IgE, immunoglobulin E; NF-κB, nuclear factor-κB; Ami, aminophylline; OVA, ovalbumin.

results showed that, compared with the control group, the mice in the aminophylline group had absolutely irregular heart rhythm, slow heart rates, but frequent premature ventricular beats. However, the aminophylline + PE group mice had regular heart rhythms, occasional premature ventricular beats, and faster ventricular rates than mice in the aminophylline-alone group (*Figure 5A, 5B*). The levels of serum myocardial enzymes, such as LDH, HBD, CK-MB, CK, and AST, were also lower than in the aminophylline-alone group (*Figure 5C-5G*). These findings indicate that PE is beneficial in reducing the cardiac toxicity induced by aminophylline.

The primary degradant of aminophylline in the human body is theophylline, which is mainly metabolized by the hepatic drug enzymes CYP1A2, CYP2E1, and CYP3A4.

CYP1A2 is involved in the decomposition of about 90–95% of theophylline. CYP1A2 catalyzes the conversion of theophylline to 1-methylxanthine (1-MX), then 1-MX is oxidized very rapidly to 1-MU by xanthine oxidase, while CYP3A4 and CYP2E1 catalyze the conversion of theophylline to 1,3-DMU. The accumulation of these metabolites is the main reason for the toxic effects of aminophylline (29). In the present study, LC-MS/MS was used to detect the accumulation of theophylline and its metabolites in asthmatic rats within 12 h. The results showed that the plasma concentrations of theophylline and 1,3-DMU in the aminophylline + PE group were significantly lower than those of the aminophylline-alone group at each time investigated. Similarly, the areas under the curve (AUC) of theophylline and 1,3-DMU were also

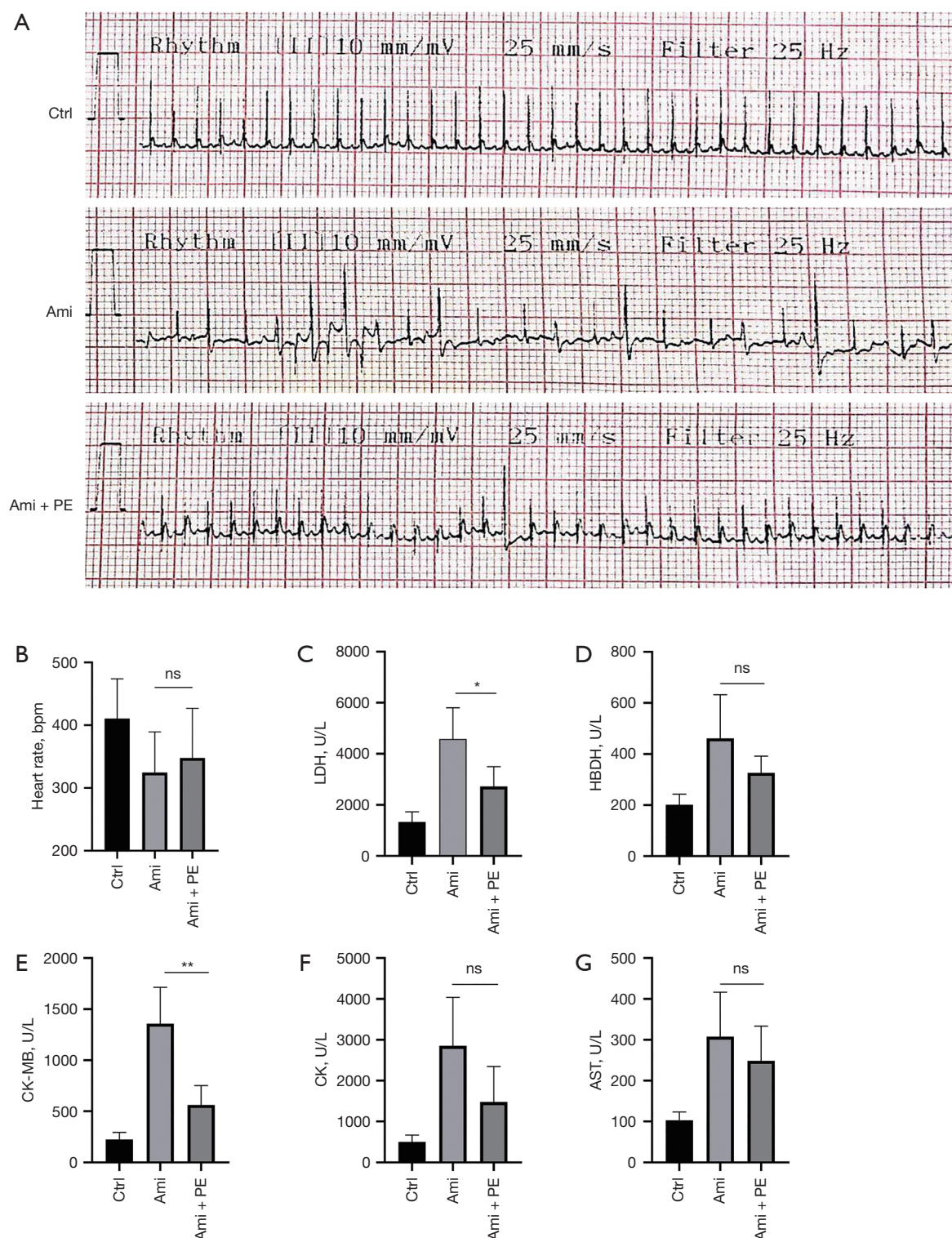


Figure 5 PE ameliorated aminophylline cardiotoxicity in mouse. (A,B) ECG was used to analyze the electrophysiology and heart rate of mice. (C-G) ELISA detection of the contents of serum myocardial enzymes, including LDH, HBD, CK-MB, CK, and AST. * $P < 0.05$, ** $P < 0.01$. PE, Praeruptorin E; ECG, electrocardiogram; ELISA, enzyme-linked immunosorbent assay; LDH, lactate dehydrogenase; HBD, hydroxybutyrate dehydrogenase; CK, creatine kinase; AST, aspartate aminotransferase; ctrl, control; Ami, aminophylline; ns, no significance.

lower than in the aminophylline group, but there was no significant difference plasma concentration of 1-MU at each time point or the AUC between the two groups (Figure 6A-6F). These results suggest that PE reduces the accumulation of some theophylline and 1,3-DMU, and the impairment of 1,3-DMU was prominent, but it still retained the blood concentration of most theophylline and played a pharmacological role.

Effect of PE on the expression of NF- κ B, PXR, and CYP3A11 in allergic airway inflammation

We previously found that PE is a potential ligand of PXR and that PE can upregulate the expression of the *CYP3A4* gene through the PXR pathway (30). Therefore, we examined the effect of PE on the expression of NF- κ B in the lungs, and PXR, CYP3A11 (the mouse homolog of CYP3A4), CYP1A2, and CYP2E1 in the liver of asthmatic mice. In lung tissue, the mRNA and protein levels of NF- κ B in the aminophylline group were obviously lower than in the OVA group. The expression of NF- κ B in the aminophylline + PE (2.5, 10, 40 mg/kg) groups was lower than in the aminophylline alone group, indicating that PE could reduce NF- κ B activity in the lungs when used in combination with aminophylline (Figure 7A-7C). In liver tissue, the levels of mRNA and the proteins of PXR and CYP3A11 in the OVA group were lower than in the control group. The levels of PXR and CYP3A11 mRNA and protein in the aminophylline treatment group increased to a certain extent, while expression in the aminophylline + PE treatment group was higher than in the aminophylline-alone group. However, there was no statistically significant difference in mRNA and protein expression levels for CYP1A2 and CYP2E1 among the treatment groups (Figure 7A-7C). These findings indicate that PE can positively regulate the expressions of NF- κ B, PXR, and CYP3A11 *in vivo* to promote the elimination of theophylline and 1,3-DMU.

PE inhibits NF- κ B activation and regulates PXR and CYP3A4 overexpression in L-02 cells

Human L-02 cells were used to clarify the mechanism of the synergistic and detoxifying effects of PE on aminophylline. First, LPS (100 ng/mL)-induced L-02 cells were treated with 60 μ M PE. The results showed that PE reduced the expression of NF- κ B, promoted the expression of PXR and CYP3A4, and reversed the inhibitory effect on inflammation

of PXR and CYP3A4 (Figure 7D,7E). Next, we established L-02 cell models under different growth states by LPS induction or siRNA interference, and further verified the mechanism of PE co-administered with aminophylline. Under the condition of only LPS-induced inflammation, the expression of NF- κ B in the aminophylline + PE group was lower than in the aminophylline group, and the expression of PXR and CYP3A4 was higher than in the aminophylline group. The findings indicated that aminophylline + PE could reduce the expression of NF- κ B and increase the expression of PXR and CYP3A4 in the inflammatory state. When the expression of PXR was knocked down by siRNA under inflammation, the effect of PE on the synergized reduction of NF- κ B expression was reduced, as well as its effects on the expression of PXR and CYP3A4 (Figure 7F-7H). Although the results indicated that inflammation could inhibit the expression of PXR, whether NF- κ B had an inhibitory effect on PXR still remains to be verified unequivocally. Therefore, ChIP experiments were conducted and the results revealed that chromatin was mainly concentrated at about 500 bp and that PXR was expressed in both the input and ChIP DNA samples, suggesting that NF- κ B was enriched at the binding site of the PXR promoter region and that PE could reduce the enrichment of the NF- κ B and PXR promoter (Figure 7I,7J). Therefore, the findings indicate that PE could reduce the toxicity and enhance the efficacy of aminophylline through actions on the NF- κ B/PXR/CYP3A4 pathway.

Discussion

As the 14th most common chronic disease in the world, asthma not only seriously affects patients' physical and mental health, but also brings a heavy economic burden to families, countries, and healthcare services (31). Multiple inflammatory cells and factors play key roles in the development of asthma. During the process of infiltration and aggregation of inflammatory cells, such as eosinophils, into the lungs and trachea, a large number of cytokines and inflammatory mediators is secreted. These are involved in chemotaxis, leading to the contraction of airway smooth muscle and increased mucus secretion and mucosal edema (32). The Th2 cytokines, IL-4, IL-5, and IL-13, and eosinophil chemokines are also major participants. IL-4 is one of the most important cytokines regulating Th2 inflammation, which can induce the maturation of B cells and promote the transformation of IgG into IgE (33).

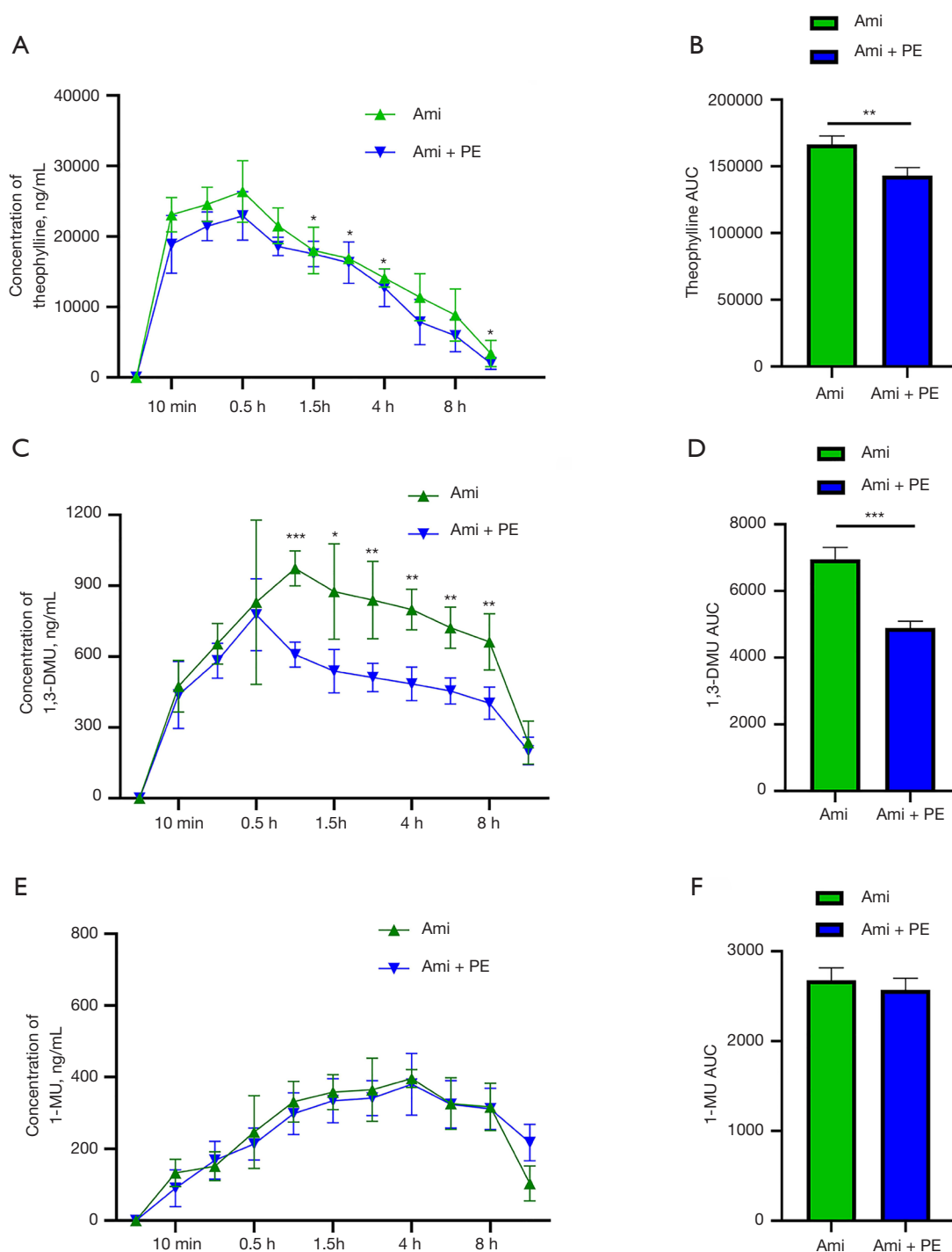
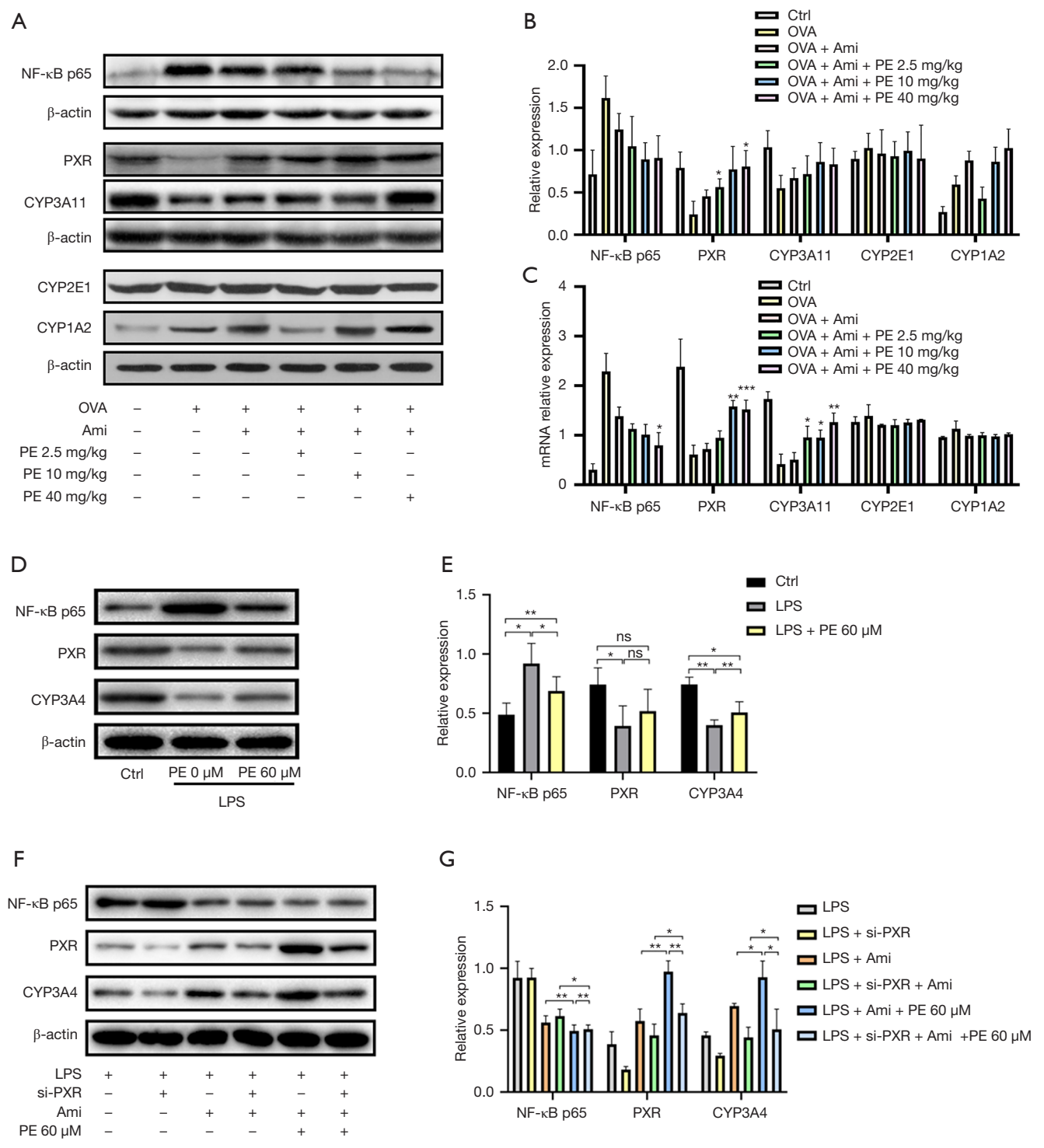


Figure 6 PE promotes metabolism of Ami metabolites in asthmatic rats. (A,C,E) Plasma concentration of theophylline, 1,3-DMU, and 1-MU in asthmatic rats in a period of 12 h determined by LC-MS/MS analysis. (B,D,F) AUC of theophylline, 1,3-DMU, and 1-MU by GraphPad Prism 8 analysis. * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$. $n = 4$. PE, Praeruptorin E; Ami, aminophylline; 1,3-DMU, 1,3-dimethyluric acid; 1-MU, 1-methyluric acid; LC-MS/MS, liquid chromatography-tandem mass spectrometry; AUC, area under the curve; SD, Sprague-Dawley; OVA, ovalbumin.



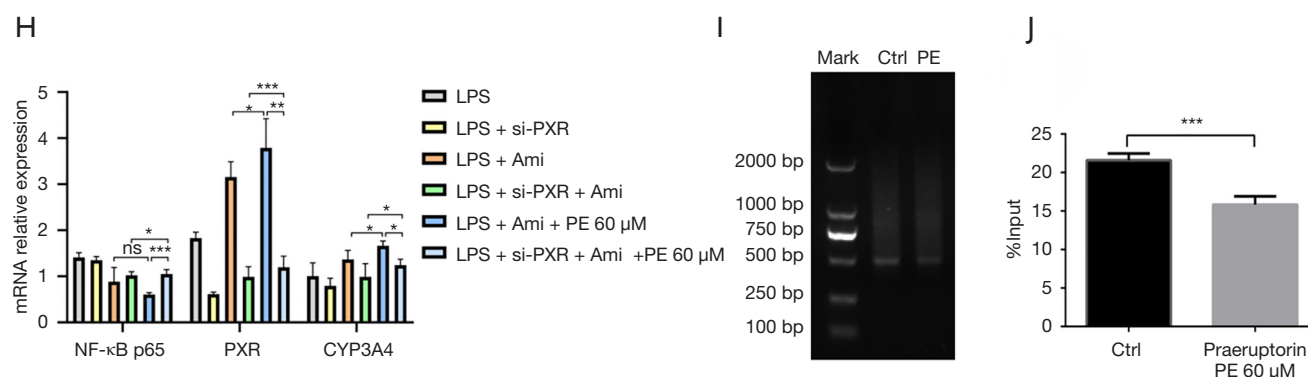


Figure 7 PE attenuates toxicity and promotes the efficacy of Ami anti-asthma activity via NF-κB/PXR/CYP3A4 pathways. (A-C) Expression levels of NF-κB p65 in the lungs and PXR, CYP3A11, CYP1A2, CYP2E1 in the liver of asthmatic mice determined by Western blotting and reverse transcription PCR. OVA + Ami + PE 2.5 mg/kg group, OVA + Ami + PE 10 mg/kg group and OVA + Ami + PE 40 mg/kg group compared with the OVA + Ami group, * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, $n = 5$. (D,E) After the LPS-induced inflammatory state of L-02 cells was established, the expression of NF-κB p65, PXR, and CYP3A4 in L-02 cells was detected by Western blotting after PE treatment for 24 h. (F-H) Under LPS-induced inflammation and si-PXR effected, the L-02 cells were treated with Ami and PE for 24 h, Western blotting and reverse transcription PCR were used to detect the expression of NF-κB p65, PXR, and CYP3A4. (I,J) Degree of enrichment of NF-κB p65 and PXR promoters detected by ChIP assay. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$. Data are representative of 3 independent experiments. PE, Praeruptorin E; Ami, aminophylline; NF-κB, nuclear factor-κB; PXR, pregnane X receptor; CYP3A4, cytochrome P450 3A; PCR, polymerase chain reaction; LPS, lipopolysaccharide; ChIP, chromatin immunoprecipitation; ctrl, control; OVA, ovalbumin; ns, no significance.

IL-5 is important for the differentiation and maturation of eosinophils and has a vital role in the allergen-induced eosinophilic airway inflammation (34). IL-5 also enhances IL-4-induced IgE production. IL-13 promotes B-cell differentiation and can directly induce airway inflammation and the altitude reaction (35).

Theophylline is a derivative of methylxanthine and has significant anti-asthmatic activity, a low price, and is suitable for administration in both children and adults. As a non-selective phosphodiesterase inhibitor and adenosine receptor antagonist, the mechanism of its anti-asthmatic action is inhibition of the production of adhesion molecules and cytokines, therefore preventing airway aggregation of inflammatory cells, the expression of inflammatory mediators and cytokines, and Th 2 transcription and translation (36). Moreover, airway smooth muscle relaxation is another pharmacodynamic effect of aminophylline. However, the plasma concentration of aminophylline should reach 10–20 mg/L before it becomes effective, making the plasma drug concentration close to its toxic concentration. When the concentration is >20 mg/L, some patients

could exhibit toxic symptoms that often present as severe cardiotoxicity and central nervous system disturbances. When the drug concentration exceeds 40 mg/L, patients develop symptoms of aminophylline toxicity, which can be life-threatening (37,38). Therefore, reducing the blood drug concentration of aminophylline and its metabolites does not only lower the incidence of toxicity and side-effects, but also exerts a significant anti-inflammatory effect.

Peucedanum praeruptorum Dunn is the main medicinal material of Chinese medicine used for the relief of cough and asthma symptoms. Previous studies have shown that various active components of *Peucedanum praeruptorum* Dunn have anti-inflammatory effects (23,39,40). Therefore, we also speculated that PE, its main active component, would enhance the role of aminophylline in the treatment of asthma. In the present study, we found that PE combined with aminophylline could promote aminophylline reduction in the infiltration of inflammatory cells, mucus secretion, and collagen cell fibrosis in the lungs and bronchi of asthmatic mice and repair their physiological structures. It also acts to inhibit the expression levels of Th2 cytokines

IL-4, IL-5, and IL-13, and IgE and NF- κ B p65. These findings indicate that PE could play a synergistic role with aminophylline. Furthermore, our results showed that PE can reduce the plasma concentration of theophylline in asthmatic rats to a significant extent and reduce the accumulation of 1,3-DMU. Interestingly, the changes in the plasma concentration of 1-MU were not significant. According to previous reports (41), it has been reported that PE does not reduce aminophylline metabolite levels through the CYP1A2 metabolic pathway.

NF- κ B is a key transcription factor in various pathological conditions. When stimulated by an antigen, inhibitor of NF- κ B(I κ B) is separated from NF- κ B p65 and phosphorylated. NF- κ B p65 is translocated from the cytoplasm into the nucleus, facilitating the synthesis and release of pro-inflammatory cytokines (42,43). NF- κ B can regulate Th2 cell differentiation and Th2 cytokine expression in allergic asthma. Inhibiting the activity of NF- κ B is an effective way to treat asthma (44,45). Our results have shown that PE in combination with aminophylline reduces the expression of NF- κ B p65 in the lungs of asthmatic rats and decreases the expression of LPS-induced NF- κ B *in vitro*. Therefore, we deduced that PE can cooperate with aminophylline to inhibit the NF- κ B pathway and reduce Th2 cytokine secretion, thereby ameliorating airway inflammation, mucus secretion, and fibrosis in asthmatic mice.

PXR belongs to the nuclear receptor superfamily of NR1I, and participates in the bioconversion of many endogenous and exogenous chemicals. PXR is composed of a ligand binding domain (LBD) and a DNA binding domain. When the exogenous molecule enters a cell, it directly binds to the intracellular LBD to form the receptor-ligand complex, and then binds to the retinoid X receptor (RXR) to form the PXR/RXR heterodimer (46-48). Asthma is a chronic inflammatory disease of the airways that involves the activity of many cytokines. Therefore, asthma is often associated with the increased expression of cytokines, such as NF- κ B. A large amount of activated NF- κ B binds to the reaction element of RXR on the heterodimer of PXR/RXR through its p65 subunit in the nucleus, thereby reducing the binding of the PXR/RXR dimer to the promoter of CYP3A4, and finally downregulates the drug-metabolizing enzymes, such as CYP3A4, by inhibiting the transcriptional activity of PXR (26,49,50). When PXR expression in cells is increased, it binds to p65 in the cytoplasm, prevents p65

nuclear translocation, and weakens the role of NF- κ B in activating downstream pro-inflammatory genes. Therefore, PXR and NF- κ B have a 2-way regulatory effect (51,52). It was found that after transfection of human PXR into mice with knocked out *PXR* genes, human specific inducers could induce CYP3A (53), indicating that the PXR/RXR dimer could bind to the PXR response element on the *CYP3A* gene promoter and induce the expression of the *CYP3A* gene. In our previous study, we also found that PE could upregulate the expression of the *CYP3A4* gene through the PXR pathway (30). In the present study, PE increased the expression of PXR and CYP3A4 in asthmatic mice, but there was no obvious effect on the metabolic enzymes, CYP1A2 and CYP2E1, indicating that CYP3A4 metabolic enzymes were the main target for PE. *In vitro* results demonstrated that PE could also reverse the inhibitory effects on inflammation of PXR and CYP3A4. When PXR expression was reduced with siRNA interference, the synergistic effect of PE was weakened. In addition, the ChIP assay verified that NF- κ B was enriched at the binding site of the PXR promoter region and that PE could reduce the enrichment of the NF- κ B and PXR promoter, weakening the inhibition of NF- κ B on PXR. In addition, some studies have found that PE can play an anti-inflammatory effect by inhibiting STAT3 signaling pathway (54), and STAT3 also has a regulatory effect on PXR (55), which means that PE may also produce attenuated and synergistic effects through STAT3/PXR signaling axis. In the future, we should also explore another role.

Conclusions

The findings of the present study confirmed that PE could enhance aminophylline activity to reduce the infiltration of inflammatory cells, mucus secretion, and fibrosis in the lungs of asthmatic mice; inhibit the expression of inflammatory factors IL-4, IL-5, IL-13, IgE, and NF- κ B; and have a synergistic effect with aminophylline to treat asthma. It was also found that PE could reduce the plasma concentration of theophylline, 1,3-DMU and play a role in reducing toxicity. During the *in vitro* experiments, PE was found to assist aminophylline in reducing the expression of NF- κ B and in reversing the expression of PXR and CYP3A4, indicating that PE is involved in reducing toxicity and enhancing its efficacy through the bidirectional regulation of NF- κ B/PXR/CYP3A4 (Figure 8).

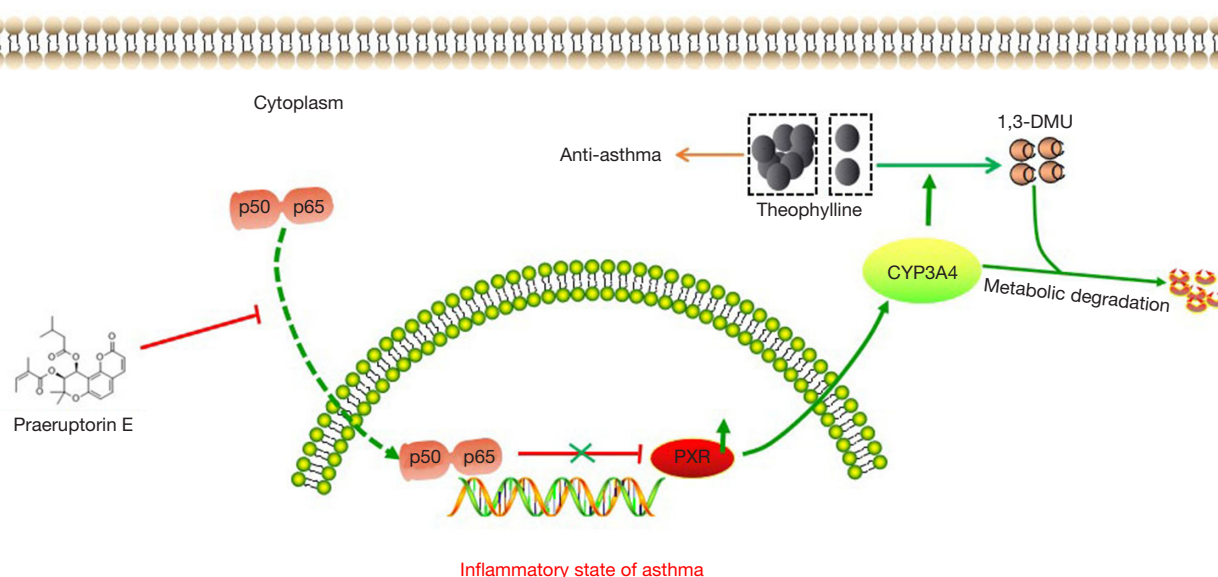


Figure 8 In the inflammatory state of asthma, PE can inhibit the expression of NF- κ B p65, reduce the inhibitory effect on PXR, and promote the expression of CYP3A4, CYP3A4 promotes the metabolism of theophylline and 1,3-DMU. PE can enhance the anti-asthma effect of aminophylline, while also reducing the toxic effect caused by theophylline by targeting the NF- κ B/PXR/CYP3A11 axis. PE, Praeruptorin E; NF- κ B, nuclear factor- κ B; PXR, pregnane X receptor; CYP3A4, cytochrome P450 3A; 1,3-DMU, 1,3-dimethyluric acid.

Taken together, these findings indicate that *Peucedanum praeruptorum* Dunn is a promising herbal medicine that will benefit asthmatics treated with theophylline. In the future, we will continue to execute in-depth studies, including studies on improving the mechanism of PE, safety experiments, pharmacological effects and primate studies.

Acknowledgments

Funding: This work was supported by grants from the National Natural Science Foundation of China (Nos. 81403006, 81760674, 82060678); Research Unit of Island Emergency Medicine, Chinese Academy of Medical Sciences (No. 2019RU013).

Footnote

Reporting Checklist: The authors have completed the ARRIVE reporting checklist. Available at <https://atm.amegroups.com/article/view/10.21037/atm-22-386/rc>

Data Sharing Statement: Available at <https://atm.amegroups.com/article/view/10.21037/atm-22-386/dss>

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <https://atm.amegroups.com/article/view/10.21037/atm-22-386/coif>). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. All experiments were approved by the Institutional Animal Care and Treatment Committee of Hainan Medical University (Ratified on October 16, 2017), in compliance with Guidelines for the Humane Treatment of Laboratory Animals (Ministry of Science and Technology of the People's Republic of China; policy No. 2006398).

Open Access Statement: This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the

formal publication through the relevant DOI and the license).
See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

References

- Kim TH, Park YM, Ryu SW, et al. Receptor Interacting Protein 2 (RIP2) Is Dispensable for OVA-Induced Airway Inflammation in Mice. *Allergy Asthma Immunol Res* 2014;6:163-8.
- Bousquet J, Mantzouranis E, Cruz AA, et al. Uniform definition of asthma severity, control, and exacerbations: document presented for the World Health Organization Consultation on Severe Asthma. *J Allergy Clin Immunol* 2010;126:926-38.
- Nakajima H, Takatsu K. Role of cytokines in allergic airway inflammation. *Int Arch Allergy Immunol* 2007;142:265-73.
- Li BW, Hendriks RW. Group 2 innate lymphoid cells in lung inflammation. *Immunology* 2013;140:281-7.
- Licona-Limón P, Kim LK, Palm NW, et al. TH2, allergy and group 2 innate lymphoid cells. *Nat Immunol* 2013;14:536-42.
- Maselli DJ, Hardin M, Christenson SA, et al. Clinical Approach to the Therapy of Asthma-COPD Overlap. *Chest* 2019;155:168-77.
- Ernst E. Spinal manipulation for asthma: a systematic review of randomised clinical trials. *Respir Med* 2009;103:1791-5.
- Eason J, Markowe HL. Aminophylline toxicity--how many hospital asthma deaths does it cause? *Respir Med* 1989;83:219-26.
- Starakis I, Starakis I, Lekkou A, et al. Drug-induced cardiotoxicity due to aminophylline treatment: a case report. *Curr Ther Res Clin Exp* 2003;64:367-74.
- Kubo M, Odajima Y, Ishizaki T, et al. Intraindividual changes in theophylline clearance during constant aminophylline infusion in children with acute asthma. *J Pediatr* 1986;108:1011-5.
- Barnes PJ. Pathophysiology of allergic inflammation. *Immunol Rev* 2011;242:31-50.
- Berkman N, Krishnan VL, Gilbey T, et al. Expression of RANTES mRNA and protein in airways of patients with mild asthma. *Am J Respir Crit Care Med* 1996;154:1804-11.
- Grilli M, Chiu JJ, Lenardo MJ. NF-kappa B and Rel: participants in a multiform transcriptional regulatory system. *Int Rev Cytol* 1993;143:1-62.
- John AE, Zhu YM, Brightling CE, et al. Human airway smooth muscle cells from asthmatic individuals have CXCL8 hypersecretion due to increased NF-kappa B p65, C/EBP beta, and RNA polymerase II binding to the CXCL8 promoter. *J Immunol* 2009;183:4682-92.
- Scarino A, Noël A, Renzi PM, et al. Impact of emerging pollutants on pulmonary inflammation in asthmatic rats: ethanol vapors and agglomerated TiO2 nanoparticles. *Inhal Toxicol* 2012;24:528-38.
- Chae JW, Baek IH, Kwon KI. Effect of decursin on the pharmacokinetics of theophylline and its metabolites in rats. *J Ethnopharmacol* 2012;144:248-54.
- Zhou C, Tabb MM, Nelson EL, et al. Mutual repression between steroid and xenobiotic receptor and NF-kappaB signaling pathways links xenobiotic metabolism and inflammation. *J Clin Invest* 2006;116:2280-9.
- Goodwin B, Redinbo MR, Klierer SA. Regulation of cyp3a gene transcription by the pregnane x receptor. *Annu Rev Pharmacol Toxicol* 2002;42:1-23.
- Shah YM, Ma X, Morimura K, et al. Pregnane X receptor activation ameliorates DSS-induced inflammatory bowel disease via inhibition of NF-kappaB target gene expression. *Am J Physiol Gastrointest Liver Physiol* 2007;292:G1114-22.
- Wenjie W, Houqing L, Liming S, et al. Effects of praeuroptin C on blood pressure and expression of phospholamban in spontaneously hypertensive rats. *Phytomedicine* 2014;21:195-8.
- Shen X, Chen G, Zhu G, et al. (+/-)-3'-O, 4'-O-dicinnamoyl-cis-khellactone, a derivative of (+/-)-praeuroptin A, reverses P-glycoprotein mediated multidrug resistance in cancer cells. *Bioorg Med Chem* 2006;14:7138-45.
- Liu X, Chin JF, Qu X, et al. The Beneficial Effect of Praeuroptin C on Osteoporotic Bone in Ovariectomized Mice via Suppression of Osteoclast Formation and Bone Resorption. *Front Pharmacol* 2017;8:627.
- Xiong Y, Wang J, Wu F, et al. Effects of (±)-praeuroptin A on airway inflammation, airway hyperresponsiveness and NF-κB signaling pathway in a mouse model of allergic airway disease. *Eur J Pharmacol* 2012;683:316-24.
- Huang L, Huang M, Li YH, et al. Up-regulation of CYP3A expression through pregnant X receptor by praeuroptin D isolated from *Peucedanum praeruptorum* Dunn. *J Ethnopharmacol* 2013;148:596-602.

25. Huang L, Bi HC, Li YH, et al. Regulation of human pregnane X receptor and its target gene cytochrome P450 3A by praeruptorin A isolated from the herbal medicine *Peucedanum praeruptorum*. *Planta Med* 2013;79:1509-15.
26. Huang L, Wu Q, Li YH, et al. PXR-Mediated Upregulation of CYP3A Expression by Herb Compound Praeruptorin C from *Peucedanum praeruptorum* Dunn. *Evid Based Complement Alternat Med* 2013;2013:156574.
27. Jin H, Luo Q, Zheng Y, et al. CD4+CD25+Foxp3+ T cells contribute to the antiasthmatic effects of *Astragalus membranaceus* extract in a rat model of asthma. *Int Immunopharmacol* 2013;15:42-9.
28. Whitehurst VE, Joseph X, Alleva FR, et al. Enhancement of acute myocardial lesions by asthma drugs in rats. *Toxicol Pathol* 1994;22:72-6.
29. Sohn JA, Kim HS, Oh J, et al. Prediction of serum theophylline concentrations and cytochrome P450 1A2 activity by analyzing urinary metabolites in preterm infants. *Br J Clin Pharmacol* 2017;83:1279-86.
30. Liu YH, Mo SL, Bi HC, et al. Regulation of human pregnane X receptor and its target gene cytochrome P450 3A4 by Chinese herbal compounds and a molecular docking study. *Xenobiotica* 2011;41:259-80.
31. Tully JE, Hoffman SM, Lahue KG, et al. Epithelial NF- κ B orchestrates house dust mite-induced airway inflammation, hyperresponsiveness, and fibrotic remodeling. *J Immunol* 2013;191:5811-21.
32. Agrawal DK, Shao Z. Pathogenesis of allergic airway inflammation. *Curr Allergy Asthma Rep* 2010;10:39-48.
33. Elias JA, Lee CG, Zheng T, et al. New insights into the pathogenesis of asthma. *J Clin Invest* 2003;111:291-7.
34. Renz H, Enssle K, Lauffer L, et al. Inhibition of allergen-induced IgE and IgG1 production by soluble IL-4 receptor. *Int Arch Allergy Immunol* 1995;106:46-54.
35. Halim TY, Krauss RH, Sun AC, et al. Lung natural helper cells are a critical source of Th2 cell-type cytokines in protease allergen-induced airway inflammation. *Immunity* 2012;36:451-63.
36. Littenberg B. Aminophylline treatment in severe, acute asthma. A meta-analysis. *JAMA* 1988;259:1678-84.
37. Barnes PJ. Theophylline. *Am J Respir Crit Care Med* 2013;188:901-6.
38. McEvoy GK. Dose adjustment in renal impairment: response from AHFS Drug Information. *BMJ* 2005;331:293.
39. Yu PJ, Ci W, Wang GF, et al. Praeruptorin A inhibits lipopolysaccharide-induced inflammatory response in murine macrophages through inhibition of NF- κ B pathway activation. *Phytother Res* 2011;25:550-6.
40. Yu PJ, Li JR, Zhu ZG, et al. Praeruptorin D and E attenuate lipopolysaccharide/hydrochloric acid induced acute lung injury in mice. *Eur J Pharmacol* 2013;710:39-48.
41. Jing WH, Song YL, Yan R, et al. Identification of cytochrome P450 isoenzymes involved in metabolism of (+)-praeruptorin A, a calcium channel blocker, by human liver microsomes using ultra high-performance liquid chromatography coupled with tandem mass spectrometry. *J Pharm Biomed Anal* 2013;77:175-88.
42. Van Antwerp DJ, Martin SJ, Kafri T, et al. Suppression of TNF- α -induced apoptosis by NF- κ B. *Science* 1996;274:787-9.
43. Li Q, Verma IM. NF- κ B regulation in the immune system. *Nat Rev Immunol* 2002;2:725-34.
44. Xia YF, Ye BQ, Li YD, et al. Andrographolide attenuates inflammation by inhibition of NF- κ B activation through covalent modification of reduced cysteine 62 of p50. *J Immunol* 2004;173:4207-17.
45. Das J, Chen CH, Yang L, et al. A critical role for NF- κ B in GATA3 expression and TH2 differentiation in allergic airway inflammation. *Nat Immunol* 2001;2:45-50.
46. Willson TM, Kliewer SA. PXR, CAR and drug metabolism. *Nat Rev Drug Discov* 2002;1:259-66.
47. Kliewer SA, Moore JT, Wade L, et al. An orphan nuclear receptor activated by pregnanes defines a novel steroid signaling pathway. *Cell* 1998;92:73-82.
48. Yan J, Xie W. A brief history of the discovery of PXR and CAR as xenobiotic receptors. *Acta Pharm Sin B* 2016;6:450-2.
49. Hogle BC, Guan X, Folan MM, et al. PXR as a mediator of herb-drug interaction. *J Food Drug Anal* 2018;26:S26-31.
50. Xie W, Barwick JL, Downes M, et al. Humanized xenobiotic response in mice expressing nuclear receptor SXR. *Nature* 2000;406:435-9.
51. Zhang J, Cao L, Wang H, et al. Ginsenosides Regulate PXR/NF- κ B Signaling and Attenuate Dextran Sulfate Sodium-Induced Colitis. *Drug Metab Dispos* 2015;43:1181-9.
52. Lv C, Huang L. Xenobiotic receptors in mediating the effect of sepsis on drug metabolism. *Acta Pharm Sin B* 2020;10:33-41.
53. Zhao Y, Zhang CW, Zhou WJ, et al. Is there a role of TNFR1 in acute lung injury cases associated with extracorporeal circulation? *J Zhejiang Univ Sci B*

- 2014;15:281-8.
54. Yu PJ, Jin H, Zhang JY, et al. Pyranocoumarins isolated from *Peucedanum praeruptorum* Dunn suppress lipopolysaccharide-induced inflammatory response in murine macrophages through inhibition of NF- κ B and STAT3 activation. *Inflammation* 2012;35:967-77.
55. Zhou Y, Nie AQ, Chen S, et al. Downregulation of Renal MRPs Transporters in Acute Lymphoblastic Leukemia Mediated by the IL-6/STAT3/PXR Signaling Pathway. *J Inflamm Res* 2021;14:2239-52.
- (English Language Editor: R. Scott)

Cite this article as: Xu R, Deng H, Gan L, Zhong L, Deng Y, Wang Q, Lv C, Huang L. Chinese herbal component, Praeruptorin E, enhances anti-asthma efficacy and prevents toxicity of aminophylline by targeting the NF- κ B/PXR/CYP3A4 pathway. *Ann Transl Med* 2022;10(4):225. doi: 10.21037/atm-22-386